Optical and chemical properties and oxidative potential of aqueous-1 phase products from OH and ³C*-initiated photooxidation of eugenol 2 Xudong Li¹, Ye Tao¹, Longwei Zhu¹, Shuaishuai Ma¹, Shipeng Luo¹, Zhuzi Zhao¹, Ning 3 Sun¹, Xinlei Ge^{2,*}, Zhaolian Ye^{1,*} 4 5 6 ¹College of Chemistry and Environmental Engineering, Jiangsu University of 7 Technology, Changzhou 213001, China 8 ²Jiangsu Key Laboratory of Atmospheric Environment Monitoring and Pollution 9 Control, Collaborative Innovation Center of Atmospheric Environment and Equipment Technology, School of Environmental Sciences and Engineering, Nanjing University 10 of Information Science and Technology, Nanjing 210044, China 11 *Correspondence: 12 Zhaolian Ye (bess ye@jsut.edu.cn) and Xinlei Ge (caxinra@163.com) 13 14 Abstract: Aqueous reactions may turn precursors into light-absorbing and toxic 15 products, leading to air quality deterioration and adverse health effects. In this study, 16 we investigated comprehensively eugenol photooxidation (a representative biomass 17 18 burning emitted, highly substituted phenolic compound) in bulk aqueous phase with direct photolysis, hydroxyl radical (OH) and an organic triplet excited state (3C*). 19 Results show that the degradation rates of eugenol followed the order of 20 21 ³C*>OH >direct photolysis. During ³C*-initiated oxidation, different reactive oxygen species (ROS) including ³C*, OH, ¹O₂ and O₂* can participate in oxidation of eugenol, 22 quenching experiments verified 3C* was the most important one; while during OH-23 24 initiated oxidation, O2* was a more important ROS than OH to oxidizedegrade eugenol.

The rate constants under saturated O₂, air and N₂ followed the order of $ko_2 > k_{Air} > k_{N_2}$

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for both direct photolysis and OH-initiated oxidation, but changed to $k_{Air} > k_{N_2} > k_{O_2}$ for ³C*-mediated oxidation. pH and dissolved oxygen (DO) levels both decreased during oxidation, indicating formation of acids and the participation of DO in oxidation. UV-vis light absorption spectra of the reaction products showed clear absorbance enhancement in the 300-400 nm range for all three sets of experiments and new fluorescence at excitation/emission=250/(400-500) nm appeared, suggesting the formation of new chromophores and fluorophores (brown carbon species); and these species were likely attributed to humic-like substances (HULIS) as shown by the increases of HULIS concentrations during oxidation. Large mass yields of products (140%-197%) after 23 hours of illumination were obtained, and high oxidation degrees of these products were also observed; correspondingly, a series of oxygenated compounds were identified, and detailed reaction mechanism with functionalization as a dominant pathway was proposed. At last, dithiothreitol (DTT) assay was applied to assess oxidation potential of the reaction products, and the end products of all three sets of experiments showed higher DDT consumption rates than that of eugenol, indicating more toxic species were produced upon aqueous oxidation. Overall, our results by using eugenol as a model compound, underscore the potential importance of aqueous processing of biomass burning emissions in secondary organic aerosol (SOA) formation.

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1 Introduction

Photochemical reactions in atmospheric aqueous phases (cloud/fog droplets and aerosol water) can affect lifetimes of many organic species, and are an important source and pathway of secondary organic aerosol (SOA) formation (Vione et al., 2006; Zhao

et al., 2012). Compared to the gasSOASOA formed via gas-phase photochemical oxidation, (gasSOA), aqueous-phase SOA (aqSOA) is often more oxidized and less volatile, therefore might play an important role in haze formation, air quality and global climate change (Ervens et al., 2011; Lim et al., 2010). However, due to complexity of the aqueous reactions and influencing factors (such as precursors, oxidants, and light intensities), detailed reaction mechanism, optical property, oxidative potential (OP) and the interplay among them remain poorly understood. Many laboratory studies have focused on aqueous-phase oxidations of low molecular weight (LMW) volatile organic compounds (VOCs), such as isoprene, terpenes (α -, β -pinene), as well as their gas-phase oxidation products (such as glyoxal, methylglyoxal, cis-pinonic acid and methyl vinyl ketone) (Faust et al., 2017; Herrmann, 2003; Herrmann et al., 2015; Huang et al., 2011; Lee et al., 2012; Zhang et al., 2010). Recently, aqueous oxidation of semi-/intermediate volatility VOCs (S/IVOCs), such as the phenolic compounds emitted from combustion or pyrolysis of lignin in biomass, were also extensively investigated (Barzaghi and Herrmann, 2002; Bonin et al., 2007; Chen et al., 2020; Gilardoni et al., 2016; He et al., 2019; Jiang et al., 2021; Li et al., 2014; Li et al, 2021; Ma et al., 2021; Mabato et al., 2022; Smith et al., 2014; Sun et al., 2010; Tang et al., 2020; Yang et al., 2021; Yu et al., 2016; Lu et al., 2019). Generally, chemical structures of precursors have profound influences on the reaction mechanisms and products, while effect of oxidants also cannot be neglected. It is evident that liquid water can contain various types of oxidants, such as singlet oxygen (¹O₂), nitrate radical (NO₃), hydroxyl radical (OH), and organic triplet excited states (³C*), and all oxidants can play crucial roles in photooxidation reactions (Kaur and Anastasio, 2018; Scharko et al., 2014). Among them, OH is a ubiquitous oxidant with concentrations of 10⁻¹³-10⁻¹⁰

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¹² mol·L⁻¹ (Arakaki et al., 2013; Gligorovski et al., 2015; Herrmann et al., 2003). Hence,

aqueous OH-induced photooxidation has been extensively studied (Chen et al., 2020; Sun et al., 2010; Yu et al., 2016). Compared to OH oxidation, ³C*-initiated aqueous oxidation (photosensitized reactions) has also attracted attentions in recent years (Ma et al., 2021; Wang et al., 2021). Several classes of organic compounds in ambient air, including non-phenolic aromatic carbonyls, quinones, aromatic ketones and nitrogencontaining heterocyclic compounds, can form ³C* after absorbing light (Alegría et al., 1999; Kaur et al., 2019; Nau and Scaiano, 1996; Rossignol et al., 2014; Chen et al., 2018). These compounds are termed photosensitizers. ³C* is capable of reacting with O₂ to produce singlet oxygen (¹O₂) and superoxide radicals (O₂··). Various reactive oxygen species (ROS) can be generated and affect greatly the ³C*-initiated aqueous-phase reactions. Despite some studies demonstrating importance of ROS in photochemical process (Ma et al., 2021; Wang et al., 2020; Wang et al., 2021), our current understanding on ³C*-initiated oxidation is still limited.

Excitation-emission matrix (EEM) fluorescence spectroscopy, as a low-cost, rapid, non-destructive and high-sensitivity technique, can offer detailed information on chromophores hence has been widely employed for studies of aquatic dissolved organic matter (Aryal et al., 2015). Nevertheless, it has not been extensively used in atmospheric aerosol research (Mladenov et al., 2011). Prior studies have investigated the relationship between the fluorescence components and chemical structures of atmospheric aerosols by using high-resolution aerosol mass spectrometry (AMS) and EEM fluorescence spectroscopy (Chen et al., 2016a; Chen et al., 2016b). An earlier report from Chang and Thompson (2010) found fluorescence spectra of aqueous-phase reaction products of phenolic compounds, which had some similarities with those of humic-like substances (HULIS), and Tang et al. (2020) reported that aqueous photooxidation of vanillic acid could be a potential source of HULIS. Chang and

Thompson (2010) also showed that light-absorbing and fluorescent substances generally had large conjugated moieties (i.e., quinones, HULIS, polycyclic aromatic hydrocarbons (PAHs)), which can damage human body (Dou et al., 2015; McWhinney et al., 2013). HULIS are considered as an important contributor to induce oxidative stress since they can serve as electron carriers to catalyze ROS formation (Dou et al., 2015; Ma et al., 2019; Huo et al., 2021; Xu et al., 2020), causing adverse health effects. Dithiothreitol (DTT) assay (Alam et al., 2013; Verma et al., 2015), as a non-cellular method, was widely employed to determine oxidation activity and OP of atmospheric PM (Chen et al., 2019; Cho et al., 2005) for the evaluation of its health effects. Some other works (Fang et al., 2016; McWhinney et al., 2013; Verma et al., 2015; Zhang et al., 2022) focused on the link between chemical components and OP in PM, and confirmed that several kinds of compounds, such as quinones, HULIS and transition metals usually had strong DTT activities. However, DTT method is rarely used to evaluate the OP of aqueous-phase oxidation products previously (Ou et al., 2021).

In the present work, we chose eugenol (ally guaiacol) as a model compound to conduct aqueous oxidation experiment. As a representative methoxyphenol emitted from biomass burning (BB) (Hawthorne et al., 1989; Simpson et al., 2005), it was widely detected in atmospheric particles. For instance, concentration and emission factor of this compound from beech wood burning were 0.032 μg/m³ and 1.534 μg/g, which were twice those of guaiacol (0.016 μg/m³ and 0.762 μg/g) (Bari et al., 2009). Eugenol is a semivolatile aromatic compound with a moderate water-solubility (2.46 g/L at 298 K). Chemical characteristics of aqueous reaction products under direct photolysis (without oxidant) and oxidations by OH radicals and ³C* triplet states, were comprehensively elucidated by a suite of analytical techniques including high-performance liquid chromatography (HPLC), ultraviolet and visible (UV-Vis)

spectrophotometry, gas chromatography mass spectrometry (GC-MS), and soot particle aerosol mass spectrometry (SP-AMS). Moreover, light absorption, fluorescent and oxidative properties of the aqueous oxidation products were also investigated.

2 Materials and methods

2.1 Chemicals and reagents

Eugenol (99%), tert-butanol (TBA, 99%), 3,4-dimethoxybenzaldehyde (DMB, 99%), para-benzoquinone (*p*-BQ, 99%), dithiothreitol (99%) and 5,5'-dithiobis-2-nitrobenzoic acid (DTNB, 99%), 2-nitro-5-thiobenzoic (99%), 5,5-dimethyl-1-pyrroline N-oxide (DMPO), 2,2,6,6-tetramethylpiperidine (TEMP) were all purchased from Sigma-Aldrich. Superoxide dismutase (SOD) was purchased from Bovine Erythrocytes BioChemika. Dichloromethane (HPLC-MS grade, 99%), methanol (HPLC-MS grade, 99%), acetonitrile (HPLC-MS grade, 98%), hydrogen peroxide (H₂O₂, 35 wt %), and 2,4,6-trimethylphenol (TMP, 99%) were all obtained from Acros Chemicals. Sodium azide (NaN₃, 98%) was purchased from J&K Scientific Ltd. (Beijing, China). All solutions were prepared using ultrapure water (Millipore) on the days of experiments.

2.2 Photochemical oxidation experiments

Aqueous-phase photochemical reactions were carried out in a Rayonet photoreactor (model RPR-200) equipped with 16 light tubes (2 RPR-3000, 7 RPR-3500 and 7 RPR-4190 tubes), which was frequently used to mimic sunlight for photochemical experiments and was described in details by several groups (George et al., 2015; Hong et al., 2015; Huang et al., 2018; Jiang et al., 2021; Zhao et al., 2014).

Pyrex tubes containing sample solutions were placed in the center and received radiation from surrounded lamps of all sides. To ensure mixing of the solution, a fan and a magnetic stir bar were placed at the bottom of the reaction tube. The solution temperature was controlled at $25\pm2^{\circ}$ C. The same photoreactor system and a normalized distribution of photon fluxes inside the reactor have been reported elsewhere (George et al., 2015), and the wavelength of light was in the range of $280\sim500$ nm. We only measured light intensity at the surface of the solution with a radiometer (Photoelectric instrument factory of Everfine Corporation, Hangzhou, China), which was determined to be $\sim\!2400~\mu\text{W/cm}^2$ in the range of 290-320 nm (UVB), lower than the sunlight intensity (6257.1 $\mu\text{W/cm}^2$).

In this work, 300 μM H₂O₂ and 15 μM DMB were added into solutions as sources of OH and 3 C*, respectively. The initial concentration of eugenol was 300 μM. For 3 C*-mediated experiments, solutions were adjusted to pH=3 by sulfuric acid in order to perform experiments under optimal conditions (Ma et al., 2021; Smith et al., 2014) since DMB triplet state is protonated to a more reactive form in acidic solution. We conducted three sets of oxidation experiments: (A) 300 μM eugenol + 300 μM H₂O₂, (B) 300 μM eugenol + 15 μM DMB, and (C) 300 μM eugenol without oxidants. In each series of experiments, a dark control experiment was performed synchronously with a Pyrex tube wrapped by aluminum foil. Results showed loss of eugenol under dark conditions were negligible (data not shown). In addition, to evaluate the roles of ROS in eugenol degradation during 3 C*-initiated oxidation, quenching experiments by using specific scavengers to capture different ROS were performed, namely TBA for OH, NaN₃ for 1 O₂, SOD for O₂-, and TMP for 3 C*, respectively (Pan et al., 2020; Chen et al., 2020). For OH-initiated oxidation, quenching experiments using *p*-BQ for O₂-. (Ma et al., 2019; Raja et al., 2005), and TBA for OH were conducted. For most experiments,

solutions were saturated by air and each experiment presented was repeated three times unless otherwise stated. Average results with one standard deviation were provided. In order to further evaluate the role of oxygen in photooxidation, experiments were also conducted by using different saturated gases (air, N₂ and O₂).

