1	Optical, and chemical properties and oxidative potential of aqueous-
2	phase products from OH and ³ C [*] -initiated photolysisphotooxidation of
3	eugenol
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16	Abstract: Aqueous reactions may turn precursors into light-absorbing and toxic
17	products, leading to air quality deterioration and adverse health effects , etc . In this study,
18	we investigated comprehensively eugenol photooxidation (a representative biomass
19	burning emitted, highly substituted phenolic compound) in bulk aqueous phase with
20	direct photolysis, hydroxyl radical (OH) and an organic triplet excited state (3C*).
21	Results show that the degradation rates of eugenol followed the order of
22	$^{3}C^{*}$ >OH >direct photolysis. Quenching experiments verified that $^{3}C^{*}$ indeed played a
23	dominant role in During ³ C* initiated oxidation, while different reactive oxygen

24	species (ROS) including ³ C [*] , OH, ¹ O ₂ and O ₂ , generated can participate in oxidation of
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25 <u>eugenol, quenching experiments verified ³C*</u> was the most important forone; while

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26	during OH-initiated oxidation. Photolysis, O2 - was a more important ROS than OH to
27	oxidize eugenol. The rate constants under saturated O2, air and N2 followed the order
28	of $k_{O_2} > k_{Air} > k_{N_2}$ for both direct photolysis and OH-initiated oxidation, but changed to
29	$k_{Air} > k_{N_2} > k_{O_2}$ for ${}^{3}C^{*}$ -mediated oxidation. pH and dissolved oxygen (DO) levels both
30	decreased during oxidation, indicating formation of acids and the participation of DO
31	in oxidation. UV-vis light absorption spectra of the reaction products showed clear
32	absorbance enhancement in the 300-400 nm range for all three easessets of experiments
33	and new fluorescence at excitation/emission=250/(400-500) nm appeared, suggesting
34	the formation of new chromophores and fluorophores (brown carbon species); and these
35	species were likely attributed to humic-like substances (HULIS) as shown by the
36	increases of HULIS concentrations during oxidation. Large mass yields of products
37	(140%-197%) after 23 hours of illumination were obtained, and high oxidation degrees
38	of these products were also observed; correspondingly, a series of oxygenated
39	compounds were identified, and detailed reaction mechanism with functionalization as
40	a dominant pathway was proposed. At last, dithiothreitol (DTT) assay was applied to
41	assess oxidation potential of the reaction products, and the end products inof all three
42	photolysis conditionssets of experiments showed higher DDT consumption rates than
43	that of the precursoreugenol, indicating more toxic species were produced upon
44	aqueous oxidation. Overall, our results by using eugenol as a model compound,
45	underscore the potential importance of aqueous processing of biomass burning
46	emissions in secondary organic aerosol (SOA) formation, as well as its impacts on
47	particulate matter concentration and toxicity, radiative balance and climate change
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1 Introduction

50 Photochemical reactions in atmospheric aqueous phases (cloud/fog droplets and 51 aerosol water) can affect lifetimes of many organic species, and are an important source and pathway of secondary organic aerosol (SOA) formation (Vione et al., 2006; Zhao 52 et al., 2012). Compared to the gasSOA formed via gas-phase photochemical oxidation, 53 54 aqueous-phase SOA (aqSOA) is often more oxidized and less volatile, therefore might play an important role in haze formation, air quality and global climate change (Ervens 55 et al., 2011; Lim et al., 2010). However, due to complexity of the aqueous reactions and 56 influencing factors (such as precursors, oxidants, and light intensities), detailed reaction 57 mechanism, optical property, oxidative potential (OP) and the interplay among them 58 remain poorly understood. 59

60 Many laboratory studies have focused on aqueous-phase oxidations of low molecular weight (LMW) volatile organic compounds (VOCs), such as isoprene, 61 terpenes (α -, β -pinene), as well as their gas-phase oxidation products (such as glyoxal, 62 63 methylglyoxal, cis-pinonic acid and methyl vinyl ketone) (Faust et al., 2017; Herrmann, 2003; Herrmann et al., 2015; Huang et al., 2011; Lee et al., 2012; Zhang et al., 2010). 64 Recently, aqueous oxidation of semi-/intermediate volatility VOCs (S/IVOCs), such as 65 66 the phenolic compounds emitted from combustion or pyrolysis of lignin in biomass, were also extensively investigated (Barzaghi and Herrmann, 2002; Bonin et al., 2007; 67 Chen et al., 2020; Gilardoni et al., 2016; He et al., 2019; Jiang et al., 2021; Li et al., 68 69 2014; Li et al, 2021; Ma et al., 2021; Mabato et al., 2022; Smith et al., 2014; Sun et al., 70 2010; Tang et al., 2020; Yang et al., 2021; Yu et al., 2016). Generally, chemical structures of precursors have profound influences on the reaction mechanisms and 71 72 products, while effect of oxidants also cannot be neglected. It is evident that liquid water can contain various types of oxidants, such as singlet oxygen (1O2), nitrate radical (NO3), 73 hydroxyl radical (OH), and organic triplet excited states (³C^{*}), and all can play crucial 74

75 roles in photooxidation reactions (Kaur and Anastasio, 2018; Scharko et al., 2014). Among them, OH is a ubiquitous oxidant with concentrations of 10⁻¹³-10⁻¹² mol·L⁻¹ 76 (Arakaki et al., 2013; Gligorovski et al., 2015; Herrmann et al., 2003). Hence, aqueous 77 78 OH-induced photooxidation has been extensively studied (Chen et al., 2020; Sun et al., 2010; Yu et al., 2016). Compared to OH oxidation, ³C*-initiated aqueous oxidation 79 (photosensitized reactionreactions) has also attracted attentions in recent years (Ma et 80 al., 2021; Wang et al., 2021). Several classes of organic compounds in ambient air, 81 including non-phenolic aromatic carbonyls, quinones, aromatic ketones and nitrogen-82 containing heterocyclic compounds, can form ³C* after absorbing light (Alegría et al., 83 1999; Kaur et al., 2019; Nau and Scaiano, 1996; Rossignol et al., 2014; Chen et al., 84 2018). These compounds are termed as photosensitizers. ${}^{3}C^{*}$ is capable of reacting with 85 O₂ to produce singlet oxygen (¹O₂) and superoxide radicals (O₂. Various reactive 86 oxygen species (ROS) can be generated and affect greatly the ³C*-initiated aqueous-87 88 phase reactions. Despite some studies demonstrating importance of ROS in photochemical process (Ma et al, 2021; Wang et al., 2020; Wang et al., 2021; Wu et al., 89 $\frac{2021}{1000}$, our current understanding on ${}^{3}C^{*}$ -initiated oxidation is still limited. 90

91 Excitation-emission matrix (EEM) fluorescence spectroscopy, as a low-cost, rapid, 92 non-destructive and high-sensitivity technique, can offer detailed information on chromophores hence has been widely employed for studies of aquatic dissolved organic 93 94 matter (Aryal et al., 2015). Nevertheless, it has not been extensively used in 95 atmospheric aerosol research (Mladenov et al., 2011). Prior studies have investigated the relationship between the fluorescence components and chemical structures of 96 atmospheric aerosols by using high-resolution aerosol mass spectrometry (AMS) and 97 EEM fluorescence spectroscopy (Chen et al., 2016a; Chen et al., 2016b). An earlier 98 report from Chang and Thompson (2010) found fluorescence spectra of aqueous-phase 99

100 reaction products of phenolic compounds, which had some similarities with those of 101 humic-like substances (HULIS), and Tang et al. (2020) reported that aqueous photooxidation of vanillic acid could be a potential source of HULIS. Chang and 102 103 Thompson (2010) also showed that light-absorbing and fluorescent substances 104 generally had large conjugated moieties (i.e., quinones, HULIS, polycyclic aromatic hydrocarbons (PAHs)), which can damage human body (Dou et al., 2015; McWhinney 105 et al., 2013). HULIS are considered as an important contributor to induce oxidative 106 107 stress since they can serve as electron carriers to catalyze ROS formation (Dou et al., 2015; Ma et al., 2019; Huo et al., 2021; Xu et al., 2020), causing adverse health effects. 108 Dithiothreitol (DTT) assay (Alam et al., 2013; Verma et al., 2015a2015), as a non-109 110 cellular method, was widely employed to determine oxidation activity and OP of atmospheric PM (Chen et al., 2019; Cho et al., 2005) for the evaluation of its health 111 effects. Some other works (Fang et al., 2016; McWhinney et al., 2013; Verma et al., 112 113 2015; Zhang et al., 2022) focused on the link between chemical 114 compositioncomponents and OP in PM, and confirmed that several kinds of compounds, such as quinones, HULIS and transition metals usually had strong DTT activities. 115 However, DTT method is rarely used to evaluate the OP of aqueous-phase oxidation 116 products previously (Ou et al., 2021). 117

In the present work, we chose eugenol (ally guaiacol) as a model compound to conduct aqueous oxidation experiment. As a representative methoxyphenol emitted from biomass burning (BB) (Hawthorne et al., 1989; Simpson et al., 2005), it was widely detected in atmospheric particles. For instance, concentration and emission factor of this compound from beech wood burning were $0.032 \ \mu g/m^3$ and $1.534 \ \mu g/g$, which were twice those of guaiacol (0.016 $\mu g/m^3$ and 0.762 $\mu g/g$) (Bari et al., 2009). Eugenol is a semivolatile aromatic compound with a moderate water-solubility (2.46 125 g/L at 298 K) too.). Chemical characteristics of aqueous reaction products under direct 126 photolysis (without oxidant) and oxidations by OH radicals and ³C* radicalstriplet 127 states, were comprehensively elucidated by a suite of analytical techniques including 128 high-performance liquid chromatography (HPLC), ultraviolet and visible (UV-Vis) 129 spectrophotometry, gas chromatography mass spectrometry (GC-MS), EEM-and soot particle aerosol mass spectrometry (SP-AMS). The relative importance of various ROS 130 131 species to eugenol degradation was explored. Moreover, light absorption, fluorescent and oxidative properties of the aqueous oxidation products were also investigated. 132

133 2 Materials and methods

134 2.1 Chemicals and reagents

Eugenol (99%), tert-butanol (TBA, 99%), 3,4-dimethoxybenzaldehyde (DMB, 135 99%), para-benzoquinone (p-BQ, 99%), dithiothreitol (99%) and 5,5'-dithiobis-2-136 nitrobenzoic acid (DTNB, 99%), 2-nitro-5-thiobenzoic (99%), 5,5-dimethyl-1-137 pyrroline N-oxide (DMPO), 2,2,6,6-tetramethylpiperidine (TEMP) were all purchased 138 from Sigma-Aldrich. Superoxide dismutase (SOD) was purchased from Bovine 139 140 Erythrocytes BioChemika. Dichloromethane (HPLC-MS grade, 99%), methanol (HPLC-MS grade, 99%), acetonitrile (HPLC-MS grade, 98%), hydrogen peroxide 141 (H2O2, 35 wt %), and 2,4,6-trimethylphenol (TMP, 99%) were all obtained from 142 Acors Acros Chemicals. Sodium azide (NaN3, 98%) was purchased from J&K Scientific 143 Ltd. (Beijing, China). All solutions were prepared using ultrapure water (Millipore) on 144 145 the days of experiments.

