Author Response for "Comparison of aqueous SOA product distributions from guaiacol oxidation by non-phenolic and phenolic methoxybenzaldehydes as photosensitizers in the absence and presence of ammonium nitrate" by Mabato et al.

We thank the Reviewer for their thorough comments. We have revised the manuscript accordingly, and below are our point-by-point responses (in blue) to the comments (in black) and changes to the manuscript (in red). In those changes that begin with line numbers, the original text is also in blue. In addition, please note that the line numbers in the responses correspond to those in the revised manuscript.

Reviewer 1

This work specically investigated the influences from photosentisized organics and nitrate on the aqSOA formation, which is rarely discussed in previous studies, and it further elucidate the product distributions under these two cases. And finally it proposes the implications of these processes to the aerol chemistry. The paper has been reviewed by a few experts and the authors have responded to their comments and revised the paper accordingly. This reviewer, as an additional one, overall recommends its publication in ACP after some minor corrections suggested below

(1) They are a few very recenly published articles regarding the chemistry induced by 3C* and its impact on the light absorption of the products, should be added, a more throughou literature search especially in 2022 should be conducted and necessary

Response: Thank you for pointing this out. We have added recently published related papers to the introduction and discussion as follows:

Lines 33–35: Photosensitized reactions involving triplet excited states of organic compounds (3 C*) are efficient pathways for the formation of secondary organic aerosol in the aqueous phase (aqSOA; Smith et al., 2014, 2015, 2016; Yu et al., 2014, 2016; Chen et al., 2018; Lu et al., 2019; Ye et al., 2019; Chen et al., 2020; Liu et al., 2020; Jiang et al., 2021; Ma et al., 2021; Misovich et al., 2021; Ou et al., 2021; F. Li et al., 2022; X. Li et al., 2022; Aregahegn et al., 2022; Mabato et al., 2022; Wang et al., 2022).

Lines 48–50: Most previous studies on aqSOA formation via photosensitized non-carbonyl phenol oxidation have examined 3,4-dimethoxybenzaldehyde (DMB), a non-phenolic methoxybenzaldehyde, as the photosensitizer (Smith et al., 2014, 2015; Yu et al., 2014, 2016; Ye et al., 2019; Chen et al., 2020; Jiang et al., 2021; Ma et al., 2021; Misovich et al., 2021; Ou et al., 2021; X. Li et al., 2022).

Lines 289–292: The absorbance enhancement of phenolic aqSOA generated via reactions with ³CDMB*/³VL* has been linked to the formation of conjugated structures due to oligomerization and functionalization (e.g., additions of hydroxyl and carbonyl groups; Yu et al., 2014, 2016; Smith et al., 2016; Ye et al., 2019; Chen et al., 2020; Jiang et al., 2021; Misovich et al., 2021; Ou et al., 2021; F. Li et al., 2022; X. Li et al., 2022; Mabato et al., 2022; Wang et al., 2022).

(2) Line 91, if DMB and VL are moderate and poor photosensitizers, why they were chosen, enen if they are relatively abudant in the real atmosphere.

Response: Comparable levels of phenolic and non-phenolic carbonyls have been reported in biomass burning smoke (Simoneit et al., 1993; Anastasio et al., 1997). Although certain phenolic carbonyls, including VL, can initiate photosensitized reactions and contribute to aqSOA formation (Smith et al., 2016; Mabato et al., 2022), photosensitization by phenolic carbonyls is less understood compared to photosensitization by DMB (a non-phenolic carbonyl), the most commonly used photosensitizer in related studies.

DMB and VL were chosen to represent non-phenolic and phenolic methoxybenzaldehydes photosensitizers, respectively, as (1) their structures differ only by one functional group (– OCH₃ for the former and –OH for the latter), (2) they are both abundant in biomass burning emissions, and (3) there is available information on their photophysical properties (e.g., quantum yield of ³C* formation and ³C* lifetime) (Felber et al., 2021). This information was already provided in lines 86-92.

(3) Are the output light intensity from the Xe lamp stable over the course of the experiments? Did you have a test on that??

Response: The Xe lamp delivered stable output light intensity during our experiments. Based on our control experiments, the output light intensity only fluctuated by, on average, 0.9% within 3 h of operation (the duration of each experiment).

(4) How do you remove the influence of light absorption from the photosensitizers from that from the products? Their light absorption varied during the experiments, so a simple deduction can not do well, can you elaborate?

Response: The increase of light absorption from 350 to 550 nm, where GUA did not initially absorb light and where DMB and VL have little absorption, suggests the formation of light-absorbing products (Smith et al., 2016). This information was already provided in the SI (lines 106-108). In this work, the change in the integrated area of absorbance from 350 to 550 nm was therefore used to represent the absorbance enhancements. For the light absorption measurements in Figure 4a, the integrated area of absorbance from 350 to 550 nm at 180 min was normalized for the absorption of the reactants (GUA + photosensitizer; GUA + photosensitizer + AN; GUA + AN) by subtracting the corresponding values at 0 min (before irradiation) from each data point. Identifying the specific chromophores responsible for the absorbance enhancements can be performed using the methods employed in this study by matching the retention time (RT) of the products detected using UHPLC-HESI-Orbitrap-MS with that in the UHPLC-PDA. However, as mentioned in line 300, the detected products did not exhibit distinct peaks in the UHPLC-PDA chromatograms, likely due to the concentration of the chromophores being below the detection limit of PDA.

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1 Comparison of aqueous SOA product distributions from guaiacol

oxidation by non-phenolic and phenolic methoxybenzaldehydes as

photosensitizers in the absence and presence of ammonium nitrate

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12 **Abstract.** Aromatic carbonyls (e.g., methoxybenzaldehydes), an important class of photosensitizers, are abundant in the 13 atmosphere. Photosensitization and nitrate-mediated photo-oxidation can occur simultaneously, yet studies about their 14 interactions, particularly for aqueous secondary organic aerosol (aqSOA) formation, remain limited. This study compared 15 non-phenolic (3,4-dimethoxybenzaldehyde, DMB) and phenolic (vanillin, VL) methoxybenzaldehydes as photosensitizers 16 for aqSOA formation via guaiacol (GUA) oxidation in the absence and presence of ammonium nitrate (AN) under 17 atmospherically relevant cloud and fog conditions. GUA oxidation by triplet excited states of DMB (³DMB*) (GUA+DMB) was ~4 times faster and exhibited greater light absorption than oxidation by 3VL* (GUA+VL). Both GUA+DMB and 18 19 GUA+VL formed aqSOA composed of oligomers, functionalized monomers, oxygenated ring-opening species, and N-20 containing products in the presence of AN. The observation of N-heterocycles such as imidazoles indicates the participation 21 of ammonium in the reactions. The majority of generated aqSOA are potential brown carbon (BrC) chromophores. 22 Oligomerization and functionalization dominated in GUA+DMB and GUA+VL, but functionalization appeared to be more 23 important in GUA+VL due to contributions from VL itself. AN did not significantly affect the oxidation kinetics, but it had 24 distinct effects on the product distributions, likely due to differences in the photosensitizing abilities and structural features 25 of DMB and VL. In particular, the more extensive fragmentation in GUA+DMB than in GUA+VL likely generated more N-26 containing products in GUA+DMB+AN. In GUA+VL+AN, the increased oligomers may be due to VL-derived phenoxy 27 radicals induced by 'OH or 'NO₂ from nitrate photolysis. Furthermore, increased nitrated products observed in the presence 28 of both DMB or VL and AN than in AN alone implies that photosensitized reactions may promote nitration. This work 29 demonstrates how the structural features of photosensitizers affect aqSOA formation via non-carbonyl phenol oxidation. 30 Potential interactions between photosensitization and AN photolysis were also elucidated. These findings facilitate a better 31 understanding of photosensitized aqSOA formation and highlight the importance of AN photolysis in these reactions.

1 Introduction

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Photosensitized reactions involving triplet excited states of organic compounds (3C*) are efficient pathways for the 33 34 formation of secondary organic aerosol in the aqueous phase (aqSOA; Smith et al., 2014, 2015, 2016; Yu et al., 2014, 2016; 35 Chen et al., 2018; Lu et al., 2019; Ye et al., 2019; Chen et al., 2020; Liu et al., 2020; Jiang et al., 2021; Ma et al., 2021; Misovich et al., 2021; Ou et al., 2021; F. Li et al., 2022; X. Li et al., 2022; Aregahegn et al., 2022; Mabato et al., 2022; 36 37 Wang et al., 2022). Upon irradiation by solar radiation, photosensitizers form an excited triplet state that directly reacts with substrates (e.g., phenols), and can generate singlet oxygen (${}^{1}O_{2}$), superoxide (O_{2}) or hydroperoxyl (${}^{*}HO_{2}$) radicals, and 38 hydroxyl radicals ('OH) upon reactions with O₂ and substrates (George et al., 2018; Chen et al., 2020), thereby facilitating 39 40 the oxidation of rather volatile species and contributing to aqSOA formation. An important class of photosensitizers is aromatic carbonyls (e.g., methoxybenzaldehydes) which are abundant in aerosol particles, cloud waters, and fog waters 41 42 (Anastasio et al., 1997; Felber et al., 2021). Aromatic carbonyls can be emitted from anthropogenic sources and biomass 43 burning (BB; Lipari et al., 1984; Edye and Richards, 1991; Hawthorne et al., 1992; Simoneit et al., 1993, 1999; Anastasio et al., 1997; Felber et al., 2021), or formed via atmospheric oxidation of aromatic hydrocarbons (Hoshino et al., 1978; Calvert 44 45 and Madronich, 1987; Anastasio et al., 1997; Felber et al., 2021). BB is also a significant source of phenols through lignin pyrolysis (Simpson et al., 2005). Phenolic carbonyls have a hydroxyl (-OH) group on the aromatic ring, whereas non-46 47 phenolic carbonyls do not. BB smoke has been reported to have comparable concentrations of phenolic and non-phenolic carbonyls (Simoneit et al., 1993; Anastasio et al., 1997). 48

Most previous studies on agSOA formation via photosensitized non-carbonyl phenol oxidation have examined 3,4dimethoxybenzaldehyde (DMB), a non-phenolic methoxybenzaldehyde, as the photosensitizer (Smith et al., 2014, 2015; Yu et al., 2014, 2016; Ye et al., 2019; Chen et al., 2020; Jiang et al., 2021; Ma et al., 2021; Misovich et al., 2021; Ou et al., 2021; X. Li et al., 2022). By contrast, phenolic carbonyls have been mainly studied as aqSOA precursors via 'OH-, nitrate-, nitrite-, and ³DMB*-mediated oxidation (Li et al., 2014; Huang et al., 2018; Pang et al., 2019; Jiang et al., 2021; Misovich et al., 2021). However, strongly light-absorbing phenolic carbonyls (e.g., molar absorptivity above 300 nm \geq 7 × 10³ M⁻¹ cm⁻¹) can also serve as photosensitizers to promote aqSOA formation (Smith et al., 2016; Mabato et al., 2022). For instance, the direct photosensitized oxidation of phenolic carbonyls (i.e., oxidation of phenolic carbonyls by their ³C* or ³C*-derived oxidants) such as vanillin (VL; another methoxybenzaldehyde) efficiently form low-volatility products, with aqSOA mass yields of up to 140% (Smith et al., 2016). Moreover, the aqSOA mass yields from the oxidation of syringol by ³DMB* and ³VL* are similar (111% and 114%, respectively; Smith et al., 2014, 2016). In addition, we recently reported that the direct photosensitized oxidation of VL and guaiacol oxidation by ³VL* yield similar products (oligomers, functionalized monomers, and oxygenated ring-opening products) as observed with ³DMB* (Yu et al., 2014; Mabato et al., 2022). Guaiacol is a non-carbonyl BB methoxyphenol with an emission rate from fireplace wood combustion in the range of 172 to 279 mg/kg (Schauer et al., 2001; Simoneit, 2002). The atmospheric reactivity of methoxyphenols has recently been reviewed (Liu et al., 2022). However, our previous experiments (Mabato et al., 2022) were performed at a concentration (0.1 mM VL) higher than what was typically used for DMB (0.005 to 0.01 mM; Smith et al., 2014, 2015; Yu et al., 2014, 2016). Therefore, direct comparisons between photosensitization by ³DMB* and ³VL* cannot be made. Despite the above findings, much is still unknown about how aqSOA formation proceeds in systems using phenolic carbonyls as photosensitizers.

BB aerosols are typically internally mixed with other aerosol components, such as ammonium nitrate (AN; Zielinski et al., 2020). Hence, aromatic carbonyls and phenols may coexist with AN in BB aerosols. Nitrate and ammonium facilitate the formation of aqSOA and brown carbon (BrC) via a number of pathways. Nitrate photolysis can produce 'OH and nitrating agents (e.g., 'NO₂; Minero et al., 2007; Huang et al., 2018; Mabato et al., 2022; Wang et al., 2022; Yang et al., 2022), and ammonium reacts with carbonyls to yield N-containing heterocycles (e.g., imidazoles) and oligomers capable of UV-Vis light absorption (De Haan et al., 2009, 2011; Nozière et al., 2009, 2010, 2018; Shapiro et al., 2009; Yu et al., 2011; Lee et al., 2013; Powelson et al., 2014; Gen et al., 2018; Grace et al., 2019; Mabato et al., 2019). Furthermore, nitrate photolysis may be an important process for SO₂ oxidation and SOA formation in the particle phase (Gen et al., 2019a, 2019b, 2022; Zhang et al., 2020, 2021, 2022), and it can potentially modify the morphology of atmospheric viscous particles (Liang et al., 2021). Yet, understanding of the effects of inorganic nitrate on aqSOA formation remains limited. In addition, aqSOA formation studies involving aromatic carbonyls and phenols have probed either photosensitization or nitrate-mediated photo-oxidation, but these reactions can occur simultaneously. For instance, we previously reported nitrated compounds, including a potential imidazole derivative from the direct photosensitized oxidation of VL in the presence of AN (Mabato et al., 2022). Accordingly, investigations on reaction systems including both photosensitizers and AN may provide further insights into the aqueous-phase processing of BB aerosols.

