Optical and chemical properties and oxidative potential of aqueous-phase products from OH and $^3C^*$-initiated photooxidation of eugenol

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Abstract: Aqueous reactions may turn precursors into light-absorbing and toxic products, leading to air quality deterioration and adverse health effects. In this study, we investigated comprehensively eugenol photooxidation (a representative biomass burning emitted, highly substituted phenolic compound) in bulk aqueous phase with direct photolysis, hydroxyl radical (OH) and an organic triplet excited state ($^3C^*$). Results show that the degradation rates of eugenol followed the order of $^3C^*$>OH >direct photolysis. During $^3C^*$-initiated oxidation, different reactive oxygen species (ROS) including $^3C^*$, OH, $^1O_2$ and $O_2^{\bullet-}$ can participate in oxidation of eugenol, quenching experiments verified $^3C^*$ was the most important one; while during OH-initiated oxidation, $O_2^{\bullet-}$ was a more important ROS than OH to oxidize eugenol. The rate constants under saturated O_2, air and N_2 followed the order of $k_{O_2} > k_{Air} > k_{N_2}$ for

both direct photolysis and OH-initiated oxidation, but changed to $k_{Air} > k_{N_2} > k_{O_2}$ for $^3C^*$-mediated oxidation. pH and dissolved oxygen (DO) levels both decreased during oxidation, indicating formation of acids and the participation of DO in oxidation. UV-vis light absorption spectra of the reaction products showed clear absorbance enhancement in the 300-400 nm range for all three sets of experiments and new fluorescence at excitation/emission=250/(400-500) nm appeared, suggesting the formation of new chromophores and fluorophores (brown carbon species); and these species were likely attributed to humic-like substances (HULIS) as shown by the increases of HULIS concentrations during oxidation. Large mass yields of products (140%-197%) after 23 hours of illumination were obtained, and high oxidation degrees of these products were also observed; correspondingly, a series of oxygenated compounds were identified, and detailed reaction mechanism with functionalization as a dominant pathway was proposed. At last, dithiothreitol (DTT) assay was applied to assess oxidation potential of the reaction products, and the end products of all three sets of experiments showed higher DDT consumption rates than that of eugenol, indicating more toxic species were produced upon aqueous oxidation. Overall, our results by using eugenol as a model compound, underscore the potential importance of aqueous processing of biomass burning emissions in secondary organic aerosol (SOA) formation.

1 Introduction

Photochemical reactions in atmospheric aqueous phases (cloud/fog droplets and aerosol water) can affect lifetimes of many organic species, and are an important source and pathway of secondary organic aerosol (SOA) formation (Vione et al., 2006; Zhao

et al., 2012). Compared to the gasSOA formed via gas-phase photochemical oxidation, aqueous-phase SOA (aqSOA) is often more oxidized and less volatile, therefore might play an important role in haze formation, air quality and global climate change (Ervens et al., 2011; Lim et al., 2010). However, due to complexity of the aqueous reactions and influencing factors (such as precursors, oxidants, and light intensities), detailed reaction mechanism, optical property, oxidative potential (OP) and the interplay among them remain poorly understood.

Many laboratory studies have focused on aqueous-phase oxidations of low molecular weight (LMW) volatile organic compounds (VOCs), such as isoprene, terpenes (α-, β-pinene), as well as their gas-phase oxidation products (such as glyoxal, methylglyoxal, *cis*-pinonic acid and methyl vinyl ketone) (Faust et al., 2017; Herrmann, 2003; Herrmann et al., 2015; Huang et al., 2011; Lee et al., 2012; Zhang et al., 2010). Recently, aqueous oxidation of semi-/intermediate volatility VOCs (S/IVOCs), such as the phenolic compounds emitted from combustion or pyrolysis of lignin in biomass, were also extensively investigated (Barzaghi and Herrmann, 2002; Bonin et al., 2007; Chen et al., 2020; Gilardoni et al., 2016; He et al., 2019; Jiang et al., 2021; Li et al., 2014; Li et al, 2021; Ma et al., 2021; Mabato et al., 2022; Smith et al., 2014; Sun et al., 2010; Tang et al., 2020; Yang et al., 2021; Yu et al., 2016). Generally, chemical structures of precursors have profound influences on the reaction mechanisms and products, while effect of oxidants also cannot be neglected. It is evident that liquid water can contain various types of oxidants, such as singlet oxygen ($^1O_2$), nitrate radical (NO_3), hydroxyl radical (OH), and organic triplet excited states ($^3C^*$), and all can play crucial roles in photooxidation reactions (Kaur and Anastasio, 2018; Scharko et al., 2014). Among them, OH is a ubiquitous oxidant with concentrations of 10^{-13}-10^{-12} mol·L^{-1} (Arakaki et al., 2013; Gligorovski et al., 2015; Herrmann et al., 2003). Hence, aqueous

75 OH-induced photooxidation has been extensively studied (Chen et al., 2020; Sun et al.,

76 2010; Yu et al., 2016). Compared to OH oxidation, $^3C^*$-initiated aqueous oxidation

77 (photosensitized reactions) has also attracted attentions in recent years (Ma et al., 2021;

78 Wang et al., 2021). Several classes of organic compounds in ambient air, including non-

79 phenolic aromatic carbonyls, quinones, aromatic ketones and nitrogen-containing

80 heterocyclic compounds, can form $^3C^*$ after absorbing light (Alegría et al., 1999; Kaur

81 et al., 2019; Nau and Scaiano, 1996; Rossignol et al., 2014; Chen et al., 2018). These

82 compounds are termed photosensitizers. $^3C^*$ is capable of reacting with O_2 to produce

83 singlet oxygen ($^1O_2$) and superoxide radicals ($O_2^{\bullet-}$). Various reactive oxygen species

84 (ROS) can be generated and affect greatly the $^3C^*$-initiated aqueous-phase reactions.

85 Despite some studies demonstrating importance of ROS in photochemical process (Ma

86 et al, 2021;Wang et al., 2020; Wang et al., 2021), our current understanding on $^3C^*$-

87 initiated oxidation is still limited.

88 Excitation-emission matrix (EEM) fluorescence spectroscopy, as a low-cost, rapid,

89 non-destructive and high-sensitivity technique, can offer detailed information on

90 chromophores hence has been widely employed for studies of aquatic dissolved organic

91 matter (Aryal et al., 2015). Nevertheless, it has not been extensively used in

92 atmospheric aerosol research (Mladenov et al., 2011). Prior studies have investigated

93 the relationship between the fluorescence components and chemical structures of

94 atmospheric aerosols by using high-resolution aerosol mass spectrometry (AMS) and

95 EEM fluorescence spectroscopy (Chen et al., 2016a; Chen et al., 2016b). An earlier

96 report from Chang and Thompson (2010) found fluorescence spectra of aqueous-phase

97 reaction products of phenolic compounds, which had some similarities with those of

98 humic-like substances (HULIS), and Tang et al. (2020) reported that aqueous

99 photooxidation of vanillic acid could be a potential source of HULIS. Chang and

Thompson (2010) also showed that light-absorbing and fluorescent substances generally had large conjugated moieties (i.e., quinones, HULIS, polycyclic aromatic hydrocarbons (PAHs)), which can damage human body (Dou et al., 2015; McWhinney et al., 2013). HULIS are considered as an important contributor to induce oxidative stress since they can serve as electron carriers to catalyze ROS formation (Dou et al., 2015; Ma et al., 2019; Huo et al., 2021; Xu et al., 2020), causing adverse health effects. Dithiothreitol (DTT) assay (Alam et al., 2013; Verma et al., 2015), as a non-cellular method, was widely employed to determine oxidation activity and OP of atmospheric PM (Chen et al., 2019; Cho et al., 2005) for the evaluation of its health effects. Some other works (Fang et al., 2016; McWhinney et al., 2013; Verma et al., 2015; Zhang et al., 2022) focused on the link between chemical components and OP in PM, and confirmed that several kinds of compounds, such as quinones, HULIS and transition metals usually had strong DTT activities. However, DTT method is rarely used to evaluate the OP of aqueous-phase oxidation products previously (Ou et al., 2021).

In the present work, we chose eugenol (ally guaiacol) as a model compound to conduct aqueous oxidation experiment. As a representative methoxyphenol emitted from biomass burning (BB) (Hawthorne et al., 1989; Simpson et al., 2005), it was widely detected in atmospheric particles. For instance, concentration and emission factor of this compound from beech wood burning were 0.032 μg/m^3 and 1.534 μg/g, which were twice those of guaiacol (0.016 μg/m^3 and 0.762 μg/g) (Bari et al., 2009). Eugenol is a semivolatile aromatic compound with a moderate water-solubility (2.46 g/L at 298 K). Chemical characteristics of aqueous reaction products under direct photolysis (without oxidant) and oxidations by OH radicals and ^{3}C* triplet states, were comprehensively elucidated by a suite of analytical techniques including high-performance liquid chromatography (HPLC), ultraviolet and visible (UV-Vis)

spectrophotometry, gas chromatography mass spectrometry (GC-MS), and soot particle aerosol mass spectrometry (SP-AMS). Moreover, light absorption, fluorescent and oxidative properties of the aqueous oxidation products were also investigated.

2 Materials and methods

2.1 Chemicals and reagents

Eugenol (99%), tert-butanol (TBA, 99%), 3,4-dimethoxybenzaldehyde (DMB, 99%), para-benzoquinone (p-BQ, 99%), dithiothreitol (99%) and 5,5′-dithiobis-2-nitrobenzoic acid (DTNB, 99%), 2-nitro-5-thiobenzoic (99%), 5,5-dimethyl-1-pyrroline N-oxide (DMPO)**,** 2,2,6,6-tetramethylpiperidine (TEMP) were all purchased from Sigma-Aldrich. Superoxide dismutase (SOD) was purchased from Bovine Erythrocytes BioChemika. Dichloromethane (HPLC-MS grade, 99%), methanol (HPLC-MS grade, 99%), acetonitrile (HPLC-MS grade, 98%), hydrogen peroxide (H_2O_2, 35 wt %), and 2,4,6-trimethylphenol (TMP, 99%) were all obtained from Acros Chemicals. Sodium azide (NaN_3, 98%) was purchased from J&K Scientific Ltd. (Beijing, China). All solutions were prepared using ultrapure water (Millipore) on the days of experiments.

2.2 Photochemical oxidation experiments

Aqueous-phase photochemical reactions were carried out in a Rayonet photoreactor (model RPR-200) equipped with 16 light tubes (2 RPR-3000, 7 RPR-3500 and 7 RPR-4190 tubes), which was frequently used to mimic sunlight for photochemical experiments and was described in details by several groups (George et al., 2015; Hong et al., 2015; Huang et al., 2018; Jiang et al., 2021; Zhao et al., 2014).

Pyrex tubes containing sample solutions were placed in the center and received radiation from surrounded lamps of all sides. To ensure mixing of the solution, a fan and a magnetic stir bar were placed at the bottom of the reaction tube. The solution temperature was controlled at 25 ± 2°C. The same photoreactor system and a normalized distribution of photon fluxes inside the reactor have been reported elsewhere (George et al., 2015), and the wavelength of light was in the range of 280~500 nm. We only measured light intensity at the surface of the solution with a radiometer (Photoelectric instrument factory of Everfine Corporation, Hangzhou, China), which was determined to be ~2400 $\mu W/cm^2$ in the range of 290-320 nm (UVB), lower than the sunlight intensity (6257.1 $\mu W/cm^2$).