146 **2.2 Photochemical oxidation experiments**

147 Aqueous-phase photochemical reactions were carried out in a Rayonet 148 photoreactor (model RPR-200) equipped with 16 light tubes (2 RPR-3000, 7 RPR-3500 and 7 RPR-4190 tubes), which was frequently used to mimic sunlight for 149 photochemical experiments and was described in details by several groups (George et 150 al., 2015; Hong et al., 2015; Huang et al., 2018; Jiang et al., 2021; Zhao et al., 2014). 151 Pyrex tubes containing sample solutions were placed in the center and received 152 radiation from surrounded lamps of all sides. To ensure mixing of the solution, a fan 153 154 and a magnetic stir bar were placed at the bottom of the reaction tube. The solution 155 temperature was controlled at 25±2°C. The same photoreactor system and a normalized distribution of photon fluxes inside the reactor have been reported elsewhere (George 156 157 et al., 2015), and the wavelength of light was in the range of 280~500 nm. We only 158 measured light intensity at the surface of the solution with a radiometer (Photoelectric instrument factory of Everfine Corporation, Hangzhou, China), which was determined 159 160 to be ~2400 μ W/cm² in the range of 290-320 nm (UVB), lower than the sunlight intensity (6257.1 µW/cm²). 161

In this work, 300 μ M H₂O₂ and 15 μ M DMB were added into solutions as sources 162 163 of OH and ${}^{3}C^{*}$ -radicals,*, respectively. The initial concentration of eugenol was 300 μ M. For ³C*-mediated experiments, solutions were adjusted to pH=3 by sulfuric acid in 164 order to perform experiments under optimal conditions (Ma et al., 2021; Smith et al., 165 166 2014) since DMB triplet state is protonated to a more reactive form in acidic solution. 167 We conducted three sets of photolysisoxidation experiments: (A) 300 µM eugenol + 300 µM H₂O₂, (B) 300 µM eugenol + 15 µM DMB, and (C) 300 µM eugenol without 168 oxidants. In each series of experiments, a dark control experiment was performed 169 synchronously with a Pyrex tube wrapped by the aluminum foil. Results showed loss 170 of eugenol under dark conditions were negligible (data not shown). In addition, to 171

172 evaluate the roles of ROS in eugenol degradation during 3C*-initiated oxidation, 173 quenching experiments by using specific scavengers to capture different ROS were performed, namely TBA for OH, NaN3 for 1O2, SOD for O2*, and TMP for 3C*, 174 respectively (Pan et al., 2020; WuChen et al., 20212020). For OH-initiated oxidation, 175 quenching experiments using p-BQ for O2. (Ma et al., 2019; Raja et al., 2005), and 176 TBA for OH were conducted. For most experiments, solutions were saturated by air 177 and each experiment presented was repeated three times unless otherwise stated. 178 Average results with one standard deviation were provided. In order to further evaluate 179 the role of oxygen in photooxidation, experiments were also conducted by using 180 different saturated gases (air, N2 and O2). 181

182 2.3 Analytical methods

183 **2.3.1 Determination of eugenol concentrations**

184 Before and during the photochemical experiment, 2 mL of reacted solution was sampled periodically and subjected to HPLC (LC-10AT, Shimadzu, Japan) analysis to 185 quantify eugenol concentration. The HPLC was equipped with an InertSustain AQ-C18 186 187 reverse phase column (4.6×250 mm, 5.0 µm, Shimadzu) and a UV-vis detector. The mobile phase was a mixture of acetonitrile/H2O (v/v: 60/40) at a flow rate of 0.6 188 mL/min, and the detection wavelength was 280 nm. The first-order kinetic rate constant 189 of eugenol degradation can be obtained from the slope of plot of $-\ln(c_1/c_0)$ versus 190 reaction time as presented in Eq.(1). 191

$$\ln(c_t/c_0) = -kt \tag{1}$$

193 Where c_0 and c_t are eugenol concentrations (in μ M) at the initial and reaction time 194 t, while k represents the pseudo first-order rate constant, (in s⁻¹).

195 2.3.2 UV-vis and fluorescent spectra

The UV-vis light absorbance spectra of reacted solutions (placed in a 1 cm path length quartz cuvette) were measured by using an UV-vis spectrophotometer (Specord 210 plus, Analytik Jena., Germany). The instrument has a dual-beam optical system with tungsten and deuterium lamps as light sources. A reference absorption spectrum of ultrapure water was carried out in the same cuvette prior to sample analysis for baseline correction.

Immediately after the UV-Vis measurement, the cuvette was transferred to a threedimensional EEM fluorescence spectrometer (FluoroMax Plus, HORIBA Scientific). The ranges of wavelength varied from 200 to 450 nm for excitation wavelengths (Ex) and from 290 to 650 nm for emission wavelength (Em). Intervals of the excitation and emission wavelengths were 5 nm and 2 nm, respectively. The reported absorbance and EEM spectra here are averages of the results from experiments in triplicate.

208 2.3.3 Determination of HULIS concentrations

209 Solid phase extraction (SPE) cartridges (CNW Poly-Sery HLB, 60mg60 210 mg/cartridge) were used to isolate HULIS from the reaction products. The SPE cartridge was first rinsed with 1 mL ultrapure water and 3 mL methanol prior to 211 212 extraction. The solution was acidified to pH ~2 using HCl and loaded on an SPE cartridge, which was rinsed with 1 mL ultrapure water again. Next, 3 mL 213 methanol/ammonia (98:2, v/v) mixture was added into the SPE cartridge to elute 214 215 HULIS, and the solution was blewblown to full dryness with high purity N2, followed 216 by dilution with ultrapure water to 25 mL for quantification of HULIS using the HPLC coupled with an evaporative light scattering detector (ELSD3000). Recovery efficiency 217

of the HULIS standard, Suwanne River Fulvic Acid (SRFA), was 75-80% with the
standard deviation of reproducibility less than 5%. More details have been described
elsewhere (Tao et al., 2021).

221 2.3.4 Oxidative potentials (OPspotential (OP) based on DTT assay

222 The OPSOP of reaction products werewas determined by the DTT method (Cho et 223 al., 2005; Lin and Yu, 2019) with slight improvements. Briefly, 1.2 mL sample solution was transferred into a 10 mL glass tube, then 6 mL phosphate buffer (0.1 M, pH 7.4) 224 and 300 µL of 2.5 mM DTT were added and mixed thoroughly. The DTT mixed 225 226 solution was placed in a 37°C water bath for incubation. Over the course of reactions 227 that lasted for 150 minutes, 1 mL aliquot of DTT mixture was taken every 30 minutes, and 100 µL of 5 mM DTNB (prepared in 0.1 mM phosphate buffer) was added and 228 229 loaded in a centrifuge tube. Next, reactions between DTNB and DTT produced bright yellow TNB, which was quantified by the UV-Vis spectrometer within 30 minutes. 230 Finally, we measured the light absorbance (At) at 412 nm at time t to indirectly quantify 231 232 the remaining DTT. Another 1.2 mL ultrapure water instead of sample solution was 233 treated in the same way and the absorbance was denoted as A as the blank value. A0 represents the initial light absorbance value. Thus, DTT concentration consumed by the 234 235 sample solution (MDTT, µM) and that by the blank solution (MDTT0, µM) can be calculated according to Eq.(2) and Eq.(3), respectively. 236

$$M_{DTT} = \frac{A_0 - A_t}{A_0} \times C_{DTT_0}$$
⁽²⁾

238
$$M_{DTT0} = \frac{A_0 - A}{A_0} \times C_{DTT_0}$$
 (3)

Here, C_{DTT0} was the initial DTT concentration in sample solution (100 μ M in this work). DTT consumption rates (R_{DTT} and R_{DTT0}) were then obtained from the slopes of plots of M_{DTT} and M_{DTT0} versus incubation times. Experiments of blanks and samples were typically run in a triplicate. The reproducibility of the whole analysis showed thatthe relative standard deviation of DTT consumption rate was 3-4%.

244 2.3.5 ProductsProduct analysis by GC-MS

Reacted solution (about 30 mL) was extracted with 10 mL dichloromethane twice. 245 246 The extract was concentrated into 1 mL by blowing N2 gently, subsequently transferred 247 to a 2 mL vial, and analyzed by a GC-MS (7890A GC/5975C MS, Agilent) with a DB-5ms capillary column (30 m×0.25 mm×0.5 µm). The operational conditions were set as 248 follows: injector was at 200°C; ion source was at 230 °C; column oven temperature was 249 250 programmed to be held at 35°C for 4 minutes, then ramped to 250 °C at a rate of 20°C/minute and held for 10 minutes. The recovery efficiency, method detection limits 251 and quality assurance/quality control have been described in our previous work (Ye et 252 253 al., 2020).

254 2.3.6 SP-AMS analysis and mass yields of reaction products

An Aerodyne SP-AMS (Onasch et al., 2012) was applied to analyze the lowvolatility organic products, similar to our previous work (Chen et al., 2020; Ge et al., 2017). SP-AMS data were acquired in V mode and analyzed by Squirrel v.1.56D and Pika v1.15D software. The organic fragments were classified into six groups: CH, CHO, CHN, CHO₂, CHON and HO. Elemental ratios (oxygen-to-carbon, O/C; hydrogento-carbon, H/C), were calculated according to the method proposed by Canagaratna et al. (2015).