In this work, we compared agSOA formation from photosensitized guaiacol (GUA) oxidation by ³C* of nonphenolic and phenolic methoxybenzaldehydes under identical conditions (simulated sunlight and concentration) relevant to cloud and fog waters. The effects of AN on photosensitized agSOA formation were also examined. In this study, the dominant aqSOA precursor is GUA (Henry's law constant of 9.2×10^2 M atm⁻¹; Sagebiel et al., 1992), and DMB and VL were used as photosensitizers to oxidize GUA. DMB and VL (Henry's law constants of 7.3×10^3 M atm⁻¹ and 4.7×10^5 M atm⁻¹, respectively; Yaws, 1994; EPI Suite version 4.1, 2012; Felber et al., 2021), which are also abundant in BB emissions (Schauer et al., 2001; Li et al., 2014; Chen et al., 2017; Pang et al., 2019; Mabato et al., 2022) and whose structures differ only by one functional group (-OCH₃ for the former and -OH for the latter, Fig. 1), represented non-phenolic and phenolic methoxybenzaldehydes, respectively. The structures of GUA, DMB, and VL are provided in Figure 1. Based on their quantum yield of ³C* formation, DMB and VL have been classified as moderate and poor photosensitizers, respectively (Felber et al., 2021). The photosensitized oxidation of GUA by ³DMB* or ³VL* in the absence (and presence) of AN are referred to as GUA+DMB(+AN) and GUA+VL(+AN), respectively. GUA photo-oxidation by AN alone (GUA+AN) was also explored for comparison with GUA+DMB+AN and GUA+VL+AN. The molar absorptivities of GUA, DMB, VL, and nitrate are shown in Figure 1. The precursor and photosensitizer decay kinetics, detected products, and absorbance enhancement were used to characterize the reactions. However, it should be noted that we mainly focused on the analyses of the reaction products and product distribution.

99 While several studies on photo-oxidation of BB emissions are available, this work focuses on the comparison 100 between non-phenolic and phenolic methoxybenzaldehydes as photosensitizers in the absence and presence of AN for 101 aqSOA formation. We found that GUA oxidation by ³DMB* was faster and exhibited greater light absorption relative to 102 GUA+VL. These are likely attributed to the stronger photosensitizing ability of DMB and the -OH group of VL, making it 103 more prone to oxidation and more reactive towards electrophilic aromatic substitution. Oligomerization and functionalization 104 dominated in GUA+DMB and GUA+VL, but functionalization appeared to be more significant in GUA+VL due to VL 105 transformation products. Although AN did not significantly influence the oxidation kinetics due to the predominant role of 106 photosensitizer chemistry compared to nitrate, AN promoted the formation of N-containing products. These include Nheterocycles (e.g., imidazoles), suggesting the participation of ammonium in the reactions. Moreover, the product 107 distributions indicate distinct interactions between photosensitization by ³DMB* and ³VL* and AN photolysis. In particular, 108 109 AN generated more N-containing products in GUA+DMB+AN than in GUA+VL+AN, and increased the oligomers in 110 GUA+VL+AN. Furthermore, increased nitrated compounds in GUA+DMB+AN and GUA+VL+AN compared to GUA+AN 111 suggest that photosensitized reactions may promote reactions by nitrate photolysis.

2 Methods

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2.1 Aqueous phase photo-oxidation experiments

Procedures for the photo-oxidation experiments are presented in detail in our previous study (Mabato et al., 2022). Experimental solutions were prepared using 0.1 mM guaiacol (GUA, Sigma Aldrich, >98.0%) and 0.01 mM 3,4dimethoxybenzaldehyde (DMB, Acros Organics, 99+%) or 0.01 mM vanillin (VL, Acros Organics, 99%, pure), in the absence and presence of ammonium nitrate (1 mM; AN, Acros Organics, 99+%, for analysis). These GUA and methoxybenzaldehydes concentrations are within the values expected in cloud or fog drops in areas with significant wood combustion (Anastasio et al., 1997; Rogge et al., 1998; Nolte et al., 2001). The AN concentration represents values usually observed in cloud and fog waters (Munger et al., 1983; Collett et al., 1998; Zhang and Anastasio, 2003; Li et al., 2011; Giulianelli et al., 2014; Bianco et al., 2020). It must be noted that this study did not intend to identify the AN concentrations that would affect the kinetics but attempted to analyze the effects of AN on photosensitized aqSOA formation. A solution composed of 0.1 mM GUA and 1 mM AN (GUA+AN) was also examined for comparison with GUA+DMB+AN and GUA+VL+AN. Sulfuric acid (H₂SO₄; Acros Organics, ACS reagent, 95% solution in water) was used to adjust the pH of the solutions to 4, which is within typical cloud pH values (2-7; Pye et al., 2020) and pH values observed in wood burningimpacted cloud and fog waters (Collett et al., 1998; Raja et al., 2008). The solutions (initial volume of 500 mL) were bubbled with synthetic air (0.5 dm³/min) for 30 min before irradiation and throughout the reactions to achieve air-saturated conditions (Du et al., 2011; Chen et al., 2020) and were continuously magnetically stirred. In this study, the reactions can generate ³DMB*/³VL* and secondary oxidants (¹O₂, O₂-/HO₂, OH) but not ozone. Solutions contained in a quartz photoreactor were irradiated using a xenon lamp (model 6258, Ozone free xenon lamp, 300 W, Newport) equipped with a

longpass filter (20CGA-305 nm cut-on filter, Newport) to eliminate light below 300 nm. The reaction temperatures were maintained at 27 ± 2 °C using cooling fans positioned around the photoreactor and lamp housing. The averaged initial photon flux in the reactor measured from 300 to 380 nm was $\sim 3 \times 10^{15}$ photons cm⁻² s⁻¹ nm⁻¹ (Fig. 1), similar to our previous work (Mabato et al., 2022). Samples were collected every 30 min for 180 min for offline analyses of (1) GUA, DMB, and VL concentrations using ultra-high-performance liquid chromatography with photodiode array detector (UHPLC-PDA) and (2) absorbance measurements using UV-Vis spectrophotometry. Moreover, the samples collected before and after irradiation (180 min) were analyzed for (3) reaction products using UHPLC coupled with heated electrospray ionization Orbitrap mass spectrometry (UHPLC-HESI-Orbitrap-MS) operated in positive and negative ion modes and (4) concentrations of small organic acids using ion chromatography (IC). Each experiment was repeated independently at least three times. The reported decay rate constants, small organic acids concentration, and absorbance enhancement were averaged from triplicate experiments, and the corresponding errors represent one standard deviation. The pseudo-first-order rate constant (k') for GUA decay was determined using the following equation (Huang et al., 2018):

$$ln\left([GUA]_t/[GUA]_0\right) = -k't \tag{Eq. 1}$$

where [GUA]_t and [GUA]₀ are GUA concentrations at time t and 0, respectively. DMB or VL decay rate constants were calculated by replacing GUA with DMB or VL in Eq. 1. The decay rate constants were normalized to the photon flux measured for each experiment through dividing k' by the measured 2-nitrobenzaldehyde (2NB; a chemical actinometer) decay rate constant, j(2NB) (Mabato et al., 2022). In addition, the decay rate constants were corrected for the internal light screening due to DMB, VL, and AN (Leifer, 1988; Zhang and Anastasio, 2003; Smith et al., 2014, 2015, 2016). The values of the internal light screening factor (S_{λ}) determined around the peak in the light absorption action spectrum (DMB: 310-335 nm, VL: 304-364 nm, nitrate: 300-331 nm) (Smith et al., 2014, 2015, 2016) for an 8.5 cm cell were 0.95 for GUA+AN, 0.51 for GUA+DMB, 0.54 for GUA+DMB+AN, 0.57 for GUA+VL, and 0.59 for GUA+VL+AN. Moreover, two independently prepared samples for each reaction condition were analyzed using UHPLC-HESI-Orbitrap-MS. Only peaks that were reproducibly detected in both sets of samples were considered. For clarity, the formulas discussed in this work correspond to neutral analytes (e.g., with H⁺ or NH₄⁺ removed from the ion formula). The details of the analytical procedures are provided in the Supplement (Sects. S1 to S4).

2.2 Calculation of- normalized abundance of products

Several recent studies have used comparisons of relative abundance of products based on peak areas from mass spectrometry (MS) results (e.g., Lee et al., 2014; Romonosky et al., 2017; Wang et al., 2017; Fleming et al., 2018; Song et al., 2018; Klodt et al., 2019; Ning et al., 2019) to show the relative importance of different types of compounds (K. Wang et al., 2021). However, comparisons of relative abundance among different compounds can be subject to uncertainties as ionization efficiencies in soft ionization, such as ESI, may significantly vary between different compounds (Kebarle, 2000; Schmidt et al., 2006; Leito et al., 2008; Perry et al., 2008; Kruve et al., 2014). In our previous work (Mabato et al., 2022), we introduced the normalized abundance of products ([P], unitless) (Eq. 2) as a semi-quantitative analysis that gives an overview of how

the signal intensities changed under different experimental conditions but not the quantification of the absolute product 164 165 concentration. The calculation assumes equal ionization efficiencies of different compounds, which is commonly used to 166 estimate O:C ratios of SOA (Bateman et al., 2012; Lin et al., 2012; Laskin et al., 2014; De Haan et al., 2019):

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[P] =
$$\frac{A_{P,t}}{A_{GUA,t}} \cdot \frac{[GUA]_t}{[GUA]_0}$$
 (Eq. 2)

where $A_{P,t}$ and $A_{GUA,t}$ are the extracted ion chromatogram (EIC) peak areas of the product P and GUA from UHPLC-HESI-Orbitrap-MS analyses at time t, respectively; $[GUA]_t$ and $[GUA]_0$ are the GUA concentrations (μ M) determined using UHPLC-PDA at time t and 0, respectively. Note that the normalized abundance of products has intrinsic uncertainties due to the variability in ionization efficiencies for various compounds. Moreover, it should be noted that the normalized abundance of products was calculated using only the positive ion mode data as the GUA signal from the negative ion mode was weak and thus may present large uncertainties during normalization. Therefore, products that may not give signals or may have weak signals in the positive ion mode were possibly underestimated in the normalized product abundance. Nevertheless, it enables the comparison of MS results among different experiments. As demonstrated in our previous work (Mabato et al., 2022) and the current study, a higher normalized abundance of products generally correlates with higher efficiency of oxidation. The reported uncertainties were propagated from the changes in [GUA] measured using UHPLC-PDA and the MS signal intensities.

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3 Results and Discussion

- 182 Using kinetics data, MS analyses, and absorbance enhancement data, we first examined the differences between GUA+DMB
- 183 and GUA+VL (Sect. 3.1). Then, we analyzed GUA+DMB+AN, GUA+VL+AN, and GUA+AN (Sect. 3.2) to explore the
- 184 effects of nitrate photolysis and ammonium on photosensitized agSOA formation.
- photosensitized GUA oxidation by non-phenolic (3DMB*) and phenolic (3VL*) 185 Comparison of 186 methoxybenzaldehydes
- 187 Prior studies have reported that photosensitized non-carbonyl phenol oxidation in the presence of 3,4-
- 188 dimethoxybenzaldehyde (DMB) and vanillin (VL) (separately) was mainly driven by ³DMB* and ³VL*, respectively (Smith
- et al., 2014; Mabato et al., 2022), while contributions from secondary oxidants such as ¹O₂ and 'OH were likely minor. 189
- 190 However, both ³DMB* and ³VL* are efficiently quenched by O₂, suggesting that energy transfer should be considered in
- 191 evaluating photosensitized processes involving these methoxybenzaldehydes (Felber et al., 2021). Moreover, it was found
- 192 that ³DMB*, ¹O₂, and O₂* were the major contributors to the photosensitized oxidation of 4-ethylguaiacol (Chen et al., 2020).
- 193 Recently, the oxidation of guaiacyl acetone (a non-conjugated phenolic carbonyl) in the presence of DMB has been reported

to be initiated by ³DMB*, ¹O₂, OH, or methoxy radical (OCH₃) (Misovich et al., 2021). Further studies are thus required to identify the specific oxidants in these reaction systems. In this study, reactions initiated in the presence of DMB or VL are collectively referred to as photosensitized reactions. The reaction conditions, initial guaiacol (GUA) and DMB or VL decay rate constants, normalized product abundance, and the chemical characteristics of aqSOA formed in this work are summarized in Table 1.