2.3 Analytical methods

2.3.1 Determination of eugenol concentrations

Before and during the photochemical experiment, 2 mL of reacted solution was sampled periodically and subjected to HPLC (LC-10AT, Shimadzu, Japan) analysis to quantify eugenol concentration. The HPLC was equipped with an InertSustain AQ-C18 reverse phase column (4.6×250 mm, 5.0 μ m, Shimadzu) and a UV-vis detector. The mobile phase was a mixture of acetonitrile/H₂O (v/v: 60/40) at a flow rate of 0.6 mL/min, and the detection wavelength was 280 nm. The first-order kinetic rate constant of eugenol degradation can be obtained from the slope of plot of -ln(c_1/c_0) versus reaction time, as presented in Eq.(1).

$$ln(c_1/c_0) = -kt$$
 (1)

Where c_0 and c_t are eugenol concentrations (in μM) at the initial and reaction time t, while k represents the pseudo first-order rate constant (in s⁻¹).

2.3.2 UV-vis and fluorescent spectra

The UV-vis light absorbanceabsorption spectra of reacted solutions (placed in a 1 cm path length quartz cuvette) were measured by using an UV-vis spectrophotometer (Specord 210 plus, Analytik Jena., Germany). The instrument has a dual-beam optical system with tungsten and deuterium lamps as light sources. A reference absorption

spectrum of ultrapure water was carried out in the same cuvette prior to sample analysis for baseline correction.

Immediately after the UV-Vis measurement, the cuvette was transferred to a three-dimensional EEM fluorescence spectrometer (FluoroMax Plus, HORIBA Scientific). The ranges of wavelength varied from 200 to 450 nm for excitation wavelengths (Ex) and from 290 to 650 nm for emission wavelength (Em). Intervals of the excitation and emission wavelengths were 5 nm and 2 nm, respectively. The reported absorbance and EEM spectra here are averages of the results from experiments in triplicate.

2.3.3 Determination of HULIS concentrations

Solid phase extraction (SPE) cartridges (CNW Poly-Sery HLB, 60 mg/cartridge) were used to isolate HULIS from the reaction products. The SPE cartridge was first rinsed with 1 mL ultrapure water and 3 mL methanol prior to extraction. The solution was acidified to pH ~2 using HCl and loaded on an SPE cartridge, which was rinsed with 1 mL ultrapure water again. Next, 3 mL methanol/ammonia (98:2, v/v) mixture was added into the SPE cartridge to elute HULIS, and the solution was blown to full dryness with high purity N₂, followed by dilution with ultrapure water to 25 mL for quantification of HULIS using the HPLC coupled with an evaporative light scattering detector (ELSD3000). Recovery efficiency of the HULIS standard, Suwanne River Fulvic Acid (SRFA), was 75-80% with the standard deviation of reproducibility less than 5%. More details have been described elsewhere (Tao et al., 2021).

2.3.4 Oxidative potential (OP) based on DTT assay

The OP of reaction products was determined by the DTT method (Cho et al., 2005; 216 Lin and Yu, 2019) with slight improvements. Briefly, 1.2 mL sample solution was transferred into a 10 mL glass tube, then 6 mL phosphate buffer (0.1 M, pH 7.4) and 300 μL of 2.5 mM DTT were added and mixed thoroughly. The DTT mixed solution was placed in a 37°C water bath for incubation. Over the course of reactions that lasted for 150 minutes, 1 mL aliquot of DTT mixture was taken every 30 minutes, and 100 μL of 5 mM DTNB (prepared in 0.1 mM phosphate buffer) was added and loaded in a centrifuge tube. Next, reactions between DTNB and DTT produced bright yellow TNB, which was quantified by the UV-Vis spectrometer within 30 minutes. Finally, we measured the light absorbance (A_t) at 412 nm to indirectly quantify the remaining DTT. Another 1.2 mL ultrapure water instead of sample solution was treated in the same way and the absorbance was denoted as A as the blank value. A₀ represents the initial light absorbance value. Thus, DTT concentration consumed by the sample solution (MDTT, μM) and that by the blank solution (MDTTO, μM) can be calculated according to Eq.(2) and Eq.(3), respectively.

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$$M_{DTT} = \frac{A_0 - A_t}{A_0} \times C_{DTT_0}$$
 (2)

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$$M_{DTT0} = \frac{A_0 - A}{A_0} \times C_{DTT_0}$$
 (3)

Here, C_{DTT0} was the initial DTT concentration in sample solution (100 μ M in this work). DTT consumption rates (R_{DTT} and R_{DTT0}) were then obtained from the slopes of plots of M_{DTT} and M_{DTT0} versus incubation times. Experiments of blanks and samples were typically run in a triplicate. The reproducibility of the whole analysis showed that the relative standard deviation of DTT consumption rate was 3-4%.

2.3.5 Product analysis by GC-MS

Reacted solution (about 30 mL) was extracted with 10 mL dichloromethane twice. The extract was concentrated into 1 mL by blowing N_2 gently, subsequently transferred to a 2 mL vial, and analyzed by a GC-MS (7890A GC/5975C MS, Agilent) with a DB-

5ms capillary column (30 m \times 0.25 mm \times 0.5 µm). The operational conditions were set as follows: injector was at 200°C; ion source was at 230 °C; column oven temperature was programmed to be held at 35°C for 4 minutes, then ramped to 250 °C at a rate of 20°C/minute and held for 10 minutes. The recovery efficiency, method detection limits and quality assurance/quality control have been described in our previous work (Ye et al., 2020).

2.3.6 SP-AMS analysis and mass yields of reaction products

An Aerodyne SP-AMS (Onasch et al., 2012) was applied to analyze the low-volatility organic products, similar to our previous work (Chen et al., 2020; Ge et al., 2017). SP-AMS data were acquired in V mode and analyzed by Squirrel v.1.56D and Pika v1.15D software. The organic fragments were classified into six groups: CH, CHO, CHN, CHO₂, CHON and HO. Elemental ratios (oxygen-to-carbon, O/C; hydrogen-to-carbon, H/C), were calculated according to the method proposed by Canagaratna et al. (2015).

Since the AMS analysis requires nebulization of sample solution into particles before determination, and quantification of organics was influenced by the atomization efficiency and carrier gas flow, we thus cannot use SP-AMS measured concentration to quantify the mass of products directly. In this case, according to Li et al. (2014), we added an internal standard (SO_4^{2-}) prior to AMS analysis, and the mass ratio of particle-phase organics to SO_4^{2-} ($\Delta Org/SO_4^{2-}$) can be used to calculate the mass concentration of products. Furthermore, the mass yield of aqueous-oxidation products ($Y_{products}$, %), which is the mass of products generated per unit mass of precursor consumed, can be calculated according to Eq. (4).

$$Y_{\text{products}}(\%) = \frac{(\Delta \text{Org/SO}_4^{2-})[SO_4^{2-}]_0}{C_0 M_{\eta}} \times 100\%$$
 (4)

Where $[SO_4^{2-}]_0$ is the SO_4^{2-} concentration (here 7.27 mg/L), C_0 is the initial eugenol concentration (in mmol/L), M is MW of the precursor (164 g/mol for eugenol), and η is the degraded fraction of eugenol.

3 Results and discussion

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3.1 Kinetics of aqueous photooxidation

Figure 1 shows unreacted eugenol concentrations (c_t) and the negative logarithm of c_t/c_0 (-ln(c_t/c_0)) as a function of reaction time, respectively. The pseudo first-order rate constants (k) obtained by Eq.(1) were also presented. As described in Fig. 1a, eugenol concentration decreased to be <20% of the initial concentration in 3 hours, suggesting photooxidation was fast under all three reaction conditions. In the presence of ³C*, eugenol was degraded nearly 100% after 3 hours. Previous study (Chen et al., 2020) on ³C*-initiated 4-ethylguaiacol oxidation reports a time of 21 hours for a complete degradation. Apart from difference of precursors, different light irradiation spectra and stronger energy of light in this work than the previous work might be responsible for the fast loss of eugenol. The bond dissociation energies (BDEs) are 340 kJ/mol for OH, 374 kJ/mol for C-H in -CH₃ group, 345 kJ/mol for C=C bond, and 403 kJ/mol for C-H in -OCH3 group, respectively (Herrmann et al., 2003; He et al., 2019). Due to influences of steric hindrance and intramolecular hydrogen bonding, the Habstraction from OH group might not be favorable and the most probable H-abstraction might take place in C=C of the allyl group. As a result, breakage of C=C into C-C at the allyl group can lead to the formation of 2-methoxy-4-propyl-phenol (Section 3.6.1). When photon energy is higher than the BDE, chemical bonds can break, leading to decomposition of compounds and possibly further mineralization. The energy of photon

of 300 nm is 412 kJ/mol and can break all major bonds in eugenol, while the energy of 350 nm is 353 kJ/mol, being able to break some of the bonds in eugenol as well. Overall, eugenol can be easily decomposed after absorbing the photons.

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As shown in Fig. 1b, the first-order rate constants were 2.43×10⁻⁴ s⁻¹, 2.73×10⁻⁴ s⁻¹ ¹, and 5.75×10⁻⁴ s⁻¹ for direct photolysis and photooxidations by OH and ³C*, respectively. 3C*-initiated photooxidation was quicker than that attacked by OH, likely due to combined contributions from reactions with ¹O₂, O₂*- and OH (Section 3.2). Similar results were found for aqueous phase reactions of three phenols against OH and $^3C^*$ by Yu et al. (2016) (Note the initial concentrations of H_2O_2 and DMB were 100 μM and 5 μ M, respectively, with the same ratio as 300 μ M H₂O₂ to 15 μ M DMB in this work)

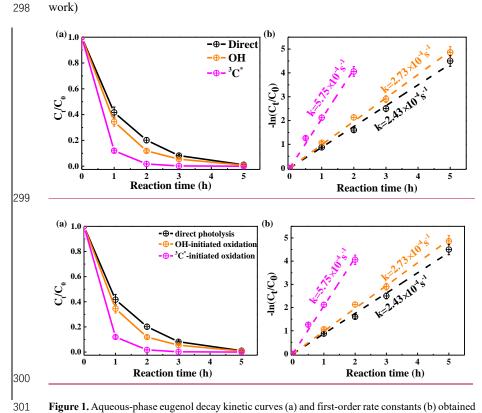


Figure 1. Aqueous-phase eugenol decay kinetic curves (a) and first-order rate constants (b) obtained

based on Equation 1 under direct photolysis, OH-initiated oxidation and ³C*-initiated oxidation.

Error bar represents one standard deviation from the measurements in triplicate.

3.2 Relative importance of ROS in photooxidation

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3.2.1 Quenching experiments in ³C*-initiated photooxidation

Relative importance of different ROS in photooxidation can be investigated by addition of scavengers/quenchers, and then be evaluated based on the different degradation efficiencies of eugenol in absence and presence of the corresponding ROS quenchers. For each quencher, we conducted several gradient experiments with varying molar ratios of eugenol to quencher. The ratios were 0.075:1, 0.15:1, 0.3:1, 0.75:1, 1.5:1 for quenchers of NaN₃, TMP and TBA, and 1.2:1, 1.6:1, 2.5:1, 5:1, 10:1 for SOD, which were all within the typical ranges of ROS quenching experiments reported previously (Zhou et al., 2018). Excess concentrations of quenchers have been added repeatedly to ensure the complete reactions between ROS and scavengers. Figure 2 displays the impacts of quenchers on eugenol degradation. All rate constants (k) with quenchers were lower than those of the quencher-free solutions. The optimum molar ratio of eugenol to quencher was selected such that the eugenol degradation did not change with the increase of added quencher (Wang et al., 2021). For example, along with the decrease of molar ratios of eugenol to NaN3 from 1.5:1 to 0.075:1, the variation of eugenol degradation was stabilized at the ratio of 0.15:1, indicating that ¹O₂ has been completely quenched at this ratio, therefore a molar ratio of 0.15:1 for NaN3 was optimal, since excess scavenger may generate other products that interfere the existing reactions. Finally, the optimal molar ratios of eugenol to quenchers of TBA, NaN3, TMP and SOD, were determined to be 1.5, 0.15, 0.075 and 2.5, respectively. Table 1 and Fig. S1 compared the rate constants determined under the ratios above and they were in an

order of TMP<NaN₃<SOD<TBA, suggesting relative importance of generated ROS to eugenol degradation was in the order of ${}^3C^*>{}^1O_2>O_2$ ${}^{\leftarrow}>$ OH. This result suggests that ${}^3C^*$ plays a major role among all ROS in the photooxidation. Previously, Laurentiis et al. (2013) reported that 4-carboxybenzophenone (70 μ M) could act as ${}^3C^*$ and the photosensitized degradation was more effective than oxidants such as OH, O₃; Misovich et al. (2021) investigated the aqueous DMB-photosensitized reaction (5 μ M, same as in this study) also demonstrated that ${}^3C^*$ was the greatest contributor to phenol or guaiacyl acetone degradation, followed by 1O_2 , while both OH and 1O_2 contributions were relatively minor.