Since the AMS analysis requires nebulization of sample solution into particles before determination, and quantification of organics was influenced by the atomization efficiency and carrier gas flow, etc., we thus cannot use SP-AMS measured concentration to quantify the mass of products directly. In this case, according to Li et al. (2014), we added an internal standard (SO4²⁻) prior to AMS analysis, and the mass ratio of particle-phase organics to SO4²⁻ (Δ Org/SO4²⁻) can be used to calculate the mass concentration of products. Furthermore, the mass yield of aqueous-oxidation products (Y_{products}, %), which is the mass of products generated per unit mass of precursor consumed, can be calculated <u>asaccording to</u> Eq. (4).

271
$$Y_{\text{products}}(\%) = \frac{(\Delta \text{Org}/So_4^{2-})[so_4^{2-}]_0}{c_0 M_{\Pi}} \times 100\%$$
(4)

Where $[SO_4^{2-}]_0$ is the SO_4^{2-} concentration (here 7.27 mg+/L⁻⁴), C₀ is the initial eugenol concentration (in mmol/L), M is MW of the precursor (164 g/mol for eugenol), and η is the degraded fraction of eugenol.

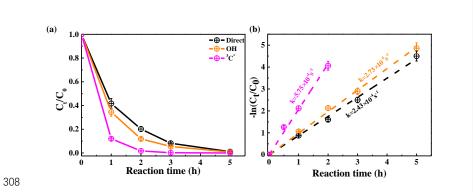
275 **3 Results and discussion**

276 **3.1 Kinetics of aqueous photooxidation**

Figure 1 shows unreacted eugenol concentrations (ct) and the negative logarithm 277 278 of c_t/c_0 (-ln(c_t/c_0)) as a function of reaction time, respectively. The pseudo first-order rate constants (k) obtained by Eq.(1) were also presented. As described in Fig. 1a, 279 eugenol concentration decreased to be <20% of the initial concentration in 3 hours, 280 281 suggesting photolysisphotooxidation was fast under all three reaction conditions. In the presence of ³C^{*}, eugenol was degraded nearly 100% after 3 hours. Previous study (Chen 282 et al., 2020) on ³C*-initiated 4-ethylguaiacol oxidation reports a time of 21 hours for a 283 complete degradation. Apart from difference of precursors, different light irradiation 284 spectra and stronger energy of light in this work than the previous work might be 285 responsible for the fast loss of eugenol. The bond dissociation energies (BDEs) are 340 286 kJ/mol for OH, 374 kJ/mol for C-H in -CH3 group, 345 kJ/mol for C=C bond, and 403 287

288 kJ/mol for C-H in -OCH3 group, respectively (Herrmann et al., 2003; He et al., 2019). The lowest BDE was found for the O-H bond and C-C bond. Due to influences of steric 289 290 hindrance and intramolecular hydrogen bonding, the H-abstraction from OH group might not be favorable and the most probable H-abstraction might take place in C=C 291 of the allyl group. As a result, breakage of C=C into C-C at the allyl group can lead to 292 the formation of 2-methoxy-4-propyl-phenol (Section 3.6.1). When photon energy is 293 higher than the BDE, chemical bonds can break, leading to decomposition of 294 compounds and possibly further mineralization. The energiesenergy of photons 295 atphoton of 300 and 350-nm areis 412 kJ/mol, and and can break all major bonds in 296 eugenol, while the energy of 350 nm is 353 kJ/mol, higher thanbeing able to break some 297 298 of the weakest BDE bonds in eugenol, therefore it as well. Overall, eugenol can be easily 299 decomposed after absorbing the photonphotons.

300 As shown in Fig. 1b, the first-order rate constants were $2.43 \times 10^{-4} \text{ s}^{-1}$, $2.73 \times 10^{-4} \text{ s}^{-1}$ ¹, and 5.75×10^{-4} s⁻¹ for direct photolysis and photooxidations by OH and ${}^{3}C^{*}$, 301 respectively. ³C*-initiated photooxidation was quicker than that attacked by OH, likely 302 due to combined contributions from reactions with ¹O₂, O₂⁻⁻ and OH (Section 3.2). 303 Similar results were found for aqueous phase reactions of three phenols against OH and 304 ${}^{3}C^{*}$ by Yu et al. (2016) (Note the initial concentrations of H₂O₂ and DMB were 100 μ M 305 and 5 μ M, respectively, with the same ratio as 300 μ M H2O2 to 15 μ M DMB in this 306 307 work)



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309 Figure 1. Aqueous-phase eugenol decay kinetic curves (a) and regressed first-order rate constants*

310 (b) obtained based on Equation 1 under three conditionsdirect photolysis, OH-initiated oxidation

311 and ³C*-initiated oxidation. Error bar represents one standard deviation from the measurements in

312 triplicate.

313 **3.2 Relative importance of ROS in photooxidation**

314 **3.2.1** Quenching experiments in ³C^{*}-initiated photooxidation

315 Relative importance of different ROS in photooxidation can be investigated by 316 addition of radical-scavengers/quenchers, and then be evaluated based on the different degradation efficiencies of eugenol in absence and presence of the corresponding ROS 317 318 quenchers. For each quencher, we conducted several gradient experiments with varying 319 molar ratios of eugenol to quencher. The ratios were 0.075:1, 0.15:1, 0.3:1, 0.75:1, 1.5:1 320 for quenchers of NaN₃, TMP and TBA, and 1.2:1, 1.6:1, 2.5:1, 5:1, 10:1 for SOD, which 321 were all within the typical ranges of ROS quenching experiments reported previously (Zhou et al., 2018). Excess concentrations of quenchers have been added repeatedly to 322 ensure the complete reactions between radicalsROS and scavengers. Figure 2 displays 323 324 the impacts of quenchers on eugenol degradation. All rate constants (k) with quenchers 325 were lower than those of the quencher-free solutions. The optimum molar ratio of 326 eugenol to quencher was chosen whenselected such that the inhibition degree of 14

327	eugenol degradation unchangeddid not change with the increase of added quencher
328	(Wang et al., 2021). For example, along with the decrease of molar ratios of eugenol to
329	NaN ₃ from 1.5:1 to 0.075:1, the inhibitory degreevariation of eugenol degradation was
330	stabilized at the ratio of 0.15:1, indicating that $^1\mathrm{O}_2$ has been completely quenched at
331	this ratio, therefore a molar ratio of 0.15:1 for NaN3 was optimal, since excess
332	scavenger may generate other products that interfere the existing reactions.
333	SimilarlyFinally, the optimal molar ratios of eugenol to quenchers of TBA, NaN ₃ , TMP
334	and SOD, were determined to be 1.5, 0.15, 0.075 and 2.5, respectively. Table 1 and Fig.
335	S1 compared the rate constants determined under the ratios above and they were in an
336	order of TMP <nan3<sod<tba, generated="" importance="" of="" relative="" ros="" suggesting="" td="" to<=""></nan3<sod<tba,>
337	eugenol degradation was in the order of ${}^{3}C^{*} > {}^{1}O_{2} > O_{2}^{\cdot} > OH$. This result suggests that
338	${}^{3}C^{*}$ plays a major role <u>among all ROS</u> in the photooxidation. Previously, Laurentiis et
339	al. (2013) reported that 4-carboxybenzophenone (70 $\mu M)$ could act as $^3C^*$ and the
340	photosensitized degradation was more effective than oxidants such as OH, O ₃ , etc.;:
341	Misovich et al. (2021) investigated the aqueous DMB-photosensitized reaction (5 μ M,
342	same as $\frac{1}{10}$ in this study) also demonstrated that ${}^{3}C^{*}$ was the greatest contributor to phenol
343	or guaiacyl acetone degradation, followed by ${}^{1}O_{2}$, while both OH and ${}^{1}O_{2}$ contributions
344	were relatively minor.
345	We To further assess the relative importance of different ROS, we propose to use
346	the following Eq.(5) to roughly assess the contribution of for a certain ROS (Ctros) to
347	eugenol degradationrough estimation:
348	$\frac{\text{Ct}_{\text{ROS}}=k_{\text{ROS}}/k_{\text{Ct}}(\underline{\text{Red}_{\text{ROS}}(\underline{\text{in \%}})=(\underline{\text{k}}-\underline{\text{k}}_{\text{quencher}})/k_{\text{Ct}}}{\frac{*100\%}{2}}$
349	(5)
350	Here knosk (in s ⁻¹) is the rate constant contributed by the ROS, which is defined as the
351	difference between the original rate constant inof 3C*-initiated oxidation (kor OH-
I	15

initiated oxidation in Section 3.2.2) and $\underline{k_{quencher}}$ (in s⁻¹) is the rate constant –after the target ROS has been completely scavenged by its corresponding quencher ($\underline{k_{quencher}}$). k and $\underline{k_{quencher}}$ in fact refer to those reported in Fig. S1b. <u>Red_{ROS} then refers to the</u> percentages of reduction due to addition of quencher for a ROS.