3.1.1 Kinetic analysis of photosensitization by ³DMB* and ³VL*

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200 No significant loss of GUA or photosensitizers was observed for dark experiments (p > 0.05). Figure S1 shows the decay of 201 GUA, DMB, and VL under different experimental conditions. Upon irradiation, the GUA decay rate constant in GUA+DMB 202 was ~4 times higher than in GUA+VL. In GUA+DMB, the decay rate constant of GUA was ~8 times higher than that of 203 DMB, consistent with a previous study (Smith et al., 2014). Contrastingly, the decay rate constant of VL was 2.4 times 204 higher than that of GUA in GUA+VL. This VL consumption was also observed in our earlier work using 0.1 mM GUA + 205 0.1 mM VL (Mabato et al., 2022). These trends could be explained by the following reasons. First, DMB has a stronger photosensitizing ability than VL based on its higher quantum yield of ³C* formation and longer lifetime of ³DMB* 206 207 compared to ³VL* (Felber et al., 2021). Second, VL is also a phenolic compound similar to GUA, and is therefore highly reactive towards oxidation. For instance, its -OH group can be oxidized by ³VL* via H-atom abstraction to form phenoxy 208 209 radicals which can undergo coupling to form oligomers (Kobayashi and Higashimura, 2003; Sun et al., 2010; Mabato et al., 210 2022). The faster consumption of VL than GUA suggests a competition between ground-state VL and GUA for reaction with 211 ³VL*. Moreover, compared to a –OCH₃ group (in DMB), an –OH group (in VL) has a stronger electron-donating ability and is thus more activating towards electrophilic aromatic substitution. It should be noted that the differences in the GUA decay 212 213 rate constants among different reaction systems are not quantitatively equivalent to photosensitizing efficiencies, and a 214 detailed quantitative analysis of which is beyond the scope of this study. Nonetheless, these results suggested that GUA 215 oxidation in GUA+DMB was overall more efficient than in GUA+VL. Our kinetic analysis focused on the decay rate 216 constants of the agSOA precursor (GUA) and the photosensitizers (DMB and VL) during photosensitization under the same 217 experimental conditions (same aqSOA precursor and concentration, same photosensitizer concentration, and same lamp 218 photon flux). The effects of other factors (e.g., intersystem crossing efficiency) on the rate constants were not examined. 219 Explicit kinetic studies (e.g., Smith et al., 2014, 2015) that measure second-order rate constants should be conducted in the 220 future to extend the applicability of the kinetic parameters to other conditions.

3.1.2 Product distributions and chemical characteristics of aqSOA from photosensitization by ³DMB* and ³VL*

The products detected using UHPLC-HESI-Orbitrap-MS were used to characterize the aqSOA formed in this work. The signal-weighted distributions of aqSOA calculated from combined positive (POS) and negative (NEG) ion modes MS results are summarized in Figure 2. The signal-weighted distributions calculated separately from POS and NEG ion modes MS results are available in Figures S2 and S3. It should be noted that in this work, the product distributions for all experiments

226 were based on the same irradiation time of 180 min. An irradiation time of 180 min was chosen as it was sufficient to show 227 the differences in the extent of reaction of GUA among the reaction systems studied. For reaction systems with precursors of 228 different reactivities, chemical analysis at a fixed reaction time may be looking at different generations of products of each 229 precursor, as Yu et al. (2014) reported. Measuring the product distribution at a fixed time might have missed the information 230 on what/how many products are formed at the similar amounts of precursors reacted. The situation could be even more 231 complicated if different precursors had major differences in pathways and dominant intermediates. However, comparing the 232 product distributions after a certain time of light exposure, as is the case for this study, is useful to evaluate what products 233 would form after a certain time of photosensitization. Oligomers and derivatives of GUA dominated both GUA+DMB and 234 GUA+VL, in agreement with pronounced oligomerization from triplet-mediated oxidation of relatively high phenol 235 concentration (e.g., 0.1 to 3 mM; Li et al., 2014; Yu et al., 2014, 2016; Slikboer et al., 2015; Ye et al., 2019; Mabato et al., 2022). Figure 3 schematically depicts the main differences between photosensitized GUA oxidation by 3DMB* and 3VL* in 236 the absence and presence of AN. As shown in Fig. 3, 3DMB* and 3VL* can oxidize GUA via H-atom abstraction to form 237 238 phenoxy radicals which undergo coupling to form oligomers (Kobayashi and Higashimura, 2003; Sun et al., 2010; Mabato et 239 al., 2022). The higher oligomer contribution in GUA+DMB is likely due to the better photosensitizing ability of DMB than VL and partly the lower abundance of ³VL* due to fast VL consumption. VL was consumed faster than DMB during GUA 240 241 oxidation ascribable to the -OH group of VL, making it more susceptible to oxidation and more reactive towards 242 electrophilic aromatic substitution. In addition, the normalized product abundance for GUA+DMB was ~4 times higher than 243 that for GUA+VL (Table 1), further suggesting more efficient photosensitized GUA oxidation by ³DMB* than by ³VL*. The 244 oxidation of GUA or transient organic intermediates by secondary oxidants (e.g., ¹O₂ and ^{*}OH) from ³DMB* or ³VL* and the 245 fragmentation of larger compounds generate highly oxidized ring-opening products (Yu et al., 2014; Huang et al., 2018; 246 Chen et al., 2020). GUA+DMB had a higher contribution of ring-opening products than GUA+VL, likely due to the greater 247 availability of secondary oxidants in the former and fast VL consumption lowering the production of these species in 248 GUA+VL. The IC analyses also indicate the formation of small organic acids (e.g., formic acid), which appeared to have 249 higher concentrations in the presence of DMB than in VL (Fig. S4). Although no data is available for the concentration 250 changes (every 30 min) of small organic acids during the reaction, it is likely that an increasing trend would be observed as 251 fragmentation, which leads to the decomposition of initially formed oligomers and the generation of smaller oxygenated 252 products, becomes important at longer irradiation times (Huang et al., 2018). This trend has also been observed in our 253 previous work on the direct photosensitized oxidation of VL (Mabato et al., 2022), as well as other studies on 254 photosensitized oxidation of non-carbonyl phenols and phenolic carbonyls (e.g., Yu et al., 2016; Jiang et al., 2021). The 255 reactions of secondary oxidants or ring-opening products with GUA can form functionalized products. Notably, the 256 contribution of monomers in GUA+VL was almost twice as high as in GUA+DMB, ascribable to VL transformation 257 products. We previously showed that for the direct photosensitized oxidation of VL, functionalization prevails over 258 oligomerization at 0.01 mM VL, the [VL] used in this work, while oligomerization dominates at higher [VL] (0.1 mM; 259 Mabato et al., 2022).

It has been reported that oligomerization could occur during the electrospray ionization process (Yasmeen et al., 2010). In this work, it was confirmed that the oligomers observed were generated in the solutions via aqueous reactions instead of being artefacts of HESI-MS. This is based on the absence of dimers and higher oligomers in the HESI mass spectra of dark control solutions acquired by direct infusion (Yu et al., 2016).

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The major GUA+DMB and GUA+VL products (Tables S1-S2) are mostly oligomers which can be formed through the coupling of phenoxy radicals (Kobayashi and Higashimura, 2003; Sun et al., 2010; Mabato et al., 2022). GUA+DMB products matched those reported in previous works on ${}^{3}DMB^{*-}$ and/or ${}^{\circ}OH$ -mediated phenol oxidation (Yu et al., 2014, 2016). These include GUA dimers and trimers (e.g., $C_{14}H_{14}O_{4}$ and $C_{21}H_{18}O_{8}$, #1 and 19; Table S1), aldehydes ($C_{7}H_{6}O_{4}$, #13; Table S1), and esters ($C_{16}H_{18}O_{6}$, #14; Table S1). Functionalized products include $C_{11}H_{12}O_{5}$ and $C_{10}H_{12}O_{3}$ (#8 and 12; Table S1). More than half of the major GUA+VL products are the same oligomers detected from GUA+DMB (e.g., $C_{13}H_{10}O_{4}$ and $C_{20}H_{18}O_{6}$, #4 and 21; Table S1). The rest are mainly functionalized species such as $C_{7}H_{8}O_{4}$ and $C_{8}H_{8}O_{5}$ (#28 and 35; Table S2), corresponding to a hydroxylated GUA and hydroxylated VL, respectively.

The average elemental ratios and elemental distribution of the products (Fig. S5a-d) were consistent with those in previous studies on similar reaction systems (Yu et al., 2014, 2016; Mabato et al., 2022). The majority of the GUA+DMB and GUA+VL products had H:C ≤1.0 and O:C ≤0.5, typical for aromatic species (Mazzoleni et al., 2012; Kourtchev et al., 2014; Jiang et al., 2021). GUA+DMB had more compounds with higher O:C (≥0.6), in agreement with higher contributions of ring-opening products than in GUA+VL (Fig. 2). The higher (OS_C) for GUA+VL than in GUA+DMB (Table 1) was probably due to the significant functionalization in the former. Moreover, the distributions of OS_C and carbon number (Fig. S6a-d) show that these aqSOA products have similar elemental composition to those of low-volatility oxygenated organic aerosols (LV-OOA), semi-volatile oxygenated organic aerosols (SV-OOA), and slightly with biomass burning organic aerosols (BBOA) (Kroll et al., 2011). Further discussions on van Krevelen diagrams (Fig. S5a-d) and OS_C vs. carbon number plots (Fig. S6a-d) for GUA+DMB and GUA+VL aqSOA are available in the Supplement (Sect. S5). In brief, ³DMB*-initiated GUA oxidation was faster and yielded higher normalized product abundance than oxidation by ³VL*. This is likely due to the stronger photosensitizing ability of DMB than VL and the -OH group of VL facilitating its rapid consumption. In addition, oligomerization and functionalization dominated in both GUA+DMB and GUA+VL, as reported in similar studies (Yu et al., 2014, 2016; Chen et al., 2020; Jiang et al., 2021; Misovich et al., 2021; Mabato et al., 2022). However, functionalization was more prominent in the latter, attributable to the transformation of VL. Nonetheless, it must be noted that for phenolic aqSOA, fragmentation will ultimately be more predominant at longer irradiation times (Huang et al., 2018; Yu et al., 2016; Mabato et al., 2022).

3.1.3 Light absorption of aqSOA from photosensitization by ³DMB* and ³VL*

The absorbance enhancement of phenolic aqSOA generated via reactions with ³CDMB*/³VL* has been linked to the formation of conjugated structures due to oligomerization and functionalization (e.g., additions of hydroxyl and carbonyl groups; Yu et al., 2014, 2016; Smith et al., 2016; Ye et al., 2019; Chen et al., 2020; Jiang et al., 2021; Misovich et al., 2021;

293 Ou et al., 2021; F. Li et al., 2022; X. Li et al., 2022; Mabato et al., 2022; Wang et al., 2022). Moreover, the aqueous-phase 294 photo-oxidation of BB emissions can enhance BrC absorbance via the formation of aromatic dimers and functionalized 295 products (Hems et al., 2020). The increase in light absorption throughout 180 min of irradiation and the change in the rate of 296 sunlight absorption (ΔR_{abs}) (Jiang et al., 2021) from 350 to 550 nm at 180 min during typical clear and haze days in Beijing, 297 China for all the reaction systems studied are provided in Figure 4. Figure S7 shows the absorption spectra after 180 min of 298 irradiation for each reaction system studied. In this work, the absorbance enhancement of GUA+DMB and GUA+VL (Fig. 299 4a) could be due to oligomers and functionalized monomers, which are the highest contributors to the product signals. 300 Identifying the chromophores responsible for the absorbance enhancement may be beneficial in understanding the impact of 301 aqSOA on the Earth's radiative balance and determining the reactions that affect light absorption by aqSOA (Mabato et al., 302 2022). However, the detected products did not exhibit distinct peaks in the UHPLC-PDA chromatograms, likely due to the 303 concentration of the chromophores being below the detection limit of PDA. Nevertheless, the higher absorbance enhancement and ΔR_{abs} for GUA+DMB than GUA+VL was probably due to the higher contribution and normalized 304 305 abundance (by ~6 times) of oligomers in the former.

Additional information about aqSOA lightabsorption can be deduced from the plots of the double bond equivalent (DBE) values vs. carbon number (n_C) (Lin et al., 2018). Figure S8 shows these plots along with the DBE reference values of fullerene-like hydrocarbons (Lobodin et al., 2012), cata-condensed polycyclic aromatic hydrocarbons (PAHs; Siegmann and Sattler, 2000), and linear conjugated polyenes with a general formula C_xH_{x+2} . The shaded area indicates a sufficient level of conjugation for visible light absorption, and species within this region are potential BrC chromophores. GUA+DMB and GUA+VL aqSOA exhibited a significant overlap in the DBE vs. n_C space; nearly all products from both systems, including the high-relative-abundance species, are potential BrC chromophores. GUA+DMB had more oligomeric products with high relative abundance ($n_C \ge 12$ and DBE ≥ 8). For GUA+VL, high-relative-abundance products also include monomeric species ($n_C = 7$ -8 and 4-5 DBE) corresponding to hydroxylated products (e.g., $C_7H_8O_4$ and $C_8H_8O_5$; 28 and 35; Table S2). These observations further indicate the importance of oligomerization and functionalization for the absorbance enhancement of aqSOA generated via photosensitization by $^3DMB^*$ and $^3VL^*$. In summary, $^3DMB^*$ and $^3VL^*$ can oxidize GUA resulting in aqSOA and BrC formation, but GUA+DMB products exhibited stronger light absorption. In GUA+VL, the extent of GUA oxidation was limited by significant VL consumption.

