

Response to the comments of Reviewer #2

The reviewed study investigates combining precursors in SOA experiments and the effect of the combination in its ability to produce SOA. Interestingly the authors observe that combining an alkane and aromatic precursor in their experiments results in an SOA yield that is greater than the weighted average of each precursor's individual SOA yield. This could be from a change in partitioning or from new chemical pathways that are activated only in the combination of dodecane and TMD. The authors state the cause is from the occurring chemistry, and not just partitioning. This is justified by looking at molecular species with ESI-TOF-MS observed in in experiments as well as the UV-Vis absorbance of the resulting SOA extract. The authors don't make the strongest case for what specifically changes in the occurring chemistry to generate more SOA, but they do point to measurable differences, such as unique ESI-TOF-MS peaks in the mixed experiments. Overall, the results are interesting and worthy of publication.

Response: We thank Anonymous Referee #2 for the review and the positive evaluation of our manuscript. We have fully considered the comments and [responded to these comments below in blue text](#). The revisions in the manuscript are highlighted in yellow color. The response and changes are listed below.

Specific Comments:

1. *In Figure 1 a and b you show that the corrected mass is substantially greater than the experimentally measured mass, by at least an order of magnitude it appears. This is concerning as there is undoubtedly some error in the correction that is used. Could the authors please estimate the error associated with the mass correction they use and how that propagates into the "mass corrected" values in Figure 1.*

[The uncertainty of the mass correction is about \$\pm 11.2\%\$, and we have added this in the main text.](#)

[Page 4, line 124-125: "The uncertainty of the mass correction here is about \$\pm 11.2\%\$ \(see Supporting Information for details\)"](#)

[Supporting Information, Page 2-3, Line 44-57: "The error associated with the mass](#)

correction here is mainly from the gas phase. For the gas phase wall-loss correction, the gas-particle partitioning timescale ($\bar{\tau}_{g-p}$) and the vapor wall-loss timescale ($\bar{\tau}_{g-w}$) are expressed as the following equation:

$$\bar{\tau}_{g-p} = \frac{1}{2\pi\bar{N}_p\bar{D}_pD_{gas}\bar{F}_{FS}}$$

$$\bar{\tau}_{g-w} = \frac{1}{k_w}$$

$$k_w = \left(\frac{A}{V}\right) \frac{a_w \bar{c}}{1.0 + \frac{\pi}{2} \left[\frac{a_w \bar{c}}{4(k_e D_{gas})^{0.5}} \right]}$$

For $\bar{\tau}_{g-w}$, the parameters in the formula are fixed values, so only $\bar{\tau}_{g-p}$ is considered here. The two variables N_p and D_p in $\bar{\tau}_{g-p}$ are independent and the uncertainty of them are 10% and 1%, respectively. Considering the propagation of uncertainty, we can obtain the variance formula:

$$\sigma_{\tau}^2 = \sigma_N^2 \left(\frac{\partial \tau}{\partial N}\right)^2 + \sigma_D^2 \left(\frac{\partial \tau}{\partial D}\right)^2$$

Then the uncertainty of $\bar{\tau}_{g-p}$ can be calculated as:

$$\frac{\sigma_{\tau}}{\tau} = \sqrt{\left(\frac{\sigma_N}{N}\right)^2 + \left(\frac{\sigma_D}{D}\right)^2} = \sqrt{0.1^2 + 0.01^2} = \sqrt{0.0101} = 10.0\%$$

The uncertainty of $\bar{\tau}_{g-p}$ is $\pm 10.0\%$, the measurement uncertainty of the precursors concentration by TD-GC applied in this work is about $\pm 5\%$, this resulting in the final uncertainty of mass correction to be about $\pm 11.2\%$.”

2. *In Figure 5 it is difficult to see the difference between peaks because of the x-axis scale. It would be more informative to make the x-axis from 200 – 300 nm, and simply note in the caption that absorbance at wavelengths >300 nm is negligible.*

Thank you for this suggestion. We have updated Figure 5 as Figure 6(a), and the figure caption is also updated.