356 According to Eq.(5), CtacRedac* was calculated to be 0.857, therefore contribution of ³C* was estimated to be as high as 85.7%. Similarly, the contributions values of ¹O₂, 357 O2⁻ and OH were 80.5%, 61.4% and 53.9%, respectively. The total contribution of the 358 four ROS largely exceeded 100%. This Note RedRos only reflects the relative important 359 of ROS and it does not corresponds to the exact contribution of that ROS in eugenol 360 degradation without quenchers. The reason is that although the addition of a ROS 361 362 scavenger can be explained eliminate oxidation by the fact that this ROS-scavengers can actually, but it also significantly interruptinterrupts the radicaloriginal chain reactions 363 364 as compared to those in the absence of scavengers. For instance, addition of TMP not 365 only scavenges ³C^{*}, but also inhibits generation of ¹O₂, O₂⁻, etc. These findings suggest 366 that we cannot directly precisely quantify the contribution of a ROS just on the basis of 367 its scavenging efficiency, therefore the contributions calculated from Eq.(5) can only 368 be used to compare the relative importance of different ROS. Onethe scavenger, and reactions with other ROS might be enhanced. In this regard, the sum of the four RedROS 369 values may exceed 100%. Therefore, one should be cautious to apply quenching 370 371 approach to quantify the role of ROS in complex reaction systemuse RedRos as a precise quantification of the ROS contribution in aqueous oxidation. Determination of ROS 372 concentrations during oxidation should be instead be an effective way to elucidate the 373 374 role of ROS. Therefore Here, we tried to detect in-situ generated OH, O2⁻ and ¹O2 during photochemical reactions using a micro electron spin resonance (ESR) spectrometer 375 (Bruker Magnettech, Berlin, Germany) with DMPO as the spin trap to form stable 376

377 DMPO-OH or DMPO-O2[•], with TEMP to capture ¹O₂ to produce TEMP-¹O₂ spinadduct (TEMPO). The radicals can be identified and quantified by the peak patterns in 378 ESR spectra, such as the quarter line with a height ratio of 1:2:2:1 for DMPO-OH, 379 1:1:1:1 for DMPO-O2⁻ and 1:1:1 for TEMP-¹O₂ (Guo et al., 2021). Unfortunately, OH 380 radical cannot be detected since its concentrationsconcentration might be lower than 381 382 the detection limit of the instrument (Fig. S2, ESR spectra of OH). In contrast, we were able to detect higher high concentrations of ³C^{*} and found the intensity of TEMP-¹O₂ 383 signal reached its maximum at 30 minutes, then decreased slowly (Fig. S2, ESR spectra 384 of ¹O₂). Combining the greatest inhibitive great quenching effect of TMP with high ¹O₂ 385 concentration from ESR method, we can conclude that ³C* and ¹O₂ play relatively 386 387 important roles in eugenol photooxidation.

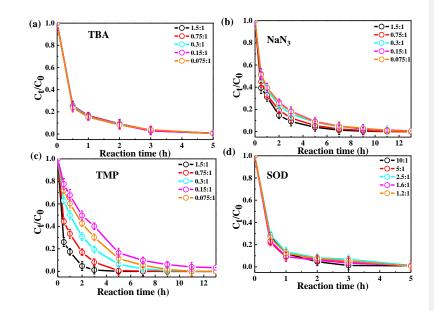


Figure 2. Ratio of unreacted eugenol concentration to its initial concentration (Ct/C0) at different
molar ratios of eugenol to quencher, as a function of reaction time: (a) TBA, (b) NaN3, (c) TMP and
(d) SOD.

393 3.2.2 Quenching experiments in OH-initiated photooxidation

394 To examine the contributions of ROS to eugenol degradation for OH-initiated oxidation, TBA and p-BQ as trapping agents were added. Similar to ³C*-initiated 395 oxidation, several gradient experiments with varying molar ratios of eugenol to 396 quenchers were conducted. The ratios were set as 6.5:1, 3.2:1, 1.6:1, 1.1:1 and 0.8:1 for 397 p-BQ and 3.0:1, 1.5:1, 0.75:1, 0.3:1 and 0.15:1 for TBA. According to Fig. S3, molar 398 ratio only had a slight influence on eugenol degradation, although degradation can be 399 inhibited effectively by quenchers. Thus, we determined the appropriate molar ratios of 400 0.8 and 0.75 for p-BQ and TBA, respectively, as excess scavengers might influence the 401 chemical reactions. 402

Variations of the rate constants for the aforementioned quenching experiments were 403 determined, in comparison with those conducted without quenchers, and the results are 404 listed in Table 1 and presented in Fig. S4. For TBA quenching tests, the rate constant 405 406 decreased by 18.7% (from 2.73×10⁻⁴ s⁻¹ to 2.22×10⁻⁴ s⁻¹), showing that OH radical played a certain role in eugenol photooxidation. Since H2O2 was mainly photolyzed at 407 wavelength <300 nm to generate OH radical, irradiation above 300 nm did not affect 408 the reaction significantly. The p-BQ could quench O2, further suppressing the 409 generation of other ROS (e.g., •HO2), therefore the rate constant decreased the most 410 (from 2.73×10⁻⁴ s⁻¹ to 1.20×10⁻⁴ s⁻¹), suggesting O2⁻⁻ was important for eugenol 411 photooxidation. This hypothesis could be further confirmed by the decline of rate 412 constant under N2-saturated solution (Section 3.2.3). However, it was difficult to detect 413 both OH and O2 directly due to their relatively short lifetimes and low concentrations 414 415 via ESR in this work.

417 **Table 1.** The first-order rate constants of eugenol in the presence of various scavengers. The initial

418 conditions were as follows: 300 µM eugenol; molar ratios of eugenol to quenchers TBA, NaN₃,

419 TMP and SOD, of 1.5, 0.15, 0.075 and 2.5, respectively; molar ratios of eugenol to quenchers *p*-BQ

420 and TBA of 0.8 and 0.75, respectively.

Quenchers	ROS	Reaction rate constant k (s^{-1})	Pearson's R ²	
no quencher	-	5.75×10 ⁻⁴	0.996	
TBA	OH	2.65×10^{-4}	0.999	
SOD	O2 ^{'-}	2.22×10^{-4}	0.995	
NaN ₃	$^{1}O_{2}$	1.12×10^{-4}	0.999	
TMP	³ C*	0.82×10^{-4}	0.999	
	OH-initi	ated quenching experiments		
Quenchers	ROS	Reaction rate constant k (s ⁻¹)	R ²	
No quencher	-	2.73×10 ⁻⁴	0.995	
TBA	OH	2.22×10^{-4}	0.998	
p-BQ	O_{2}^{-}	1.20×10^{-4}	0.995	

421

422 3.2.3 Influences of different saturated gases

423 In order to assess the role of O₂ in eugenol photolysisdegradation, a series of experiments were performed under both O2-saturated and N2-saturated conditions in 424 425 addition to air. N2 gas was purged into reaction solution for ~30 minutes before 426 experiment to achieve the O₂-free condition. Figure 3 compared the changes of eugenol 427 concentrations and rate constants (see insets) under the three gas conditions for direct photolysis, OH-initiated and 3C*-initiated oxidations, respectively. The rate constants 428 followed the order of $k_{D_2} > k_{Air} > k_{N_2}$ under both direct photolysis and OH oxidation, 429 providing evidence in support of O2 being significant for eugenol degradation. This 430 might be explained by the fact that O₂ can act as an electron acceptor to generate O2. 431 and •HO₂, and subsequently form H₂O₂ and OH. For direct photolysis, rate constant 432 under O2-saturated condition increased 14.4% while it decreased 19.3% under N2 433 saturation from that under saturated air. For OH-initiated oxidation, the difference of 434 19

435 rate constants under three saturated gases became more distinct.

On the contrary, rate constants followed the order of $k_{\rm Air}$ > $k_{\rm N_2}$ > $k_{\rm O_2}$ in ${}^3C^*\text{-}$ 436 initiated oxidation. There are two possible explanations. On one hand, under N2-437 saturated condition without oxygen, DMB would involve in reactions (R1-R4), leading 438 to a more effective generation of ³DMB* therefore a higher degradation efficiency than 439 under O2-saturated condition. On the other hand, for air/O2-saturated solutions, 440 irradiation of DMB and eugenol would involve also reactions (R5-R8) in addition to 441 442 (R1-R4), and as a result, the amount of ³DMB* radical-decreased, due to formation of other ROS (1O2, O2-, OH, etc) with relatively weak oxidative capacities. In summary, 443 quenching of ³DMB* by ground state molecular oxygen could account for the low 444 degradation efficiency in O2-saturated condition. 445

446	$DMB + h\nu \rightarrow {}^{1}DMB^{*} \rightarrow {}^{3}DMB^{*}$	(R1)	
447	$^{3}\text{DMB}^{*} \rightarrow \text{DMB}$	(R2)	
448	$^{3}\text{DMB}^{*} \rightarrow \text{Products}$	(R3)	
449	$^{3}\text{DMB}^{*} + ^{1}\text{DMB}^{*} \rightarrow \text{DMB}^{*+/-}(\text{DMB}^{*+} + \text{DMB}^{*-})$	(R4)	
450	$^{3}\text{DMB}^{*} + \text{O}_{2} \rightarrow \text{DMB} + {}^{1}\text{O}_{2}$	(R5)	
451	$DMB^{-} + O_2 \rightarrow DOM^{+} + O_2^{-}$	(R6)	
452	$O_2 - + 2H^+ \rightarrow H_2O_2 + O_2$	(R7)	
453	$H_2O_2 \rightarrow 2OH$	_(R8) •	带格式的: 左
454			

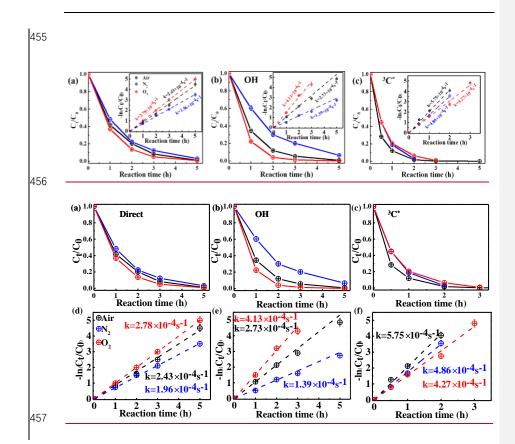
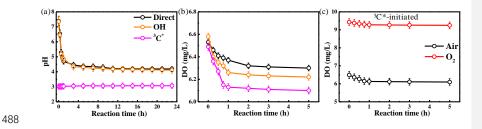


Figure 3. Ratio of unreacted eugenol concentration to its initial concentration (C_t/C_0) as a function of reaction time at different saturated gases under (a) direct photolysis (b) OH-initiated oxidation and (c) ${}^{3}C^{*}$ -initiated oxidation. The insets in (a c) show the corresponding rate<u>Rate</u> constants<u>- of (a-</u> <u>c) are presented in (d-f) correspondingly.</u>

462 3.2.4 Variations of pH and dissolved oxygen

The initial pH values of reaction solutions for direct photolysis and OH-initiated oxidation were unadjusted, while those that for the ${}^{3}C^{*}$ -oxidation was adjusted to 3. The variation of solution pH is presented in Fig. 4a. The pH values decreased dramatically quickly at the beginning of illumination (from 7.4 to ~5.0 in the first 1 hour) then tended to be flat for both direct photolysis and OH-initiated oxidation. However, little change of pH (less than 0.1 unit) was observed for the ³C*-initiated photooxidation
throughout the oxidation, <u>which can be</u> likely <u>ascribingascribed</u> to its low initial pH of
3. Note a small amount of acids can change solution pH significantly when original pH
is high, but cannot change pH remarkably when<u>Since</u> the original solution pH was low.
Therefore, acidic (pH=3), we cannot rule out formation of acidic products (such as
organic acids) during ³C*-initiated oxidation as during direct photolysis and OHinitiated oxidation.