319 3.2 Comparison of photosensitized GUA oxidation by non-phenolic (3DMB*) and phenolic (3VL*)

320 methoxybenzaldehydes in the presence of AN

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321 3.2.1 Kinetic analysis of photosensitization by ³DMB* and ³VL* in the presence of AN

- 322 Ammonium nitrate (AN) did not significantly affect (p > 0.05) the decay rate constants of GUA, DMB, and VL for both
- 323 GUA+DMB+AN and GUA+VL+AN (Table 1), likely due to the higher molar absorptivities of the photosensitizers
- 324 compared to that of nitrate. This implies that the chemistry of ³DMB* and ³VL* dominated that of nitrate. In this work, the
- 325 GUA decay rate constants decreased in the order of GUA+DMB/GUA+DMB+AN > GUA+VL/GUA+VL+AN > GUA+AN

326 (Table 1). Note that as the molar absorptivities of the photosensitizers are higher than that of nitrate, the kinetics data were 327 also analyzed on a per-photon-absorbed basis for a more appropriate comparison of reaction efficiency (Sect. S6). The 328 apparent quantum efficiency of GUA photodegradation (ϕ_{GUA}) in the presence of nitrate (GUA+AN: $0.17 \pm 3.8 \times 10^{-2}$) was 329 ~2 and ~7 times higher than that in the presence of DMB ($0.10 \pm 2.9 \times 10^{-3}$) or VL ($0.026 \pm 7.2 \times 10^{-3}$), respectively. This 330 suggests that nitrate-mediated GUA photo-oxidation is more efficient than photosensitization by ³DMB* or ³VL* on a per-331 photon-absorbed basis.

3.2.2 Product distributions and chemical characteristics of aqSOA from photosensitization by ³DMB* and ³VL* in the presence of AN

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For both GUA+DMB+AN and GUA+VL+AN, AN had no significant effect on the normalized product abundance (Table 1), but it induced the formation of N-containing products composed of N-heterocycles (e.g., imidazoles and pyridines) and oligomers, as well as nitrated species. Similarly, we previously reported a potential imidazole derivative from the direct photosensitized oxidation of VL in the presence of AN, which was attributed to the reaction of ring-opening products with dissolved ammonia (Mabato et al., 2022). Oligomers remained the highest signal contributors in the presence of AN (Fig. 2), but interactions between photosensitization by ³DMB* and ³VL* and AN photolysis were distinct. First, nitrated species had similar contributions in both cases, but the contribution and normalized abundance of all N-containing products in GUA+DMB+AN were 2 and ~14 times higher, respectively, than in GUA+VL+AN. This difference can be attributed to the higher contribution of N-heterocycles and N-containing oligomers in GUA+DMB+AN. Compared to GUA+VL, GUA+DMB had a higher contribution of ring-opening products which can react with ammonia, as discussed earlier (Figs. 2 and 3). Second, the decrease in oligomers in GUA+DMB+AN may be due to their fragmentation induced by 'OH from nitrate photolysis, then conversion to N-containing products. Correspondingly, the contribution of possibly ring-retaining Ncontaining products in GUA+DMB+AN (18.6%) was ~3 times higher than that in GUA+VL+AN (6.5%). While fragmentation of oligomers likely occurred in GUA+VL+AN as well, the increase in oligomers suggests that other reactions have taken place. For GUA+VL+AN, 'OH or 'NO₂ from nitrate photolysis may have initiated H-atom abstraction from the -OH group of VL, generating phenoxy radicals which can undergo coupling to form more oligomers (Kobayashi and Higashimura, 2003; Sun et al., 2010; Mabato et al., 2022). This may also explain the more significant decrease of monomers in GUA+VL+AN (~3 times) compared to GUA+DMB+AN (~2 times). Similarly, we previously observed an increase in oligomers during the direct photosensitized oxidation of 0.01 mM VL (Mabato et al., 2022), the [VL] used in this work, upon adding 1 mM AN. These findings indicate that photosensitization by non-phenolic and phenolic methoxybenzaldehydes may interact differently with AN photolysis.

GUA+AN mainly formed oligomers analogous to 'OH-mediated phenol oxidation (Yu et al., 2014, 2016), followed by N-containing products. The normalized product abundance of GUA+AN was the lowest among all experiments, likely due to the lower GUA decay constant relative to photosensitized oxidation. Moreover, the normalized abundance of N-containing products in GUA+AN was ~12 times lower than that in GUA+DMB+AN but comparable to that in

GUA+VL+AN. This discrepancy for GUA+VL+AN might be due to the weaker signals of its N-containing products in the positive compared to the negative ion mode. As previously mentioned, the normalized product abundance was calculated using only the positive ion mode data as the GUA signal from the negative ion mode was weak and thus may present large uncertainties during normalization. Interestingly, the contributions from nitrated species in GUA+DMB+AN and GUA+VL+AN were higher than in GUA+AN, suggesting possible enhancement of nitration reactions. This is likely due to the increased formation of 'NO₂, for instance, via the reactions of 'OH and O₂*- (from ³DMB* or ³VL*) with NO₂- (Pang et al., 2019; Mabato et al., 2022). Similarly, we previously reported enhanced nitration via the direct photosensitized oxidation of VL in the presence of AN under air-saturated conditions (O₂ is present) relative to nitrogen-saturated conditions (Mabato et al., 2022). These imply that photosensitization may promote reactions induced by nitrate photolysis.

The major products from GUA+DMB+AN, GUA+VL+AN, and GUA+AN (Tables S3–S5) include oligomers and functionalized monomers detected in GUA+DMB and GUA+VL (Tables S1–S2). The N-heterocycles from GUA+DMB+AN include C₆H₆N₄ (#41; Table S3), which may be 2,2'-biimidazole (BI), a reaction product from glyoxal + reduced nitrogenous compounds (e.g., ammonium salts) (De Haan et al., 2009; Galloway et al., 2009; Nozière et al., 2009; Shapiro et al., 2009; Yu et al., 2011; Kampf et al., 2012; Gen et al., 2018; Mabato et al., 2019). The nitrated products include C₁₂H₁₁N₃O₃ and C₁₅H₁₀N₄O₃ (#42 and 49; Table S3), which possibly have a nitrated imidazole moiety and a nitrophenol moiety, respectively. For GUA+VL+AN, oligomers (C₁₄H₁₂O₆ and C₂₀H₁₆O₇; #55 and 59, Table S4) which were not among the major products in GUA+VL were noted. C₁₀H₈O₂ likely has a furanone group (#50; Table S4); furanones are the primary products of the reaction of 'OH with toluene and other aromatic hydrocarbons (Smith et al., 1999). Moreover, C₁₁H₉N₃O₃ (#57; Table S4) has a nitrated imidazole moiety. Among the N-containing compounds in GUA+AN is C₄H₃N₃O₃ (#69; Table S5), which may be a nitrated imidazole-2-carboxaldehyde. Imidazole-2-carboxaldehyde is also a reaction product from glyoxal + reduced nitrogenous compounds (e.g., ammonium salts) (De Haan et al., 2009; Galloway et al., 2009; Nozière et al., 2009; Shapiro et al., 2009; Yu et al., 2011; Kampf et al., 2012; Gen et al., 2018; Mabato et al., 2019).

The $\langle O:C \rangle$ for GUA+DMB+AN and GUA+VL+AN were lower than those in the absence of AN (Table 1), possibly due to the formation of N-heterocycles, altering the elemental ratios. The $\langle O:C \rangle$ and $\langle H:C \rangle$ were comparable in GUA+DMB+AN and GUA+VL+AN, but the $\langle N:C \rangle$ for the former was higher, implying a greater extent of reactions involving AN. Relative to GUA+DMB+AN and GUA+VL+AN, GUA+AN had a higher $\langle N:C \rangle$, as can be expected given that AN was the only oxidant source. The lower $\langle OS_C \rangle$ of GUA+DMB+AN and GUA+VL+AN compared to GUA+AN may be attributed to triplet-initiated oxidation generating higher-molecular-weight products with less fragmentation compared to 'OH-mediated oxidation (Yu et al., 2014; Chen et al., 2020). Nonetheless, AN generally increased the $\langle OS_C \rangle$ for both GUA+DMB and GUA+VL, with a more noticeable increase for the former, suggesting more oxidized products. Similarly, in a previous work, the more oxygenated and oxidized aqSOA from the photo-oxidation of phenolic carbonyls in AN solutions than in ammonium sulfate solutions has been ascribed to nitrate photolytic products promoting the reactions (Huang et al., 2018). Furthermore, GUA+DMB+AN and GUA+VL+AN aqSOA had mainly similar features in the OS_C vs. n_C plots as those observed in the absence of AN (Fig. S6). More information on van Krevelen diagrams (Figs. S5e-h and S9) and OS_C

vs. n_C plots (Figs. S6e-h and S10) for GUA+DMB+AN, GUA+VL+AN, and GUA+AN aqSOA are provided in the Supplement (Sect. S7). In essence, AN had no significant effect on the decay kinetics ascribable to photosensitizer chemistry prevailing over nitrate, but it induced the formation of N-containing products. Moreover, AN modified the product distributions, albeit in different ways (Figs. 2 and 3). In particular, N-containing products were more abundant in GUA+DMB+AN, probably due to more extensive fragmentation in GUA+DMB than in GUA+VL. In GUA+VL+AN, AN promoted oligomer formation likely via the -OH group of VL. Furthermore, GUA+DMB+AN and GUA+VL+AN had more nitrated products than GUA+AN, suggesting that photosensitized reactions may promote nitrate photolysis-initiated reactions.

3.2.3 Light absorption of aqSOA from photosensitization by ³DMB* and ³VL* in the presence of AN

The presence of AN also did not appreciably affect the absorbance enhancement and ΔR_{abs} for both GUA+DMB+AN and GUA+VL+AN (Fig. 4). For GUA+DMB+AN, the N-containing products may have offset the decrease in oligomers to maintain the absorbance enhancement observed from GUA+DMB. Wang et al. (2022) reported that nitration might contribute significantly to absorbance enhancement for methoxyphenols in sodium nitrate. In GUA+VL+AN, the decrease in monomers may have counteracted the increased oligomers and the generated N-containing products. Compared to GUA+DMB+AN, the N-containing products from GUA+VL+AN probably had less impact on the absorbance enhancement based on their smaller signal contribution.

Similar to experiments without AN, CHO species from GUA+DMB+AN and GUA+VL+AN were mainly overlapped in the DBE vs. n_C space (Fig. S8c,d) and were mostly potential BrC chromophores. In both systems, GUA dimers were the products with the highest relative abundance. For GUA+DMB+AN, products with high relative abundance also include a CHN species, while two CHON species had high n_C (18,20) and DBE (16,14) values. In GUA+VL+AN, products with high relative abundance include a CHON species (n_C = 11 and 9 DBE). Approximately 30% and 43% of the N-containing products for GUA+DMB+AN and GUA+VL+AN, respectively, were among the potential BrC chromophores. This suggests the possible significance of N-containing products for light absorption of aqSOA from photosensitization by methoxybenzaldehydes and AN photolysis. Correspondingly, nitroaromatic compounds and N-heterocycles are frequently noted in BBOA (Iinuma et al., 2010; Kitanovski et al., 2012; Kourtchev et al., 2016) and have been proposed to be potential contributors to BrC light absorption (Laskin et al., 2015). Relative to GUA+DMB+AN and GUA+VL+AN, only 19% of the N-containing products in GUA+AN were potential BrC chromophores (Fig. S8e,f), and these did not include CHN species. These indicate that the N-containing products formed in the presence of both photosensitizers and AN may be more significant contributors to the light absorption of phenolic aqSOA than those formed in AN only.

4 Conclusions and atmospheric implications

The photosensitized oxidation of guaiacol (GUA) by triplet excited states of 3,4-dimethoxybenzaldehyde (³DMB*) and vanillin (³VL*) (separately) in the absence and presence of ammonium nitrate (AN) were compared under identical conditions (simulated sunlight and concentration) relevant to atmospheric cloud and fog waters. Compared to GUA+VL, faster GUA oxidation and stronger light absorption by the products were observed in GUA+DMB. Moreover, VL was consumed faster relative to DMB, limiting the extent of GUA oxidation in GUA+VL. These differences are rooted in DMB having a better photosensitizing ability than VL and the –OH group of VL, making it more susceptible to oxidation and more reactive towards electrophilic aromatic substitution. Both GUA+DMB and GUA+VL generated aqSOA (including potential BrC chromophores) composed of oligomers, functionalized monomers, oxygenated ring-opening products, and N-containing products in the presence of AN. The major aqSOA formation processes for GUA+DMB and GUA+VL were oligomerization and functionalization, but functionalization appeared to be more significant in GUA+VL due to VL transformation products. The photochemical evolution of aqSOA from GUA+DMB has been reported by Yu et al. (2016). Similar experiments for aqSOA from GUA+VL should be conducted in the future to better understand photosensitized reactions involving phenolic carbonyl photosensitizers.

AN did not significantly affect the decay kinetics due to the predominant effect of ³DMB* and ³VL* chemistry compared to nitrate, but it promoted the formation of N-containing products; these are composed of N-heterocycles (e.g., imidazoles) and oligomers and nitrated species. The observation of N-heterocycles agrees with our previous findings that ammonium participates in photosensitized oxidation of phenolic compounds in the presence of AN (Mabato et al., 2022). These results also suggest that photosensitized oxidation of phenolic compounds in the presence of AN might be an important source of N-heterocycles and nitrated products. Identifying the sources of N-heterocycles and nitrated compounds is important due to their environmental and health impacts (Laskin et al., 2009). Moreover, photosensitized reactions by nonphenolic and phenolic methoxybenzaldehydes may be differently influenced by AN photolysis. For instance, the more extensive fragmentation in GUA+DMB than in GUA+VL possibly resulted in more N-containing products in GUA+DMB+AN. Furthermore, the increased oligomers in GUA+VL+AN may be due to VL-derived phenoxy radicals induced by 'OH or 'NO₂ from nitrate photolysis. In addition, more nitrated compounds observed in GUA+DMB+AN and GUA+VL+AN than in GUA+AN imply that photosensitized reactions may promote nitrate-mediated photolytic reactions. On a related note, the significance of photosensitization by BrC (via formation of solvated electrons; Y. Wang et al., 2021) and marine dissolved organic matter (via O₂ formation; Garcia et al., 2021) in enhanced nitrite production from nitrate photolysis have been reported. A recent study from our group has shown that glyoxal photo-oxidation mediated by both nitrate photolysis and photosensitization can significantly enhance the atmospheric sink of glyoxal (Zhang et al., 2022). Further studies are needed to improve our understanding of the interplay between photosensitized reactions and nitrate photolysis.

