

Reply to Anonymous Referee #2

We thank referee #2 for the constructive comments. The original comments are in black font, our replies to the comments appear below each comment in blue font, and changes in the manuscript are in red font.

Specific Comments 1. In the experimental it is stated that “Pure organic solvents provided the higher extraction efficiency than mixtures with water and similar ones to mixtures of acetonitrile and methanol.” How was the extraction efficiency quantified and was it compared across different types of molecules, or only across the ones that are extracted well with the solvent that was chosen? What were the different pure organic solvents that were tested?

In the revised paper we explain our extraction tests more in detail. The extraction tests were based on real ambient UFP filter samples extracted with different types of solvents. We minimized the consumption of our valuable filter samples, by only testing four different solvent combinations. The solvents were evaluated based on the non-target-software`s identifications and corresponding signal intensities.

Line 131-134: Pure organic solvents provided the higher extraction efficiency than mixtures with water and similar efficiencies to mixtures of acetonitrile and methanol. The tests were based on UFP filter extractions with different solvents (100% methanol; 50% acetonitrile 50% methanol; 60% methanol 40% water; 60% acetonitrile 40% water), and evaluated based on the non-target-analysis-generated identifications and corresponding signal intensities.

2. In the experimental it is stated that “A circular section with a diameter: : .according to the array of the nozzles of each impactor stage”. Can you please clarify what this means? Were specific areas of the foil targeted?

For the extraction we cut out a section in the middle of the filters which is located below the array of nozzles of each impactor stage. By this approach we minimized the surface for extraction (e.g. the borders of the foil which were not exposed to particle deposition). We phrase this more clearly as follows:

Line 136-137: A circular section with a diameter of 2.5 cm was cut out of each foil sample located below the array of the nozzles of each impactor stage.

3. In the discussion of the UHPLC/HRMS method it is noted that the two standards that were tested had a small linear response range “likely due to their adsorptive behavior on glass surfaces”. If these are

representative of the types of molecules found in this work, how likely is it that the other chemicals may have been influenced by this as well? I recognize that quantification was not attempted for the other compounds, but I would suggest adding a note to this effect given that qualitative comparisons of peak areas were carried out.

We agree- it is possible that the other compounds are influenced by adsorption to glass surfaces as well. As we do not quantify and just refer to detected compounds, this effect should be negligible regarding the given results. Since we compare the intensities of the different base stock esters, we will add a note that presumably all ester compounds show a similar adsorption tendency as the used ester standard.

Line 183-184: Presumably, all the detected pentaerythritol esters and trimethylpropane esters show a similar adsorption tendency like the used ester standard.

4. In the results and discussion it is noted that “the majority of these compounds does not distinguish from the blank”. Does this mean that the majority of the compounds were also measured in the field blank? Were laboratory blanks also run and were these clean of the chemicals? Is this contamination occurring in the field, or is this coming during the sample handling/processing?

In the non-target approach it is common to use field blanks to compare with collected samples, and it is not uncommon that the majority of detected compounds emerges from the procedure (especially in organic trace analysis). We did not analyse sample preparation laboratory blanks, as many different analysis steps would have to be taken into account. We did measure with each run a solvent blank, but this serves only to characterize the state of the instrument. The field blanks are the ones that were handled in the same way like the UFP samples, and therefore should represent different possible sources of contaminants (e.g. from sampling, transport, storage, filter preparation- and extraction procedure, and instrument). All compounds which do not show a signal-to-blank ratio larger than five were filtered out, and we observed no significant accumulation of these compounds during sampling, sample preparation and measurement. As all different contaminant processes are represented by the field blanks, we cannot identify from which step each contaminant emerges.

5. It is noted that the program provides a false assignment for the pentaerythritol [sic] esters. On page 10 it is written that: “The native molecular fingerprint is displayed in Figure S4”. What does this mean? Are these the results using the false assignments? If so, why is this being shown? If not, please clarify what this means and what is being plotted in Figure S4.

Yes the fingerprint in Figure S4 is the depiction of the results using the original non-target software results. With this plot in the supplementary information we wanted to highlight that software-generated results should not be trusted blindly without further evaluation, especially as non-target analysis of HR-MS data is a growing discipline that is used by many PhD students who are likely not familiar with potential cluster formation in (+)ESI-MS.

As we corrected the results of the software-generated analysis, we wanted to provide full transparency by also showing the native result.

6. It is noted that the O/C ratios are below 0.6 and thus that the UFP do not become oxidized during transport. Please provide the O/C range for the starting material to support this (from Figures S6-S10).

We will add a note regarding the O/C range of the purchased jet engine oils.

Line 282-283: The analysed jet engine lubrication oils show O/C ratios within the same range.

7. On page 15: “Although no tri-ortho isomer of TCP was detected, it is still to consider that isomers with only one ortho-methyl group feature possibly a higher toxicity than the meta- and para-isomers: : :”. I am unsure what is being communicated here and suggest rephrasing.

As tricresyl phosphate (TCP) is composed of three aromatic moieties, six combinations regarding the position of the methyl groups are possible with at least one *ortho*-methyl group (o-m-m) (o-p-p) (o-m-p) (o-o-p) (o-o-m) (o-o-o). We will formulate the sentence more clearly.

Line 356-358: Although no tri-*ortho* isomer of TCP was detected, it is still to consider that isomers with only one *ortho*-methyl group feature possibly a higher toxicity than isomers having methyl groups only in *meta*- and *para*-position (Hanhela et al., 2005).

Minor comments:

8. Please add a note that this is positive ion mode in the caption for Figure 2 (unless it includes both positive and negative ion mode, in which case please clarify that).

Thank you for the remark we will add the ionization mode in the caption of Figure 2.

Figure 1. The sample-to-blank ratios of the detected compounds, averaged over all samples belonging to a certain size fraction (0.010-0.018 μm ; 0.018-0.032 μm ; 0.032-0.056 μm). Detection of the

compounds was accomplished in positive ionisation mode. The sample-to-blank ratio of 5 was specified for data filtering and is displayed as a dashed horizontal line.

9. There is a darker purple in Figure 3 A and D that is not present in Figure 3 B and C. I think this is just a shading issue, but I recommend correcting it so that all the colors match the key.

We have checked Figure 3 (A-D), every colored circle should be included in all plots. It is possible that a circle is visible in one plot (A) and not in another due to overlaying of isomers (appears naturally in B, C and D, while separated by chromatography in A). We did not use shading by creating the plots.