475 As discussed in Section 3.2.3, oxygen can take part in photochemical reaction to form ROS, which may in turn destroy the structure of precursor. Here we measured the 476 oxygen consumption during oxidation through determination of dissolved oxygen (DO) 477 478 contents by a dissolved oxygen meter (Seven2Go Pro S9, Zurich, Switzerland). DO was consumed mainly at the first 1 hour and remained stable afterwards with the increase 479 480 of reaction time (Figs. 4b-c and Fig. S5). The amounts of consumed DO followed the order of ³C*>OH>direct photolysis. The maximum consumed DO was found in ³C*-481 482 initiated oxidation, which canmight be explained by the transferconsumption of electrons from O2 that reacts with 3C* to O2-to-form 1O2, a major contributor to eugenol 483 degradation. (R5). Obviously, a steady-state DO level was reached when the 484 485 consumption rate was equal to the diffusion of O2 into the solution (Pan et al., 2020). Overall, these results re-emphasize that O2 can influence eugenol degradation and 486 487 radicalchemical transformation via induction of radical chain reactions.



489 **Figure 4.** (a) pH values and (b) dissolved issolved oxygen (DO) contents against reaction time under

490 three<u>direct</u> photolysis-conditions, OH-initiated oxidation, ³C*-initiated oxidation, and (c) DO

491 contents during ³C*-initiated oxidation under air or O₂-saturated conditions.

492

493 **3.3 Optical properties of reaction products**

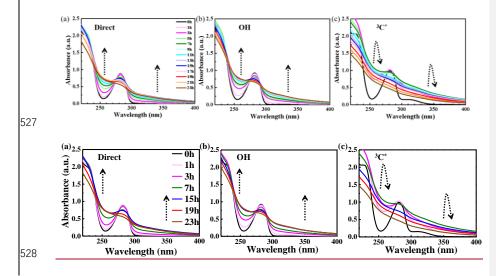
494 **3.3.1 Light-absorbing properties**

495 The UV-vis light absorption spectra of the solutions at different reaction times are presented in Fig. 5. The light absorption by eugenol itself mainly occurs in the range of 496 260-300 nm (n $\rightarrow\pi^*$ electronic transition, 270-350 nm), which overlaps with the major 497 photon fluxes (280 and 500 nm) of our lamp for photooxidations.photooxidation. 498 499 Therefore, we can clearly observe that the characteristic absorption peak at 280 nm of precursor decreased with the propagation of direct photolysis (Fig. 5a), similar to that 500 in OH-initiated photooxidation (Fig. 5b). However, the reaction was quick in the 501 502 presence of ³C*, and the characteristic absorption peak at 280 nm after 3 hours of 503 illumination almost disappeared, suggesting nearly a complete loss of eugenol, consistent with the results in Section 3.1 that more than 99% eugenol was degraded in 504 3 hours. Additionally, there was an obvious absorption enhancement at longer 505 wavelengths (300-400 nm) during the photooxidation, whereas eugenol itself did not 506 absorb light in this range, indicating some light-absorbing products (e.g., brown carbon 507 (BrC) species) were generated. Aqueous photooxidation of some phenolic compounds 508 509 (e.g., vanillic acid) also presented long-wavelength (300-400nm) light absorbance, with intensity increasing with illumination time (Tang et al, 2020; Zhao et al., 2015). In 510 addition, there were some differences for light absorbance at wavelength of 300-400 511 512 nm in the three eases.reaction conditions. For direct photolysis and OH-initiated oxidation, light absorbance increased during the first 15 hours, then remained at a 513 plateau until 23 hours. However, for 3C*-initiated oxidation, light absorbance increased 514 23

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515 during the first 7 hours, then decreased slowly afterwards. The different shapes of UV-516 vis spectra between OH and ${}^{3}C^{*}$ photooxidations indicate formations of different 517 products.

Compared to the light spectrum of eugenol, there were also increases of light 518 absorbance at ~260 nm ($\pi \rightarrow \pi^*$ electronic transitions) upon aqueous 519 520 photolysisoxidation in all three reaction conditions (Fig. 5), demonstrating the generation of new substances likely with both aromatic C=C and carbonyl (C=O) 521 522 functional groups (Tang et al., 2020). The enhancement at 300-400 nm may point to products with high MWs and conjugated structures, possibly linking with HULIS or 523 oligomers. Unfortunately, we were unable to quantify the relative contributions of 524 individual products to the overall light absorbance between 300 to 400 nm due to lack 525 of thea full speciation of the products and their light absorption spectra. 526



529 **Figure 5.** UV–vis light absorption spectra of reacted solutions at different reaction times under (a)

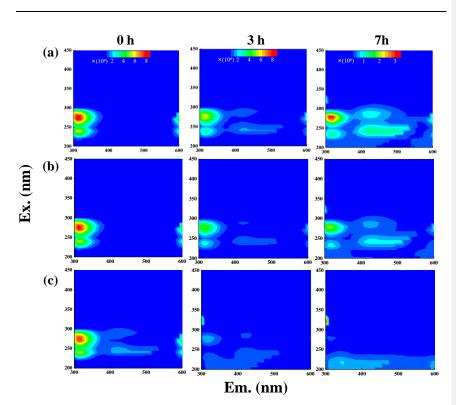
530 direct photolysis, (b) OH-initiated oxidation, and $(c)^{3}C^{*}$ -initiated oxidation.

531 3.3.2 Fluorescence properties

532 The fluorescence properties of solutions before (0 hour) and during 533 photolysisphotooxidation (3 and 7 hours) were investigated via the EEM technique, as shown in Fig. 6. For comparison, we also presented EEM profiles of pure eugenol (non-534 535 irradiated), pure DMB, and the end solutions (23 hours) of direct photolysis and OH-536 initiated oxidation in Fig. S6. The peaks at Excitation/Emission (Ex/Em)=275/313 nm can be attributed to fluorescence of the phenolic structure of parent substance (eugenol 537 here), as suggested by Laurentiis et al. (2013). As shown in both Fig. 6 and Fig. S6, the 538 fluorescence intensity decreased after photolysisoxidation due to eugenol decay, and 539 the reduction was very fast for 3C*-initiated oxidation. This finding matches with the 540 fast photolysisdegradation and large rate constant for 3C*-initiated oxidation. The EEM 541 542 plots for direct photolysis and OH-initiated oxidation had similar contour patterns as shown in Figs. 6a and b, although EEM profiles changed significantly with irradiation 543 time. We also observed distinct fluorescent peaks at Ex/Em=235/(400-500) nm, 544 545 indicating that illumination can cause a red shift in fluorescence emission wavelength. As suggested by Chang et al. (2010), fluorophores at Ex/Em=240/400 nm are linked 546 547 with aromatic structures and condensed saturated bonds including polycyclic aromatic 548 hydrocarbons. Another work (Li et al., 2021) showed that red shift in the fluorescence spectra was usually related to an increase in the size of ring system and an increase in 549 the degree of conjugation. Previous studies (Chen et al., 2016a; Chen et al., 2019; 550 551 Laurentiis et al., 2013; Wu et al., 2019) have reported that fluorescent compounds with 552 emission wavelength at 400-500 nm may be highly oxygenated species such aswere likely linked with HULIS. Additionally, HULIS have two typical fluorescent peaks in 553 EEM profile at Ex/Em=(200-300)/(400-500) nm and Ex/Em=350/(400-500) nm with 554 the former one having a higher intensity (Graber and Rudich, 2006; Laurentiis et al., 555 2013; Vione et al., 2019; Wu et al., 2021). There was also evidence that direct photolysis 556

557 of tyrosine and 4-phenoxyphenol generated HULIS with new fluorescence signals at 558 Ex/Em=(200-250)/(400-450) nm and 300/(400-450) nm (Bianco et al., 2014). In this regard, we inferred that new peak at Ex/Em=235/(400-500) nm here was likely 559 attributed to HULIS. For the ³C*-initiated photolysisoxidation, extra fluorescent peaks 560 at Ex/Em=(220-300)/(400-500) nm appeared in the first 1 hour (data not shown), but 561 intensities weakened and gradually disappeared upon prolonged 562 their photolysisreactions (3 hours). Nevertheless, EEM results should be interpreted with 563 caveatscaution because many-complicated substances might contribute to absorption 564 and emission at a certain wavelength, and it is hard to distinguish and isolate fluorescent 565 and nonfluorescent constituents simply via the EEM technique. 566

567 Another interesting finding was that a small fluorescence peak appeared at Ex/Em=(300-350)/(300-350) nm in some of the EEM profiles. Specifically, it appeared 568 earlier for ³C*-oxidation (at 3 hours) than the other two systems, yet its intensity seemed 569 570 to be a bit stronger in the end solutions of direct photolysis and OH-oxidation (Fig. S6). 571 EEM fluorescence spectra of HULIS from fog water are reported to have peaks at 572 shorter excitation and emission wavelengths than those of terrestrial fulvic acids 573 (Graber and Rudich, 2006). Moreover, as suggested by Leenheer and Croue (2003), 574 fluorescence peak position of the maximum Ex/Em for HULIS with lower MWs would shift towards lower wavelengths, thus, we inferred fluorescence peak at Ex/Em=(300-575 576 350)/(300-350) nm might be in part attributed to the organic acids with only a few 577 carbon atoms- (probably C1-C6). Nevertheless, large uncertainties still exist in using EEM fluorescence technique to identify molecular compositions of the products due to 578 lack of standard EEM profiles for specific compounds from aqueous phase oxidation 579 and clearly more studies are needed in future. 580



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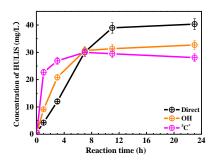
582 Figure 6. EEM fluorescence spectra of the initial solution (0 hour) and those at different reaction

583 time (3 and 7 hours) under (a) direct photolysis, (b) OH-initiated oxidation, and (c) ${}^{3}C^{*}$ -initiated

584 oxidation.