This study demonstrates that the structural features of photosensitizers affect aqSOA formation via non-carbonyl phenol oxidation. The VL results are broadly relevant to other phenolic carbonyls, but the effects of different functional groups should still be considered. For instance, the aldehyde/ketone pair of syringaldehyde and acetosyringone, both phenolic carbonyls, have been reported to have equal reactivity towards direct photosensitized oxidation. This is due to the greater light absorption by the aldehyde form but higher quantum efficiency for loss for the ketone form (Smith et al. 2016). However, more aqSOA was observed from syringaldehyde than acetosyringone (in either AN or ammonium sulfate; Huang et al., 2018). Our findings also imply that while the contributions of photosensitization by ³VL* (and other phenolic carbonyls) to aqSOA formation would be relatively less compared to that of ³DMB* (and other non-phenolic carbonyls), these are not negligible. As both non-phenolic and phenolic carbonyls such as the methoxybenzaldehydes examined in this work are emitted in large amounts from biomass burning, future experiments should probe the aqSOA contribution of a wider variety of photosensitizers. Moreover, further experiments on photosensitized reactions in authentic particulate matter (PM) samples should be conducted in the future. Multicomponent reactions such as GUA+DMB+AN and GUA+VL+AN should also be explored for a more accurate simulation of ambient conditions. These would be useful in assessing the overall impact of photosensitized reactions and AN photolysis on aqSOA formation in areas impacted by biomass burning and high AN concentrations, and for their better representation in aqSOA models.

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- 470 Data availability.
- 471 The data used in this publication are available to the community and can be accessed by request to the corresponding author.
- 472 Author contributions.
- 473 BRGM designed and conducted the experiments; BRGM and CKC wrote the paper. All co-authors contributed to the
- 474 discussion of the manuscript.
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Table 1. Reaction conditions, initial GUA (and DMB or VL) decay rate constants, normalized abundance of products, average elemental ratios, and average carbon oxidation state ($\langle OS_C \rangle$) in each experiment. The reaction systems consisted of GUA (0.1 mM), DMB (0.01 mM), VL (0.01 mM), and AN (1 mM) under air-saturated conditions after 180 min of simulated sunlight irradiation. The UHPLC-HESI-Orbitrap-MS data were obtained in both positive (POS) and negative (NEG) ion modes.

Exp no.	Reaction conditions	Initial GUA (and DMB or VL) decay rate constants (min ⁻¹ /s ⁻¹) ^a	Normalized abundance of products ^b	Normalized abundance of N- containing compounds ^b	(O:C) ^c	⟨H:C⟩ ^c	⟨N:C⟩ ^c	⟨OSc⟩ ^c
1	GUA+DMB	GUA: 6.3 ± 0.25 DMB: 0.78 ± 0.10	376 ± 22	NA	POS: 0.34	0.91	NA	-0.22
		DIVID: 0.78 ± 0.10			NEG: 0.40	0.94	NA	-0.15
2	GUA+ DMB+AN	GUA: 5.3 ± 0.50	310 ± 4	114	POS: 0.28	0.94	0.12	-0.03
		DMB: 0.69 ± 0.052			NEG: 0.37	0.91	0.04	-0.05
3	GUA+VL	GUA: 1.5 ± 0.14 VL: 3.6 ± 0.55	94 ± 5	NA	POS: 0.41	0.91	NA	-0.10
					NEG: 0.40	0.94	NA	-0.14
4	GUA+	GUA: 1.6 ± 0.12	100 ± 2	8	POS: 0.31	1.02	0.02	-0.34
	VL+AN	VL: 2.9 ± 0.032			NEG: 0.39	0.91	0.03	-0.02
5	GUA+AN	0.57 ± 0.036	23 ± 1	9	POS: 0.35	0.99	0.16	0.19
					NEG: 0.38	1.01	0.05	-0.08

a The data fitting was performed in the initial linear region. Each value is the average of results from triplicate experiments, corrected for internal light screening due to DMB, VL, and AN, and normalized to the experimental photon flux. Errors represent one standard deviation. The normalized product abundance was calculated using the data from UHPLC-HESI-Orbitrap-MS in the positive (POS) ion mode as the GUA signal from the negative (NEG) ion mode was weak, which may introduce significant uncertainties during normalization. The uncertainties were propagated from the changes in [GUA] measured using UHPLC-PDA and the MS signal intensities. The samples for experiments without AN (marked with NA) were not analyzed for N-containing compounds. The average elemental ratios (⟨O:C⟩, ⟨H:C⟩, and ⟨N:C⟩) and ⟨OSc⟩ were based on the UHPLC-HESI-Orbitrap-MS results and estimated using the signal-weighted method (Bateman et al., 2012). The OS_C of GUA, DMB, and VL are -0.57, -0.44, and -0.25, respectively.

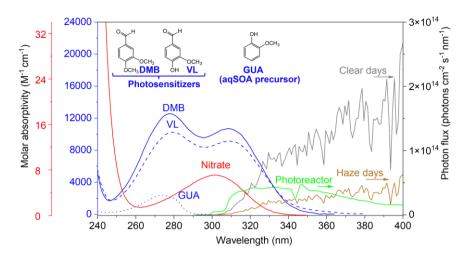


Figure 1. The base-10 molar absorptivities (M⁻¹ cm⁻¹) of 3,4-dimethoxybenzaldehyde (DMB, blue solid line), vanillin (VL, blue dashed line), guaiacol (GUA, blue dotted line), and nitrate (red solid line). The green line is the photon flux in the aqueous photoreactor. The gray and brown lines are the photon fluxes on typical clear and haze days, respectively, in Beijing, China (Mabato et al., 2022). The top of the figure also shows the structures of DMB, VL, and GUA.

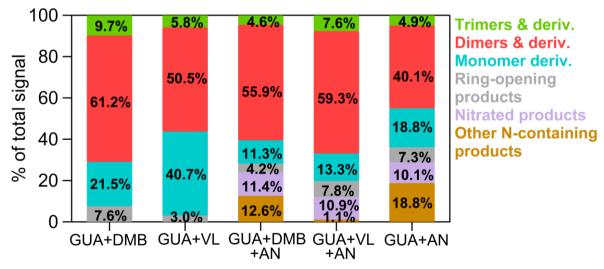


Figure 2. Signal-weighted distributions of aqSOA from GUA+DMB, GUA+VL, GUA+DMB+AN, GUA+VL+AN, and GUA+AN. These product distributions were calculated from combined UHPLC-HESI-Orbitrap-MS data obtained in positive (POS) and negative (NEG) ion modes. The values indicate the contribution of different product classifications to the total signals for each reaction condition.

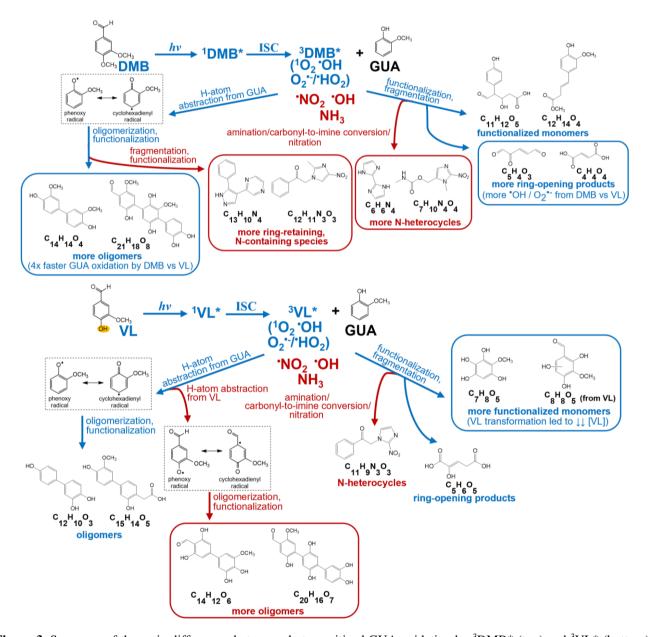


Figure 3. Summary of the main differences between photosensitized GUA oxidation by ³DMB* (top) and ³VL* (bottom) in the absence (blue labels and boxes) and presence (red labels and boxes) of ammonium nitrate at pH 4 under air-saturated conditions. Boxed structures indicate product classifications with notable differences. DMB and VL absorb light and are promoted to their singlet excited states (¹DMB* and ¹VL*), which then undergo intersystem crossing (ISC) to form ³DMB* and ³VL*. Secondary oxidants (¹O₂, O₂-/HO₂, OH) can be formed from ³DMB* and ³VL* upon reactions with O₂ and GUA (George et al., 2018; Chen et al., 2020; Misovich et al., 2021; Mabato et al., 2022). The structures shown are examples of the major products (Tables S1 to S4) for different product classifications.



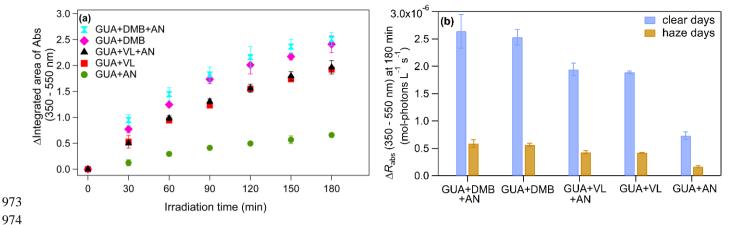


Figure 4. (a) Increase in light absorption throughout 180 min of irradiation for all reaction systems studied and (b) Change in the rate of sunlight absorption (ΔR_{abs}) from 350-550 nm at 180 min during typical clear and haze days in Beijing, China for aqSOA from GUA+DMB+AN, GUA+DMB, GUA+VL+AN, GUA+VL, and GUA+AN. Error bars represent one standard deviation of triplicate experiments.

Supplementary material Comparison of aqueous SOA product distributions from guaiacol oxidation by non-phenolic and methoxybenzaldehydes as photosensitizers in the absence and presence of ammonium nitrate

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phenolic

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Section S1. UHPLC-PDA analyses

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An ultra-high performance liquid chromatography system (UHPLC, Waters Acquity H-Class, 33 Waters, Milford, USA) coupled to a photodiode array (PDA) detector (Waters, Milford, USA) 34 was used for the quantification of GUA, DMB, and VL concentrations. The samples were first 35 filtered through a 0.2 µm Chromafil® Xtra PTFE filter (Macherey-Nagel GmbH & Co. KG, 36 Germany). The separation of products was conducted using an Acquity HSS T3 column (1.8 37 38 μm, 2.1 mm × 100 mm; Waters Corp.). The column oven was held at 30 °C, and the autosampler was cooled at 4 °C. The injection volume was set to 5 µL. The binary mobile 39 40 phase was composed of water (A) and acetonitrile (B). The gradient elution was performed at a flow rate of 0.2 mL/min: 0–1 min, 10% eluent B; 1–25 min, linear increase to 90% eluent B; 41 25-29.9 min, hold 90% eluent B; 29.9-30 min, decrease to 10% eluent B; 30-35 min, re-42 43 equilibrate at 10% eluent B for 5 min. GUA, DMB, and VL were analyzed using the channels 44 with UV absorption at 274, 274, and 300 nm, respectively.

Section S2. UHPLC-HESI-Orbitrap-MS analyses

A Thermo Orbitrap Fusion Lumos Mass Spectrometry (Thermo Fisher Scientific, Waltham, 46 MA, USA) connected to a Thermo Scientific UltiMate 3000 UHPLC system (Thermo Fisher 47 Scientific, Waltham, MA, USA) via heated electrospray ionization (HESI) as the interface 48 49 (UHPLC-HESI-Orbitrap-MS) was used to characterize the reaction products. The mobile 50 phases used were 0.1% (v/v) formic acid (in milli-Q water) (A) and acetonitrile (B). The same 51 settings (e.g., column, gradient, oven temperature) used in the UHPLC-PDA (Sect. S1) were applied in the UHPLC-HESI-Orbitrap-MS system. The HESI-MS spectra were acquired in 52 both positive and negative ion modes. The HESI parameters were as follows: Spray voltage, 53 54 2500 V for both positive and negative HESI; sheath gas, 35 arbitrary units; nebulizer auxiliary gas, 10 arbitrary units; sweep gas, 3 arbitrary units. General instrumental parameters were set 55 as follows: ion transfer tube temperature, 320 °C; vaporizer temperature, 350 °C. The mass 56

range for full scan MS was set at 50-1000 m/z with a mass resolution of 60,000 at 200 m/z. The automatic gain control (AGC) target was 4.0×10^5 with a maximum injection time of 50 ms. The UHPLC-HESI-Orbitrap-MS data obtained in positive and negative ion modes were pretreated using Progenesis QI (version 2.4; Nonlinear Dynamics) for peak picking and alignment. Most peaks detected in the blank (~99% for all experiments) were excluded from the samples except for peaks with a minimum of 2.5 times greater intensity in the sample spectrum than in the blank (Laskin et al., 2014). In addition, a peak was considered a product if the difference in the peak area between the samples before and after irradiation is ≥ 10 times. In this work, two independently prepared samples for each reaction condition were analyzed using the UHPLC-HESI-Orbitrap-MS. Only peaks that were reproducibly detected in both sets of samples were retained. The formula assignments were carried out using the MIDAS molecular formula calculator (http://magnet.fsu.edu/~midas/) with the following constraints: C \leq 100, H \leq 150, O \leq 30, and N \leq 10, and mass error of 10 ppm. The nitrogen atom was excluded in the constraints for experiments without AN. The ChemSpider database (Royal Society of Chemistry) was also queried to return valid molecules that may be useful for proposing product structures. Overall, the proposed structures in this work are based on the molecular formulas, DBE values, and structural and mechanistic information provided in earlier similar works on methoxyphenols (Yee et al., 2013; Li et al., 2014; Yu et al., 2014, 2016; He et al., 2019; Chen et al., 2020; Jiang et al., 2021; Misovich et al., 2021; Mabato et al., 2022). For clarity, the formulas discussed in this work correspond to neutral analytes (e.g., with H⁺ or NH₄⁺ removed from the ion formula).