To further assess the relative importance of different ROS, we propose to use the following Eq.(5) for a rough estimation:

337 Redros (in %)=
$$(k-k_{quencher})/k*100\%$$
 (5)

Here k (in s⁻¹) is the original rate constant of ³C*-initiated oxidation (or OH-initiated oxidation in Section 3.2.2) and k_{quencher} (in s⁻¹) is the rate constant after the target ROS has been completely scavenged by its quencher. k and k_{quencher} in fact refer to those reported in Fig. S1b. Redros then refers to the percentages of reduction due to addition of quencher for a ROS.

According to Eq.(5), Red_{3C*} was calculated to be 85.7%. Similarly, the values of ${}^{1}\text{O}_{2}$, O₂. and OH were 80.5%, 61.4% and 53.9%, respectively. Note Red_{ROS} only reflects the relative important of ROS and it does not corresponds to the exact contribution of that ROS in eugenol degradation without quenchers. The reason is that although the addition of a ROS scavenger can eliminate oxidation by this ROS, but it also significantly interrupts the original chain reactions as compared to those in the absence of the scavenger, and reactions with other ROS might be enhanced. In this regard, the sum of the four Red_{ROS} values may exceed 100%. Therefore, one should be

cautious to use Red_{ROS} as a precise quantification of the ROS contribution in aqueous oxidation. Determination of ROS concentrations during oxidation should be instead be an effective way to elucidate the role of ROS. Here, we tried to detect in-situ generated OH, O2*- and ¹O2 during photochemical reactions using a micro electron spin resonance (ESR) spectrometer (Bruker Magnettech, Berlin, Germany) with DMPO as the spin trap to form stable DMPO-OH or DMPO-O2*-, with TEMP to capture ¹O₂ to produce TEMP-¹O₂ spin-adduct (TEMPO). The radicals can be identified and quantified by the peak patterns in ESR spectra, such as the quarter line with a height ratio of 1:2:2:1 for DMPO-OH, 1:1:1:1 for DMPO-O2*- and 1:1:1 for TEMP-1O2 (Guo et al., 2021). Unfortunately, OH radical cannot be detected since its concentration might be lower than the detection limit of the instrument (Fig. S2, ESR spectra of OH). In contrast, we were able to detect high concentrations of ³C* and found the intensity of TEMP-¹O₂ signal reached its maximum at 30 minutes, then decreased slowly (Fig. S2, ESR spectra of ¹O₂). Combining the great quenching effect of TMP with high ¹O₂ concentration from ESR method, we can conclude that ³C* and ¹O₂ play relatively important roles in eugenol photooxidation.

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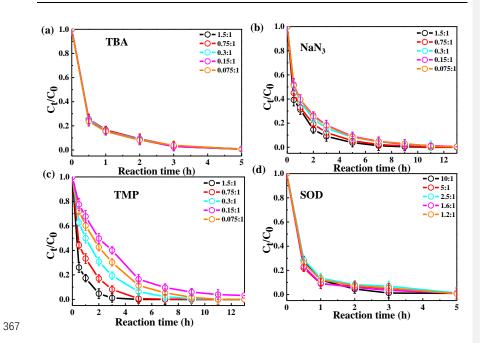


Figure 2. Ratio of unreacted eugenol concentration to its initial concentration (C_t/C_0) at different molar ratios of eugenol to quencher, as a function of reaction time: (a) TBA, (b) NaN₃, (c) TMP and (d) SOD.

3.2.2 Quenching experiments in OH-initiated photooxidation

To examine the contributions of ROS to eugenol degradation for OH-initiated oxidation, TBA and p-BQ as trapping agents were added. Similar to ${}^{3}C^{*}$ -initiated oxidation, several gradient experiments with varying molar ratios of eugenol to quenchers were conducted. The ratios were set as 6.5:1, 3.2:1, 1.6:1, 1.1:1 and 0.8:1 for p-BQ and 3.0:1, 1.5:1, 0.75:1, 0.3:1 and 0.15:1 for TBA. According to Fig. S3, molar ratio only had a slight influence on eugenol degradation, although degradation can be inhibited effectively by quenchers. Thus, we determined the appropriate molar ratios of 0.8 and 0.75 for p-BQ and TBA, respectively, as excess scavengers might influence the

chemical reactions.

Variations of the rate constants for the aforementioned quenching experiments were determined, in comparison with those conducted without quenchers, and results are listed in Table 1 and presented in Fig. S4. For TBA quenching tests, the rate constant decreased by 18.7% (from 2.73×10⁻⁴ s⁻¹ to 2.22×10⁻⁴ s⁻¹), showing that OH radical played a certain role in eugenol photooxidation. Since H₂O₂ was mainly photolyzed at wavelength <300 nm to generate OH radical, irradiation above 300 nm did not affect the reaction significantly. The *p*-BQ could quench O₂-, further suppressing the generation of other ROS (e.g., •HO₂), therefore the rate constant decreased the most (from 2.73×10⁻⁴ s⁻¹ to 1.20×10⁻⁴ s⁻¹), suggesting O₂- was important for eugenol photooxidation. This hypothesis could be further confirmed by the decline of rate constant under N₂-saturated solution (Section 3.2.3). However, it was difficult to detect both OH and O₂-directly due to their relatively short lifetimes and low concentrations via ESR in this work.

³ C*-initiated quenching experiments							
Quenchers	ROS	Reaction rate constant k (s ⁻¹) Pearson's R					
no quencher	-	5.75×10^{-4}	0.996				
TBA	ОН	2.65×10^{-4}	0.999				
SOD	O_2 ·-	2.22×10^{-4}	0.995				
NaN_3	$^{1}\mathrm{O}_{2}$	1.12×10^{-4}	0.999				
TMP	${}^{3}C^{*}$	0.82×10^{-4}	0.999				
OH-initiated quenching experiments							
Quenchers	ROS	Reaction rate constant k (s ⁻¹)	R ²				
No quencher	-	2.73×10 ⁻⁴	0.995				
TBA	OH	2.22×10^{-4}	0.998				
p-BQ	O ₂ ·-	1.20×10^{-4}	0.995				

3.2.3 Influences of different saturated gases

In order to assess the role of O_2 in eugenol degradation, a series of experiments were performed under both O_2 -saturated and N_2 -saturated conditions in addition to air. N_2 gas was purged into reaction solution for ~ 30 minutes before experiment to achieve the O_2 -free condition. Figure 3 compared the changes of eugenol concentrations and rate constants under the three gas conditions for direct photolysis, OH-initiated and ${}^3C^*$ -initiated oxidations, respectively. The rate constants followed the order of $ko_2 > k_{Air} > k_{N_2}$ under both direct photolysis and OH oxidation, providing evidence in support of O_2 being significant for eugenol degradation. This might be explained by the fact that O_2 can act as an electron acceptor to generate $O_2^{\bullet -}$ and $\bullet HO_2$, and subsequently form H_2O_2 and OH. For direct photolysis, rate constant under O_2 -saturated condition increased 14.4% while it decreased 19.3% under N_2 saturation from that under saturated air. For

OH-initiated oxidation, the difference of rate constants under three saturated gases
became more distinct.

On the contrary, rate constants followed the order of $k_{Air} > k_{N_2} > k_{O_2}$ in ${}^3C^*$ initiated oxidation. There are two possible explanations. On one hand, under N₂saturated condition without oxygen, DMB would involve in reactions (R1-R4), leading
to a more effective generation of ${}^3DMB^*$ therefore a higher degradation efficiency than
under O₂-saturated condition. On the other hand, for air/O₂-saturated solutions,
irradiation of DMB and eugenol would involve also reactions (R5-R8) in addition to
(R1-R4), and as a result, the amount of ${}^3DMB^*$ decreased, due to formation of other
ROS (1O_2 , ${}^2O_2^*$, OH, etc) with relatively weak oxidative capacities. In summary,
quenching of ${}^3DMB^*$ by ground state molecular oxygen could account for the low
degradation efficiency in O₂-saturated condition.

425
$$DMB + hv \rightarrow {}^{1}DMB^* \rightarrow {}^{3}DMB^*$$
 (R1)

$$^{3}DMB^{*} \rightarrow DMB \tag{R2}$$

$$^{3}DMB^{*} \rightarrow Products \tag{R3}$$

428
$${}^{3}DMB^{*} + {}^{1}DMB^{*} \rightarrow DMB^{\bullet+/\bullet-}(DMB^{\bullet+} + DMB^{\bullet-})$$
 (R4)

429
$${}^{3}DMB^{*} + O_{2} \rightarrow DMB + {}^{1}O_{2}$$
 (R5)

430
$$DMB^{-} + O_2 \rightarrow DOM^{+} + O_2^{-}$$
 (R6)

431
$$O_2^{-+} + 2H^+ \rightarrow H_2O_2 + O_2$$
 (R7)

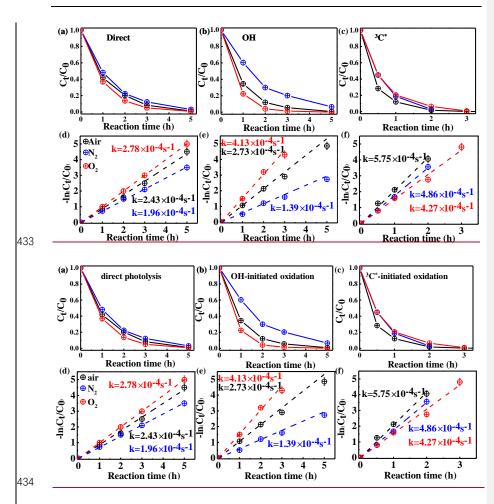


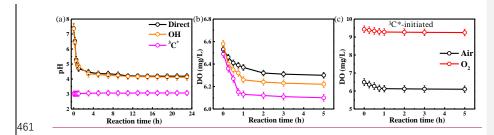
Figure 3. Ratio of unreacted eugenol concentration to its initial concentration (C_t/C_0) as a function of reaction time at different saturated gases under (a) direct photolysis (b) OH-initiated oxidation and (c) ${}^3C^*$ -initiated oxidation. Rate constants of (a-c) are presented in (d-f) correspondingly.

3.2.4 Variations of pH and dissolved oxygen

The initial pH values of reaction solutions for direct photolysis and OH-initiated oxidation were unadjusted, while that for the ${}^{3}C^{*}$ -oxidation was adjusted to 3. The variation of solution pH is presented in Fig. 4a. The pH values decreased quickly at the beginning of illumination (from 7.4 to \sim 5.0 in the first 1 hour) then tended to be flat for

both direct photolysis and OH-initiated oxidation. However, little change of pH (less than 0.1 unit) was observed for the ${}^{3}C^{*}$ -initiated photooxidation throughout the oxidation, which can be likely ascribed to its low initial pH of 3. Since the solution pH was acidic (pH=3), we cannot rule out formation of acidic products (such as organic acids) during ${}^{3}C^{*}$ -initiated oxidation as during direct photolysis and OH-initiated oxidation.

As discussed in Section 3.2.3, oxygen can take part in photochemical reaction to form ROS, which may in turn destroy the structure of precursor. Here we measured the oxygen consumption during oxidation through determination of dissolved oxygen (DO) contents by a dissolved oxygen meter (Seven2Go Pro S9, Zurich, Switzerland). DO was consumed mainly at the first 1 hour and remained stable afterwards (Figs. 4b-c and Fig. S5). The amounts of consumed DO followed the order of ${}^{3}C^{*}$ >OH>direct photolysis. The maximum consumed DO was found in ${}^{3}C^{*}$ -initiated oxidation, which might be explained by the consumption of O₂ that reacts with ${}^{3}C^{*}$ form ${}^{1}O_{2}$ (R5). Obviously, a steady-state DO level was reached when the consumption rate was equal to the diffusion of O₂ into the solution (Pan et al., 2020). Overall, these results re-emphasize that O₂ can influence eugenol degradation and chemical transformation via induction of radical chain reactions.



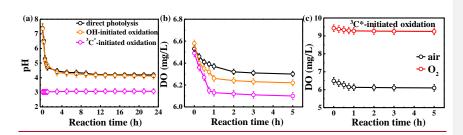


Figure 4. (a) pH values and (b) dissolved oxygen (DO) contents against reaction time under direct photolysis, OH-initiated oxidation, ³C*-initiated oxidation, and (c) DO contents during ³C*-initiated oxidation under air or O₂-saturated conditions.