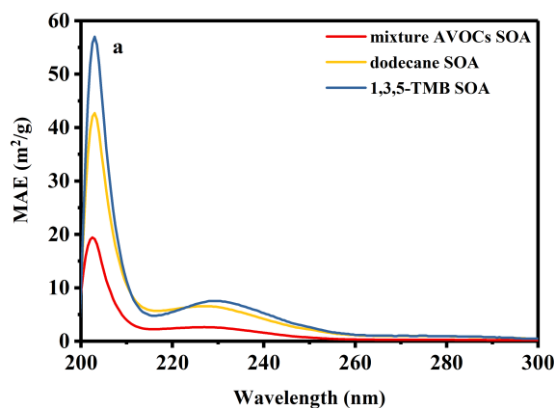


Figure 6. (a) UV-Vis spectra (MAE) of the n-dodecane, 1,3,5-TMB, and mixture AVOCs SOA. As the absorbance at wavelengths >300 nm is negligible, we only show the range from 200 to 300 nm. (Page 27, line 650-652)

3. *It seems erroneous to say the observed peaks around 205 nm are strictly due to carboxyl. Given the presence of NO in your experiments, you will have nitrate functionality in your products. Nitrate absorbs strongly around 210 nm, but this can be shifted from neighboring functional groups, etc.*

We have modified the description in the manuscript. To further determine the potential functional groups in SOA extracts, the IR spectra are acquired by the Attenuated Total Internal Reflection Infrared (ATR-IR) analysis. We have added this in the manuscript.

Page 9, line 259-267: “To further determine the potential functional groups in SOA extracts, ATR-IR spectra were acquired (Figure 6b). In order to eliminate the influence of water, experiments were conducted under dry conditions. As shown in Figure 6 (b) and Table S3, the bold peak at 3360 cm^{-1} corresponds to the characteristic peak of C-OH in alcohol. The peak at 3192 cm^{-1} originates from O-H stretching vibration of carboxylic acid. The two characteristic peaks at 2921 cm^{-1} and 2850 cm^{-1} corresponds to the C-H stretching vibration of alkane. The peaks at 1660 cm^{-1} and 1633 cm^{-1} originate from C=O stretching vibrations. The signal at 1465 cm^{-1} and 1415 cm^{-1} represent the deformation vibrations of methyl and methylene groups. The peak around 1268 cm^{-1} corresponds to the vibration of nitrate groups in nitrate ester. The results above suggest that the SOA extracts are dominantly composed of carbonyl compounds, carboxylic acid, nitrate ester, and alcohol. This is consistent with previous studies (Huang et al., 2015; Fahnstock et al., 2015).”

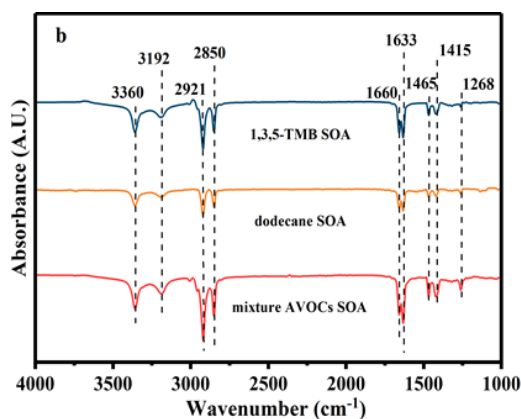


Figure 6. (b) ATR-IR spectra for the n-dodecane, 1,3,5-TMB, and mixture AVOCs SOA by using a background spectrum obtained without a sample as the reference. (Page 27, line 650-652)

Table S3. List of functional groups in SOA extracts. (Page 4, line 70, Supporting Information)

Functional groups	Wavenumber (cm ⁻¹)
C-OH in alcohol	3360
O-H stretching vibration of carboxylic acid	3192
C-H stretching vibration of alkane	2921, 2850
C=O stretching vibrations	1660, 1633
deformation vibrations of methyl and methylene groups	1465, 1415
nitrate groups in nitrate ester	1268

4. Based on figure 5, you cannot say (line 243) that the mixed AVOCs forms more carboxyl compounds. Because the mixed experiments contain more aerosol mass, you would expect the absorbance to be greater even with the same yield of carboxyls (or nitrate, see comment 3) relative to the non-mixed experiments.