The double bond equivalent (DBE) values (Koch and Dittmar, 2006) and carbon oxidation state (OS_C; Kroll et al., 2011, 2015; Lv et al., 2016) of the neutral formulas were calculated using the following equations:

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82 DBE =
$$C - H/2 + N/2 + 1$$
 (Eq. S1)

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$$OS_C = 2 \times O/C + 3 \times N/C - H/C$$
 (Eq. S2)

- 84 where C, H, O, and N correspond to the number of carbon, hydrogen, oxygen, and nitrogen
- atoms in the neutral formula. Moreover, the average oxygen to carbon (O:C) ratios, (O:C):
- 86 $((0: C) = \sum_{i} (abundance_{i}) O_{i} / \sum_{i} (abundance_{i}) C_{i})$, average nitrogen to carbon (N:C) ratios,
- 87 $\langle N:C \rangle$: $(\langle N:C \rangle = \sum_i (abundance_i) N_i / \sum_i (abundance_i) C_i)$, and average hydrogen to carbon
- 88 (H:C) ratios, $\langle \text{H:C} \rangle$: ($\langle \text{H:C} \rangle = \sum_{i} (\text{abundance}_{i}) H_{i} / \sum_{i} (\text{abundance}_{i}) C_{i}$) after the reactions
- 89 were further estimated using the signal-weighted method (Bateman et al., 2012). The average
- 90 OS_C , $\langle OS_C \rangle$ was also calculated as follows:

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$$\langle OSc \rangle = 2 \times \langle O:C \rangle + 3 \times \langle N:C \rangle - \langle H:C \rangle$$
 (Eq. S3)

- 92 Section S3. IC analyses of small organic acids
- 93 An ion chromatography system (IC, Dionex ICS-1100, Sunnyvale, CA) equipped with a
- 94 Dionex AS-DV autosampler (Sunnyvale, CA) enabled the analyses of small organic acids. The
- separation was achieved using an IonPacTM AS11 column (4×250 mm) with an IonPacTM
- AG11 guard column (4×50 mm). The isocratic elution was applied at a 1.0 mL/min flow rate
- 97 with 12 mM sodium hydroxide (NaOH) as the eluent. The total run time was set at 10 min. The
- 98 standard solutions (1–50 μM) of formic, succinic, and oxalic acid were analyzed three times
- 99 along with the samples and water blank. Formic, succinic, and oxalic acid had retention times
- of 1.9 min, 3.7 min, and 5.9 min, respectively.
- 101 Section S4. UV-Vis spectrophotometric analyses
- 102 A UV-Vis spectrophotometer (UV-3600, Shimadzu Corp., Japan) was used to measure the
- absorbance changes for the samples. The absorbance values from 200 to 700 nm were measured
- instantly after sample collection, and measurements were done in triplicate. The change in the
- integrated area of absorbance from 350 to 550 nm was used to represent the absorbance
- enhancements. The increase of light absorption at this wavelength range, where GUA, DMB,

and VL did not initially absorb light and where DMB and VL have little absorption, suggests the formation of light-absorbing products (Smith et al., 2016).

Section S5. Further discussions on van Krevelen diagrams and OSc vs. nc plots for

GUA+DMB and GUA+VL aqSOA

Consistent with the higher contribution of ring-opening species, GUA+DMB had more products with H:C ≥1.5 and O:C ≤0.5 (Fig. S5a–b), possibly due to more oxygenated aliphatic species. GUA+VL (Fig. S5c–d) also had high-relative-abundance products with H:C of ~1 and O:C ≥0.5. Similar to our previous work (0.1 mM GUA + 0.1 mM VL; Mabato et al., 2022), the two high-relative-abundance species with O:C ≥0.5 were associated with hydroxylated products (C₇H₈O₄ and C₈H₈O₅, #28 and 35; Table S2) that were also observed in earlier works on ³DMB* and 'OH-mediated oxidation (Yu et al., 2014, 2016). These hydroxylated products were also present in GUA+DMB but with lower relative abundance. Triplet-mediated phenol oxidation can generate H₂O₂ (Anastasio et al., 1997), a photolytic source of 'OH. Indeed, hydroxylation is significant in aqueous-phase phenol oxidation (Li et al., 2014; Yu et al., 2014, 2016; Chen et al., 2020; Jiang et al., 2021; Misovich et al., 2021; Mabato et al., 2022).

The OS_C vs. n_C plots for both GUA+DMB and GUA+VL display high-relative-abundance species clustered at n_C of 12 to 15 and OS_C >-1, which can be ascribed to dimers and derivatives (Fig. S6a–d). The species with n_C >15 had the highest DBE values and can be attributed to trimers. These compounds were more abundant in GUA+DMB, likely due to the greater extent of photosensitized reactions by $^3DMB^*$ compared to $^3VL^*$. Indeed, oligomerization is an important process in aqSOA formation via triplet-mediated oxidation (Yu et al., 2014, 2016; Chen et al., 2020; Jiang et al., 2021; Misovich et al., 2021; Mabato et al., 2022). As indicated by the higher quantity of low DBE species, ring-opening and fragmentation pathways were more extensive in GUA+DMB. In GUA+VL, there were also high-relative-

- abundance products with $n_C < 10$, $OS_C \ge 0$, and DBE < 5, corresponding to the hydroxylated
- products mentioned earlier.
- 133 Section S6. Estimation of the apparent quantum efficiency of guaiacol photodegradation
- 134 The apparent quantum efficiency of GUA photodegradation (φ_{GUA}) in the presence of DMB,
- VL, or nitrate during simulated sunlight illumination can be defined as (Anastasio et al., 1997;
- 136 Smith et al., 2014, 2016):

137
$$\Phi_{\text{GUA}} = \frac{\text{mol GUA destroyed}}{\text{mol photons absorbed}}$$
 (Eq. S4)

- Φ_{GUA} was calculated using the measured rate of GUA decay and rate of light absorption by
- DMB, VL, or nitrate through the following equation:

140
$$\Phi_{\text{GUA}} = \frac{\text{rate of GUA decay}}{\text{rate of light absorption by DMB or VL or nitrate}} = \frac{k l_{\text{GUA}} \times [\text{GUA}]}{\sum [(1-10^{-\varepsilon} \lambda^{[\text{C}]l}) \times l_{\lambda}']}$$
(Eq. S5)

- where k'_{GUA} is the pseudo-first-order rate constant for GUA decay, [GUA] is the concentration
- of GUA (M), ε_{λ} is the base-10 molar absorptivity (M⁻¹ cm⁻¹) of DMB, VL, or nitrate at
- wavelength λ , [C] is the concentration of DMB, VL, or nitrate (M), l is the pathlength of the
- illumination cell (cm), and Γ_{λ} is the volume-averaged photon flux (mol-photons L⁻¹ s⁻¹ nm⁻¹)
- determined from 2NB actinometry:

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$$j(\text{2NB}) = 2.303 \times \Phi_{\text{2NB}} \times l \times \Sigma_{300 \text{ }nm}^{350 \text{ }nm} (\varepsilon_{\text{2NB},\lambda} \times l'_{\lambda} \times \Delta \lambda)$$
 (Eq. S6)

- where j(2NB) is the decay rate constant of 2-nitrobenzaldehyde (2NB), the chemical
- actinometer used to determine the photon flux in the aqueous photoreactor, $\Phi_{2NB,\lambda}$ and $\varepsilon_{2NB,\lambda}$
- are the quantum yield (molecule photon⁻¹) and base-10 molar absorptivity (M⁻¹ cm⁻¹) for 2NB,
- respectively, and $\Delta\lambda$ is the wavelength interval between actinic flux data points (nm).
- 151 Section S7. Further discussions on van Krevelen diagrams and OSc vs. nc plots for
- 152 GUA+DMB+AN, GUA+VL+AN, and GUA+AN aqSOA
- 153 The position of the CHO, CHON, and CHN species in the van Krevelen diagrams for
- 154 GUA+DMB+AN and GUA+VL+AN broadly resembled those of CHO species in the absence

of AN (Fig. S5). The CHON species for GUA+DMB+AN and GUA+VL+AN mostly had O:C ratios <0.7, consistent with previous studies on BBOA e.g., wheat straw burning in K-Puszta in the Great Hungarian Plain of Hungary, biomass burning at Canadian rural sites such as Saint Anicet, and BBOA from Amazonia (Schmitt-Kopplin et al., 2010; Claeys et al., 2012; Kourtchev et al., 2017).

The CHN species in GUA+DMB+AN and GUA+VL+AN appeared to have analogous H:C ratios. GUA+DMB+AN had ~2 times more CHON and CHN species than GUA+VL+AN, and there were more of these species with higher abundance in the former, indicating a greater extent of reactions with AN. The high-relative-abundance products for GUA+DMB+AN and GUA+VL+AN were similar to those in the absence of AN, except the hydroxylated products (e.g., C₇H₈O₄; #28; Table S2) previously mentioned for GUA+VL. Among the high-relative-abundance products for GUA+DMB+AN was a CHN species with H:C of ~0.8. For GUA+VL+AN, the high-relative-abundance products include two CHON species with O:C and H:C ratios of 0.3-0.6 and 0.6-0.8. The major difference between GUA+AN and GUA+DMB+AN/GUA+VL+AN was the presence of more high-relative-abundance CHON and CHN species (Fig. S9) in GUA+AN which can be expected given that AN was the only source of oxidants in this case. Compared to GUA+AN, more species (CHO, CHON, and CHN) were observed for GUA+DMB+AN and GUA+VL+AN, attributable to contributions from both photosensitization and (ammonium) nitrate photolysis.

Moreover, GUA+DMB+AN and GUA+VL+AN aqSOA had mainly similar features in the OS_C vs. n_C plots as those observed in the absence of AN (Fig. S6). GUA+DMB+AN and GUA+VL+AN aqSOA also had more CHON and CHN species with higher OS_C, n_C, and DBE (Fig. S6e–h) relative to GUA+AN (Fig. S10), indicating more conjugated N-containing compounds. For GUA+DMB+AN and GUA+VL+AN, the CHON and CHN species had a wider range of OS_C compared to CHO species (Fig. S6e–h). The high-relative-abundance

species (n_C of 12 to 15 and $OS_C >-1$) corresponded to dimers and trimers similar to those noted in the absence of AN, along with some N-containing species. These include a CHN species with n_C of 13, $OS_C \sim 0$, and 11 DBE for GUA+DMB+AN, and 2 CHON species with n_C of 5 and 11, OS_C of 2.5 and 1, and 6 and 9 DBE for GUA+VL+AN.