3.3 Optical properties of reaction products

3.3.1 Light-absorbing properties

The UV-vis light absorption spectra of the solutions at different reaction times are presented in Fig. 5. The light absorption by eugenol itself mainly occurs in the range of 260-300 nm ($n\rightarrow\pi^*$ electronic transition, 270-350 nm), which overlaps with the major photon fluxes (280 and 500 nm) of our lamp for photooxidation. Therefore, we can clearly observe that the characteristic absorption peak at 280 nm of precursor decreased with the propagation of direct photolysis (Fig. 5a), similar to that in OH-initiated photooxidation (Fig. 5b). However, the reaction was quick in the presence of ${}^{3}C^{*}$, and the characteristic absorption peak at 280 nm after 3 hours of illumination almost disappeared, suggesting nearly a complete loss of eugenol, consistent with the results in Section 3.1 that more than 99% eugenol was degraded in 3 hours. Additionally, there was an obvious absorption enhancement at longer wavelengths (300-400 nm) during the photooxidation, whereas eugenol itself did not absorb light in this range, indicating some light-absorbing products (e.g., brown carbon (BrC) species) were generated. Aqueous photooxidation of some phenolic compounds (e.g., vanillic acid) also presented long-wavelength (300-400 nm) light absorbance, with intensity increasing

with illumination time (Tang et al, 2020; Zhao et al., 2015). In addition, there were some differences for light absorbance at wavelength of 300-400 nm in the three reaction conditions. For direct photolysis and OH-initiated oxidation, light absorbance increased during the first 15 hours, then remained at a plateau until 23 hours. However, for ${}^{3}C^{*}$ -initiated oxidation, light absorbance increased during the first 7 hours, then decreased slowly afterwards. The different shapes of UV-vis spectra between OH and ${}^{3}C^{*}$ photooxidations indicate formations of different products.

Compared to the light spectrum of eugenol, there were also increases of light absorbance at ~260 nm ($\pi\rightarrow\pi^*$ electronic transitions) upon aqueous oxidation in all three reaction conditions (Fig. 5), demonstrating the generation of new substances likely with both aromatic C=C and carbonyl (C=O) functional groups (Tang et al., 2020). The enhancement at 300-400 nm may point to products with high MWs and conjugated structures. Unfortunately, we were unable to quantify the relative contributions of individual products to the overall light absorbance between 300 to 400 nm due to lack of a full speciation of the products and their light absorption spectra.

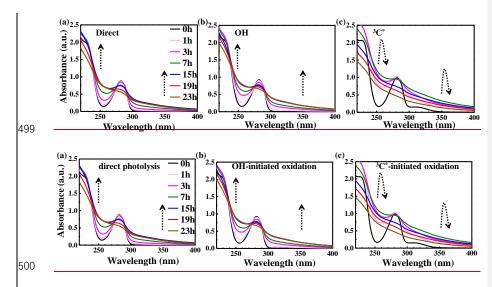


Figure 5. UV-vis light absorption spectra of reacted solutions at different reaction times under (a)

direct photolysis, (b) OH-initiated oxidation, and (c)³C*-initiated oxidation.

3.3.2 Fluorescence properties

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The fluorescence properties of solutions before (0 hour) and during photooxidation (3 and 7 hours) were investigated via the EEM technique, as shown in Fig. 6. For comparison, we also presented EEM profiles of pure eugenol (non-irradiated), pure DMB, and the end solutions (23 hours) of direct photolysis and OH-initiated oxidation in Fig. S6. The peaks at Excitation/Emission (Ex/Em)=275/313 nm can be attributed to fluorescence of the phenolic structure of parent substance (eugenol here), as suggested by Laurentiis et al. (2013). As shown in both Fig. 6 and Fig. S6, the fluorescence intensity decreased after oxidation due to eugenol decay, and the reduction was very fast for ³C*-initiated oxidation. This finding matches with the fast degradation and large rate constant for ³C*-initiated oxidation. The EEM plots for direct photolysis and OHinitiated oxidation had similar contour patterns as shown in Figs. 6a and b, although EEM profiles changed significantly with irradiation time. We also observed distinct fluorescent peaks at Ex/Em=235/(400-500) nm, indicating that illumination can cause a red shift in fluorescence emission wavelength. As suggested by Chang et al. (2010), fluorophores at Ex/Em=240/400 nm are linked with aromatic structures and condensed saturated bonds including polycyclic aromatic hydrocarbons. Another work (Li et al., 2021) showed that red shift in the fluorescence spectra was usually related to an increase in the size of ring system and an increase in the degree of conjugation. Previous studies (Chen et al., 2016a; Chen et al., 2019) have reported that fluorescent compounds with emission wavelength at 400-500 nm were likely linked with HULIS. Additionally, HULIS have two typical fluorescent peaks in EEM profile at Ex/Em=(200-300)/(400-500) nm and Ex/Em=350/(400-500) nm with the former one having a higher intensity

(Graber and Rudich, 2006; Laurentiis et al., 2013; Vione et al., 2019). There was also evidence that direct photolysis of tyrosine and 4-phenoxyphenol generated HULIS with new fluorescence signals at Ex/Em=(200-250)/(400-450) nm and 300/(400-450) nm (Bianco et al., 2014). In this regard, we inferred that new peak at Ex/Em=235/(400-500) nm here was likely attributed to HULIS. For the ³C*-initiated oxidation, extra fluorescent peaks at Ex/Em=(220-300)/(400-500) nm appeared in the first 1 hour (data not shown), but their intensities weakened and gradually disappeared upon prolonged reactions (3 hours). Nevertheless, EEM results should be interpreted with caution because many substances might contribute to absorption and emission at a certain wavelength, and it is hard to distinguish and isolate fluorescent and nonfluorescent constituents simply via the EEM technique.

Another interesting finding was that a small fluorescence peak appeared at Ex/Em=(300-350)/(300-350) nm in some of the EEM profiles. Specifically, it appeared earlier for ${}^{3}C^{*}$ -oxidation (at 3 hours) than the other two systems, yet its intensity seemed to be a bit stronger in the end solutions of direct photolysis and OH-oxidation (Fig. S6). Moreover, as suggested by Leenheer and Croue (2003), fluorescence peak position of the maximum Ex/Em for HULIS with lower MWs would shift towards lower wavelengths, thus, we inferred fluorescence peak at Ex/Em=(300-350)/(300-350) nm might be in part attributed to the organic acids with a few carbon atoms (probably C1-C6). Nevertheless, large uncertainties still exist in using EEM fluorescence technique to identify molecular compositions of the products due to lack of standard EEM profiles for specific compounds from aqueous phase oxidation and clearly more studies are needed in future.

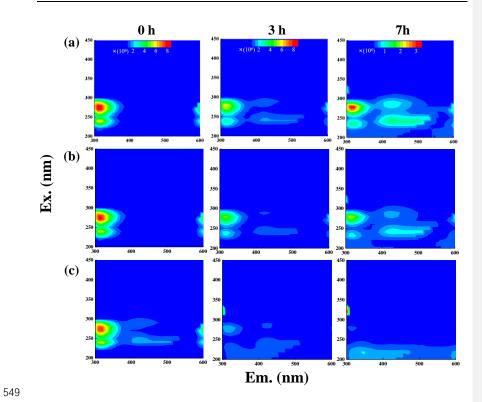


Figure 6. EEM fluorescence spectra of the initial solution (0 hour) and those at different reaction time (3 and 7 hours) under (a) direct photolysis, (b) OH-initiated oxidation, and (c) ${}^3C^*$ -initiated oxidation.

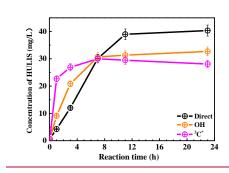
3.4 Characteristics of HULIS

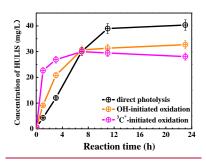
The EEM spectra revealed new prominent fluorescent peak at Ex/Em=250/(400-500) nm, which was likely owing to HULIS. HULIS can be divided into fulvic acid (water soluble at all pHs), humic acid (base soluble, acid insoluble) and humin (insoluble at all pHs). In principle, extracted HULIS in this work with polymer-based HLB SPE packing include LMW organic acids, fulvic acids and other humic substances.

Figure 7 presents the measured HULIS concentrations against the reaction time.

The results show clearly that aqueous-phase eugenol oxidation is a source of HULIS,

and the amount increased gradually in the first 7 hours, then remained at a similar level (about 30 mg/L) for the OH-initiated oxidation. For direct photolysis, HULIS concentration increased until 11 hours and then became steady at a level around 40 mg/L. For the ${}^{3}\text{C}^{*}$ -oxidation, HULIS concentration increased to a maximum at 7 hours, then declined slightly afterwards. A plausible reason of such variabilities is that generated HULIS was capable of further taking part in photochemical reactions since it can act as photosensitizer. Moreover, Yu et al. (2016) characterized the products from aqueous oxidations of phenols by ${}^{3}\text{C}^{*}$ triplet states and OH radicals, and found both could produce oligomers and hydroxylated species but the ${}^{3}\text{C}^{*}$ -oxidation could produce more of these compounds when 50% of the precursor was reacted. Considering the large increases of HULIS in the first 7 hours and the much faster increase of ${}^{3}\text{C}^{*}$ -oxidation in the first 3 hours shown in Fig. 7, we postulate that HULIS species might be some of the high MW oligomers, which can in turn eontributed contribute to fluorescence at emission of ~400 nm (Barsotti et al., 2016).





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Figure 7. HULIS concentrations as a function of reaction time under direct photolysis, OH-initiated oxidation and ${}^{3}C^{*}$ -initiated oxidation.

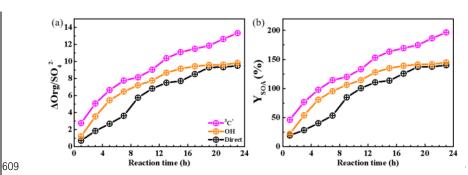
3.5 Mass yield and oxidation degree of reaction products

3.5.1 Mass yields

HULIS is only a subset of the products from aqueous oxidation, and here we used AMS to further quantify the total reaction products. Figure 8a shows SP-AMS measured organic mass profiles (normalized by sulfate mass, ΔOrg/SO₄²⁻) against the reaction time. As the reaction propagated, ΔOrg/SO₄²⁻ increased continuously in ³C*-initiated system. Nevertheless it arose stepwise and reached a maximum at 19 hours, then remained at a plateau for the direct photolysis and OH-mediated oxidation. Figure 8b illustrates the calculated mass yields at different reaction times. The mass yields were in the ranges of 46.2%-196.5%, 22.1%-144.9%, 19.3%-140.1% for ³C*-oxidation, OH-oxidation and direct photolysis, respectively. For the same oxidation time, mass yield from ³C*-oxidation was generally higher than those from OH-oxidation and direct photolysis. There are two plausible reasons for high mass yield of ³C*-initiated oxidation. First, oxidation by ³C* was more efficient to form oligomers and functionalized/oxygenated products (Richards-Henderson et al., 2014; Yu et al., 2016). Higher oxidative degree of products from ³C*-initiated photooxidation (see Sec.3.5.2)

supports this hypothesis. Secondly, more light-absorbing products formed during initial stage of ³C*-oxidation (Fig. 5c) may accelerate oxidation by acting as photosensitizers (Tsui et al., 2018).

The product mass yields obtained in this work (~20%-197%) overall agree with those reported previously for phenolic compounds. For examples, Huang et al. (2018) reported mass yields of 30-120% for syringaldehyde and acetosyringone; Smith et al. (2014) found that mass yields of aqSOA from three phenols with ³C* were nearly 100%, and Ma et al. (2021) reported a yield ranging from 59 to 99% for six highly substituted phenols with ³C*; Mass yields of SOA from three benzene-diols were near 100% with both OH and ³C* oxidants (Smith et al., 2015); Direct photolysis of phenolic carbonyls, and oxidation of syringol by ³C*, had SOA mass yields ranging from 80 to 140% (Smith et al., 2016). Our previous study on eugenol OH oxidation illuminated by a 500 W Xe lamp reported a mass yield of ~180% (Ye et al., 2020), slightly higher than the value determined here owing to different light wavelengths/intensities.



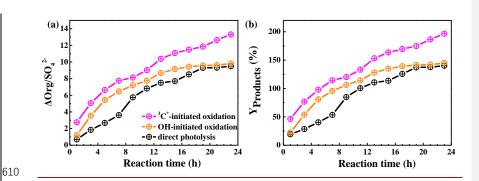


Figure 8. Variations of the organic mass normalized by sulfate (a) (ΔOrg/SO₄²⁻) and (b) mass yields of reaction products with reaction time under direct photolysis, OH-initiated oxidation and ³C*-initiated oxidation.

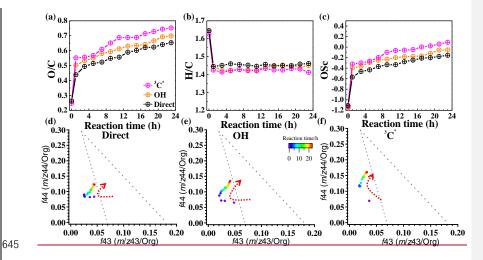
3.5.2 Oxidation degree

In order to further probe oxidation levels of the reaction products, O/C derived from SP-AMS mass spectrum of the organics was used to represent the oxidation degree of products. In addition, carbon oxidation state (OSc, defined as 2*O/C - H/C) (Kroll et al., 2011) was also calculated. Figures 9a-c depict variations of the elemental ratios (O/C and H/C) and OSc during oxidations. DramaticRapid increases of O/C and OSc during the initial stage of oxidation (within 1 hour) were observed, with O/C changing from 0.26 to 0.65, from 0.26 to 0.70, from 0.25 to 0.75, as well as OSc from -1.11 to -0.15, from -1.16 to -0.05, from -1.13 to 0.09 for direct photolysis, OH-oxidation and ³C*-oxidation, respectively. The O/C was lower than those of other phenolic aqSOA (Yu et al., 2014) due to different substituted groups in aromatic ring of the precursors. Both O/C and OSc gradually increased, while H/C changed little after 1 hour. The enhancements of OSc in the end were 1.22, 1.11 and 0.86 for ³C*-initiated oxidation, OH-initiated oxidation and direct photolysis, respectively.