The description in the manuscript has been modified, and the mass absorption efficiency (MAE) is applied to better present the light absorption properties of SOA extracts.

Page 8-9, line 251-258: “Based on the light absorption spectra, the mass absorption efficiency (MAE, m²/g) of the SOA in the extracts is calculated using the following equation (Chen et al., 2016):

$$MAE(\lambda) = \ln(10) Abs(\lambda)/C_{OM} \quad (7)$$

where $Abs(\lambda)$ is the light absorption coefficient (m^{-1}), and C_{OM} is the SOA mass concentrations in the extracts. The MAE of the SOA extracts in Figure 6 (a) was calculated from 200 to 300 nm. The MAE at 205 nm were in the order: 1,3,5-TMB SOA ($56.8 m^2/g$) > dodecane SOA ($42.5 m^2/g$) > mixture AVOCs SOA ($19.5 m^2/g$). The MAE in the 210-250 nm band also show the same pattern. This indicates that the SOA generated by the mixture AVOCs contains less light-absorbing substance per unit mass relative to dodecane SOA and 1,3,5-TMB SOA.”

5. The authors note a handful of peaks from ESI that only appear in mixed experiments, but in Figure 4 it is not especially clear that there are many peaks in panel C that are not in panel A and B. Does the ESI have similar sensitivity for all observed species? Or can sensitivity vary greatly between species? Please address this, it would help the reader to interpret figure 4.

Although the sensitivity of ESI-ToF-MS to different species may be different, our purpose is not to quantify their intensity or contribution, but to analyze their differences between experiments. Therefore, we believe that the difference in sensitivity does not affect the results and conclusion in this study.

In order to clearly see the magnitude of the peaks unique to the mixed experiments, Figure 4 has been updated as follows:

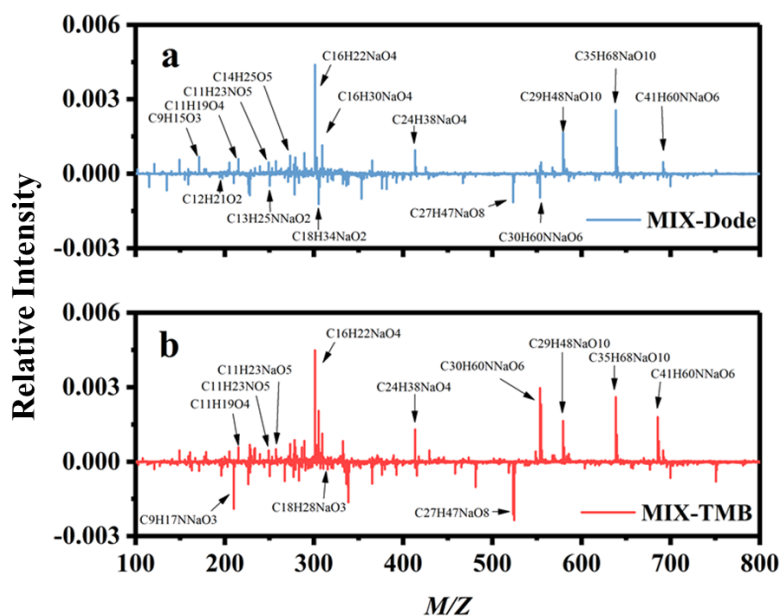


Figure 4. Mass spectra difference in (a) mixed AVOCs SOA minus *n*-dodecane SOA, and (b) mixed

AVOCs SOA minus 1,3,5-TMB SOA. The Y-axis is the relative intensity normalized by dividing by the total signal strength of the mass spectra. (Page 25, line 642)

6. Also, as it is difficult to see the magnitude of the peaks unique to the mixed experiments in panel C - is it possible that the unique peaks are present in unmixed experiments but just under detectable limits, and upon producing greater aerosol in the mixed experiments, the unique peaks were elevated to above detectable limits? Perhaps a useful way to address this would be give the intensities of the unique peak relative to the largest peak of the spectrum, in addition to addressing comment 5.

In order to clearly see the magnitude of the peaks, the relative intensity of the observed species have been added in Table 2.