586 3.4 Characteristics of HULIS

587 The EEM spectra revealed new prominent fluorescent peak at Ex/Em=250/(400-500) nm, which was likely owing to HULIS. HULIS can be divided into fulvic acid 588 (water soluble at all pHs), humic acid (base soluble, acid insoluble) and humin 589 (insoluble at all pHs). In principle, extracted HULIS in this work with polymer-based 590 HLB SPE packing include LMW organic acids, fulvic acids and other humic substances. 591 592 Figure 7 presents the measured HULIS concentrations against the reaction time. The results show clearly that aqueous-phase eugenol oxidation is a source of HULIS, 593 and the amount increased gradually in the first 7 hours, then remained at a similar level 594 (about 30 mg/L) for the OH-initiated oxidation. For direct photolysis, HULIS 595 concentration increased until 11 hours and then became steady at a level around 40 596 mg/L. For the ³C*-oxidation, HULIS concentration increased to a maximum at 7 hours, 597 then declined slightly afterwards. A plausible reason of such variabilities is that 598 generated HULIS was capable of further taking part in photochemical reactions since 599 it can act as photosensitizer. Moreover, Yu et al. (2016) characterized the products from 600 601 aqueous oxidations of phenols by 3C* triplet states and OH radicals, and found both could produce oligomers and hydroxylated species but the 3C*-oxidation could produce 602 more of these compounds when 50% of the precursor was reacted. Considering the 603 large increases of HULIS in the first 7 hours and the much faster increase of ³C*-604 oxidation in the first 3 hours shown in Fig. 7, we postulate that HULIS species might 605 overlap withbe some of the products of high MW oligomers, which can in turn 606 contributed to fluorescence at emission of ~400 nm (Barsotti et al., 2016). 607



608

Figure 7. HULIS concentrations as a function of reaction time for the three conditions<u>under direct</u>
 photolysis, OH-initiated oxidation and ³C*-initiated oxidation.

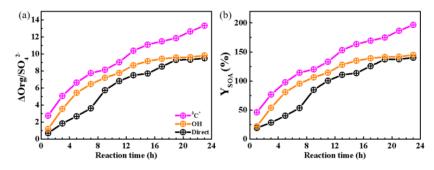
611 3.5 Mass yield and oxidation degree of reaction products

612 3.5.1 Mass yields

613 HULIS is only a subset of the products from aqueous oxidation, and here we used AMS to further quantify the total reaction products. Figure 8a shows SP-AMS 614 measured organic mass profiles (normalized by sulfate mass, $\Delta Org/SO_4^{2-}$) against the 615 reaction time. As the reaction propagated, $\Delta Org/SO_4^{2-}$ increased continuously in ${}^{3}C^{*}$ -616 initiated system. Nevertheless it arose stepwise and reached a maximum at 19 hours, 617 618 then remained at a plateau for the direct photolysis and OH-mediated oxidation. Figure 8b illustrates the calculated mass yields at different reaction times. The mass yields 619 620 after 1 hour of illumination were in the ranges of 46.2%-196.5%, 22.1%-144.9%, 19.3%-140.1% for ³C*-oxidation, OH-oxidation and direct photolysis, respectively. For 621 the same oxidation time, mass yield from ${}^{3}C^{*}$ -oxidation was generally higher than those 622 623 from OH-oxidation and direct photolysis. There are two plausible reasons for high mass yield of ³C*-initiated oxidation. First, oxidation by ³C* was more efficient to form 624 oligomers and functionalized/oxygenated products (Richards-Henderson et al., 2014; 625

Yu et al., 2016). Higher oxidative degree of products from ${}^{3}C^{*}$ -initiated photooxidation (see Sec.3.5.2) supports this hypothesis. Secondly, more light-absorbing products formed during initial stage of ${}^{3}C^{*}$ -oxidation (Fig. 5c) may accelerate oxidation by acting as photosensitizers (Tsui et al., 2018).

The product mass yields obtained in this work (~20%-197%) overall agree with 630 those reported previously for phenolic compounds. For examples, Huang et al. (2018) 631 reported mass yields of 30-120% for syringaldehyde and acetosyringone; Smith et al. 632 (2014) found that mass yields of aqSOA from three phenols with ³C* were nearly 100%, 633 and Ma et al. (2021) reported a yield ranging from 59 to 99% for six highly substituted 634 phenols with ³C*; Mass yields of SOA from three benzene-diols were near 100% with 635 both OH and ³C* oxidants (Smith et al., 2015); Direct photolysis of phenolic carbonyls, 636 and oxidation of syringol by ³C*, had SOA mass yields ranging from 80 to 140% (Smith 637 et al., 2016). Our previous study on eugenol OH oxidation illuminated by a 500 W Xe 638 lamp reported a mass yield of ~180% (Ye et al., 2020), slightly higher than the value 639 determined here owing to different simulated lightslight wavelengths/intensities. 640



641

Figure 8. Variations of the organic mass normalized by sulfate (a) ($\Delta Org/SO_4^{2-}$) and (b) mass yields of reaction products with reaction time under three-conditionsdirect photolysis, OH-initiated oxidation and ³C*-initiated oxidation.

645

646 **3.5.2 Oxidation degree**

647 In order to further probe oxidation levels of the reaction products, O/C derived 648 from SP-AMS mass spectrum of the organics was used to represent the oxidation degree 649 of products. In addition, carbon oxidation state (OSc, defined as 2*O/C - H/C) (Kroll 650 et al., 2011) was also calculated (Kroll et al., 2011). Figures 9a-c depict variations of the elemental ratios (O/C and H/C) and OSc during oxidations. Dramatic increases of 651 O/C and OSc during the initial stage of oxidation (within 1 hour) were observed, with 652 O/C changing from 0.26 to 0.65, from 0.26 to 0.70, from 0.25 to 0.75, as well as OSc 653 from -1.11 to -0.15, from -1.16 to -0.05, from -1.13 to 0.09 for direct photolysis, OH-654 oxidation and 3C*-oxidation, respectively. The O/C was lower than those of other 655 phenolic aqSOA (Yu et al., 2014) due to different substituted groups in aromatic ring of 656 657 the precursors. Both O/C and OSc gradually increased, while H/C changed little after 1 hour. The enhancements of OSc in the end were 1.22, 1.11 and 0.86 for ³C*-initiated 658 oxidation, OH-initiated oxidation and direct photolysis, respectively. 659

Furthermore, the f44 vs. f43 diagram ("triangle plot") can be used to demonstrate 660 the evolution of SOA during oxidation (Ng et al., 2010). The f44 and f43 are defined as 661 the ratios of signal intensities of m/z 44 (mainly CO₂⁺) and 43 (mainly C₂H₃O⁺) to the 662 663 total organics. The results that the f44 increased continuously (moved upwards) during both OH and ³C* oxidations, indicating persistent formation of highly oxygenated 664 compounds including organic acids, such as formic acid and oxalic acid (Sun et al., 665 666 2010). Concentrations of organic acids increased with photochemical reactions can 667 support this assumption (data not shown). Note the f_{44} enhancement was much more significant for ³C* oxidation (from 0.07 to 0.16) than direct photolysis (from 0.07 to 668 0.12) and OH oxidation (from 0.07 to 0.13), consistent with the behaviors of its higher 669 O/C and OSc. The f_{43} value decreased in the first stage (1-3 hours) and then increased 670 at later stages. The final f_{43} values were almost the same as those of the initial solutions-671

672 and were small. As a result, all data points located outside the f_{44} vs. f_{43} -space f_{43} region

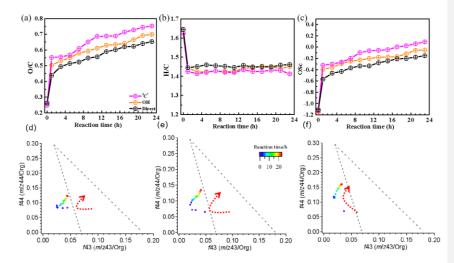
673 (bounded by the two dash lines in Figs. 9d-f) for ambient aerosols established by Ng et

674 al. (2010) for ambient aerosols, owing to the relatively low f_{43} values.).

675 In summary, our results shown here demonstrate that aqueous phase eugenol

676 photochemical oxidation can generate highly oxygenated products and hence increase

677 the degree of oxygenation of overall SOA.



679 **Figure 9.** Variations of <u>the</u> elemental ratios of (a) O/C, (b) H/C and (c) the-oxidation state (OSc) as 680 a function of reaction time, and the: f_{44} vs. f_{43} plots of reaction products under (ad) direct photolysis, 681 (be) OH-initiated oxidation, and (ef) ${}^{3}C^{*}$ -initiated oxidation.