In this work, 300 μM H_2O_2 and 15 μM DMB were added into solutions as sources of OH and $^3C^*$, respectively. The initial concentration of eugenol was 300 μM. For $^3C^*$-mediated experiments, solutions were adjusted to pH=3 by sulfuric acid in order to perform experiments under optimal conditions (Ma et al., 2021; Smith et al., 2014) since DMB triplet state is protonated to a more reactive form in acidic solution. We conducted three sets of oxidation experiments: (A) 300 μM eugenol + 300 μM H_2O_2, (B) 300 μM eugenol + 15 μM DMB, and (C) 300 μM eugenol without oxidants. In each series of experiments, a dark control experiment was performed synchronously with a Pyrex tube wrapped by aluminum foil. Results showed loss of eugenol under dark conditions were negligible (data not shown). In addition, to evaluate the roles of ROS in eugenol degradation during $^3C^*$-initiated oxidation, quenching experiments by using specific scavengers to capture different ROS were performed, namely TBA for OH, NaN_3 for $^1O_2$, SOD for $O_2^{\bullet-}$, and TMP for $^3C^*$, respectively (Pan et al., 2020; Chen et al., 2020). For OH-initiated oxidation, quenching experiments using p-BQ for $O_2^{\bullet-}$ (Ma et al., 2019; Raja et al., 2005), and TBA for OH were conducted. For most experiments,

172 solutions were saturated by air and each experiment presented was repeated three times

173 unless otherwise stated. Average results with one standard deviation were provided. In

174 order to further evaluate the role of oxygen in photooxidation, experiments were also

175 conducted by using different saturated gases (air, N_2 and O_2).

176 **2.3 Analytical methods**

177 **2.3.1 Determination of eugenol concentrations**

178 Before and during the photochemical experiment, 2 mL of reacted solution was

179 sampled periodically and subjected to HPLC (LC-10AT, Shimadzu, Japan) analysis to

180 quantify eugenol concentration. The HPLC was equipped with an InertSustain AQ-C18

181 reverse phase column (4.6×250 mm, 5.0 μm, Shimadzu) and a UV-vis detector. The

182 mobile phase was a mixture of acetonitrile/H_2O (v/v: 60/40) at a flow rate of 0.6

183 mL/min, and the detection wavelength was 280 nm. The first-order kinetic rate constant

184 of eugenol degradation can be obtained from the slope of plot of -$\ln(c_t/c_0)$ versus

185 reaction time as presented in Eq.(1).

$$\ln(c_t/c_0)=-kt \qquad\qquad (1)$$

186

187 Where c_0 and c_t are eugenol concentrations (in μM) at the initial and reaction time

188 t, while k represents the pseudo first-order rate constant (in s^{-1}).

189 **2.3.2 UV-vis and fluorescent spectra**

190 The UV-vis light absorbance spectra of reacted solutions (placed in a 1 cm path

191 length quartz cuvette) were measured by using an UV-vis spectrophotometer (Specord

192 210 plus, Analytik Jena., Germany). The instrument has a dual-beam optical system

193 with tungsten and deuterium lamps as light sources. A reference absorption spectrum

194 of ultrapure water was carried out in the same cuvette prior to sample analysis for

195 baseline correction.

196   Immediately after the UV-Vis measurement, the cuvette was transferred to a three-

197 dimensional EEM fluorescence spectrometer (FluoroMax Plus,HORIBA Scientific).

198 The ranges of wavelength varied from 200 to 450 nm for excitation wavelengths (Ex)

199 and from 290 to 650 nm for emission wavelength (Em). Intervals of the excitation and

200 emission wavelengths were 5 nm and 2 nm, respectively. The reported absorbance and

201 EEM spectra here are averages of the results from experiments in triplicate.

202 **2.3.3 Determination of HULIS concentrations**

203   Solid phase extraction (SPE) cartridges (CNW Poly-Sery HLB, 60 mg/cartridge)

204 were used to isolate HULIS from the reaction products. The SPE cartridge was first

205 rinsed with 1 mL ultrapure water and 3 mL methanol prior to extraction. The solution

206 was acidified to pH ~2 using HCl and loaded on an SPE cartridge, which was rinsed

207 with 1 mL ultrapure water again. Next, 3 mL methanol/ammonia (98:2, v/v) mixture

208 was added into the SPE cartridge to elute HULIS, and the solution was blown to full

209 dryness with high purity N_2, followed by dilution with ultrapure water to 25 mL for

210 quantification of HULIS using the HPLC coupled with an evaporative light scattering

211 detector (ELSD3000). Recovery efficiency of the HULIS standard, Suwanne River

212 Fulvic Acid (SRFA), was 75-80% with the standard deviation of reproducibility less

213 than 5%. More details have been described elsewhere (Tao et al., 2021).

214 **2.3.4 Oxidative potential (OP) based on DTT assay**

215   The OP of reaction products was determined by the DTT method (Cho et al., 2005;

216 Lin and Yu, 2019) with slight improvements. Briefly, 1.2 mL sample solution was

transferred into a 10 mL glass tube, then 6 mL phosphate buffer (0.1 M, pH 7.4) and

300 µL of 2.5 mM DTT were added and mixed thoroughly. The DTT mixed solution

was placed in a 37°C water bath for incubation. Over the course of reactions that lasted

for 150 minutes, 1 mL aliquot of DTT mixture was taken every 30 minutes, and 100 µL

of 5 mM DTNB (prepared in 0.1 mM phosphate buffer) was added and loaded in a

centrifuge tube. Next, reactions between DTNB and DTT produced bright yellow TNB,

which was quantified by the UV-Vis spectrometer within 30 minutes. Finally, we

measured the light absorbance (A_t) at 412 nm to indirectly quantify the remaining DTT.

Another 1.2 mL ultrapure water instead of sample solution was treated in the same way

and the absorbance was denoted as A as the blank value. A_0 represents the initial light

absorbance value. Thus, DTT concentration consumed by the sample solution (M_{DTT},

µM) and that by the blank solution (M_{DTT0}, µM) can be calculated according to Eq.(2)

and Eq.(3), respectively.

$$M_{DTT} = \frac{A_0 - A_t}{A_0} \times C_{DTT_0} \tag{2}$$

$$M_{DTT0} = \frac{A_0 - A}{A_0} \times C_{DTT_0} \tag{3}$$

Here, C_{DTT0} was the initial DTT concentration in sample solution (100 µM in this

work). DTT consumption rates (R_{DTT} and R_{DTT0}) were then obtained from the slopes of

plots of M_{DTT} and M_{DTT0} versus incubation times. Experiments of blanks and samples

were typically run in a triplicate. The reproducibility of the whole analysis showed that

the relative standard deviation of DTT consumption rate was 3-4%.

2.3.5 Product analysis by GC-MS

Reacted solution (about 30 mL) was extracted with 10 mL dichloromethane twice.

The extract was concentrated into 1 mL by blowing N_2 gently, subsequently transferred

to a 2 mL vial, and analyzed by a GC-MS (7890A GC/5975C MS, Agilent) with a DB-

5ms capillary column (30 m×0.25 mm×0.5 µm). The operational conditions were set as follows: injector was at 200℃; ion source was at 230 ℃; column oven temperature was programmed to be held at 35℃ for 4 minutes, then ramped to 250 ℃ at a rate of 20℃/minute and held for 10 minutes. The recovery efficiency, method detection limits and quality assurance/quality control have been described in our previous work (Ye et al., 2020).

2.3.6 SP-AMS analysis and mass yields of reaction products

An Aerodyne SP-AMS (Onasch et al., 2012) was applied to analyze the low-volatility organic products, similar to our previous work (Chen et al., 2020; Ge et al., 2017). SP-AMS data were acquired in V mode and analyzed by Squirrel v.1.56D and Pika v1.15D software. The organic fragments were classified into six groups: CH, CHO, CHN, CHO_2, CHON and HO. Elemental ratios (oxygen-to-carbon, O/C; hydrogen-to-carbon, H/C), were calculated according to the method proposed by Canagaratna et al. (2015).

Since the AMS analysis requires nebulization of sample solution into particles before determination, and quantification of organics was influenced by the atomization efficiency and carrier gas flow, we thus cannot use SP-AMS measured concentration to quantify the mass of products directly. In this case, according to Li et al. (2014), we added an internal standard (SO_4^{2-}) prior to AMS analysis, and the mass ratio of particle-phase organics to SO_4^{2-} ($\Delta Org/SO_4^{2-}$) can be used to calculate the mass concentration of products. Furthermore, the mass yield of aqueous-oxidation products ($Y_{products}$, %), which is the mass of products generated per unit mass of precursor consumed, can be calculated according to Eq. (4).

$$Y_{products}(\%)=\frac{(\Delta Org/SO_4^{2-})[SO_4^{2-}]_0}{c_0 M\eta} \times 100\% \tag{4}$$

265 Where $[SO_4^{2-}]_0$ is the SO_4^{2-} concentration (here 7.27 mg/L), C_0 is the initial eugenol

266 concentration (in mmol/L), M is MW of the precursor (164 g/mol for eugenol) , and η

267 is the degraded fraction of eugenol.

3 Results and discussion

3.1 Kinetics of aqueous photooxidation

270   Figure 1 shows unreacted eugenol concentrations (c_t) and the negative logarithm

271 of c_t/c_0 ($-\ln(c_t/c_0)$) as a function of reaction time, respectively. The pseudo first-order

272 rate constants (k) obtained by Eq.(1) were also presented. As described in Fig. 1a,

273 eugenol concentration decreased to be <20% of the initial concentration in 3 hours,

274 suggesting photooxidation was fast under all three reaction conditions. In the presence

275 of $^3C^*$, eugenol was degraded nearly 100% after 3 hours. Previous study (Chen et al.,

276 2020) on $^3C^*$-initiated 4-ethylguaiacol oxidation reports a time of 21 hours for a

277 complete degradation. Apart from difference of precursors, different light irradiation

278 spectra and stronger energy of light in this work than the previous work might be

279 responsible for the fast loss of eugenol. The bond dissociation energies (BDEs) are 340

280 kJ/mol for OH, 374 kJ/mol for C-H in $-CH_3$ group, 345 kJ/mol for C=C bond, and 403

281 kJ/mol for C-H in $-OCH_3$ group, respectively (Herrmann et al., 2003; He et al., 2019).

282 Due to influences of steric hindrance and intramolecular hydrogen bonding, the H-

283 abstraction from OH group might not be favorable and the most probable H-abstraction

284 might take place in C=C of the allyl group. As a result, breakage of C=C into C-C at

285 the allyl group can lead to the formation of 2-methoxy-4-propyl-phenol (Section 3.6.1).

286 When photon energy is higher than the BDE, chemical bonds can break, leading to

287 decomposition of compounds and possibly further mineralization. The energy of photon

of 300 nm is 412 kJ/mol and can break all major bonds in eugenol, while the energy of 350 nm is 353 kJ/mol, being able to break some of the bonds in eugenol as well. Overall, eugenol can be easily decomposed after absorbing the photons.