Furthermore, the f_{44} vs. f_{43} diagram ("triangle plot") can be used to demonstrate

the evolution of SOA during oxidation (Ng et al., 2010). The f_{44} and f_{43} are defined as the ratios of signal intensities of m/z 44 (mainly CO₂⁺) and 43 (mainly C₂H₃O⁺) to the total organics. The results that the f_{44} increased continuously (moved upwards) during both OH and 3 C* oxidations, indicating persistent formation of highly oxygenated compounds including organic acids, such as formic acid and oxalic acid (Sun et al., 2010). Note the f_{44} enhancement was much more significant for 3 C* oxidation (from 0.07 to 0.16) than direct photolysis (from 0.07 to 0.12) and OH oxidation (from 0.07 to 0.13), consistent with the behaviors of its higher O/C and OSc. The f_{43} value decreased in the first stage (1-3 hours) and then increased at later stages. The final f_{43} values were almost the same as those of the initial solutions and were small. As a result, all data points located outside the f_{44} vs. f_{43} region (bounded by the two dash lines in Figs. 9d-f) for ambient aerosols established by Ng et al. (2010).

In summary, our results shown here demonstrate that aqueous phase eugenol photochemical oxidation can generate highly oxygenated products and hence increase the degree of oxygenation of overall SOA.



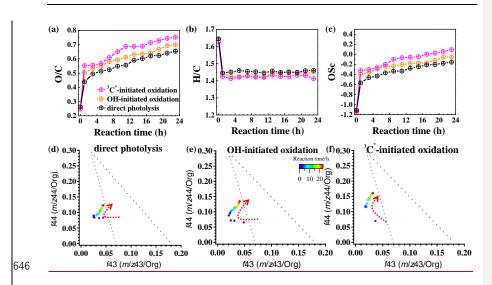


Figure 9. Variations of the elemental ratios of (a) O/C, (b) H/C and (c)oxidation state (OSc) as a function of reaction time; f_{44} vs. f_{43} plots of reaction products under (d) direct photolysis, (e) OH-initiated oxidation, and (f) ${}^{3}C^{*}$ -initiated oxidation.

3.6 Molecular characterization of reaction products and proposed reaction mechanism

3.6.1 Major products identified by GC-MS

SP-AMS was limited to probe bulk composition of low-volatility oxidation products, thus the molecular-level characterization of products was performed by using GC-MS here. The total ion chromatograph (TIC) of GC-MS on the solutions before illumination (0 hour) and at illumination times of 11 and 23 hours for the ${}^{3}C^{*}$ -initiated photooxidation is shown in Fig. S7. As shown in Fig.S7, eugenol (retention time (RT) at 11.50 min) loss was more than 90% at 11 hours, which could be confirmed by the experimental data reported in Section 3.1. Comparison of products at 11 hours and 23 hours showed no significant difference. Similar to aqueous photochemical oxidation

with OH (Ye et al., 2020), a series of products were identified and listed in Table 2. Except 5-ally-3-methoxybenzene-1,2-diol (MW 180, RT=12.59 min), the other eight products were detected for both OH and ${}^{3}C^{*}$ -initiated photooxidations. Some of them (Eugenol, DMB, product 1, 2, 5) were identified by using certified reference materials, some of them (product 3, 4, 6, 7, 8, 9) were inferred according to the molecular ion peaks and fragments from GC-MS, based on spectra from the NIST database (Stein, 2014) and on the reactants and reaction conditions.

We also found 4-(1-hydroxypropyl)-2-methoxyphenol (product 8) was relatively abundant (Fig.S7), suggesting functionalization might dominates dominate as compared to oligomerization and fragmentation. Products were mainly from addition/elimination of hydroxyl (-OH), methoxyl (-OCH₃) to benzene ring or allyl group and further oxidized to carbonyl or carboxyl compounds. As suggested by Bonin et al. (2007), the OH-addition to the aromatic ring of phenol preferentially takes place at the ortho (48%) and the para (36%) positions, leading to the formation of OH-adduct product 6 (5-allyl-3-methoxybenzene-1,2-diol). Notably, dimers and ring-opening products were not observed, but they cannot be excluded since they would be probably out of the detection of GC-MS technique (Vione et al., 2014).

Table 2. Major reaction products identified via GC-MS

	RT	Name*	Proposed chemical	Chemical	Nominal
	(min)		structure	formula	MW
					(g/mol)
Product 1	10.68	4-allylphenol	NU C	C ₉ H ₁₀ O	134
Precursor	11.50	Eugenol	OH O	$C_{10}H_{12}O_2$	164

Product 2	11.81	4-hydroxy-3-	_o	C ₈ H ₈ O ₃	152
		methoxybenzaldehy	но		
		de	O .		
Product 3	12.06	(E)-2-methoxy-4-	но	$C_{10}H_{12}O_2$	164
		(prop-1-en-1-			
		yl)phenol			
Product 4	12.11	4-(hydroxymethyl)-	oн I	$C_8H_{10}O_3$	154
		2-methoxyphenol	ò		
		71	ОН		
Product 5	12.18	2-methoxy-4-	НО	$C_{10}H_{14}O_2$	166
		propylphenol			
Photosensi	12.29	3,4-	0	C ₉ H ₁₀ O ₃	166
tizer	12.2)	dimethoxybenzaldeh		0,111,003	100
		yde(DMB)			
Product	12.59	5-allyl-3-	ОН	$C_{10}H_{12}O_3$	180
6**	12.37	methoxybenzene-	но	010111203	100
Ü		1,2-diol	0 0 0		
Product 7	12.65	4-(1-hydroxyallyl)-	OH .	$C_{10}H_{12}O_3$	180
Troduct /	12.03	2-methoxyphenol	0	C ₁₀ 11 ₁₂ O ₃	100
		z-methoxyphenoi			
			ОН		
Product 8	12.79	4-(1-	OH	$C_{10}H_{14}O_3$	182
		hydroxypropyl)-2-	но		
		methoxyphenol			
Product 9	12.91	(E)-4-(3-	ОН	$C_{10}H_{12}O_3$	180
		hydroxyprop-1-en-1-			
		yl)-2-methoxyphenol			
			HO'		

*Precursor (eugenol) and triplet precursor (DMB) are also shown.

**This compound was only identified in ³C*-oxidation solution.

3.6.2 Reaction mechanism

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The reaction pathways of 3C*-initiated photooxidation of eugenol are demonstrated in Scheme 1 based on the products identified by GC-MS. The other intermediates and the potential pathways were proposed according to the identified products and the reaction rationality from the starting reactant. To better depict the mechanism, DMB was expressed as [RCHO] and eugenol as Ph-R for simplicity. [RCHO] absorbs light and undergoes excitation to ¹[RCHO]*, then experiences the intersystem crossing (ISC) to form ³[RCHO]*. ³[RCHO]* can participate in subsequent reactions via three channels. First, it can react with O₂ to form ¹O₂ via energy transfer. Secondly, it can transform to [RCHO]*-, subsequently reacts with O₂ to generate O₂- via electron transfer, which can disproportionate to H₂O₂. The decomposition of H₂O₂ can generate OH radical. Thirdly, the ³[RCHO]* can react with Ph-R to from [Ph-R•] via H-abstraction. The cleavage of [Ph-R•] to free radical segment (such as CH2CH• or CH3O•) takes place, then an additional hydrogen transfer could occur, resulting in a 2H-addition to the new intermediate to form 4-allyl-phenol (product 1). Similarly, when the CH₂CH• is lost from [Ph-R•], an addition of H₂O would happen on the new compound (product 4) and further oxidized to 4-hydroxy-3methoxybenzaldehyde (product 2). Another possibility is the intermediate [Ph-R•] can resonate to several different isoelectronic species, the radical position changes to aromatic ring or allyl group site, which would couple with HO-OH to form hydroxylated eugenol monomer (product 6, 7, 9 MW=180). Consequently, the isoelectronic species at allyl group site could also abstract a hydrogen to form isoeugenol (product 3 MW=164). Also, breakage of C=C into C-C and 2H-addition at

allyl group site could form 2-methoxy-4-propyl-phenol (product 5, MW=166). Besides, the C=C breaking intermediate can couple with HO-OH to form 4-(1-hydroxypropyl)-2-methoxyphenol (product 8, MW=182). In conclusion, ³C* can directly oxidize eugenol to form SOA products or small molecular compounds, or indirectly oxidize eugenol via energy transfer, electron transfer, hydrogen abstraction, proton-coupled electron transfer or other radical chain reactions.

The organic groups, such as methoxy, allyl groups can be eliminated from aromatic ring, which then participate in photochemical reaction, resulting in generation of dimers, small organic acids, CO_2 and H_2O , etc. Dimers previously reported from aqueous reaction of 4-methylsyringol with OH were not detected via GC-MS in the present work but dimer fragment ions ($C_{20}H_{22}O_4^+$) were detected by SP-AMS with trace amounts. Functionalization due to the additions of hydroxyl, carbonyl functional groups to the aromatic rings could account for the enhancement of light absorption at wavelength of 300-400 nm. However, polar high MW organic acids were not detected likely due to the limitation of GC-MS technique.

Scheme 1. Proposed reaction mechanism of ${}^3C^*$ -initiated photooxidation of eugenol. The red text represents the precursor, and the compounds labeled by Product 1-9 are those identified by GC-MS (Table 2).

3.7 Oxidative potential (OP) of reaction products

The OP of oxidation products can be represented by the consumption rate of DTT concentration, defined as $R_{\rm DTT}$. Figure 10a shows the DTT consumed mass ($M_{\rm DTT}$) as a function of incubation times (0, 30, 60, 90, 120 and 150 min) for a triplicate sample

(300 μ M eugenol) and blank (ultrapure water). M_{DTT} values for both blank and eugenol were proportional to incubation time, indicating that ROS-generating substances in reaction solution act only as catalyst and itself was not consumed. The slopes represent DTT consumption rates, which are also illustrated in Fig. 10a. Average R_{DTT0} (blank) was 0.31 μ M/min and R_{DTT} for initial 300 μ M eugenol (before experiment) was 0.52 μ M/min. Since self-oxidation of DTT might lead to the consumption of DTT in ultrapure water, final DTT consumption rate of reacted solution after oxidation was then blank-corrected by subtracting the average R_{DTT0}.

Figure 10b shows changes of blank-corrected R_{DTT} with reaction time for direct photolysis, OH-initiated oxidation and ³C*-initiated oxidation, respectively. The R_{DTT} value of ³C*-oxidation products increased quickly and reached the maximum (0.9) at 7 hours, then decreased slowly and its end value was lower than that from OH-oxidation. The R_{DTT} value of OH-oxidation products on the other hand increased slowly and reached the maximum at 21 hours. The R_{DTT} value of products from direct photolysis increased continuously but also slowly to ~0.36 till the end of oxidation. Nevertheless, we can see that the final R_{DTT} values were all higher than that of eugenol, proving that aqueous-phase processing can generate products with higher OP, resulting in more health hazards than the precursor does. The DTT consumption rates are comparable to those using the same DTT method (Charrier and Anastasio, 2012; Lin and Yu, 2019). The weak correlation was found between HULIS concentration and R_{DTT}, implying that OP was not only dependent upon HULIS. Moreover, HULIS with diverse molecular structures also exhibit different ROS-generation potentials (Kramer et al., 2016), therefore the HULIS as an ensemble may not correlate well with OP.

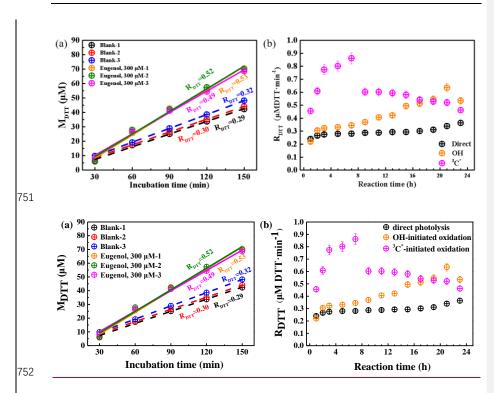


Figure 10. (a) DTT consumed mass versus incubation times for blank (ultrapure water) and 300 μ M eugenol solutions in a triplicate, and (b) blank-corrected DTT consumption rates versus reaction time for direct photolysis, OH-initiated oxidation and ${}^3C^*$ -induced oxidation.