3.6 Molecular characterization of reaction products and proposed reaction mechanism

684 3.6.1 Major products identified by GC-MS

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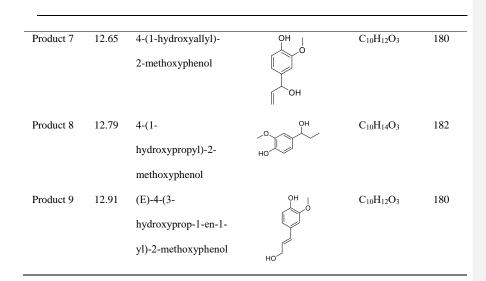
685 SP-AMS was limited to probe bulk composition of low-volatility oxidation 686 products, thus the molecular-level characterization of products was performed by using 687 GC-MS here. The total ion chromatograph (TIC) of GC-MS on the solutions before 32

688 illumination (0 hour) and at illumination times of 11 and 23 hours for the ³C*-initiated 689 photooxidation is shown in Fig. S7. As shown in Fig.S7, eugenol (retention time (RT) at 11.50 min) loss was more than 90% at 11 hours, which could be confirmed by the 690 691 experimental data reported in Section 3.1. Comparison of products at 11 hours and 23 692 hours showed no significant difference. Similar to aqueous photochemical oxidation with OH (Ye et al., 2020), a series of products were identified and listed in Table 2. 693 Except 5-ally-3-methoxybenzene-1,2-diol (MW 180, RT=12.59 min), the other eight 694 products were detected for both OH and 3C*-initiated photooxidations. Some of them 695 (Eugenol, DMB, product 1, 2, 5) were identified by using certified reference materials, 696 some of them (product 3, 4, 6, 7, 8, 9) were inferred according to the molecular ion 697 698 peaks and fragments from GC-MS, based on spectra from the NIST database (Stein, 2014) and on the reactants and reaction conditions. 699

We also found 4-(1-hydroxypropyl)-2-methoxyphenol (product 8) was relatively 700 701 abundant (Fig.S7), suggesting functionalization might dominates as compared to 702 oligomerization and fragmentation. Products were mainly from addition/elimination of 703 hydroxyl (-OH), methoxyl (-OCH3) to benzene ring or allyl group and further oxidized 704 to carbonyl or carboxyl compounds. As suggested by Bonin et al. (2007), the OHaddition to the aromatic ring of phenol preferentially takes place at the ortho (48%) and 705 the para (36%) positions, leading to the formation of OH-adduct product 6 (5-allyl-3-706 707 methoxybenzene-1,2-diol). Notably, dimers and ring-opening products were not 708 observed, but they cannot be excluded since they would be probably out of the detection of GC-MS technique (Vione et al., 2014). 709

	RT	Name*	Proposed chemical	Chemical	Nominal
	(min)		structure	formula	MW
					(g/mol)
Product 1	10.68	4-allylphenol		C ₉ H ₁₀ O	134
Precursor	11.50	Eugenol	OH C	$C_{10}H_{12}O_2$	164
Product 2	11.81	4-hydroxy-3- methoxybenzaldehy	но-	C ₈ H ₈ O ₃	152
Product 3	12.06	de (E)-2-methoxy-4- (prop-1-en-1-	HO	$C_{10}H_{12}O_2$	164
Product 4	12.11	yl)phenol 4-(hydroxymethyl)- 2-methoxyphenol	OH OH	C ₈ H ₁₀ O ₃	154
Product 5	12.18	2-methoxy-4- propylphenol	HO	$C_{10}H_{14}O_2$	166
Photosensi tizer	12.29	3,4- dimethoxybenzaldeh yde(DMB)		C ₉ H ₁₀ O ₃	166
Product 6**	12.59	5-ally1-3- methoxybenzene- 1,2-diol	HO O	$C_{10}H_{12}O_3$	180

711 Table 2. Major reaction products identified via GC-MS



712 *Precursor (eugenol) and triplet precursor (DMB) are also shown.

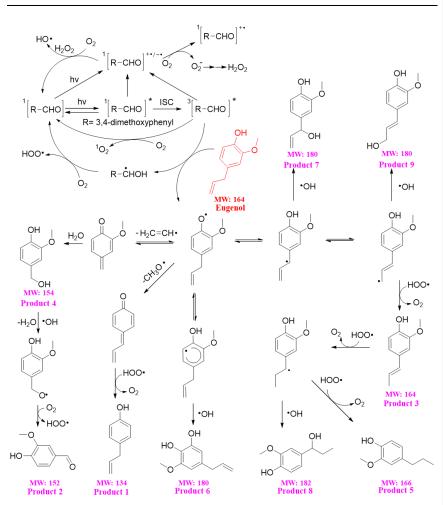
713 **This compound was only identified in ³C*-oxidation solution.

714 **3.6.2 Reaction mechanism**

The reaction pathways of ³C*-initiated photooxidation of eugenol are 715 716 demonstrated in Scheme 1 based on the products identified by GC-MS. The other intermediates and the potential pathways were proposed according to the 717 identified products and the reaction rationality from the starting reactant. To better 718 depict the mechanism, DMB was expressed as [RCHO] and eugenol as Ph-R for 719 720 simplicity. First, [RCHO] absorbs light and undergoes excitation to ¹[RCHO]^{*}, then experiences the intersystem crossing (ISC) to form 3[RCHO]*. 3[RCHO]* can 721 participate in subsequent reactions via three channels. First, it can react with O2 to form 722 ¹O₂ via energy transfer. Secondly, it can transform to [RCHO]⁻, subsequently reacts 723 724 with O₂ to generate O₂ • via electron transfer, which can disproportionate to H₂O₂. The 725 decomposition of H₂O₂ can generate OH radical. Thirdly, the ³[RCHO]^{*} can react with Ph-R to from [Ph-R•] via H-abstraction. The cleavage of [Ph-R•] to free radical segment 726 35

727 (such as CH₂CH• or CH₃O•) takes place, then an additional hydrogen transfer could 728 occur, resulting in a 2H-addition to the new intermediate to form 4-allyl-phenol (product 1). Similarly, when the CH₂CH• is lost from [Ph-R•], an addition of H₂O 729 730 would happen on the new compound (product 4) and further oxidized to 4-hydroxy-3-731 methoxybenzaldehyde (product 2). Another possibility is the intermediate [Ph-R•] can resonate to several different isoelectronic species, the radical position changes to 732 aromatic ring or allyl group site, which would couple with HO• to form hydroxylated 733 eugenol monomer (product 6, 7, 9 MW=180). Consequently, the isoelectronic species 734 735 at allyl group site could also abstract a hydrogen to form isoeugenol (product 3 MW=164). Also, breakage of C=C into C-C and 2H-addition at allyl group site could 736 737 form 2-methoxy-4-propyl-phenol (product 5, MW=166). Besides, the C=C breaking intermediate can couple with HO• to form 4-(1-hydroxypropyl)-2-methoxyphenol 738 (product 8, MW=182). In conclusion, ³C^{*} can directly oxidize eugenol to form SOA 739 740 products or small molecular compounds, or indirectly oxidize eugenol via energy transfer, electron transfer, hydrogen abstraction, proton-coupled electron transfer or 741 742 other radical chain reactions.

The organic groups, such as methoxy, allyl groups can be eliminated from 743 aromatic ring, which then participate in photochemical reaction, resulting in generation 744 of dimers, small organic acids, CO2 and H2O, etc. Dimers previously reported from 745 746 aqueous reaction of 4-methylsyringol with OH were not detected via GC-MS in the 747 present work but dimer fragment ions (C20H22O4+) were detected by SP-AMS with trace amounts. Functionalization due to the additions of hydroxyl, carbonyl functional groups 748 to the aromatic rings could account for the enhancement of light absorption at 749 wavelength of 300-400 nm. However, polar high MW organic acids were not detected 750 likely due to the limitation of GC-MS technique. 751





Scheme 1. Proposed reaction mechanism of ³C*-initiated photooxidation of eugenol. The red texts
 representtext represents the products listed in Table 2precursor, and the compounds labeled by
 Product 1-9 are those identified by GC-MS-(Table 2).

756 **3.7 Oxidative potential (OP) of reaction products**

Previous laboratory studies (Verma et al., 2015b; Xu et al., 2020) have confirmed
 that HULIS is a major constituent contributing to ROS-generation potential. As HULIS
 is an important fraction of the products from aqueous photooxidation of eugenol in this

760 work (Fig. 7), here we investigated the links between the OPs and reaction products. 761 The OP of oxidation products can be represented by the consumption rate of DTT 762 concentration-per minute, defined as R_{DTT}. Figure 10a shows the DTT consumed mass (MDTT) as a function of incubation times (0, 30, 60, 90, 120 and 150 min) for a triplicate 763 764 sample (300 µM eugenol) and blank (ultrapure water). MDTT values for both blank and eugenol were proportional to incubation time, indicating that ROS-generating 765 substances in reaction solution act only as catalyst and itself was not consumed. The 766 767 slopes represent DTT consumption rates, which are also illustrated in Fig. 10a. Average RDTT0 (blank) was 0.31 µM/min and RDTT for initial 300 µM eugenol (before experiment) 768 was 0.52 µM/min. Since self-oxidation of DTT might lead to the consumption of DTT 769 770 in ultrapure water, final DTT consumption rate of reacted solution after photolysisoxidation was then blank-corrected by subtracting the average RDTTO. 771

772 Figure 10b shows changes of blank-corrected RDTT with photolysisreaction time 773 for direct photolysis, OH-initiated oxidation and ³C*-initiated oxidation, respectively. 774 The RDTT value of ³C*-oxidation products increased quickly and reached the maximum 775 (0.9) at 7 hours, then decreased slowly and its end value was lower than that from OH-776 oxidation. The RDTT value of OH-oxidation products on the other hand increased slowly and reached the maximum at 21 hours. The RDTT value of products from direct 777 photolysis increased continuously but also slowly to ~0.36 till the end of oxidation. 778 779 Nevertheless, we can see that the final RDTT values were all higher than that of initial 780 eugenol, proving that aqueous-phase processing can generate products with higher OP, resulting in more health hazards than the precursor does. The DTT consumption rates 781 are comparable to those using the same DTT method (Charrier and Anastasio, 2012; 782 Lin and Yu, 2019). This finding further indicates the effectiveness of DTT method to 783 represent OP. The weak correlation was found between HULIS concentration and RDTT, 784

785 implying that OP was not only dependent upon HULIS. Moreover, HULIS with diverse

786 molecular structures also exhibit different ROS-generation potentials (Kramer et al.,

787 2016), therefore the HULIS as an ensemble may not correlate well with OP.

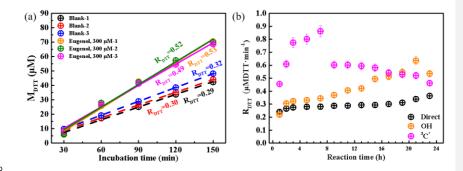




Figure 10. (a) DTT consumed mass versus incubation times for blank (ultrapure water) and 300 μM
eugenol solutions in a triplicate, and (b) blank-corrected DTT consumption rates versus reaction
time for direct photolysis, OH-initiated oxidation and ³C*-induced oxidation.