Table 2. Representative identified mass spectral peaks, molecular formulas, molecular weights, and relative intensity of *n*-dodecane, 1,3,5-TMB, and mixture AVOCs-derived SOA. (Page 21, line 615)

Molecular Formula	M+H	M+Na	MIX-Relative Intensity ($\times 10^{-3}$)	TMB-Relative Intensity ($\times 10^{-3}$)	<i>n</i> -dodecane Relative Intensity ($\times 10^{-3}$)
C ₉ H ₁₄ O ₃	171.099		1.41	1.73	0.733
C ₉ H ₁₇ NO ₃		210.111	0.484	2.38	0.864
C ₁₁ H ₁₈ O ₄	215.126		0.641	0.0563	0.0489
C ₁₄ H ₂₂ O ₃	239.166		0.217	0.0526	0.0489
C ₁₄ H ₂₀ O ₂		243.134	0.113	0.0966	0.110
C ₁₁ H ₂₂ NO ₅	249.158		0.489	0.0583	0.0718
C ₁₄ H ₂₆ O ₂		249.183	0.503	0.0495	0.0459
C ₁₃ H ₂₅ NO ₂		250.177	1.22	1.79	1.69
C ₈ H ₁₂ O ₉	253.056		0.220	0.0308	0.0109
C ₁₁ H ₂₂ O ₅		257.135	0.535	0.0265	0.0223
C ₁₂ H ₂₀ O ₆	261.131		0.387	0.284	0.393
C ₁₅ H ₂₈ O ₂		263.199	0.250	0.0808	0.0427
C ₁₄ H ₂₄ O ₅	273.167		0.799	0.0872	0.0643
C ₁₆ H ₂₂ O ₄	279.159		0.728	0.139	0.0724
C ₁₄ H ₂₆ O ₄		281.172	0.305	0.0685	0.0427
C ₁₈ H ₂₈ O		283.207	1.14	1.83	1.46
C ₁₃ H ₂₂ NO ₆	289.153		0.890	0.0513	0.0565
C ₁₆ H ₂₂ O ₄		301.146	4.77	1.09	1.41
C ₁₈ H ₃₄ O ₂		305.263	3.26	1.22	0.004.4
C ₁₆ H ₃₀ O ₄		309.202	1.28	0.149	0.135
C ₁₈ H ₂₈ O ₃		315.194	0.936	1.25	1.26

C ₁₆ H ₂₄ O ₅		319.151	2.09	0.0290	0.0197
C ₂₀ H ₃₄ O ₂		329.246	0.319	0.0927	0.133
C ₁₉ H ₃₈ O ₄		353.267	1.42	1.46	2.42
C ₂₄ H ₃₈ O ₄		413.266	2.69	1.39	1.74
C ₂₀ H ₃₄ O ₈		425.214	0.297	0.395	0.0333
C ₂₄ H ₃₆ NaO ₅		427.245	0.107	0.108	0.0393
C ₂₇ H ₄₈ O ₈		523.325	0.183	1.54	1.33
C ₃₀ H ₆₀ NO ₆		553.459	3.08	0.145	3.32
C ₂₈ H ₄₈ O ₁₀		567.307	0.272	0.0195	0.0215
C ₂₉ H ₄₈ O ₁₀		579.296	1.54	0.0150	0.0167
C ₃₅ H ₆₈ O ₁₀		639.480	1.2	0.109	0.134
C ₄₁ H ₆₀ NO ₆		685.434	2.01	0.209	2.11

Technical corrections:

7. Line 84: Add “The” before “OH precursor”

We have added this in the manuscript.

Page 3, line 85: “OH precursor” → “The OH precursor”

8. Line 84 and 86: Correct subscripts in molecular formulas.

We have corrected the subscripts in the molecular formulas.

Page 3, line 86: “NaNO2” → “NaNO₂”

Page 3, line 88: “NO2” → “NO₂”

9. Line 219: *m/z* is commonly italicized, *m/z*, this should be corrected throughout document.

We have corrected this throughout this manuscript.

Page 7, line 218-219; Figure 4; Figure S2: “m/z” → “*m/z*”

10. Line 243: “formes” should be forms

We have corrected this in the manuscript.