4. Atmospheric implications

The high mass yields of aqueous-phase photooxidation of eugenol (exceeding 100% after 23 hours of illumination) found here are similar or even higher than those previously reported yields of a number of phenolic compounds (e.g., Smith et al., 2014, 2015, 2016; Ma et al., 2021), which re-emphasizes the importance of biomass burning (BB) to SOA budget (Gilardoni et al., 2016), particularly in regions or periods with significant BB activities. In addition, our study here used 300 μ M H₂O₂ and 15 μ M DMB as sources of OH and 3 C*, and 3 C*-mediated oxidation appeared to be faster than

OH-initiated oxidation of eugenol. Of course, whether or not ³C* is more important than OH in real atmosphere depends upon their concentrations. OH and ³C* are difficult to measure and concentrations vary greatly in real atmospheric samples. Herrmann et al. (2010) estimated an average OH level of 0.35x10⁻¹⁴ M in urban fog water; Kaur and Anastasio (2018) measured ³C* concentration to be (0.70-15) x10⁻¹⁴ M, 10-100 times higher than the co-existing OH in ambient fog waters; Kaur et al. (2019) determined both OH and ³C* concentrations in PM extracts, OH steady-state concentration was $4.4(\pm 2.3) \times 10^{-16} \text{ M}$, similar to its level in fog, cloud and rain, while ${}^{3}\text{C*}$ concentration was 1.0(±0.4) x10⁻¹³ M, a few hundred times higher than OH and nearly double its average value in fog. Therefore, together with these measurements, our findings signify a likely more important role of ³C* than OH in aqueous-phase (especially aerosol water) reactions. However, the liquid water content of aerosol is typically ~10000 times smaller than that of cloud (for instance, ~50 μg m⁻³ versus 0.5 g m⁻³). Even if the reaction rates in aerosol water were 10 times higher than those in cloud water, the overall importance of aqueous reactions initiated by the same oxidant in aerosol phase would be still ~1000 times smaller than it in cloud water. Moreover, quenching experiments reveal that O2 can inhibit eugenol degradation by effectively scavenging ³C* while it can promote degradation by fostering chain reactions in OH-induced oxidation, which offer insights to the control of reaction pathways by regulating ROS generations; of course, such operation calls for application of highly sensitive EPR method. Eugenol has a strong light absorption peak around 280 nm, therefore it can undergo direct photolysis, and addition of OH or other photosensitizers (³C*) can gradually

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direct photolysis, and addition of OH or other photosensitizers (³C*) can gradually diminish its light absorption around 280 nm, but increase the absorption in visible light range (>300 nm). In the meantime, HULIS was generated continuously, and GC-MS

identified a number of high MW organic products, in line with those detected in earlier aqueous photooxidation of phenolic compounds (Jiang et al., 2021; Misovich et al., 2021; Tang et al., 2020; Yu et al., 2014). Overall, our work demonstrates that aqueous oxidation of BB emissions is a source of BrC, and this BrC may act as photosensitizer to oxidize other species; a portion of this BrC might be HULIS, and some high MW aromatic compounds are a subset of this HULIS. However, a recent study by Wang et al. (2021) shows that fossil foilfuel derived OA (FFOA) can be an effective precursor of aqSOA, but the aqSOA became less light-absorbing than the FFOA. These contrasting results indicate that contribution of aqueous oxidation to BrC is largely dependent upon the precursors; molecular structures of major chromophores, changes of the structures upon oxidation as well as their interplay with light absorptivity should be carefully investigated to achieve a full understanding of the impacts of aqueous processing on air quality, radiative forcing and climate change.

Investigations on the OPs of reaction products from eugenol photooxidation show that aqueous processing can produce more toxic products than the precursor. This result is in agreement with our previous work on resorcinol, hydroquinone and methoxyhydroquinone (Ou et al., 2021). Although more studies on a broad spectrum of atmospherically relevant species and multiple indicators of toxicity are clearly needed, our findings here underscore the potential of aqueous processing on the enhancement of particle toxicity.

5 Conclusions

This study comprehensively investigated the aqueous photooxidation of eugenol upon direct photolysis and attacks by OH radicals and ³C* triplet states. By using a suite of techniques, the decay kinetics of eugenol, chemical, optical properties as well

as toxicity of reaction products were studied. The first-order rate constants followed the order of ${}^{3}C^{*}>OH>$ direct photolysis (300 μ M H₂O₂ and 15 μ M DMB as sources of OH and ${}^{3}C^{*}$). Further quenching experiments on different ROS during ${}^{3}C^{*}$ -mediated oxidation showed that ${}^{3}C^{*}$ was the major contributor, followed by ${}^{1}O_{2}$, O₂. and OH; O₂. played a more important role than OH during OH-initiated oxidation. The rate constants under saturated O₂, air and N₂ followed the order of $k_{O_2} > k_{Air} > k_{N_2}$ for both direct photolysis and OH-initiated oxidation, but changed to $k_{Air} > k_{N_2} > k_{O_2}$ for ${}^{3}C^{*}$ -mediated oxidation. O₂ appeared to be a scavenger of ${}^{3}C^{*}$ therefore suppressing ${}^{3}C^{*}$ oxidation while it could promote generation of OH thus accelerate OH-mediated oxidation. pH and DO levels both decreased during oxidation, indicating formation of acids and a certain role of DO in oxidation.

Eugenol itself can absorbs lights significantly around 280 nm, and aqueous oxidation gradually decrease this absorption of UV light but enhanced the absorbance in the visible light range (mainly 300-400 nm), indicative of the generation of BrC species. These species were likely linked with HULIS, as HULIS concentration increased during the course of oxidation, in particular for the initial stage of ³C*-mediated reactions. The final mass yields of reaction products (after 23 hours of irradiation) were 140.1%, 144.9% and 196.5% for direct photolysis, OH-oxidation and ³C*-oxidation, respectively. Oxidation degrees of the products increased continuously with the illumination time, indicating persistent formation of highly oxygenated compounds, especially during ³C*-mediated reactions. Molecular characterization by GC-MS identified a series of oxygenated compounds, allowing us to propose the detailed oxidation mechanism. Functionalization appeared to be a dominant pathway to form the observed species.

DTT method was used to assess OP of the reaction products. The end products in

all three sets of experiments showed higher DDT consumption rates than that of the precursor; products from ³C*-oxidation showed particularly fast increase in the first few hours of reactions. This result demonstrates that species that are more toxic than its precursors could be produced upon aqueous oxidation, indicative of the potential toxic effects induced by aqueous processing. Data availability. The relevant data of this study are available at: http://nuistairquality.com/eugenol data and figure Supplement. The supplement related to this article is available on line at: XXX Author Contributions: XDL, YT, LWZ, SSM, SPL, ZZZ and NS conducted the experiments. XDL and YT analyzed the data. XDL and ZLY prepared and wrote the paper with contributions from all co-authors. ZLY and XLG reviewed and commented on the paper. Competing interests. The authors declare that they have no conflict of interest. Acknowledgements. The authors acknowledge support from the National Natural Science Foundation of China (21976093 and 42021004), the Natural Science Foundation of Jiangsu Province (BK20181476), open fund by Jiangsu Key Laboratory of Atmospheric Environment Monitoring and Pollution Control (KHK1904) and the Postgraduate Research & Practice Innovation Program of Jiangsu Province (SJCX21_1332, SJCX20_1030) and of Jiangsu University of Technology

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882 883

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887 888

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890	https://doi.org/10.1039/B201652D, 2002.
891	Barsotti, F., Ghigo, G., and Vione, D. Computational assessment of the fluorescence emission of
892	phenol oligomers: A possible insight into the fluorescence properties of humic-like Substances
893	(HULIS), J. Photochem. Photobio. A, 315, 87-93
894	https://doi.org/10.1016/j.jphotochem.2015.09.012, 2016.
895	Bianco, A., Minella, M., De Laurentiis, E., Maurino, V., Minero, C., and Vione, D. Photochemical
896	generation of photoactive compounds with fulvic-like and humic-like fluorescence in aqueous
897	solution, Chemosphere, 111, 529-536, https://dx.doi.org/10.1016/j.chemosphere.2014.04.035,
898	2014.
899	Bonin, J., Janik, I., Janik, D. and Bartels, D. M.: Reaction of the hydroxyl radical with phenol in water
900	up to supercritical conditions, J. Phys. Chem. A, 111(10), 1869-1878,
901	https://doi.org/10.1021/jp0665325, 2007.
902	Canagaratna, M. R., Jimenez, J. L., Kroll, J. H., Chen, Q., Kessler, S. H., Massoli, P., Hildebrandt Ruiz
903	L., Fortner, E., Williams, L. R., Wilson, K. R., Surratt, J. D., Donahue, N. M., Jayne, J. T., and
904	Worsnop, D. R.: Elemental ratio measurements of organic compounds using aerosol mass
905	spectrometry: characterization, improved calibration, and implications, Atmos. Chem. Phys.
906	15, 253-272, https://doi.org/10.5194/acp-15-253-2015, 2015.
907	Chang, J. L., and Thompson, J. E.: Characterization of colored products formed during irradiation o
908	aqueous solutions containing $\mathrm{H}_{2}\mathrm{O}_{2}$ and phenolic compounds, Atmos. Environ., 44, 541-551
909	https://doi.org/10.1016/j.atmosenv.2009.10.042, 2010.
910	Charrier, J. G., and Anastasio, C.: On dithiothreitol (DTT) as a measure of oxidative potential for ambien
911	particles: evidence for the importance of soluble transition metals, Atmos. Chem. Phys. 12
912	9321-9333, https://doi.org/10.5194/acp-12-9321-2012, 2012.
913	Chen, H., Ge, X., Ye, Z.: Aqueous-phase secondary organic aerosol formation via reactions with organic
914	triplet excited states—a short review. Curr. Pollut. Rep., 4, 8-12
915	https://doi.org/10.1007/s40726-018-0079-7, 2018.
916	Chen, Q., Ikemori, F., and Mochida, M.: Light Absorption and excitation-emission fluorescence of urban
917	organic aerosol components and their relationship to chemical structure, Environ. Sci. Technol.
918	50, 10859-10868, https://doi.org/10.1021/acs.est.6b02541, 2016a.
919	Chen, Q., Miyazaki, Y., Kawamura, K., Matsumoto, K., Coburn, S., Volkamer, R., Iwamoto, Y., Kagami

920	S., Deng, Y., Ogawa, S., Ramasamy, S., Kato, S., Ida, A., Kajii, Y., and Mochida, M.:
921	Characterization of chromophoric water-soluble organic matter in urban, forest, and marine
922	aerosols by HR-ToF-AMS analysis and excitation-emission matrix spectroscopy, Environ. Sci.
923	Technol., 50, 10351-10360, https://doi.org/10.1021/acs.est.6b01643, 2016b.
924	Chen, Q., Wang, M., Wang, Y., Zhang, L., Li, Y., and Han, Y.: Oxidative potential of water-soluble matter
925	associated with chromophoric substances in $PM_{2.5}$ over Xi'an, China, Environ. Sci. Technol., 53,
926	8574-8584, https://doi.org/10.1021/acs.est.9b01976, 2019.
927	Chen, Y., Li, N., Li, X., Tao, Y., Luo, S., Zhao, Z., Ma, S., Huang, H., Chen, Y., Ye, Z., and Ge, X.:
928	Secondary organic aerosol formation from ³ C*-initiated oxidation of 4-ethylguaiacol in
929	atmospheric aqueous-phase, Sci. Total. Environ., 723, 137953,
930	https://doi.org/10.1016/j.scitotenv.2020.137953, 2020.
931	Cho, A. K., Sioutas, C., Miguel, A. H., Kumagai, Y., Schmitz, D. A., Singh, M., Eiguren-Fernandez, A.,
932	and Froines, J. R.: Redox activity of airborne particulate matter at different sites in the Los
933	Angeles Basin, Environ. Res., 99, 40-7, https://doi.org/10.1016/j.envres.2005.01.003, 2005.
934	De Laurentiis, E., Sur, B., Pazzi, M., Maurino, V., Minero, C., Mailhot, G., Brigante, M., and Vione, D.:
935	Phenol transformation and dimerisation, photosensitised by the triplet state of 1-
936	nitronaphthalene: A possible pathway to humic-like substances (HULIS) in atmospheric waters,
937	Atmos. Environ., 70, 318-327, https://doi.org/10.1016/j.atmosenv.2013.01.014, 2013.
938	Dou, J., Lin, P., Kuang, B. Y., and Yu, J.: Reactive oxygen species production mediated by humic-like
939	substances in atmospheric aerosols: enhancement effects by pyridine, imidazole, and their
940	derivatives, Environ. Sci. Technol., 49(11), 6457-6465, https://doi.org/10.1021/es5059378,
941	2015.
942	Ervens, B., Turpin, B. J., and Weber, R. J.: Secondary organic aerosol formation in cloud droplets and
943	aqueous particles (aqSOA): a review of laboratory, field and model studies, Atmos. Chem. Phys.,
944	11, 11069-11102, https://doi.org/10.5194/acp-11-11069-2011, 2011.
945	Fang, T., Verma, V., Bates, J. T., Abrams, J., Klein, M., Strickland, M. J., Sarnat, S. E., Chang, H. H.,
946	Mulholland, J. A., Tolbert, P. E., Russell, A. G., and Weber, R. J.: Oxidative potential of ambient
947	water-soluble PM _{2.5} in the southeastern United States: contrasts in sources and health
948	associations between ascorbic acid (AA) and dithiothreitol (DTT) assays, Atmos. Chem. Phys.,
949	16, 3865-3879, https://doi.org/10.5194/acp-16-3865-2016, 2016.