As shown in Fig. 1b, the first-order rate constants were 2.43×10^{-4} s^{-1}, 2.73×10^{-4} s^{-1}, and 5.75×10^{-4} s^{-1} for direct photolysis and photooxidations by OH and ^{3}C*, respectively. ^{3}C*-initiated photooxidation was quicker than that attacked by OH, likely due to combined contributions from reactions with ^{1}O$_2$, O$_2$$^{\bullet-}$ and OH (Section 3.2). Similar results were found for aqueous phase reactions of three phenols against OH and ^{3}C* by Yu et al. (2016) (Note the initial concentrations of H$_2$O$_2$ and DMB were 100 µM and 5 µM, respectively, with the same ratio as 300 µM H$_2$O$_2$ to 15 µM DMB in this work)

Figure 1. Aqueous-phase eugenol decay kinetic curves (a) and first-order rate constants (b) obtained based on Equation 1 under direct photolysis, OH-initiated oxidation and ^{3}C*-initiated oxidation. Error bar represents one standard deviation from the measurements in triplicate.

3.2 Relative importance of ROS in photooxidation

3.2.1 Quenching experiments in ^{3}C*-initiated photooxidation

Relative importance of different ROS in photooxidation can be investigated by

addition of scavengers/quenchers, and then be evaluated based on the different degradation efficiencies of eugenol in absence and presence of the corresponding ROS quenchers. For each quencher, we conducted several gradient experiments with varying molar ratios of eugenol to quencher. The ratios were 0.075:1, 0.15:1, 0.3:1, 0.75:1, 1.5:1 for quenchers of NaN_3, TMP and TBA, and 1.2:1, 1.6:1, 2.5:1, 5:1, 10:1 for SOD, which were all within the typical ranges of ROS quenching experiments reported previously (Zhou et al., 2018). Excess concentrations of quenchers have been added repeatedly to ensure the complete reactions between ROS and scavengers. Figure 2 displays the impacts of quenchers on eugenol degradation. All rate constants (k) with quenchers were lower than those of the quencher-free solutions. The optimum molar ratio of eugenol to quencher was selected such that the eugenol degradation did not change with the increase of added quencher (Wang et al., 2021). For example, along with the decrease of molar ratios of eugenol to NaN_3 from 1.5:1 to 0.075:1, the variation of eugenol degradation was stabilized at the ratio of 0.15:1, indicating that $^1O_2$ has been completely quenched at this ratio, therefore a molar ratio of 0.15:1 for NaN_3 was optimal, since excess scavenger may generate other products that interfere the existing reactions. Finally, the optimal molar ratios of eugenol to quenchers of TBA, NaN_3, TMP and SOD, were determined to be 1.5, 0.15, 0.075 and 2.5, respectively. Table 1 and Fig. S1 compared the rate constants determined under the ratios above and they were in an order of TMP<NaN_3<SOD<TBA, suggesting relative importance of generated ROS to eugenol degradation was in the order of $^3C^* > {}^1O_2 > O_2^{\bullet-} > OH$. This result suggests that $^3C^*$ plays a major role among all ROS in the photooxidation. Previously, Laurentiis et al. (2013) reported that 4-carboxybenzophenone (70 μM) could act as $^3C^*$ and the photosensitized degradation was more effective than oxidants such as OH, O_3; Misovich et al. (2021) investigated the aqueous DMB-photosensitized reaction (5 μM,

same as in this study) also demonstrated that $^3C^*$ was the greatest contributor to phenol

or guaiacyl acetone degradation, followed by $^1O_2$, while both OH and $^1O_2$ contributions

were relatively minor.

To further assess the relative importance of different ROS, we propose to use the

following Eq.(5) for a rough estimation:

$$Red_{ROS} \text{ (in \%)} = (k - k_{quencher})/k * 100\% \tag{5}$$

Here k (in s^{-1}) is the original rate constant of $^3C^*$-initiated oxidation (or OH-initiated

oxidation in Section 3.2.2) and $k_{quencher}$ (in s^{-1}) is the rate constant after the target ROS

has been completely scavenged by its quencher. k and $k_{quencher}$ in fact refer to those

reported in Fig. S1b. Red_{ROS} then refers to the percentages of reduction due to addition

of quencher for a ROS.

According to Eq.(5), Red_{3C^*} was calculated to be 85.7%. Similarly, the values of

$^1O_2$, $O_2^{\bullet-}$ and OH were 80.5%, 61.4% and 53.9%, respectively. Note Red_{ROS} only

reflects the relative important of ROS and it does not corresponds to the exact

contribution of that ROS in eugenol degradation without quenchers. The reason is that

although the addition of a ROS scavenger can eliminate oxidation by this ROS, but it

also significantly interrupts the original chain reactions as compared to those in the

absence of the scavenger, and reactions with other ROS might be enhanced. In this

regard, the sum of the four Red_{ROS} values may exceed 100%. Therefore, one should be

cautious to use Red_{ROS} as a precise quantification of the ROS contribution in aqueous

oxidation. Determination of ROS concentrations during oxidation should be instead be

an effective way to elucidate the role of ROS. Here, we tried to detect in-situ generated

OH, $O_2^{\bullet-}$ and $^1O_2$ during photochemical reactions using a micro electron spin resonance

(ESR) spectrometer (Bruker Magnettech, Berlin, Germany) with DMPO as the spin trap

to form stable DMPO-OH or DMPO-$O_2^{\bullet-}$, with TEMP to capture $^1O_2$ to produce TEMP-

356 $^1O_2$ spin-adduct (TEMPO). The radicals can be identified and quantified by the peak

357 patterns in ESR spectra, such as the quarter line with a height ratio of 1:2:2:1 for

358 DMPO-OH, 1:1:1:1 for DMPO-$O_2^{\cdot-}$ and 1:1:1 for TEMP-$^1O_2$ (Guo et al., 2021).

359 Unfortunately, OH radical cannot be detected since its concentration might be lower

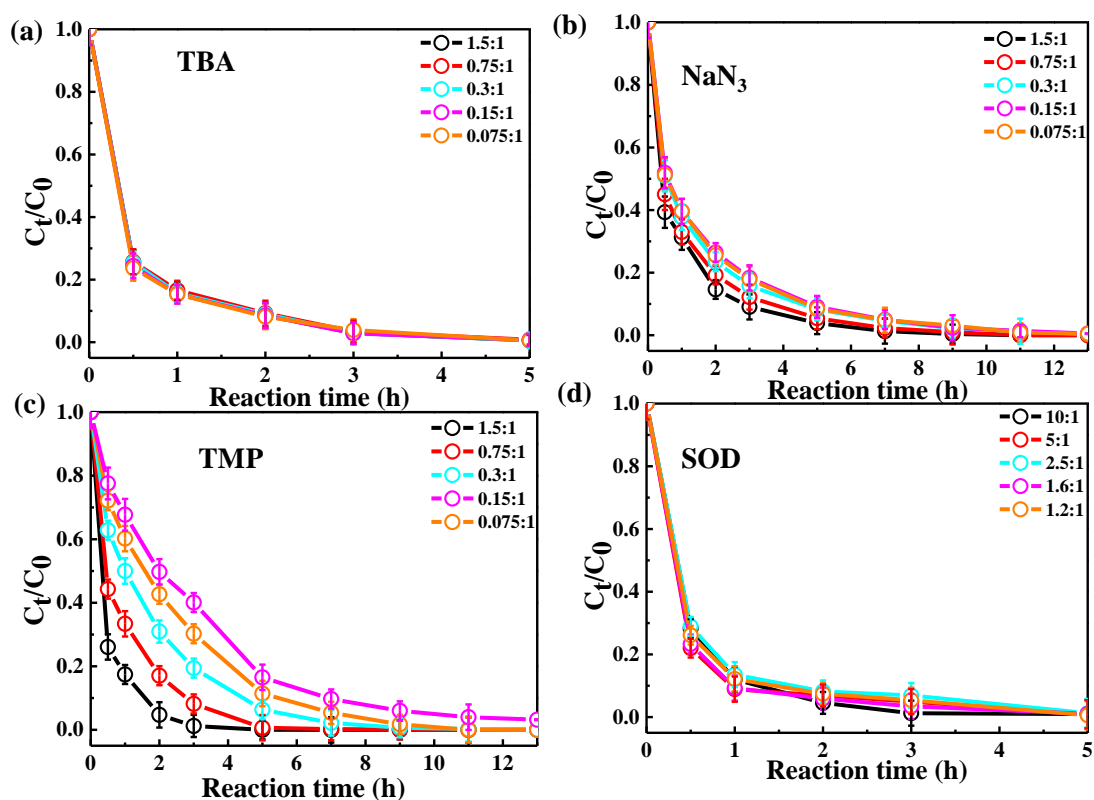
360 than the detection limit of the instrument (Fig. S2, ESR spectra of OH). In contrast, we

361 were able to detect high concentrations of $^3C^*$ and found the intensity of TEMP-$^1O_2$

362 signal reached its maximum at 30 minutes, then decreased slowly (Fig. S2, ESR spectra

363 of $^1O_2$). Combining the great quenching effect of TMP with high $^1O_2$ concentration from

364 ESR method, we can conclude that $^3C^*$ and $^1O_2$ play relatively important roles in

365 eugenol photooxidation.

366

367 **Figure 2.** Ratio of unreacted eugenol concentration to its initial concentration (C_t/C_0) at different

368 molar ratios of eugenol to quencher, as a function of reaction time: (a) TBA, (b) NaN$_3$, (c) TMP and

369 (d) SOD.

370

3.2.2 Quenching experiments in OH-initiated photooxidation

To examine the contributions of ROS to eugenol degradation for OH-initiated oxidation, TBA and p-BQ as trapping agents were added. Similar to $^3C^*$-initiated oxidation, several gradient experiments with varying molar ratios of eugenol to quenchers were conducted. The ratios were set as 6.5:1, 3.2:1, 1.6:1, 1.1:1 and 0.8:1 for p-BQ and 3.0:1, 1.5:1, 0.75:1, 0.3:1 and 0.15:1 for TBA. According to Fig. S3, molar ratio only had a slight influence on eugenol degradation, although degradation can be inhibited effectively by quenchers. Thus, we determined the appropriate molar ratios of 0.8 and 0.75 for p-BQ and TBA, respectively, as excess scavengers might influence the chemical reactions.

Variations of the rate constants for the aforementioned quenching experiments were determined, in comparison with those conducted without quenchers, and results are listed in Table 1 and presented in Fig. S4. For TBA quenching tests, the rate constant decreased by 18.7% (from 2.73×10^{-4} s^{-1} to 2.22×10^{-4} s^{-1}), showing that OH radical played a certain role in eugenol photooxidation. Since H_2O_2 was mainly photolyzed at wavelength <300 nm to generate OH radical, irradiation above 300 nm did not affect the reaction significantly. The p-BQ could quench $O_2^{\bullet-}$, further suppressing the generation of other ROS (e.g., $\bullet HO_2$), therefore the rate constant decreased the most (from 2.73×10^{-4} s^{-1} to 1.20×10^{-4} s^{-1}), suggesting $O_2^{\bullet-}$ was important for eugenol photooxidation. This hypothesis could be further confirmed by the decline of rate constant under N_2-saturated solution (Section 3.2.3). However, it was difficult to detect both OH and $O_2^{\bullet-}$ directly due to their relatively short lifetimes and low concentrations via ESR in this work.