792 4. Atmospheric implications

793 The high mass yields of aqueous-phase photooxidation of eugenol (exceeding 100% after 23 hours of illumination) studied found here are similar or even higher than those 794 795 previously reported yields of a number of phenolic compounds (e.g., Smith et al., 2014, 796 2015, 2016; Ma et al., 2021), which re-emphasizes the importance of biomass burning (BB) to SOA budget (Gilardoni et al., 2016), particularly in regions or periods with 797 798 significant BB activities. Compared to simple phenols (such as syringol) that are only present in cloud/fog waters, the highly substituted phenols are able to significantly 799 800 partition into aerosol water too (Ma et al., 2021). Since the highly substituted phenols can take up roughly 30-45% of total phenols emitted from wood burning (Schauer et 801 al., 2001), our results further imply that aqueous production of SOA from BB emissions 802 803 only occur in fog/cloud conditions but also in common humid weather 39

804	conditions, highlighting the general importance of aqueous oxidation pathway to SOA.
805	OurIn addition, our study here used 300 μM H2O2 and 15 μM DMB as sources of
806	OH and ³ C*, and ³ C*-mediated oxidation appeared to be faster than OH-initiated
807	oxidation of eugenol. Of course, whether or not ${}^{3}C^{*}$ is more important than OH in real
808	atmosphere depends upon their concentrations. OH and ${}^{3}C^{*}$ are difficult to measure and
809	concentrations vary greatly in real atmospheric samples. Herrmann et al. (2010)
810	estimated an average OH level of 0.35x10 ⁻¹⁴ M in urban fog water; Kaur and Anastasio
811	(2018) measured ${}^{3}C^{*}$ concentration to be (0.70-15) x10 ⁻¹⁴ M, 10-100 times higher than
812	the co-existing OH in ambient fog waters; Kaur et al. (2019) determined both OH and
813	$^{3}C^{*}$ concentrations in PM extracts, OH steady-state concentration was 4.4(±2.3) x10 ⁻¹⁶
814	M, similar to its level in fog, cloud and rain, while $^3\mathrm{C*}$ concentration was 1.0(±0.4)
815	$x10^{-13}$ M, a few hundred times higher than OH and nearly double its average value in
816	fog. Therefore, together with these measurements, our findings signify a likely more
816 817	fog. Therefore, together with these measurements, our findings signify a likely more important role of ${}^{3}C^{*}$ than OH in aqueous-phase (especially aerosol water) reactions.
817	important role of ${}^{3}C^{*}$ than OH in aqueous-phase (especially aerosol water) reactions.
817 818	important role of ³ C* than OH in aqueous-phase (especially aerosol water) reactions. In additionHowever, the liquid water content of aerosol is typically ~10000 times
817 818 819	important role of ${}^{3}C^{*}$ than OH in aqueous-phase (especially aerosol water) reactions. In addition <u>However, the liquid water content of aerosol is typically ~10000 times</u> smaller than that of cloud (for instance, ~50 µg m ⁻³ versus 0.5 g m ⁻³). Even if the
817 818 819 820	important role of ${}^{3}C^{*}$ than OH in aqueous-phase (especially aerosol water) reactions. In additionHowever, the liquid water content of aerosol is typically ~10000 times smaller than that of cloud (for instance, ~50 µg m ⁻³ versus 0.5 g m ⁻³). Even if the reaction rates in aerosol water were 10 times higher than those in cloud water, the
817 818 819 820 821	important role of ${}^{3}C^{*}$ than OH in aqueous-phase (especially aerosol water) reactions. In additionHowever, the liquid water content of aerosol is typically ~10000 times smaller than that of cloud (for instance, ~50 µg m ⁻³ versus 0.5 g m ⁻³). Even if the reaction rates in aerosol water were 10 times higher than those in cloud water, the overall importance of aqueous reactions initiated by the same oxidant in aerosol phase
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 817 818 819 820 821 822 823 	important role of ${}^{3}C^{*}$ than OH in aqueous-phase (especially aerosol water) reactions. In additionHowever, the liquid water content of aerosol is typically ~10000 times smaller than that of cloud (for instance, ~50 µg m ⁻³ versus 0.5 g m ⁻³). Even if the reaction rates in aerosol water were 10 times higher than those in cloud water, the overall importance of aqueous reactions initiated by the same oxidant in aerosol phase would be still ~1000 times smaller than it in cloud water. Moreover, quenching experiments reveal that O ₂ can inhibit eugenol degradation by effectively scavenging
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 817 818 819 820 821 822 823 824 825 	important role of ${}^{3}C^{*}$ than OH in aqueous-phase (especially aerosol water) reactions. In additionHowever, the liquid water content of aerosol is typically ~10000 times smaller than that of cloud (for instance, ~50 µg m ⁻³ versus 0.5 g m ⁻³). Even if the reaction rates in aerosol water were 10 times higher than those in cloud water, the overall importance of aqueous reactions initiated by the same oxidant in aerosol phase would be still ~1000 times smaller than it in cloud water. Moreover, quenching experiments reveal that O ₂ can inhibit eugenol degradation by effectively scavenging ${}^{3}C^{*}$ radical while it can promote degradation by fostering radical chain reactions in OH- induced oxidation, which offer insights to the_control of reaction pathways by

Eugenol has a strong light absorption peak around 280 nm, therefore it can

829 photolyze itselfundergo direct photolysis, and addition of OH or other photosensitizers 830 (³C*) can gradually diminish its light absorption around 280 nm, but increase the 831 absorption in visible light range (>300 nm). In the meantime, HULIS was generated continuously, and GC-MS identified a number of high MW organic products, in line 832 833 with those detected in earlier aqueous photooxidation of phenolic compounds (Jiang et al., 2021; Misovich et al., 2021; Tang et al., 2020; Yu et al., 2014). Overall, our work 834 demonstrates that aqueous oxidation of BB emissions is a source of BrC, and this BrC 835 may act as photosensitizer to oxidize other species; a portion of this BrC might be 836 HULIS, and some high MW aromatic compounds are a subset of this HULIS. However, 837 a recent study by Wang et al. (2021) shows that fossil foil derived OA (FFOA) can be 838 839 an effective precursor of aqSOA, but the aqSOA became less light-absorbing than the FFOA. Aqueous oxidation of 4-nitrophenol with OH can lead to a photobleaching effect 840 too (Witkowski et al., 2022). These contrasting results indicate that contribution of 841 842 aqueous oxidation to BrC is largely dependent upon the precursors; molecular 843 structures of major chromophores, changes of the structures upon oxidation as well as 844 their interplay with light absorptivity should be carefully investigated to achieve a full 845 understanding of the impacts of aqueous processing on air quality, radiative forcing and climate change. 846

Investigations on the OPs of reaction products from eugenol photooxidation in all three conditions show that aqueous processing can produce more toxic products than itsthe precursor. This result is in agreement with our previous work on resorcinol, hydroquinone and methoxyhydroquinone (Ou et al., 2021). Although more studies on a broad spectrum of atmospherically relevant species and multiple indicators of toxicity are clearly needed, our findings here underscore the potential of aqueous processing on the enhancement of particle toxicity. Considering high PM concentration is often 854 accompanied with cold and humid weather conditions, the additional adverse health

855 effects caused by aqueous oxidation may amplify the health hazards of PM pollution.

856 5 Conclusions

This study comprehensively investigated the aqueous photooxidation of eugenol 857 upon direct photolysis and attacks by OH radicals and ³C* radicalstriplet states. By 858 using a suite of techniques, the decay kinetics of eugenol, chemical, optical properties 859 860 as well as toxicity of reaction products were systematically-studied. The first-order 861 photolysis rate constants followed the order of ³C*>OH >direct photolysis (300 µM H₂O₂ and 15 µM DMB as sources of OH and ³C*). Further quenching experiments on 862 different ROS during 3C*-mediated oxidation showed that 3C* was the major 863 864 contributor, followed by 1O2, O2- and OH; O2- played a more important role than OH 865 during OH-initiated oxidation. Photolysis The rate constants under saturated O2, air and N_2 followed the order of $k_{O_2} > k_{Air} > k_{N_2}$ for both direct photolysis and OH-initiated 866 oxidation, but changed to $k_{\rm Air}$ > $k_{\rm N_2}$ > $k_{\rm O_2}$ for $^3C^*$ -mediated oxidation. O2 appeared to 867 be a scavenger of ³C^{*} therefore suppressing ³C^{*} oxidation while it could promote 868 869 generation of OH thus accelerate OH-mediated oxidation. pH and DO levels both decreased during oxidation, indicating formation of acids and a certain role of DO in 870 871 oxidation.

Eugenol itself can absorbs lights significantly around 280 nm, and aqueous oxidation gradually decrease this absorption of UV light but enhanced the absorbance in the visible light range (mainly 300-400 nm), indicative of the generation of BrC species. These species were likely linked with HULIS, as HULIS concentration increased during the course of oxidation, in particular for the initial stage of ${}^{3}C^{*}$ mediated reactions. The final mass yields of reaction products (after 23 hours of 42 878 irradiation) were 140.1%, 144.9% and 196.5% for direct photolysis, OH-oxidation and 879 ³C*-oxidation, respectively. Oxidation degrees of the products increased continuously 880 with the illumination time, indicating persistent formation of highly oxygenated 881 compounds, especially during ³C*-mediated reactions. Molecular characterization by 882 GC-MS identified a series of oxygenated compounds, allowing us to propose the 883 detailed oxidation mechanism. Functionalization appeared to be a dominant pathway to 884 form the observed species.

DTT method was used to assess OPsOP of the reaction products. The end products 885 in all three easessets of experiments showed higher DDT consumption rates than that 886 of the precursor; products from ³C*-oxidation showed particularly fast increase in the 887 888 first few hours of reactions. This result demonstrates that species that are more toxic than its precursors could be produced upon aqueous oxidation, indicative of the 889 890 potential toxic effects induced by aqueous processing. Overall, by using eugenol as a 891 model compound of BB emissions, our findings highlight the importance of aqueous 892 oxidation of BB emissions to SOA formation, its potentially important role in affecting radiative balance and climate through formation of BrC, as well as possible additional 893 adverse health effects. Such effects should be considered in air quality or climate 894 895 models to better assess the influence of BB emissions.

 B97
 Data availability. The relevant data inof this study are available from the authors upon

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 request
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 899
 http://nuistairquality.com/eugenol_data_and_figure

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901 Supplement. The supplement related to this article is available on line at: XXX
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Author Contributions: XDL, YT, LWZ, SSM, SPL, ZZZ and NS conducted the
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paper with contributions from all co-authors. ZLY and XLG reviewed and commented
on the paper.

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908 *Competing interests.* The authors declare that they have no conflict of interest.

909

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923 Review statement. This paper was xxx

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