950	Faust, J. A., Wong, J. P., Lee, A. K., and Abbatt, J. P.: Role of aerosol liquid water in secondary organic
951	aerosol formation from volatile organic compounds, Environ. Sci. Technol., 51, 1405-1413,
952	https://doi.org/10.1021/acs.est.6b04700, 2017.
953	Ge, X., Li, L., Chen, Y., Chen, H., Wu, D., Wang, J., Xie, X., Ge, S., Ye, Z., Xu, J., and Chen, M. Aerosol
954	characteristics and sources in Yangzhou, China resolved by offline aerosol mass spectrometry
955	and other techniques. Environ. Pollut., 225, 74-85, https://doi.org/
956	10.1016/j.envpol.2017.03.044, 2017.
957	George, K. M., Ruthenburg, T. C., Smith, J., Yu, L., Zhang, Q., Anastasio, C., and Dillner, A. M.: FT-IR
958	quantification of the carbonyl functional group in aqueous-phase secondary organic aerosol from
959	phenols, Atmos. Environ., 100, 230-237, https://doi.org/10.1016/j.atmosenv.2014.11.011, 2015.
960	Gilardoni, S., Massoli, P., Paglione, M., Giulianelli, L., Carbone, C., Rinaldi, M., Decesari, S., Sandrini,
961	S., Costabile, F., Gobbi, G. P., Pietrogrande, M. C., Visentin, M., Scotto, F., Fuzzi, S., and
962	Facchini, M. C.: Direct observation of aqueous secondary organic aerosol from biomass-
963	burning emissions, Proc. Natl. Acad. Sci. USA., 113, 10013-10018,
964	https://doi.org/10.1073/pnas.1602212113, 2016.
965	Gligorovski, S., Strekowski, R., Barbati, S., and Vinoe, D.: Environmental implications of hydroxyl
966	radicals (OH), Chem. Rev., 115(24), 13051-13092, https://doi.org/10.1021/cr500310b, 2015.
967	Graber, E. R., and Rudich, Y.: Atmospheric HULIS: how humic-like are they? A comprehensive and
968	critical review, Atmos. Chem. Phys., 6, 729-753, https://doi.org/10.5194/acp-6-729-2006,
969	2006.
970	Guo, Y., Zhang, Y., Yu, G., and Wang, Y., Revisiting the role of reactive oxygen species for pollutant
971	abatement during catalytic ozonation: the probe approach versus the scavenger approach, Appl.
972	Catal. B Environ., 280, 119418, https://doi.org/10.1016/j.apcatb.2020.119418, 2021.
973	Hawthorne, S.B., Krieger M.S., Miller D.J., and Mathiason M.B. Collection and quantitation of
974	methoxylated phenol tracers for atmospheric pollution from residential wood stoves, Environ. Sci.
975	Technol., 23,470-475, https://doi.org/10.1021/es00181a013, 1989.
976	He, L., Schaefer, T., Otto, T., Kroflic, A., and Herrmann, H.: Kinetic and theoretical study of the
977	atmospheric aqueous-phase reactions of OH radicals with methoxyphenolic compounds, J. Phys.
978	Chem. A, 123, 7828-7838, https://doi.org/10.1021/acs.jpca.9b05696, 2019.

Herrmann, H.: Kinetics of aqueous phase reaction relevant for atmospheric chemistry, Chem. Rev., 103,

980	4691-4716, https://doi.org/10.1021/cr020658q, 2003.
981	Herrmann, H., Hoffmann, D., Schaefer, T., Bräuer, P. and Tilgner, A.: Tropospheric aqueous-phase free-
982	radical chemistry: Radical sources, spectra, reaction kinetics and prediction tools.
983	ChemPhysChem, 11, 3796-3822, https://doi.org/10.1002/ephc.201000533, 2010.
984	Herrmann, H., Schaefer, T., Tilgner, A., Styler, S. A., Weller, C., Teich, M. and Otto, T.: Tropospheric
985	aqueous-phase chemistry: kinetics, mechanisms, and its coupling to a changing gas phase,
986	Chem. Rev., 115(10), 4259-4334, https://doi.org/10.1021/cr500447k, 2015.
987	Hong, J., Han, B., Yuan, N., and Gu, J.: The roles of active species in photo-decomposition of organic
988	compounds by microwave powered electrodeless discharge lamps, J. Environ. Sci. (China), 33,
989	60-68, https://doi.org/10.1016/j.jes.2014.12.016, 2015.
990	Huang, D., Zhang, X., Chen, Z. M., Zhao, Y., and Shen, X. L.: The kinetics and mechanism of an aqueous
991	phase isoprene reaction with hydroxyl radical, Atmos. Chem. Phys., 11, 7399-7415,
992	https://doi.org/10.5194/acp-11-7399-2011, 2011.
993	Huang, D., Zhang, Q., Cheung, H. H. Y., Yu, L., Zhou, S., Anastasio, C., Smith, J. D., and Chan, C. K.:
994	Formation and evolution of aqSOA from aqueous-phase reactions of phenolic carbonyls:
995	comparison between ammonium sulfate and ammonium nitrate solutions, Environ. Sci.
996	Technol., 52, 9215-9224, https://doi.org/10.1021/acs.est.8b03441, 2018.
997	Huo, Y., Guo, Z., Li, Q., Wu, D., Ding, X., Liu, A., Huang, D., Qiu, G., Wu, M., Zhao, Z., Sun, H., Song,
998	W., Li, X., Chen, Y., Wu, T., and Chen, J. Chemical fingerprinting of HULIS in particulate
999	matters emitted from residential coal and biomass combustion, Environ. Sci. Technol., 55, 3593-
1000	3603. https://doi.org/10.1021/acs. est.0c08518, 2021.
1001	Jiang, W., Misovich, M. V., Hettiyadura, A. P. S., Laskin, A., McFall, A. S., Anastasio, C., and Zhang, Q.:
1002	Photosensitized reactions of a phenolic carbonyl from wood combustion in the aqueous phase-
1003	chemical evolution and light absorption properties of aqSOA, Environ. Sci. Technol., 55, 5199-
1004	5211, https://doi.org/10.1021/acs.est.0c07581, 2021.
1005	Kaur, R., and Anastasio, C.: First measurements of organic triplet excited states in atmospheric waters,
1006	Environ. Sci. Technol., 52, 5218-5226, https://doi.org/10.1021/acs.est.7b06699, 2018.
1007	Kaur, R., Labins, J. R., Helbock, S. S., Jiang, W., Bein, K. J., Zhang, Q., and Anastasio, C.: Photooxidants
1008	from brown carbon and other chromophores in illuminated particle extracts, Atmos. Chem.
1009	Phys., 19, 6579-6594, https://doi.org/10.5194/acp-19-6579-2019, 2019.

1010	Kramer, A.J., Rattanavaraha, W., Zhang, Z., Gold, A., Surratt, J.D., and Lin, YH. Assessing the oxidative
1011	potential of isoprene-derived epoxides and secondary organic aerosol, Atmos. Environ., 130
1012	211-218, https://dx.doi.org/10.1016/j.atmosenv.2015.10.018, 2016.
1013	Kroll, J. H., Donahue, N. M., Jimenez, J. L., Kessler, S. H., Canagaratna, M. R., Wilson, K. R., Altieri
1014	K. E., Mazzoleni, L. R., Wozniak, A. S., Bluhm, H., Mysak, E. R., Smith, J. D., Kolb, C. E., and
1015	Worsnop, D. R.: Carbon oxidation state as a metric for describing the chemistry of atmospheric
1016	organic aerosol, Nat. Chem., 3, 133-9, https://doi.org/10.1038/nchem.948, 2011.
1017	Laurentiis, E. D., Socorro, J., Vione, D., Quivet, E., Brigante, M., Mailhot, G., Wortham, H., and
1018	Gligorovski, S.: Phototransformation of 4-phenoxyphenol sensitised by 4
1019	carboxybenzophenone: evidence of new photochemical pathways in the bulk aqueous phase and
1020	on the surface of aerosol deliquescent particles, Atmos. Environ., 8, 569-578
1021	https://doi.org/10.1016/j.atmosenv.2013.09.036, 2013.
1022	Lee, A. K. Y., Hayden, K. L., Herckes, P., Leaitch, W. R., Liggio, J., Macdonald, A. M., and Abbatt, J. P.
1023	D.: Characterization of aerosol and cloud water at a mountain site during WACS 2010: secondary
1024	organic aerosol formation through oxidative cloud processing, Atmos. Chem. Phys., 12, 7103-7116
1025	https://doi.org/10.5194/acp-12-7103-2012, 2012.
1026	Leenheer, J. A., and Croue, J. P. Characterizing aquatic dissolved organic matter, Environ. Sci
1027	Technol., 37, 18A-26A, https://doi.org/10.1021/es032333c, 2003.
1028	Li, F., Tsona, N. T., Li, J., and Du, L.: Aqueous-phase oxidation of syringic acid emitted from biomass
1029	burning: formation of light-absorbing compounds, Sci. Total Environ., 765, 144239
1030	https://doi.org/10.1016/j.scitotenv.2020.144239, 2021.
1031	Li, Y. J., Huang, D. D., Cheung, H. Y., Lee, A. K. Y., and Chan, C. K.: Aqueous-phase photochemical
1032	oxidation and direct photolysis of vanillin-a model compound of methoxy phenols from biomass
1033	burning, Atmos. Chem. Phys., 14, 2871-2885, https://doi.org/10.5194/acp-14-2871-2014, 2014
1034	Lim, Y. B., Tan, Y., Perri, M. J., Seitzinger, S. P., and Turpin, B. J.: Aqueous chemistry and its role in
1035	secondary organic aerosol (SOA) formation, Atmos. Chem. Phys., 10, 10521-10539
1036	https://doi.org/10.5194/acpd-10-14161-2010, 2010.
1037	Lin, M., and Yu, J. Z.: Dithiothreitol (DTT) concentration effect and its implications on the applicability
1038	of DTT assay to evaluate the oxidative potential of atmospheric aerosol samples, Environ
1039	Pollut., 251, 938-944, https://doi.org/10.1016/j.envpol.2019.05.074, 2019.

0 <u>Lu, .</u>	J., Ge, X., Liu, Y., Chen, Y., Xie, X., Ou, Y., Ye, Z., Chen, M. Significant secondary organic aerosol
1	production from aqueous-phase processing of two intermediate volatility organic compounds.
-2	Atmos. Environ., 211, 63-68, https://doi.org/10.1016/j.atmosenv.2019.05.014, 2019.
3 Ma,	L., Guzman, C., Niedek, C., Tran, T., Zhang, Q. and Anastasio, C.: Kinetics and mass yields of
4	aqueous secondary organic aerosol from highly substituted phenols reacting with a triplet excited
j	state, Environ. Sci. Technol., 55(9), 5772-5781, doi:10.1021/acs.est.1c00575, 2021.
Ma,	Y., Cheng, Y., Qiu, X., Cao, G., Kuang, B., Yu, J.Z., and Hu, D. Optical properties, source
	apportionment and redox activity of Humic-Like Substances (HULIS) in airborne fine
	particulates in Hong Kong, Environ. Pollut., 255,113087,
	https://doi.org/10.1016/j.envpol.2019.113087.https://doi.org/10.1016/j.envpol.2019.113087.
	2019.
Mab	ato, B. R. G., Lyu, Y., Ji, Y., Li, Y., Huang, D., Li, X., Nah, T., Lam, C. H., and Chan, C. K.: Aqueous
	secondary organic aerosol formation from the direct photosensitized oxidation of vanillin in the
	absence and presence of ammonium nitrate, Atmos. Chem. Phys., 22, 273-293,
	https://doi.org/10.5194/acp-22-273-2022, 2022.
McV	Whinney, R. D., Zhou, S., and Abbatt, J. P. D.: Naphthalene SOA: redox activity and naphthoquinone
	gas-particle partitioning, Atmos. Chem. Phys., 13, 9731-9744, https://doi.org/10.5194/acp-13-
	9731-2013, 2013.
Miso	ovich, M. V., Hettiyadura, A. P. S., Jiang, W. Q., and Zhang, Q. Molecular-level study of the photo-
	oxidation of aqueous-phase guaiacyl acetone in the presence of ³ C*: formation of brown carbon
	products, ACS Earth Space Chem., 5, 1983-1996,
	https://doi.org/10.1021/acsearthspacechem.1c00103, 2021.
Mla	denov, N, Alados-Arboledas, L., Olmo, F.J., Lyamani, H., Delgado, A., Molina, A., and Reche, I.:
	Applications of optical spectroscopy and stable isotope analyses to organic aerosol source
	discrimination in an urban area, Atmos. Environ., 45, 1960-1969, https://doi.org/
	10.1016/j.atmosenv.2011.01.029, 2011.
Nau	W. M., and Scaiano, J. C.: Oxygen quenching of excited aliphatic ketones and diketones, J. Phys.
	Chem., 100, 11360-11367, https://doi.org/10.1021/jp960932i, 1996.
Ng,	N. L., Canagaratna, M. R., Zhang, Q., Jimenez, J. L., Tian, J., Ulbrich, I. M., Kroll, J. H., Docherty,
	K. S., Chhabra, P. S., Bahreini, R., Murphy, S. M., Seinfeld, J. H., Hildebrandt, L., Donahue, N.