Table 1. The first-order rate constants of eugenol in the presence of various scavengers. The initial conditions were as follows: 300 μM eugenol; molar ratios of eugenol to quenchers TBA, NaN$_3$, TMP and SOD, of 1.5, 0.15, 0.075 and 2.5, respectively; molar ratios of eugenol to quenchers p-BQ and TBA of 0.8 and 0.75, respectively.

$^3C^*$-initiated quenching experiments			
Quenchers	ROS	Reaction rate constant k (s^{-1})	Pearson's R^2
no quencher	-	5.75×10^{-4}	0.996
TBA	OH	2.65×10^{-4}	0.999
SOD	O$_2^{-}$	2.22×10^{-4}	0.995
NaN$_3$	$^1O_2$	1.12×10^{-4}	0.999
TMP	$^3C^*$	0.82×10^{-4}	0.999
OH-initiated quenching experiments			
Quenchers	ROS	Reaction rate constant k (s^{-1})	R^2
No quencher	-	2.73×10^{-4}	0.995
TBA	OH	2.22×10^{-4}	0.998
p-BQ	O$_2^{-}$	1.20×10^{-4}	0.995

3.2.3 Influences of different saturated gases

In order to assess the role of O$_2$ in eugenol degradation, a series of experiments were performed under both O$_2$-saturated and N$_2$-saturated conditions in addition to air. N$_2$ gas was purged into reaction solution for ~30 minutes before experiment to achieve the O$_2$-free condition. Figure 3 compared the changes of eugenol concentrations and rate constants under the three gas conditions for direct photolysis, OH-initiated and $^3C^*$-initiated oxidations, respectively. The rate constants followed the order of $k_{O_2} > k_{Air} > k_{N_2}$ under both direct photolysis and OH oxidation, providing evidence in support of O$_2$ being significant for eugenol degradation. This might be explained by the fact that O$_2$ can act as an electron acceptor to generate O$_2^{\bullet-}$ and $\bullet$HO$_2$, and subsequently form H$_2$O$_2$ and OH. For direct photolysis, rate constant under O$_2$-saturated condition increased 14.4% while it decreased 19.3% under N$_2$ saturation from that under saturated air. For

412   OH-initiated oxidation, the difference of rate constants under three saturated gases

413   became more distinct.

414   On the contrary, rate constants followed the order of $k_{Air} > k_{N_2} > k_{O_2}$ in $^3C^*$-

415   initiated oxidation. There are two possible explanations. On one hand, under N_2-

416   saturated condition without oxygen, DMB would involve in reactions (R1-R4), leading

417   to a more effective generation of $^3DMB^*$ therefore a higher degradation efficiency than

418   under O_2-saturated condition. On the other hand, for air/O_2-saturated solutions,

419   irradiation of DMB and eugenol would involve also reactions (R5-R8) in addition to

420   (R1-R4), and as a result, the amount of $^3DMB^*$ decreased, due to formation of other

421   ROS ($^1O_2$, $O_2^{\bullet-}$, OH, etc) with relatively weak oxidative capacities. In summary,

422   quenching of $^3DMB^*$ by ground state molecular oxygen could account for the low

423   degradation efficiency in O_2-saturated condition.

424      $DMB + h\nu \rightarrow {}^1DMB^* \rightarrow {}^3DMB^*$ (R1)

425      $^3DMB^* \rightarrow DMB$ (R2)

426      $^3DMB^* \rightarrow Products$ (R3)

427      $^3DMB^* + {}^1DMB^* \rightarrow DMB^{\bullet+/\bullet-}(DMB^{\bullet+} + DMB^{\bullet-})$ (R4)

428      $^3DMB^* + O_2 \rightarrow DMB + {}^1O_2$ (R5)

429      $DMB^{\bullet-} + O_2 \rightarrow \cdot DOM^+ + O_2^{\bullet-}$ (R6)

430      $O_2^{\bullet-} + 2H^+ \rightarrow H_2O_2 + O_2$ (R7)

431      $H_2O_2 \rightarrow 2OH$ (R8)

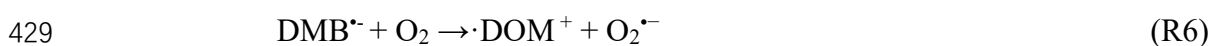
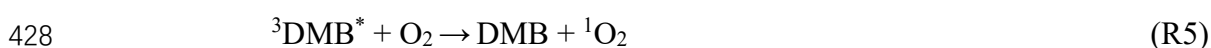
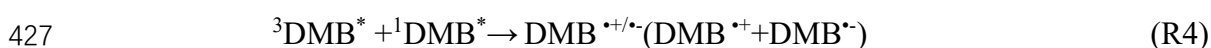
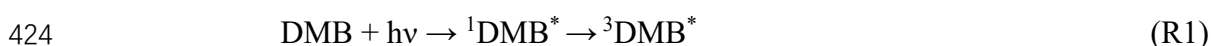
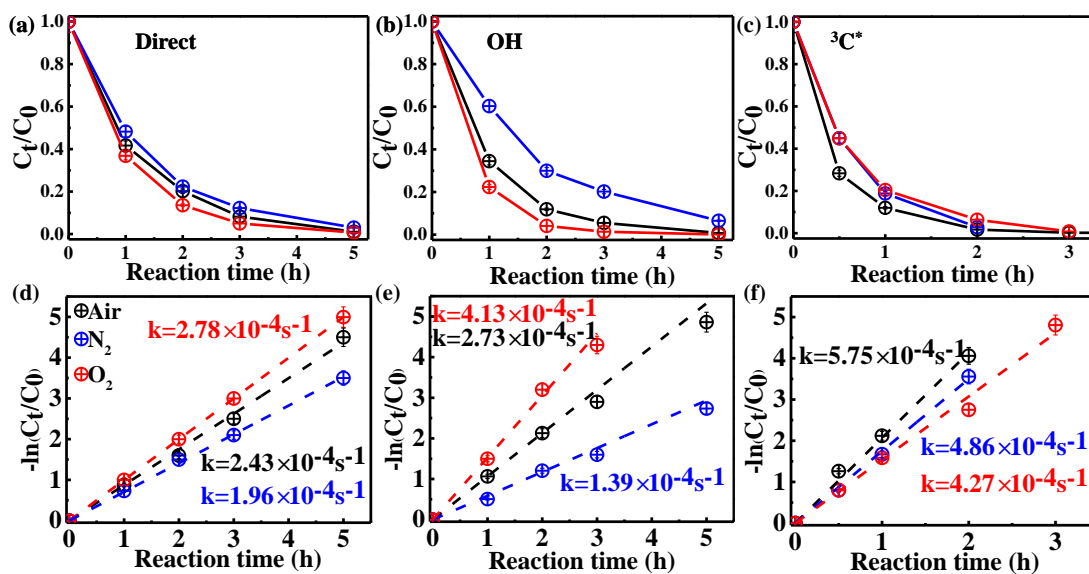



Figure 3. Ratio of unreacted eugenol concentration to its initial concentration (C_t/C_0) as a function of reaction time at different saturated gases under (a) direct photolysis (b) OH-initiated oxidation and (c) $^3C^*$-initiated oxidation. Rate constants of (a-c) are presented in (d-f) correspondingly.

3.2.4 Variations of pH and dissolved oxygen

The initial pH values of reaction solutions for direct photolysis and OH-initiated oxidation were unadjusted, while that for the $^3C^*$-oxidation was adjusted to 3. The variation of solution pH is presented in Fig. 4a. The pH values decreased quickly at the beginning of illumination (from 7.4 to ~5.0 in the first 1 hour) then tended to be flat for both direct photolysis and OH-initiated oxidation. However, little change of pH (less than 0.1 unit) was observed for the $^3C^*$-initiated photooxidation throughout the oxidation, which can be likely ascribed to its low initial pH of 3. Since the solution pH was acidic (pH=3), we cannot rule out formation of acidic products (such as organic acids) during $^3C^*$-initiated oxidation as during direct photolysis and OH-initiated oxidation.

As discussed in Section 3.2.3, oxygen can take part in photochemical reaction to form ROS, which may in turn destroy the structure of precursor. Here we measured the

449 oxygen consumption during oxidation through determination of dissolved oxygen (DO)

450 contents by a dissolved oxygen meter (Seven2Go Pro S9, Zurich, Switzerland). DO was

451 consumed mainly at the first 1 hour and remained stable afterwards (Figs. 4b-c and Fig.

452 S5). The amounts of consumed DO followed the order of $^3C^*$>OH>direct photolysis.

453 The maximum consumed DO was found in $^3C^*$-initiated oxidation, which might be

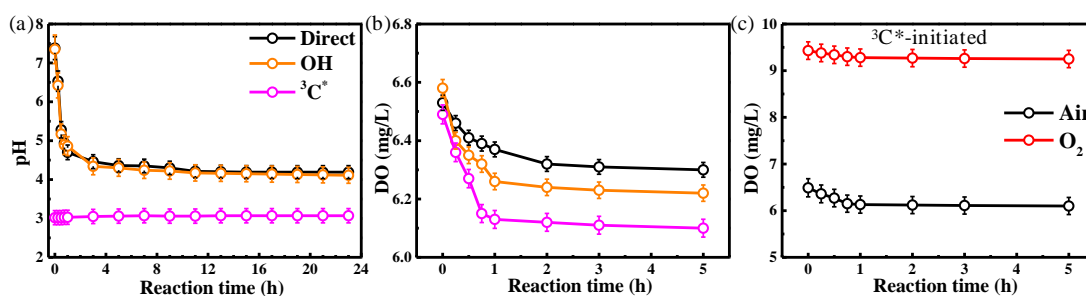
454 explained by the consumption of O_2 that reacts with $^3C^*$ form $^1O_2$ (R5). Obviously, a

455 steady-state DO level was reached when the consumption rate was equal to the diffusion

456 of O_2 into the solution (Pan et al., 2020). Overall, these results re-emphasize that O_2

457 can influence eugenol degradation and chemical transformation via induction of radical

458 chain reactions.

459

460 **Figure 4.** (a) pH values and (b) dissolved oxygen (DO) contents against reaction time under direct

461 photolysis, OH-initiated oxidation, $^3C^*$-initiated oxidation, and (c) DO contents during $^3C^*$-initiated

462 oxidation under air or O_2-saturated conditions.