No.	GUA+DMB POS Molecular formula and exact mass	DBE	Possible structure	No.	GUA+DMB NEG Molecular formula and exact mass	DBE	Possible structure
1	C ₁₄ H ₁₄ O ₄ (246.0892)	8	HO OCH ₃ OCH ₃ OCH ₃		C ₁₄ H ₁₄ O ₄ (246.0892) (No. 1; GUA+DMB POS)		
2	C ₁₃ H ₁₀ O ₃ (214.0630)	9	ОН	16	C ₁₄ H ₁₄ O ₆ (278.0790)	8	HO OCH ₃
3	C ₁₄ H ₁₂ O ₄ (244.0736)	9	HO OCH ₃	17	C ₁₂ H ₁₀ O ₄ (218.0579)	8	ОН
4	C ₁₃ H ₁₀ O ₄ (230.0579)	9	но	C ₁₃ H ₁₂ O ₄ (232.0736) (No. 6; GUA+DMB POS)			
5	C ₁₃ H ₁₀ O ₅ (246.0528)	9	ОН	18	C ₇ H ₁₀ O ₅ (174.0528)	3	H ₃ CO OCH ₃
6	C ₁₃ H ₁₂ O ₄ (232.0736)	8	HO OCH ₃	19	C ₂₁ H ₁₈ O ₈ (398.1002)	13	O OCH ₃ OH OCH ₃ OH OH
7	C ₁₄ H ₁₂ O ₅ (260.0685)	9	HO OCH ₃	20	C ₁₃ H ₁₂ O ₆ (264.0634)	8	HO OH OH
8	C ₁₁ H ₁₂ O ₅ (224.0685)	6	OH OH OH	21	C ₂₀ H ₁₈ O ₆ (354.1103)	12	OCH ₃ OH OCH ₃ OH OH OH
9	C ₁₄ H ₁₂ O ₇ (292.0583)	9	OH OCH3	22	C ₁₄ H ₁₄ O ₇ (294.0740)	8	HO OH OH
10	C ₁₁ H ₁₄ O ₆ (242.0790)	5	OH OCH ₃	23	C ₁₂ H ₁₄ O ₄ (222.0892)	6	OH OCH ₃

11	C ₁₈ H ₁₈ O ₇ (346.1053)	10	HO OCH3 OCH3 OCH3	24	C ₁₃ H ₁₀ O ₆ (262.0477)	9	OH OH OH
12	C ₁₀ H ₁₂ O ₃ (180.0786)	5	ОН	25	C ₁₃ H ₁₄ O ₄ (234.0892)	7	OCH ₃
13	C ₇ H ₆ O ₄ (154.0266)	5	HO OH	26	C ₁₄ H ₁₄ O ₅ (262.0841)	8	HO OCH ₃ OCH ₃ OH
14	C ₁₆ H ₁₈ O ₆ (306.1103)	8	H ₃ CO OCH ₃ OH	C ₁₃ H ₁₀ O ₅ (246.0528) (No. 5; GUA+DMB POS)			
15	C ₇ H ₆ O ₅ (170.0215)	5	HO OH	27	C ₁₉ H ₁₆ O ₆ (340.0947)	12	HO OH OH OH

No.	GUA+VL POS Molecular formula and exact mass	DBE	Possible structure	No.	GUA+VL NEG Molecular formula and exact mass	DBE	Possible structure	
28	C ₇ H ₈ O ₄ (156.0423)	4	HO OCH ₃	35	C ₈ H ₈ O ₅ (184.0372)	5	OOH OCH3	
	C ₁₃ H ₁₀ O ₄ (`	,	C ₁₃ H ₁₂ O ₄ (232.0736) (No. 6; GUA+DMB, Table S1)				
	(No. 4; GUA+I C ₁₃ H ₁₂ O ₄ ((No. 6; GUA+1 C ₁₄ H ₁₄ O ₄ (
	(No. 6; GUA+I	•	*		(No. 1; GUA+I	`	,	
	$C_{13}H_{10}O_5$ ($C_{14}H_{14}O_{6}$	•	•	
	(No. 5; GUA+I	OMB, Tal	ole S1)		(No. 16; GUA+	DMB, Tal	ole S1)	
29	$C_7H_8O_5$ (172.0372)	4	HO OCH ₃		C ₂₀ H ₁₈ O ₆ (No. 21; GUA+	•	•	
30	$C_6H_6O_2$ (110.0368)	4	OH	C ₁₂ H ₁₀ O ₄ (218.0579) (No. 17; GUA+DMB, Table S1)				
31	$C_{10}H_{10}O_4$ (194.0579)	6	H ₃ CO OH	C ₆ H ₆ O ₂ (110.0368) (No. 30; GUA+VL POS)				
32	C ₁₁ H ₈ O ₄ (204.0423)	8	ОПО	C ₇ H ₁₀ O ₅ (174.0528) (No. 18; GUA+DMB, Table S1)				
33	C ₁₂ H ₁₀ O ₃ (202.0630)	8	НООНОН	36	C ₁₅ H ₁₄ O ₅ (274.0841)	9	OCH ₃	
	C ₁₄ H ₁₂ O ₅ ((No. 7; GUA+I		*		C ₁₃ H ₁₂ O ₆ (No. 20; GUA+	•	•	
	C ₁₃ H ₁₄ O ₄ (234.0892) (No. 25; GUA+DMB, Table S1) 37 C ₈ H ₈ O ₄ (168.0423) 5				ОСН₃			
34	C ₁₁ H ₁₀ O ₆ (238.0477)	7	OH OCH ₃		C ₁₉ H ₁₆ O ₆ (No. 27; GUA+			
	C ₁₃ H ₁₀ O ₆ (262.0477) (No. 24; GUA+DMB, Table S1)				C ₁₁ H ₁₀ O ₆ (238.0477) (No. 34; GUA+VL POS)			
	C ₁₃ H ₁₂ O ₆ (264.0634) (No. 20; GUA+DMB, Table S1)				C ₅ H ₆ O ₅ (146.0215)	3	но он	
	C ₇ H ₆ O ₄ (1) (No. 13; GUA+1)			39	C ₆ H ₄ O ₄ (140.0110)	5	ОН	

No.	GUA+DMB+AN POS Molecular formula and exact mass	DBE	Possible structure	No.	GUA+DMB +AN NEG Molecular formula and exact mass	DBE	Possible structure	
$C_{14}H_{14}$	4O ₄ (246.0892) (No. 1;	GUA+D	MB, Table S1)	C ₁₃ H	12O4 (232.0736) (No.	6; GUA+	-DMB, Table S1)	
40	$C_{13}H_{10}N_4$ (222.0905)	11	Z H	C ₁₄ H ₁₄ O ₆ (278.0790) (No. 16; GUA+DMB, Table S1)				
$C_{13}H_{1}$	O ₅ (246.0528) (No. 5;	GUA+D	MB, Table S1)	C ₁₄ H	₁₄ O ₄ (246.0892) (No.	1; GUA+	-DMB, Table S1)	
$C_{13}H_1$	₀ O ₄ (230.0579) (No. 4;	GUA+D	MB, Table S1)	$C_{12}H_1$	₀ O ₄ (218.0579) (No.	17; GUA-	+DMB, Table S1)	
41	C ₆ H ₆ N ₄ (134.0592)	6	Z Z Z		C ₂₁ H ₁₈ O ₈ (No. 19; GUA+	•	*	
$C_{13}H_{12}$	₂ O ₄ (232.0736) (No. 6;	GUA+D	MB, Table S1)	C ₇ H ₁₀	O ₅ (174.0528) (No.	18; GUA+	-DMB, Table S1)	
42	$C_{12}H_{11}N_3O_3 \\ (245.0800)$	9	O N NO2		$C_{13}H_{12}O_{6}$ (No. 20; GUA+			
43	C ₁₀ H ₈ N ₄ O (200.0698)	9	N - 2 N N N N N N N N N N N N N N N N N	48	C ₁₆ H ₁₄ N ₆ O ₄ (354.1076)	13	H ₂ N N N N N N N N N N N N N N N N N N N	
44	C ₆ H ₆ N ₄ O (150.0542)	6	HN NH	49	C ₁₅ H ₁₀ N ₄ O ₃ (294.0753)	13	NO ₂	
45	C ₁₀ H ₁₄ N ₄ O ₄ (245.1015)	6	NH O ₂ N NO ₂		C ₁₃ H ₁₀ O ₆ (262.0477) (No. 24; GUA+DMB, Table S1)			
46	C ₁₃ H ₁₀ N ₄ O (238.0855)	11	N N N N N N N N N N N N N N N N N N N		C ₁₀ H ₁₀ O ₄ (194.0579) (No. 31; GUA+VL, Table S2)			
	C ₁₃ H ₁₂ O ₆ (264.0634) (No. 20; GUA+DMB, Table S1)				C ₇ H ₈ O ₄ (156.0423) (No. 28; GUA+VL, Table S2)			
	C ₇ H ₆ O ₄ (154.0266) (No. 13; GUA+DMB, Table S1)				4O ₄ (234.0892) (No.		·	
$C_{12}H_1$	₀ O ₃ (202.0630) (No. 33	3; GUA+`	VL, Table S2)	$C_{13}H_1$	₁₀ O ₅ (246.0528) (No.	5; GUA+	-DMB, Table S1)	
47	$C_{13}H_8O_4$ (228.0423)	10	но		C ₁₄ H ₁₄ O ₅ (No. 26; GUA+	•	*	

No.	GUA+VL+AN POS Molecular formula and exact mass	DBE	Possible structure	No.	GUA+VL+AN NEG Molecular formula and exact mass	DBE	Possible structure	
	$C_{14}H_{14}O_4$	`	′		$C_{13}H_{12}O_4$ (′	
	(No. 1; GUA+	DMB, T	able S1)		(No. 6; GUA+D		· · · · · · · · · · · · · · · · · · ·	
50	$C_{10}H_8O_2$ (160.0524)	7		C ₁₄ H ₁₄ O ₆ (278.0790) (No.16; GUA+DMB, Table S1)				
51	C ₁₆ H ₁₈ O ₄ (274.1205)	8	OCH ₃	C ₁₂ H ₁₀ O ₄ (218.0579) (No.17; GUA+DMB, Table S1)				
	C ₁₁ H ₁₂ O ₅ (No. 8; GUA+		′	57	C ₁₁ H ₉ N ₃ O ₃ (231.0644)	9	N NO2	
	$C_{14}H_{12}O_{12}$	- (,		C ₇ H ₁₀ O ₅ (,	
	(No. 7; GUA+ C ₁₂ H ₁₄ O ₄		/		(No.18; GUA+I	•	·	
	(No. 23; GUA-	•	· ·	C ₁₅ H ₁₄ O ₅ (274.0841) (No. 36; GUA+VL, Table S2)				
52	C ₁₁ H ₁₂ O ₄ (208.0736)	6		C ₁₃ H ₁₂ O ₆ (264.0634) (No. 20; GUA+DMB, Table S1)				
	C ₆ H ₆ N ₄ O (No. 44; GUA+D	•	,	58	C ₅ H ₆ O ₂ (98.0368)	3	ОН	
	C ₁₃ H ₁₂ O ₄ (No. 6; GUA+	(232.07	36)	$C_{19}H_{16}O_6$ (340.0947) (No. 27; GUA+DMB, Table S1)				
53	C ₁₂ H ₈ N ₂ O ₃ (228.0535)	10	N NO ₂	59	C ₂₀ H ₁₆ O ₇ (368.0896)	13	OCH ₃ OH OH OH	
54	C ₁₁ H ₁₄ O ₄ (210.0892)	5	OH O	C ₂₁ H ₁₈ O ₈ (398.1002) (No. 19; GUA+DMB, Table S1)				
	C ₇ H ₆ O ₄ (No. 13; GUA+			C ₇ H ₆ O ₄ (154.0266) (No. 13; GUA+DMB, Table S1)				
55	C ₁₄ H ₁₂ O ₆ (276.0634)	9	OCH ₃ OH OH OH	C ₁₅ H ₁₀ N ₄ O ₃ (294.0753) (No. 49; GUA+DMB+AN, Table S3)				
56	C ₁₄ H ₁₀ N ₄ O ₇ (346.0550)	12	O ₂ N HO NH	C ₁₃ H ₁₀ O ₆ (262.0477) (No. 24; GUA+DMB, Table S1)				
	C ₁₃ H ₁₂ O ₆ (No. 20; GUA+	*	*	C ₅ H ₆ O ₅ (146.0215) (No. 38; GUA+VL, Table S2)				

No.	GUA+AN POS Molecular formula and exact mass	DBE	Possible structure	No.	GUA+AN NEG Molecular formula and exact mass	DBE	Possible structure	
	C ₁₃ H ₁₀ O ₄ (No. 4; GUA+I	•	*		C ₁₄ H ₁₄ O ₆ (No. 16; GUA+	•	<i>'</i>	
(C ₆ H ₆ N ₄ O (No. 44; GUA+DM	(150.0542	2)	68	C ₁₂ H ₁₉ N ₃ O (221.1528)	5	H N N N N N N N N N N N N N N N N N N N	
	C ₁₁ H ₁₂ O ₅ (No. 8; GUA+l	•	<i>'</i>		C ₁₂ H ₁₀ O ₄ (No. 17; GUA-	•	<i>'</i>	
	C ₇ H ₈ O ₄ ((No. 28; GUA-				C ₁₄ H ₁₄ O ₄ (No. 1; GUA+	*	*	
60	C ₆ H ₄ N ₄ (132.0436)	7	Z= Z= Z= Z= Z= Z= Z= Z= Z= Z= Z= Z= Z= Z		C ₂₀ H ₁₈ O ₆ (No. 21; GUA+	`	,	
61	C ₁₂ H ₁₄ O ₅ (238.0841)	6	HO OCH3	C ₇ H ₁₀ O ₅ (174.0528 (No. 18; GUA+DMB, Table S1)				
62	C ₁₃ H ₁₂ N ₄ O ₅ (304.0808)	10	NO ₂ H NH NH	69	C ₄ H ₃ N ₃ O ₃ (141.0174)	5	O N NO ₂	
	C ₁₃ H ₁₂ O ₆ (No. 20; GUA+	•	<i>'</i>		C ₁₃ H ₁₂ O ₆ (No. 20; GUA+	•	*	
	C ₁₃ H ₁₂ O ₄ (No. 6; GUA+I	•	*	70	C ₁₂ H ₆ N ₄ O ₅ (286.0338)	12	NO ₂	
63	C ₈ H ₁₀ N ₄ O (178.0855)	6	HO Z Z	71	$C_{13}H_{12}O_5 \\ (248.0685)$	8	HO OCH3	
64	C ₉ H ₁₄ N ₄ O (194.1168)	5	NH ₂ H	72	C ₆ H ₆ O ₄ (142.0266)	4	НО ОН	
65	C ₈ H ₄ N ₄ (156.0436)	9	N N N	C ₁₂ H ₁₀ O ₃ (202.0630) (No. 33; GUA+VL, Table S2)				
66	C ₁₅ H ₁₉ N ₅ O ₂ (301.1539)	9	NH ₂ N N N N N N N N N N N N N N N N N N N	73	C ₁₂ H ₁₂ O ₄ (220.0736)	7		
67	C ₇ H ₁₀ N ₄ O ₄ (214.0702)	5	$ \bigwedge_{N=1}^{N} \bigcap_{N=1}^{N} NO_{2} $	$C_7H_6O_5$ (170.0215) (No. 15; GUA+DMB, Table S1)				
	C ₇ H ₈ O ₅ ((No. 29; GUA			C ₇ H ₈ O ₄ (156.0423) (No. 28; GUA+VL, Table S2)				

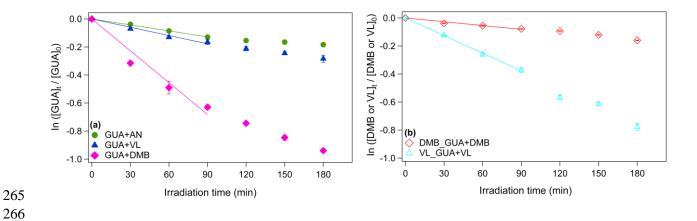


Figure S1. (a) The decay of GUA during (ammonium) nitrate-mediated photo-oxidation (GUA+AN) and photosensitized oxidation by ${}^{3}VL^{*}$ (GUA+VL) or ${}^{3}DMB^{*}$ (GUA+DMB). (b) The decay of DMB or VL during GUA photo-oxidation in GUA+DMB and GUA+VL, respectively. No statistically significant difference (p > 0.05) was noted between GUA+DMB and GUA+DMB+AN and between GUA+VL and GUA+VL+AN. Error bars represent 1 standard deviation; most error bars are smaller than the markers.