1070	M., DeCarlo, P. F., Lanz, V. A., Prevot, A. S. H., Dinar, E., Rudich, Y., and Worsnop, D. R.:
1071	Organic aerosol components observed in Northern Hemispheric datasets from aerosol mass
1072	spectrometry, Atmos. Chem. Phys., 10, 4625-4641, https://doi.org/10.5194/acp-10-4625-2010,
1073	2010.
1074	Onasch, T. B., Trimborn, A., Fortner, E. C., Jayne, J. T., Kok, G. L., Williams, L. R., Davidovits, P., and
1075	Worsnop, D. R. Soot particle aerosol mass spectrometer: Development, validation, and initial
1076	application. Aerosol Sci. Tech., 46, 804-817, http://dx.doi.org/10.1080/02786826.2012.663948,
1077	2012.
1078	Ou, Y., Nie, D., Chen, H., Ye, Z., Ge, X.: Characterization of products from the aqueous-phase
1079	photochemical oxidation of benzene-diols. Atmosphere, 12, 534,
1080	https://doi.org/10.3390/atmos12050534, 2021.
1081	Pan, Y., Ma, H., Li, Z., Du, Y., Liu, Y., Yang, J., and Li, G.: Selective conversion of lignin model veratryl
1082	alcohol by photosynthetic pigment via photo-generated reactive oxygen species, Chem. Eng. J.,
1083	393, 124772, https://doi.org/10.1016/j.cej.2020.124772, 2020.
1084	Raja, P., Bozzi, A., Mansilla, H., and Kiwi, J.: Evidence for superoxide-radical anion, singlet oxygen and
1085	OH-radical intervention during the degradation of the lignin model compound (3-methoxy-4-
1086	hydroxyphenylmethylcarbinol), J. Photochem. Photobiol. Chem., 169, 271-278,
1087	https://doi.org/10.1016/j.jphotochem.2004.07.009, 2005.
1088	Richards-Henderson, N. K., Hansel, A. K., Valsaraj, K. T., and Anastasio, C. Aqueous oxidation of green
1089	leaf volatiles by hydroxyl radical as a source of SOA: Kinetics and SOA yields, Atmos. Environ.
1090	95, 105-112, http://dx.doi.org/10.1016/j.atmosenv.2014.06.026 , 2014.
1091	Rossignol, S., Aregahegn, K. Z., Tinel, L., Fine, L., Nozière, B., and George, C.: Glyoxal induced
1092	atmospheric photosensitized chemistry leading to organic aerosol growth, Environ. Sci.
1093	Technol., 48, 3218-3227, https://doi.org/10.1021/es405581g, 2014.
1094	Scharko, N. K., Berke, A. E., and Raff, J. D.: Release of nitrous acid and nitrogen dioxide from nitrate
1095	photolysis in acidic aqueous solutions, Environ. Sci. Technol., 48, 11991-2001,
1096	https://doi.org/10.1021/es503088x, 2014.
1097	Simpson, C.D., Paulsen, M., Dills, R. L., Liu, LJ.S., and Kalman, A.A. Determination of
1098	methoxyphenols in ambient atmospheric particulate matter: Tracers for wood combustion,
1099	Environ. Sci. Technol., 39, 631-637, https://doi.org/10.1021/es0486871, 2005.

1100	Smith, J. D., Kinney, H., and Anastasio, C.: Aqueous benzene-diols react with an organic triplet excited
1101	state and hydroxyl radical to form secondary organic aerosol. Phys. Chem. Chem. Phys., 17,
1102	10227, https://doi.10.1039/c4cp06095d, 2015.
1103	Smith, J. D., Kinney, H, and Anastasio, C.: Phenolic carbonyls undergo rapid aqueous photodegradation
1104	to form low-volatility, light-absorbing products, Atmos. Environ., 126, 36-44,
1105	https://doi.org/10.1016/j.atmosenv.2015.11.035, 2016.
1106	Smith, J. D., Sio, V., Yu, L., Zhang, Q., and Anastasio, C.: Secondary organic aerosol production from
1107	aqueous reactions of atmospheric phenols with an organic triplet excited state, Environ. Sci.
1108	Technol., 48, 1049-1057, https://doi.org/10.1021/es4045715, 2014.
1109	Stephen E. Stein (2014), NIST/EPA/NIH Mass Spectral Library with Search Program - SRD 1a, National
1110	Institute of Standards and Technology, https://doi.org/10.18434/T4H594 (Accessed 2022-04-29)
1111	Sun, Y., Zhang, Q., Anastasio, C., and Sun, J.: Insights into secondary organic aerosol formed via
1112	aqueous-phase reactions of phenolic compounds based on high resolution mass spectrometry,
1113	Atmos. Chem. Phys., 10, 4809–4822, https://doi.org/10.5194/acp-10-4809-2010, 2010.
1114	Tang, S., Li, F., Tsona, N.T., Lu, C., Wang, X., and Du, L.: Aqueous-phase photooxidation of vanillic
1115	acid: a potential source of humic-like substances (HULIS), ACS Earth Space Chem., 4, 862-
1116	872, https://doi.org/10.1021/acsearthspacechem.0c00070, 2020.
1117	Tsui, W. G., and McNeill, V. F. Modeling secondary organic aerosol production from photosensitized
1118	humic-like substances (HULIS), Environ. Sci. Technol. Lett., 5, 255-259.
1119	https://doi.org/10.1021/acs.estlett.8b00101, 2018.
1120	Verma, V., Fang, T., Xu, L., Peltier, R. E., Russell, A. G., Ng, N. L., and Weber, R. J.: Organic aerosols
1121	associated with the generation of reactive oxygen species (ROS) by water-soluble PM2.5,
1122	Environ. Sci. Technol., 49, 4646-56, https://doi.org/10.1021/es505577w, 2015.
1123	Vione, D., Albinet, A., Barsotti, F., Mekic, M., Jiang, B., Minero, C., Brigante, M., and Gligorovski, S.:
1124	Formation of substances with humic-like fluorescence properties, upon photoinduced
1125	oligomerization of typical phenolic compounds emitted by biomass burning, Atmos. Environ.,
1126	206, 197-207, https://doi.org/10.1016/j.atmosenv.2019.03.005, 2019.
1127	Vione, D., Maurino, V., Minero, C., Pelizzetti, E., Harrison, M. A., Olariu, R. I., and Arsene, C.:
1128	Photochemical reactions in the tropospheric aqueous phase and on particulate matter, Chem.
1129	Soc. Rev., 35, 441-53, https://doi.org/10.1039/b510796m, 2006.

1130	Vione, D., Maurino, V., and Minero, C.: Photosensitised humic-like substances (HULIS) formation
1131	processes of atmospheric significance: a review, Environ. Sci. Pollut. Res., 21, 11614-11622,
1132	https://doi.org/10.1007/s11356-013-2319-0, 2014.
1133	Wang, J., and Wang, S. Reactive species in advanced oxidation processes: Formation, identification and
1134	reaction mechanism, Chem. Eng.J., 401, 126158, https://doi.org/10.1016/j.cej.2020.126158,
1135	2020.
1136	Wang, J., Ye, J., Zhang, Q., Zhao, J., Wu, Y., Li, J., Liu, D., Li, W., Zhang, Y., Wu, C., Xie, C., Qin, Y.,
1137	Lei, Y., Huang, X., Guo, J., Liu, P., Fu, P., Li, Y., Lee, H. C., Choi, H., Zhang, J., Liao, H., Chen,
1138	M., Sun, Y., Ge, X., Martin, S. T., and Jacob, D. J.: Aqueous production of secondary organic
1139	aerosol from fossil-fuel emissions in winter Beijing haze. Proc. Natl. Acad. Sci. USA., 118,
1140	e2022179118, https://doi.org/10.1073/pnas.2022179118, 2021.
1141	Wang, L., Lan, X., Peng, W., and Wang, Z.: Uncertainty and misinterpretation over identification,
1142	quantification and transformation of reactive species generated in catalytic oxidation processes:
1143	A review, J Hazard. Mater., 408, 124436, https://doi.org/10.1016/j.jhazmat.2020.124436, 2021.
1144	Xu, X., Lu, X., Li, X., Liu, Y., Wang, X., Chen, H., Chen, J., Yang, X., Fu, T., Zhao, Q., and Fu, Q. ROS-
1145	generation potential of Humic-like substances (HULIS) in ambient PM _{2.5} in urban Shanghai:
1146	Association with HULIS concentration and light absorbance, Chemosphere, 256, 127050,
1147	https://doi.org/10.1016/j.chemosphere.2020.127050 0045-6535, 2020.
1148	Yang, J., Au, W. C., Law, H., Lam, C. H., and Nah, T.: Formation and evolution of brown carbon during
1149	aqueous-phase nitrate-mediated photooxidation of guaiacol and 5-nitroguaiacol, Atmos.
1150	Environ., 254, 118401, https://doi.org/10.1016/j.atmosenv.2021.118401, 2021.
1151	Ye, Z., Zhuang, Y., Chen, Y., Zhao, Z., Ma, S., Huang, H., Chen, Y., and Ge, X.: Aqueous-phase oxidation
1152	of three phenolic compounds by hydroxyl radical: Insight into secondary organic aerosol
1153	formation yields, mechanisms, products and optical properties, Atmos. Environ., 223, 117240,
1154	https://doi.org/10.1016/j.atmosenv.2019.117240, 2020.
1155	Yu, L., Smith, J., Laskin, A., Anastasio, C., Laskin, J., and Zhang, Q.: Chemical characterization of SOA
1156	formed from aqueous-phase reactions of phenols with the triplet excited state of carbonyl and
1157	hydroxyl radical, Atmos. Chem. Phys., 14, 13801-13816, https://doi.org/10.5194/acp-14-
1158	13801-2014, 2014.
1159	Yu, L., Smith, J., Laskin, A., George, K. M., Anastasio, C., Laskin, J., Dillner, A. M., and Zhang, Q.:

1160	Molecular transformations of phenolic SOA during photochemical aging in the aqueous phase:
1161	$competition\ among\ oligomerization,\ functionalization,\ and\ fragmentation,\ Atmos.\ Chem.\ Phys.,$
1162	16, 4511-4527, https://doi.org/10.5194/acp-16-4511-2016, 2016.
1163	Zhang, T., Huang, S., Wang, D., Sun, J., Zhang, Q., Xu, H., Ho, S., Cao, J., and Shen, Z. Seasonal and
1164	diurnal variation of $PM_{2.5}$ HULIS over Xi'an in Northwest China: Optical properties, chemical
1165	functional group, and relationship with reactive oxygen species (ROS), Atmos. Environ., 268,
1166	118782, https://doi.org/10.1016/j.atmosenv.2021.118782, 2022.
1167	Zhang, X., Chen, Z. M., and Zhao, Y.: Laboratory simulation for the aqueous OH-oxidation of methyl
1168	vinyl ketone and methacrolein: significance to the in-cloud SOA production, Atmos. Chem.
1169	Phys., 10, 9551-9561, https://doi.org/10.5194/acp-10-9551-2010, 2010.
1170	Zhao, R., Lee, A. K., and Abbatt, J. P.: Investigation of aqueous-phase photooxidation of glyoxal and
1171	methylglyoxal by aerosol chemical ionization mass spectrometry: observation of
1172	hydroxyhydroperoxide formation, J. Phys. Chem. A., 116, 6253-63,
1173	https://doi.org/10.1021/jp211528d, 2012.
1174	Zhao, R., Mungall, E. L., Lee, A. K. Y., Aljawhary, D., and Abbatt, J. P. D.: Aqueous-phase
1175	photooxidation of levoglucosan-a mechanistic study using aerosol time of flight chemical
1176	ionization mass spectrometry (Aerosol ToF-CIMS), Atmos. Chem. Phys., 14, 9695-9706,
1177	https://doi.org/10.5194/acpd-14-8819-2014, 2014.
1178	Zhao, R., Lee, A.K.Y., Huang, L., Li, X., Yang, F., and Abbat, J.P.D. Photochemical processing of aqueous and the processing of the proc
1179	atmospheric brown carbon, Atmos. Chem. Phys., 15, 6087-6100, https://doi.org/10.5194/acpd-
1180	15-2957-2015, 2015.
1181	Zhou, Z., Chen, B., Qu, X., Fu, H., and Zhu, D.: Dissolved black carbon as an efficient sensitizer in the
1182	photochemical transformation of 17β -estradiol in aqueous solution, Environ. Sci. Technol., 52,
1183	10391-10399, https://doi.org/10.1021/acs.est.8b01928, 2018.