463

464 **3.3 Optical properties of reaction products**

465 **3.3.1 Light-absorbing properties**

466   The UV-vis light absorption spectra of the solutions at different reaction times are

467 presented in Fig. 5. The light absorption by eugenol itself mainly occurs in the range of

468 260-300 nm (n→π* electronic transition, 270-350 nm), which overlaps with the major

469 photon fluxes (280 and 500 nm) of our lamp for photooxidation. Therefore, we can

470 clearly observe that the characteristic absorption peak at 280 nm of precursor decreased

with the propagation of direct photolysis (Fig. 5a), similar to that in OH-initiated photooxidation (Fig. 5b). However, the reaction was quick in the presence of $^3C^*$, and the characteristic absorption peak at 280 nm after 3 hours of illumination almost disappeared, suggesting nearly a complete loss of eugenol, consistent with the results in Section 3.1 that more than 99% eugenol was degraded in 3 hours. Additionally, there was an obvious absorption enhancement at longer wavelengths (300-400 nm) during the photooxidation, whereas eugenol itself did not absorb light in this range, indicating some light-absorbing products (e.g., brown carbon (BrC) species) were generated. Aqueous photooxidation of some phenolic compounds (e.g., vanillic acid) also presented long-wavelength (300-400nm) light absorbance, with intensity increasing with illumination time (Tang et al, 2020; Zhao et al., 2015). In addition, there were some differences for light absorbance at wavelength of 300-400 nm in the three reaction conditions. For direct photolysis and OH-initiated oxidation, light absorbance increased during the first 15 hours, then remained at a plateau until 23 hours. However, for $^3C^*$-initiated oxidation, light absorbance increased during the first 7 hours, then decreased slowly afterwards. The different shapes of UV-vis spectra between OH and $^3C^*$ photooxidations indicate formations of different products.

Compared to the light spectrum of eugenol, there were also increases of light absorbance at ~260 nm ($\pi \rightarrow \pi^*$ electronic transitions) upon aqueous oxidation in all three reaction conditions (Fig. 5), demonstrating the generation of new substances likely with both aromatic C=C and carbonyl (C=O) functional groups (Tang et al., 2020). The enhancement at 300-400 nm may point to products with high MWs and conjugated structures. Unfortunately, we were unable to quantify the relative contributions of individual products to the overall light absorbance between 300 to 400 nm due to lack of a full speciation of the products and their light absorption spectra.

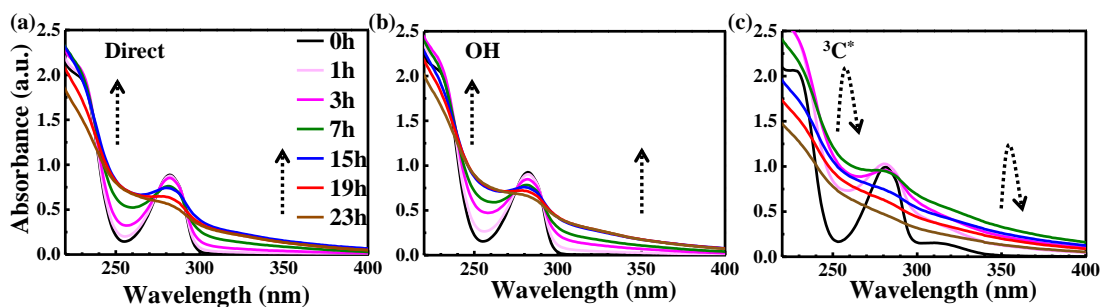



Figure 5. UV–vis light absorption spectra of reacted solutions at different reaction times under (a) direct photolysis, (b) OH-initiated oxidation, and (c)^{3}C*-initiated oxidation.

3.3.2 Fluorescence properties

The fluorescence properties of solutions before (0 hour) and during photooxidation (3 and 7 hours) were investigated via the EEM technique, as shown in Fig. 6. For comparison, we also presented EEM profiles of pure eugenol (non-irradiated), pure DMB, and the end solutions (23 hours) of direct photolysis and OH-initiated oxidation in Fig. S6. The peaks at Excitation/Emission (Ex/Em)=275/313 nm can be attributed to fluorescence of the phenolic structure of parent substance (eugenol here), as suggested by Laurentiis et al. (2013). As shown in both Fig. 6 and Fig. S6, the fluorescence intensity decreased after oxidation due to eugenol decay, and the reduction was very fast for ^{3}C*-initiated oxidation. This finding matches with the fast degradation and large rate constant for ^{3}C*-initiated oxidation. The EEM plots for direct photolysis and OH-initiated oxidation had similar contour patterns as shown in Figs. 6a and b, although EEM profiles changed significantly with irradiation time. We also observed distinct fluorescent peaks at Ex/Em=235/(400-500) nm, indicating that illumination can cause a red shift in fluorescence emission wavelength. As suggested by Chang et al. (2010), fluorophores at Ex/Em=240/400 nm are linked with aromatic structures and condensed saturated bonds including polycyclic aromatic hydrocarbons. Another work (Li et al., 2021) showed that red shift in the fluorescence spectra was usually related to an

increase in the size of ring system and an increase in the degree of conjugation. Previous studies (Chen et al., 2016a; Chen et al., 2019) have reported that fluorescent compounds with emission wavelength at 400-500 nm were likely linked with HULIS. Additionally, HULIS have two typical fluorescent peaks in EEM profile at Ex/Em=(200-300)/(400-500) nm and Ex/Em=350/(400-500) nm with the former one having a higher intensity (Graber and Rudich, 2006; Laurentiis et al., 2013; Vione et al., 2019). There was also evidence that direct photolysis of tyrosine and 4-phenoxyphenol generated HULIS with new fluorescence signals at Ex/Em=(200-250)/(400-450) nm and 300/(400-450) nm (Bianco et al., 2014). In this regard, we inferred that new peak at Ex/Em=235/(400-500) nm here was likely attributed to HULIS. For the $^3C^*$-initiated oxidation, extra fluorescent peaks at Ex/Em=(220-300)/(400-500) nm appeared in the first 1 hour (data not shown), but their intensities weakened and gradually disappeared upon prolonged reactions (3 hours). Nevertheless, EEM results should be interpreted with caution because many substances might contribute to absorption and emission at a certain wavelength, and it is hard to distinguish and isolate fluorescent and nonfluorescent constituents simply via the EEM technique.

Another interesting finding was that a small fluorescence peak appeared at Ex/Em=(300-350)/(300-350) nm in some of the EEM profiles. Specifically, it appeared earlier for $^3C^*$-oxidation (at 3 hours) than the other two systems, yet its intensity seemed to be a bit stronger in the end solutions of direct photolysis and OH-oxidation (Fig. S6). Moreover, as suggested by Leenheer and Croue (2003), fluorescence peak position of the maximum Ex/Em for HULIS with lower MWs would shift towards lower wavelengths, thus, we inferred fluorescence peak at Ex/Em=(300-350)/(300-350) nm might be in part attributed to the organic acids with a few carbon atoms (probably C_1-C_6). Nevertheless, large uncertainties still exist in using EEM fluorescence technique

to identify molecular compositions of the products due to lack of standard EEM profiles for specific compounds from aqueous phase oxidation and clearly more studies are needed in future.

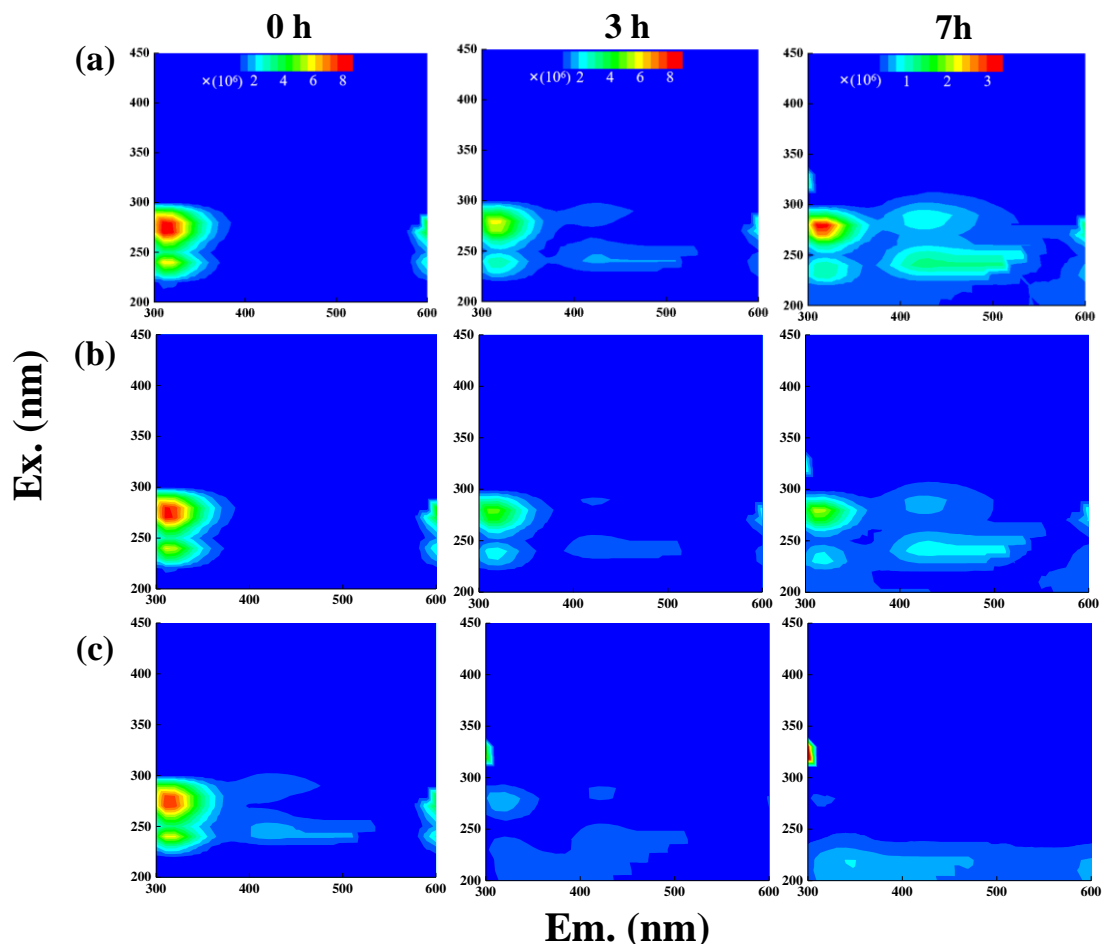



Figure 6. EEM fluorescence spectra of the initial solution (0 hour) and those at different reaction time (3 and 7 hours) under (a) direct photolysis, (b) OH-initiated oxidation, and (c) $^3C^*$-initiated oxidation.

3.4 Characteristics of HULIS

The EEM spectra revealed new prominent fluorescent peak at Ex/Em=250/(400-500) nm, which was likely owing to HULIS. HULIS can be divided into fulvic acid (water soluble at all pHs), humic acid (base soluble, acid insoluble) and humin (insoluble at all pHs). In principle, extracted HULIS in this work with polymer-based

HLB SPE packing include LMW organic acids, fulvic acids and other humic substances.

Figure 7 presents the measured HULIS concentrations against the reaction time. The results show clearly that aqueous-phase eugenol oxidation is a source of HULIS, and the amount increased gradually in the first 7 hours, then remained at a similar level (about 30 mg/L) for the OH-initiated oxidation. For direct photolysis, HULIS concentration increased until 11 hours and then became steady at a level around 40 mg/L. For the $^3C^*$-oxidation, HULIS concentration increased to a maximum at 7 hours, then declined slightly afterwards. A plausible reason of such variabilities is that generated HULIS was capable of further taking part in photochemical reactions since it can act as photosensitizer. Moreover, Yu et al. (2016) characterized the products from aqueous oxidations of phenols by $^3C^*$ triplet states and OH radicals, and found both could produce oligomers and hydroxylated species but the $^3C^*$-oxidation could produce more of these compounds when 50% of the precursor was reacted. Considering the large increases of HULIS in the first 7 hours and the much faster increase of $^3C^*$-oxidation in the first 3 hours shown in Fig. 7, we postulate that HULIS species might be some of the high MW oligomers, which can in turn contributed to fluorescence at emission of ~400 nm (Barsotti et al., 2016).