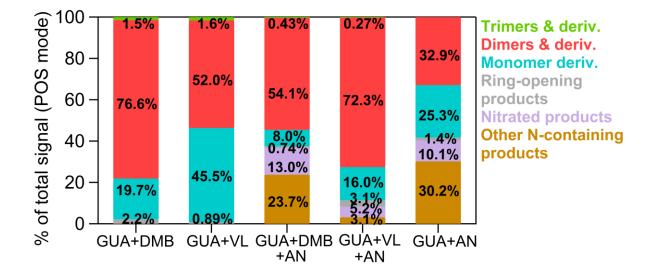


Figure S2. Signal-weighted distributions of aqSOA from GUA+DMB, GUA+VL, GUA+DMB+AN, GUA+VL+AN, and GUA+AN. These product distributions were calculated from UHPLC-HESI-Orbitrap-MS data obtained in the positive (POS) ion mode. The values indicate the contribution of different product classifications to the total signals for each reaction condition.

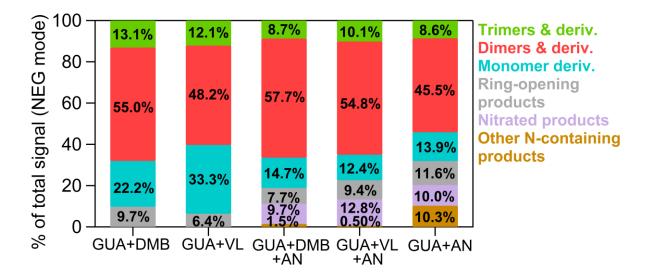


Figure S3. Signal-weighted distributions of aqSOA from GUA+DMB, GUA+VL, GUA+DMB+AN, GUA+VL+AN, and GUA+AN. These product distributions were calculated from UPLC-HESI-Orbitrap-MS data obtained in the negative (NEG) ion mode. The values indicate the contribution of different product classifications to the total signals for each reaction condition.

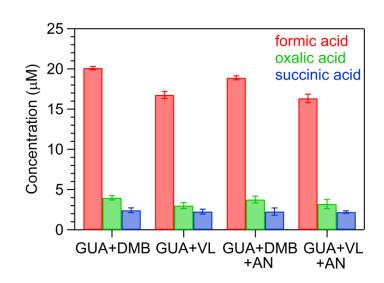


Figure S4. The concentration of formic, oxalic, and succinic acid for GUA+DMB, GUA+VL, GUA+DMB+AN, and GUA+VL+AN aqSOA. Error bars represent one standard deviation of triplicate experiments.



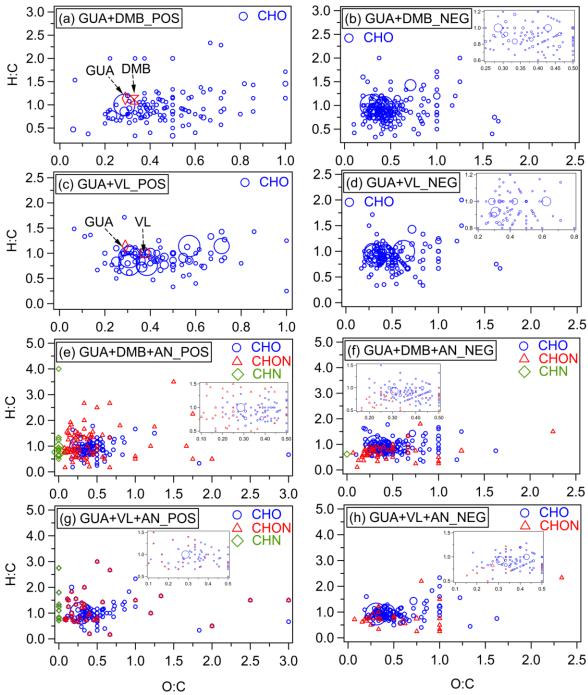


Figure S5. Van Krevelen diagrams of aqSOA from (a, b) GUA+DMB, (c, d) GUA+VL, (e, f) GUA+DMB+AN, and (g, h) GUA+VL+AN for positive (POS) and negative (NEG) ion modes. The blue circle markers indicate CHO classes, red triangle indicate CHON classes, and green diamond indicate CHN classes. The marker size reflects the relative abundance in the sample. The location of GUA, DMB, and VL in the plots are indicated only in panels a and c (red markers). The insets are expanded views of the crowded sections of the van Krevelen diagrams. Note the different scales on the axes.

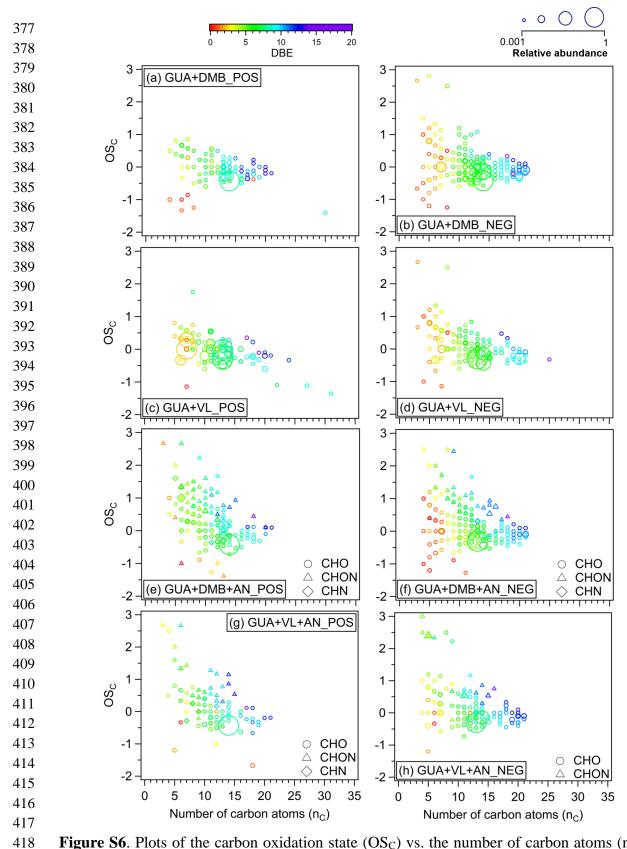


Figure S6. Plots of the carbon oxidation state (OS_C) vs. the number of carbon atoms (n_C) of aqSOA from (a, b) GUA+DMB, (c, d) GUA+VL, (e, f) GUA+DMB+AN, and (g, h) GUA+VL+AN for positive (POS) and negative (NEG) ion modes, colored by the double bond equivalent (DBE) values. The circle, triangle, and diamond markers indicate CHO, CHON and CHN classes, respectively. The marker size reflects the relative abundance in the sample.

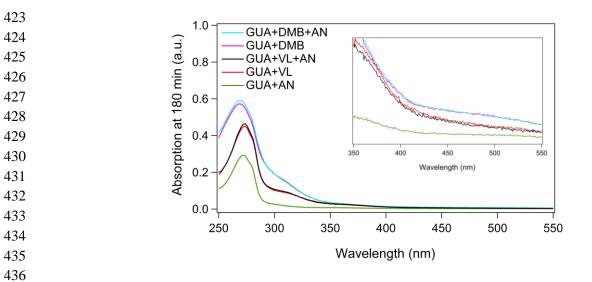


 Figure S7. UV-Vis absorption spectra of GUA+DMB+AN, GUA+DMB, GUA+VL+AN, GUA+VL, and GUA+AN after 180 min of irradiation. The inset is the expanded view from 350 to 550 nm.

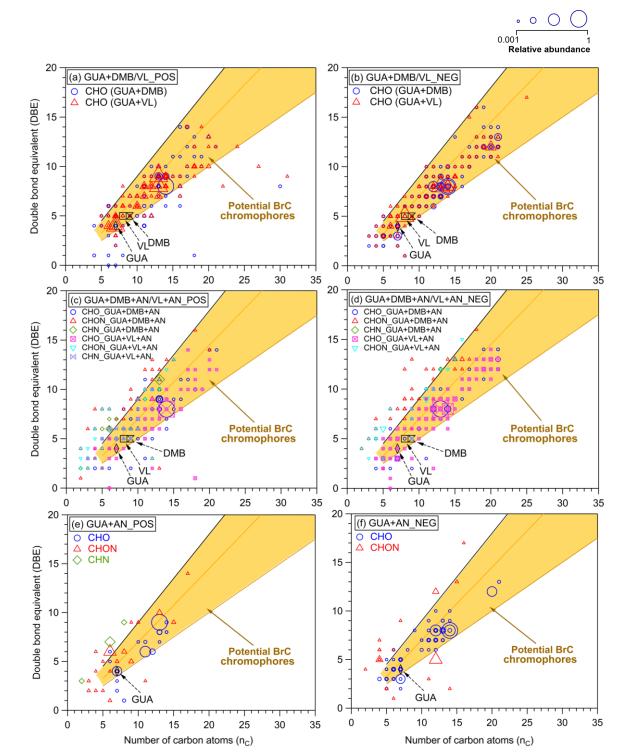


Figure S8. Plots of the double bond equivalent (DBE) values vs. the number of carbon atoms (n_C) (Lin et al., 2018) of aqSOA from (a, b) GUA+DMB and GUA+VL, (c, d) GUA+DMB+AN and GUA+VL+AN, and (e, f) GUA+AN for positive (POS) and negative (NEG) ion modes. For a and b, the blue markers indicate CHO classes for GUA+DMB and red indicate CHO classes for GUA+VL. For c and d, the blue markers indicate CHO classes, red indicate CHON classes, and green indicate CHON classes, cyan indicate CHON classes, and purple indicate CHN classes for GUA+VL+AN. For e and f, the blue markers indicate CHO classes, red indicate CHON classes, and green indicate

CHN classes for GUA+AN. The marker size reflects the relative abundance in the sample. The three lines indicate DBE reference values of fullerene-like hydrocarbons (top, black solid line; Lobodin et al, 2012), cata-condensed polycyclic aromatic hydrocarbons (PAHs; Siegmann and Sattler, 2000) (middle, orange solid line), and linear conjugated polyenes (general formula $C_{\times}H_{\times+2}$) (bottom, brown solid line). Species within the shaded area are potential BrC chromophores.

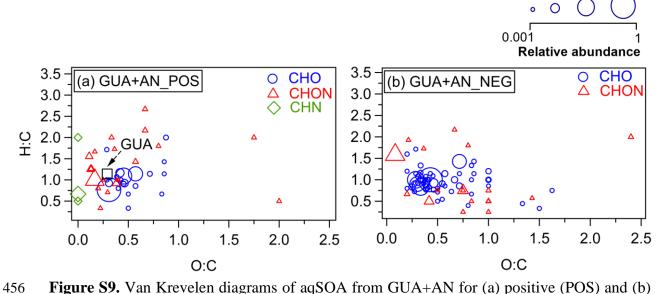


Figure S9. Van Krevelen diagrams of aqSOA from GUA+AN for (a) positive (POS) and (b) negative (NEG) ion modes. The blue markers indicate CHO classes, red indicate CHON classes, and green indicate CHN classes. The marker size reflects the relative abundance in the sample. The location of GUA is indicated only in panel a (black marker).

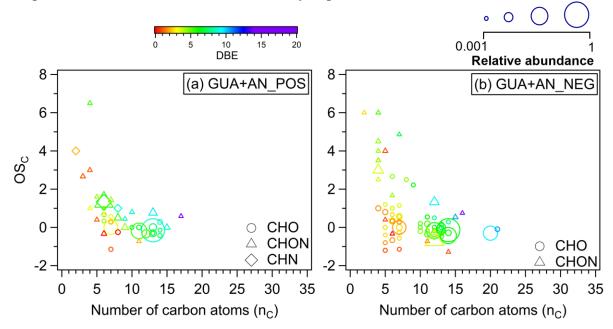


Figure S10. Plots of the carbon oxidation state (OS_C) vs. the number of carbon atoms (n_C) of aqSOA from GUA+AN for (a) positive (POS) and (b) negative (NEG) ion modes, colored by the double bond equivalent (DBE) values. The circle, triangle, and diamond markers indicate CHO, CHON and CHN classes, respectively. The marker size reflects the relative abundance in the sample.

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