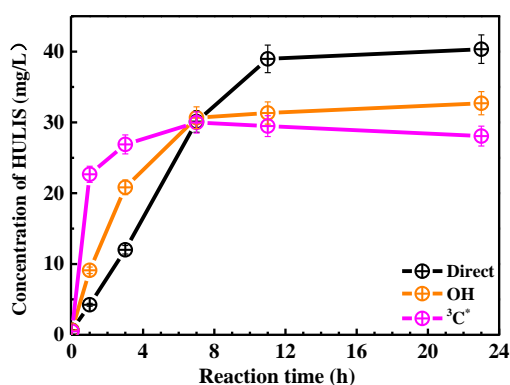



Figure 7. HULIS concentrations as a function of reaction time under direct photolysis, OH-initiated oxidation and $^3C^*$-initiated oxidation.

3.5 Mass yield and oxidation degree of reaction products

3.5.1 Mass yields

HULIS is only a subset of the products from aqueous oxidation, and here we used AMS to further quantify the total reaction products. Figure 8a shows SP-AMS measured organic mass profiles (normalized by sulfate mass, $\Delta Org/SO_4^{2-}$) against the reaction time. As the reaction propagated, $\Delta Org/SO_4^{2-}$ increased continuously in $^3C^*$-initiated system. Nevertheless it arose stepwise and reached a maximum at 19 hours, then remained at a plateau for the direct photolysis and OH-mediated oxidation. Figure 8b illustrates the calculated mass yields at different reaction times. The mass yields were in the ranges of 46.2%-196.5%, 22.1%-144.9%, 19.3%-140.1% for $^3C^*$-oxidation, OH-oxidation and direct photolysis, respectively. For the same oxidation time, mass yield from $^3C^*$-oxidation was generally higher than those from OH-oxidation and direct photolysis. There are two plausible reasons for high mass yield of $^3C^*$-initiated oxidation. First, oxidation by $^3C^*$ was more efficient to form oligomers and functionalized/oxygenated products (Richards-Henderson et al., 2014; Yu et al., 2016). Higher oxidative degree of products from $^3C^*$-initiated photooxidation (see Sec.3.5.2) supports this hypothesis. Secondly, more light-absorbing products formed during initial stage of $^3C^*$-oxidation (Fig. 5c) may accelerate oxidation by acting as photosensitizers (Tsui et al., 2018).

The product mass yields obtained in this work (~20%-197%) overall agree with those reported previously for phenolic compounds. For examples, Huang et al. (2018) reported mass yields of 30-120% for syringaldehyde and acetosyringone; Smith et al. (2014) found that mass yields of aqSOA from three phenols with $^3C^*$ were nearly 100%, and Ma et al. (2021) reported a yield ranging from 59 to 99% for six highly substituted

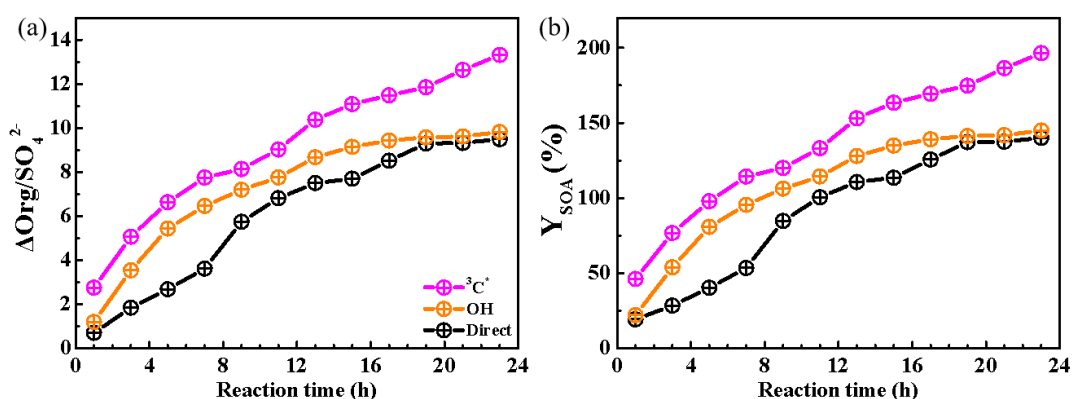
598 phenols with ^{3}C*; Mass yields of SOA from three benzene-diols were near 100% with

599 both OH and ^{3}C* oxidants (Smith et al., 2015); Direct photolysis of phenolic carbonyls,

600 and oxidation of syringol by ^{3}C*, had SOA mass yields ranging from 80 to 140% (Smith

601 et al., 2016). Our previous study on eugenol OH oxidation illuminated by a 500 W Xe

602 lamp reported a mass yield of ~180% (Ye et al., 2020), slightly higher than the value

603 determined here owing to different light wavelengths/intensities.

605 **Figure 8.** Variations of the organic mass normalized by sulfate (a) (ΔOrg/SO$_4$$^{2-}$) and (b) mass yields

606 of reaction products with reaction time under direct photolysis, OH-initiated oxidation and ^{3}C*-

607 initiated oxidation.

608

609 **3.5.2 Oxidation degree**

610 In order to further probe oxidation levels of the reaction products, O/C derived

611 from SP-AMS mass spectrum of the organics was used to represent the oxidation degree

612 of products. In addition, carbon oxidation state (OSc, defined as 2*O/C - H/C) (Kroll

613 et al., 2011) was also calculated. Figures 9a-c depict variations of the elemental ratios

614 (O/C and H/C) and OSc during oxidations. Dramatic increases of O/C and OSc during

615 the initial stage of oxidation (within 1 hour) were observed, with O/C changing from

616 0.26 to 0.65, from 0.26 to 0.70, from 0.25 to 0.75, as well as OSc from -1.11 to -0.15,

617 from -1.16 to -0.05, from -1.13 to 0.09 for direct photolysis, OH-oxidation and ^{3}C*-

oxidation, respectively. The O/C was lower than those of other phenolic aqSOA (Yu et al., 2014) due to different substituted groups in aromatic ring of the precursors. Both O/C and OSc gradually increased, while H/C changed little after 1 hour. The enhancements of OSc in the end were 1.22, 1.11 and 0.86 for $^3C^*$-initiated oxidation, OH-initiated oxidation and direct photolysis, respectively.

Furthermore, the f_{44} vs. f_{43} diagram ("triangle plot") can be used to demonstrate the evolution of SOA during oxidation (Ng et al., 2010). The f_{44} and f_{43} are defined as the ratios of signal intensities of m/z 44 (mainly CO_2^+) and 43 (mainly $C_2H_3O^+$) to the total organics. The results that the f_{44} increased continuously (moved upwards) during both OH and $^3C^*$ oxidations, indicating persistent formation of highly oxygenated compounds including organic acids, such as formic acid and oxalic acid (Sun et al., 2010). Note the f_{44} enhancement was much more significant for $^3C^*$ oxidation (from 0.07 to 0.16) than direct photolysis (from 0.07 to 0.12) and OH oxidation (from 0.07 to 0.13), consistent with the behaviors of its higher O/C and OSc. The f_{43} value decreased in the first stage (1-3 hours) and then increased at later stages. The final f_{43} values were almost the same as those of the initial solutions and were small. As a result, all data points located outside the f_{44} vs. f_{43} region (bounded by the two dash lines in Figs. 9d-f) for ambient aerosols established by Ng et al. (2010).

In summary, our results shown here demonstrate that aqueous phase eugenol photochemical oxidation can generate highly oxygenated products and hence increase the degree of oxygenation of overall SOA.

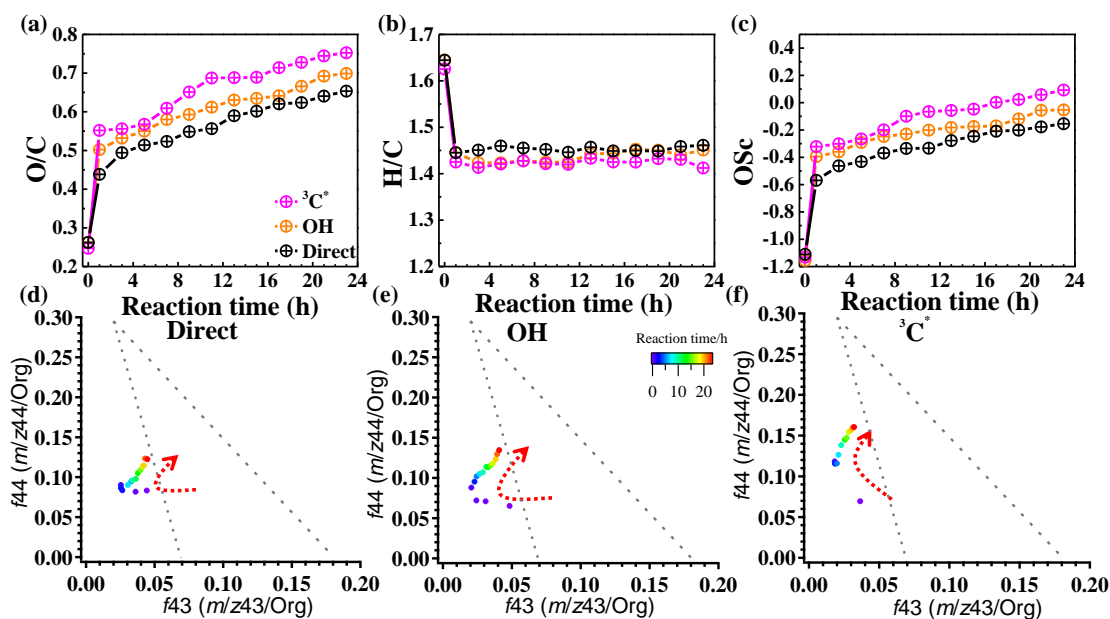
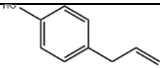
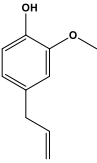
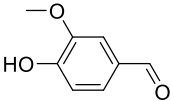
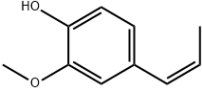
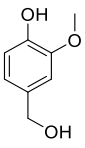
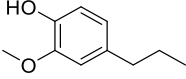
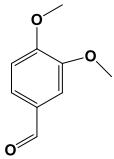
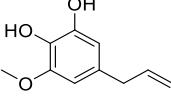
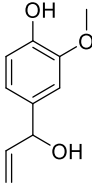
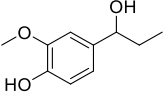
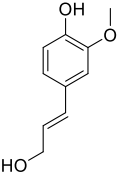



Figure 9. Variations of the elemental ratios of (a) O/C, (b) H/C and (c) oxidation state (OSc) as a function of reaction time; f_{44} vs. f_{43} plots of reaction products under (d) direct photolysis, (e) OH-initiated oxidation, and (f) $^3C^*$-initiated oxidation.

3.6 Molecular characterization of reaction products and proposed reaction mechanism

3.6.1 Major products identified by GC-MS

SP-AMS was limited to probe bulk composition of low-volatility oxidation products, thus the molecular-level characterization of products was performed by using GC-MS here. The total ion chromatograph (TIC) of GC-MS on the solutions before illumination (0 hour) and at illumination times of 11 and 23 hours for the $^3C^*$-initiated photooxidation is shown in Fig. S7. As shown in Fig.S7, eugenol (retention time (RT) at 11.50 min) loss was more than 90% at 11 hours, which could be confirmed by the experimental data reported in Section 3.1. Comparison of products at 11 hours and 23 hours showed no significant difference. Similar to aqueous photochemical oxidation with OH (Ye et al., 2020), a series of products were identified and listed in Table 2.

655 Except 5-ally-3-methoxybenzene-1,2-diol (MW 180, RT=12.59 min), the other eight

656 products were detected for both OH and $^3C^*$-initiated photooxidations. Some of them

657 (Eugenol, DMB, product 1, 2, 5) were identified by using certified reference materials,

658 some of them (product 3, 4, 6, 7, 8, 9) were inferred according to the molecular ion

659 peaks and fragments from GC-MS, based on spectra from the NIST database (Stein,

660 2014) and on the reactants and reaction conditions.

661 We also found 4-(1-hydroxypropyl)-2-methoxyphenol (product 8) was relatively

662 abundant (Fig.S7), suggesting functionalization might dominates as compared to

663 oligomerization and fragmentation. Products were mainly from addition/elimination of

664 hydroxyl (-OH), methoxyl (-OCH$_3$) to benzene ring or allyl group and further oxidized

665 to carbonyl or carboxyl compounds. As suggested by Bonin et al. (2007), the OH-

666 addition to the aromatic ring of phenol preferentially takes place at the ortho (48%) and

667 the para (36%) positions, leading to the formation of OH-adduct product 6 (5-allyl-3-

668 methoxybenzene-1,2-diol). Notably, dimers and ring-opening products were not

669 observed, but they cannot be excluded since they would be probably out of the detection

670 of GC-MS technique (Vione et al., 2014).

671
672 **Table 2.** Major reaction products identified via GC-MS

	RT (min)	Name*	Proposed chemical structure	Chemical formula	Nominal MW (g/mol)
Product 1	10.68	4-allylphenol		C$_9$H$_{10}$O	134
Precursor	11.50	Eugenol		C$_{10}$H$_{12}$O$_2$	164

Product 2	11.81	4-hydroxy-3-methoxybenzaldehyde		$C_8H_8O_3$	152
Product 3	12.06	(E)-2-methoxy-4-(prop-1-en-1-yl)phenol		$C_{10}H_{12}O_2$	164
Product 4	12.11	4-(hydroxymethyl)-2-methoxyphenol		$C_8H_{10}O_3$	154
Product 5	12.18	2-methoxy-4-propylphenol		$C_{10}H_{14}O_2$	166
Photosensitizer	12.29	3,4-dimethoxybenzaldehyde(DMB)		$C_9H_{10}O_3$	166
Product 6**	12.59	5-allyl-3-methoxybenzene-1,2-diol		$C_{10}H_{12}O_3$	180
Product 7	12.65	4-(1-hydroxyallyl)-2-methoxyphenol		$C_{10}H_{12}O_3$	180
Product 8	12.79	4-(1-hydroxypropyl)-2-methoxyphenol		$C_{10}H_{14}O_3$	182
Product 9	12.91	(E)-4-(3-hydroxyprop-1-en-1-yl)-2-methoxyphenol		$C_{10}H_{12}O_3$	180

673   *Precursor (eugenol) and triplet precursor (DMB) are also shown.

 **This compound was only identified in ^{3}C*-oxidation solution.

3.6.2 Reaction mechanism

The reaction pathways of ^{3}C*-initiated photooxidation of eugenol are demonstrated in Scheme 1 based on the products identified by GC-MS. The other intermediates and the potential pathways were proposed according to the identified products and the reaction rationality from the starting reactant. To better depict the mechanism, DMB was expressed as [RCHO] and eugenol as Ph-R for simplicity. [RCHO] absorbs light and undergoes excitation to ^{1}[RCHO]*, then experiences the intersystem crossing (ISC) to form ^{3}[RCHO]*. ^{3}[RCHO]* can participate in subsequent reactions via three channels. First, it can react with O_2 to form $^1O_2$ via energy transfer. Secondly, it can transform to [RCHO]$^{\bullet-}$, subsequently reacts with O_2 to generate $O_2^{\bullet-}$ via electron transfer, which can disproportionate to H_2O_2. The decomposition of H_2O_2 can generate OH radical. Thirdly, the ^{3}[RCHO]* can react with Ph-R to from [Ph-R•] via H-abstraction. The cleavage of [Ph-R•] to free radical segment (such as CH$_2$CH• or CH$_3$O•) takes place, then an additional hydrogen transfer could occur, resulting in a 2H-addition to the new intermediate to form 4-allyl-phenol (product 1). Similarly, when the CH$_2$CH• is lost from [Ph-R•], an addition of H_2O would happen on the new compound (product 4) and further oxidized to 4-hydroxy-3-methoxybenzaldehyde (product 2). Another possibility is the intermediate [Ph-R•] can resonate to several different isoelectronic species, the radical position changes to aromatic ring or allyl group site, which would couple with HO• to form hydroxylated eugenol monomer (product 6, 7, 9 MW=180). Consequently, the isoelectronic species at allyl group site could also abstract a hydrogen to form isoeugenol (product 3 MW=164). Also, breakage of C=C into C-C and 2H-addition at allyl group site could

form 2-methoxy-4-propyl-phenol (product 5, MW=166). Besides, the C=C breaking

intermediate can couple with HO• to form 4-(1-hydroxypropyl)-2-methoxyphenol

(product 8, MW=182). In conclusion, $^3C^*$ can directly oxidize eugenol to form SOA

products or small molecular compounds, or indirectly oxidize eugenol via energy

transfer, electron transfer, hydrogen abstraction, proton-coupled electron transfer or

other radical chain reactions.

The organic groups, such as methoxy, allyl groups can be eliminated from

aromatic ring, which then participate in photochemical reaction, resulting in generation

of dimers, small organic acids, CO_2 and H_2O, etc. Dimers previously reported from

aqueous reaction of 4-methylsyringol with OH were not detected via GC-MS in the

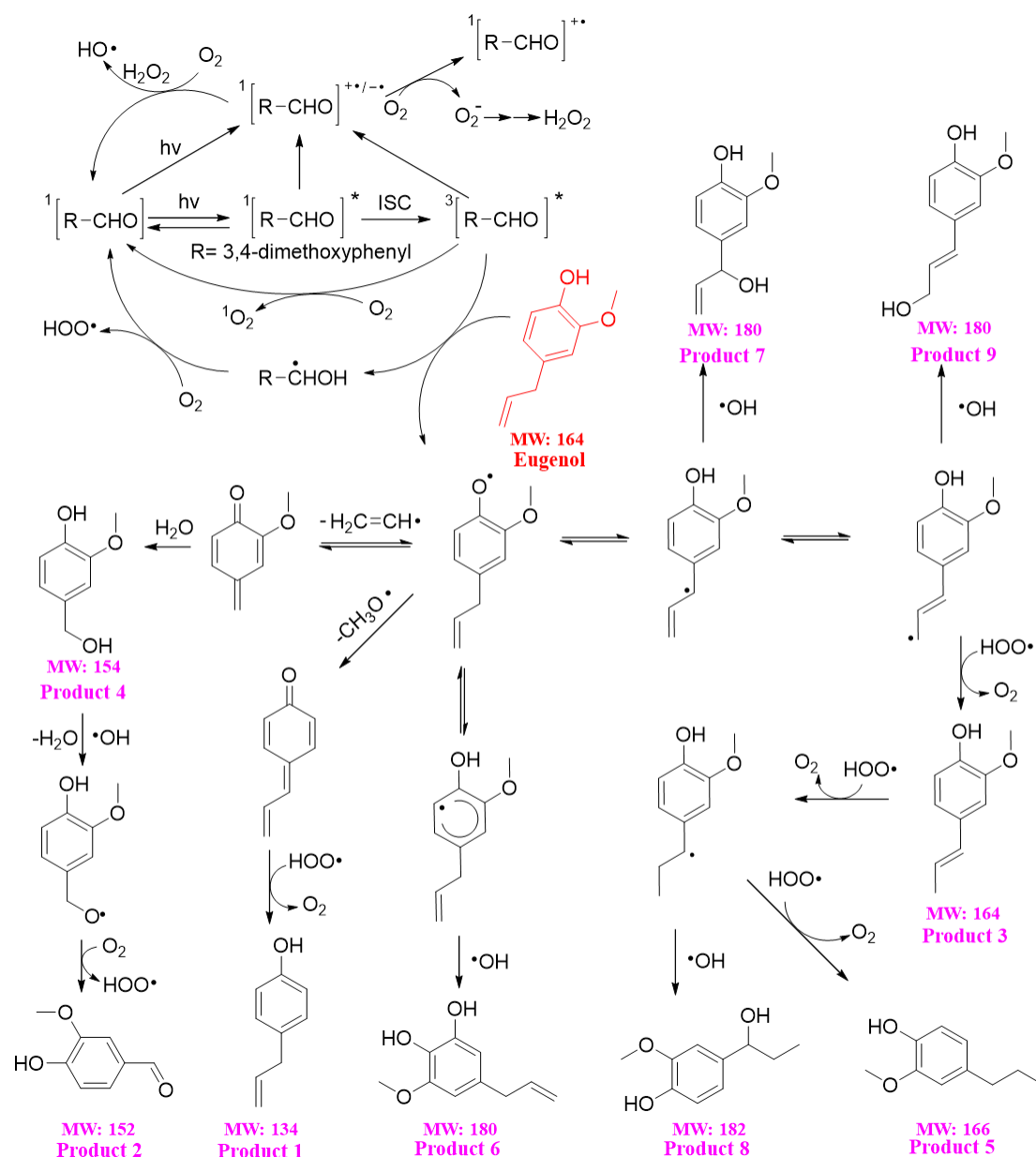
present work but dimer fragment ions ($C_{20}H_{22}O_4^+$) were detected by SP-AMS with trace

amounts. Functionalization due to the additions of hydroxyl, carbonyl functional groups

to the aromatic rings could account for the enhancement of light absorption at

wavelength of 300-400 nm. However, polar high MW organic acids were not detected

likely due to the limitation of GC-MS technique.

713

Scheme 1. Proposed reaction mechanism of $^3C^*$-initiated photooxidation of eugenol. The red text

represents the precursor, and the compounds labeled by Product 1-9 are those identified by GC-

MS (Table 2).

3.7 Oxidative potential (OP) of reaction products

The OP of oxidation products can be represented by the consumption rate of DTT

concentration, defined as R_{DTT}. Figure 10a shows the DTT consumed mass (M_{DTT}) as

a function of incubation times (0, 30, 60, 90, 120 and 150 min) for a triplicate sample

721 (300 µM eugenol) and blank (ultrapure water). M_{DTT} values for both blank and eugenol

722 were proportional to incubation time, indicating that ROS-generating substances in

723 reaction solution act only as catalyst and itself was not consumed. The slopes represent

724 DTT consumption rates, which are also illustrated in Fig. 10a. Average R_{DTT0} (blank)

725 was 0.31 µM/min and R_{DTT} for initial 300 µM eugenol (before experiment) was 0.52

726 µM/min. Since self-oxidation of DTT might lead to the consumption of DTT in

727 ultrapure water, final DTT consumption rate of reacted solution after oxidation was then

728 blank-corrected by subtracting the average R_{DTT0}.

729   Figure 10b shows changes of blank-corrected R_{DTT} with reaction time for direct

730 photolysis, OH-initiated oxidation and $^3C^*$-initiated oxidation, respectively. The R_{DTT}

731 value of $^3C^*$-oxidation products increased quickly and reached the maximum (0.9) at 7

732 hours, then decreased slowly and its end value was lower than that from OH-oxidation.

733 The R_{DTT} value of OH-oxidation products on the other hand increased slowly and

734 reached the maximum at 21 hours. The R_{DTT} value of products from direct photolysis

735 increased continuously but also slowly to ~0.36 till the end of oxidation. Nevertheless,

736 we can see that the final R_{DTT} values were all higher than that of eugenol, proving that

737 aqueous-phase processing can generate products with higher OP, resulting in more

738 health hazards than the precursor does. The DTT consumption rates are comparable to

739 those using the same DTT method (Charrier and Anastasio, 2012; Lin and Yu, 2019).

740 The weak correlation was found between HULIS concentration and R_{DTT}, implying that

741 OP was not only dependent upon HULIS. Moreover, HULIS with diverse molecular

742 structures also exhibit different ROS-generation potentials (Kramer et al., 2016),

743 therefore the HULIS as an ensemble may not correlate well with OP.

744

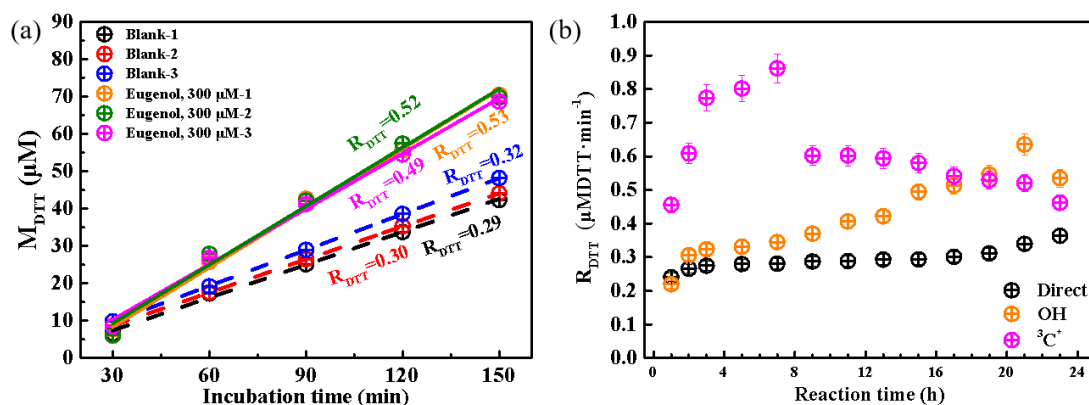



Figure 10. (a) DTT consumed mass versus incubation times for blank (ultrapure water) and 300 μM

eugenol solutions in a triplicate, and (b) blank-corrected DTT consumption rates versus reaction

time for direct photolysis, OH-initiated oxidation and $^3C^*$-induced oxidation.

4. Atmospheric implications

The high mass yields of aqueous-phase photooxidation of eugenol (exceeding 100%

after 23 hours of illumination) found here are similar or even higher than those

previously reported yields of a number of phenolic compounds (e.g., Smith et al., 2014,

2015, 2016; Ma et al., 2021), which re-emphasizes the importance of biomass burning

(BB) to SOA budget (Gilardoni et al., 2016), particularly in regions or periods with

significant BB activities. In addition, our study here used 300 μM H_2O_2 and 15 μM

DMB as sources of OH and $^3C^*$, and $^3C^*$-mediated oxidation appeared to be faster than

OH-initiated oxidation of eugenol. Of course, whether or not $^3C^*$ is more important

than OH in real atmosphere depends upon their concentrations. OH and $^3C^*$ are difficult

to measure and concentrations vary greatly in real atmospheric samples. Herrmann et

al. (2010) estimated an average OH level of 0.35×10^{-14} M in urban fog water; Kaur and

Anastasio (2018) measured $^3C^*$ concentration to be $(0.70\text{-}15) \times 10^{-14}$ M, 10-100 times

higher than the co-existing OH in ambient fog waters; Kaur et al. (2019) determined

both OH and $^3C^*$ concentrations in PM extracts, OH steady-state concentration was

763   4.4(±2.3) x10^{-16} M, similar to its level in fog, cloud and rain, while ^{3}C* concentration

764   was 1.0(±0.4) x10^{-13} M, a few hundred times higher than OH and nearly double its

765   average value in fog. Therefore, together with these measurements, our findings signify

766   a likely more important role of ^{3}C* than OH in aqueous-phase (especially aerosol water)

767   reactions. However, the liquid water content of aerosol is typically ~10000 times

768   smaller than that of cloud (for instance, ~50 μg m^{-3} versus 0.5 g m^{-3}). Even if the

769   reaction rates in aerosol water were 10 times higher than those in cloud water, the

770   overall importance of aqueous reactions initiated by the same oxidant in aerosol phase

771   would be still ~1000 times smaller than it in cloud water. Moreover, quenching

772   experiments reveal that O$_2$ can inhibit eugenol degradation by effectively scavenging

773   ^{3}C* while it can promote degradation by fostering chain reactions in OH-induced

774   oxidation, which offer insights to the control of reaction pathways by regulating ROS

775   generations; of course, such operation calls for application of highly sensitive EPR

776   method.

777   Eugenol has a strong light absorption peak around 280 nm, therefore it can undergo

778   direct photolysis, and addition of OH or other photosensitizers (^{3}C*) can gradually

779   diminish its light absorption around 280 nm, but increase the absorption in visible light

780   range (>300 nm). In the meantime, HULIS was generated continuously, and GC-MS

781   identified a number of high MW organic products, in line with those detected in earlier

782   aqueous photooxidation of phenolic compounds (Jiang et al., 2021; Misovich et al.,

783   2021; Tang et al., 2020; Yu et al., 2014). Overall, our work demonstrates that aqueous

784   oxidation of BB emissions is a source of BrC, and this BrC may act as photosensitizer

785   to oxidize other species; a portion of this BrC might be HULIS, and some high MW

786   aromatic compounds are a subset of this HULIS. However, a recent study by Wang et

787   al. (2021) shows that fossil foil derived OA (FFOA) can be an effective precursor of

788 aqSOA, but the aqSOA became less light-absorbing than the FFOA. These contrasting

789 results indicate that contribution of aqueous oxidation to BrC is largely dependent upon

790 the precursors; molecular structures of major chromophores, changes of the structures

791 upon oxidation as well as their interplay with light absorptivity should be carefully

792 investigated to achieve a full understanding of the impacts of aqueous processing on air

793 quality, radiative forcing and climate change.

794   Investigations on the OPs of reaction products from eugenol photooxidation show

795 that aqueous processing can produce more toxic products than the precursor. This result

796 is in agreement with our previous work on resorcinol, hydroquinone and

797 methoxyhydroquinone (Ou et al., 2021). Although more studies on a broad spectrum of

798 atmospherically relevant species and multiple indicators of toxicity are clearly needed,

799 our findings here underscore the potential of aqueous processing on the enhancement

800 of particle toxicity.

801 **5 Conclusions**

802   This study comprehensively investigated the aqueous photooxidation of eugenol

803 upon direct photolysis and attacks by OH radicals and $^3C^*$ triplet states. By using a

804 suite of techniques, the decay kinetics of eugenol, chemical, optical properties as well

805 as toxicity of reaction products were studied. The first-order rate constants followed the

806 order of $^3C^*$>OH >direct photolysis (300 μM H_2O_2 and 15 μM DMB as sources of OH

807 and $^3C^*$). Further quenching experiments on different ROS during $^3C^*$-mediated

808 oxidation showed that $^3C^*$ was the major contributor, followed by $^1O_2$, $O_2^{\bullet-}$ and OH;

809 $O_2^{\bullet-}$ played a more important role than OH during OH-initiated oxidation. The rate

810 constants under saturated O_2, air and N_2 followed the order of $k_{O_2} > k_{Air} > k_{N_2}$ for both

811 direct photolysis and OH-initiated oxidation, but changed to $k_{Air} > k_{N_2} > k_{O_2}$ for $^3C^*$-

812 mediated oxidation. O_2 appeared to be a scavenger of $^3C^*$ therefore suppressing $^3C^*$

813 oxidation while it could promote generation of OH thus accelerate OH-mediated

814 oxidation. pH and DO levels both decreased during oxidation, indicating formation of

815 acids and a certain role of DO in oxidation.

816   Eugenol itself can absorbs lights significantly around 280 nm, and aqueous

817 oxidation gradually decrease this absorption of UV light but enhanced the absorbance

818 in the visible light range (mainly 300-400 nm), indicative of the generation of BrC

819 species. These species were likely linked with HULIS, as HULIS concentration

820 increased during the course of oxidation, in particular for the initial stage of $^3C^*$-

821 mediated reactions. The final mass yields of reaction products (after 23 hours of

822 irradiation) were 140.1%, 144.9% and 196.5% for direct photolysis, OH-oxidation and

823 $^3C^*$-oxidation, respectively. Oxidation degrees of the products increased continuously

824 with the illumination time, indicating persistent formation of highly oxygenated

825 compounds, especially during $^3C^*$-mediated reactions. Molecular characterization by

826 GC-MS identified a series of oxygenated compounds, allowing us to propose the

827 detailed oxidation mechanism. Functionalization appeared to be a dominant pathway to

828 form the observed species.

829   DTT method was used to assess OP of the reaction products. The end products in

830 all three sets of experiments showed higher DDT consumption rates than that of the

831 precursor; products from $^3C^*$-oxidation showed particularly fast increase in the first

832 few hours of reactions. This result demonstrates that species that are more toxic than its

833 precursors could be produced upon aqueous oxidation, indicative of the potential toxic

834 effects induced by aqueous processing.

835

836 ***Data availability.*** The relevant data of this study are available at:

837   http://nuistairquality.com/eugenol_data_and_figure

838

839   *Supplement.* The supplement related to this article is available on line at: XXX

840

841   *Author Contributions:* XDL, YT, LWZ, SSM, SPL, ZZZ and NS conducted the

842   experiments. XDL and YT analyzed the data. XDL and ZLY prepared and wrote the

843   paper with contributions from all co-authors. ZLY and XLG reviewed and commented

844   on the paper.

845

846   *Competing interests.* The authors declare that they have no conflict of interest.

847

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860

861   *Review statement.* This paper was xxx

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