Shipborne measurements of Antarctic submicron organic aerosols: an NMR perspective linking multiple sources and bioregions

Stefano Decesari^{1*}, Marco Paglione¹; Matteo Rinaldi¹; Manuel Dall'Osto², Rafel Simó²; Nicola Zanca¹; Francesca Volpi¹; Maria Cristina Facchini¹; Thorsten Hoffmann³; Sven Götz³; Christopher Johannes Kampf³; Colin O'Dowd⁴; Jurgita Ovadnevaite⁴; Darius Ceburnis⁴; Emilio Tagliavini⁵.

¹Institute of Atmospheric and Climate Sciences, National Research Council of Italy (CNR), I-40129, Bologna, Italy.

²Institut de Ciències del Mar, Consejo Superior de Investigaciones Científicas (CSIC), ES-08003, Barcelona, Spain.

³Department of Chemistry, Johannes Gutenberg University of Mainz, 55128, Mainz, Germany.

⁴School of Physics and C-CAPS, National University of Ireland Galway, H91 CF50, Galway, Ireland.

⁵Department of Chemistry, University of Bologna, 40126, Bologna, Italy.

*corresponding author.

Keywords: Antarctic aerosols, natural sources of aerosol in polar regions, marginal ice zones, organic aerosol, NMR spectroscopy, primary marine particles, secondary organic aerosol, atmospheric amines

1 Abstract

2 The concentrations of submicron aerosol particles in maritime regions around Antarctica are influenced by 3 the extent of sea ice. This effect is two ways: on one side, sea ice regulates the production of particles by 4 sea spray (primary aerosols) while, on the other side, it hosts complex communities of organisms emitting 5 precursors for secondary particles. Past studies documenting the chemical composition of fine aerosols in 6 Antarctica indicate various potential primary and secondary sources active in coastal areas, in offshore 7 marine regions as well as in the sea ice itself. In particular, beside the well-known sources of organic and 8 sulfur material originating from the oxidation of dimethyl-sulfide (DMS) produced by microalgae, recent 9 findings obtained during the 2015 PEGASO cruise suggest that nitrogen-containing organic compounds are 10 also produced by the microbiota colonizing the marginal ice zone. To complement the aerosol source 11 apportionment performed using online mass spectrometric techniques, here we discuss the outcomes of 12 offline spectroscopic analysis performed by nuclear magnetic resonance (NMR) spectroscopy. In this study 13 we (i) present the composition of ambient aerosols over open ocean waters across bioregions, and 14 compared it to the composition of (ii) seawater samples and (iii) bubble bursting aerosols produced in a sea 15 spray chamber on board the ship. Our results show that the process of aerosolization in the tank enriches 16 primary marine particles with lipids and sugars while depleting them of free aminoacids, providing an 17 explanation for why aminoacids occurred only at trace concentrations in the marine aerosol samples 18 analyzed. The analysis of water-soluble organic carbon (WSOC) in ambient submicron aerosol samples 19 shows distinct NMR fingerprints for three bioregions: 1) the open Southern Ocean pelagic environments, in

which aerosols are enriched with primary marine particles containing lipids and sugars; 2) sympagic areas in the Weddell Sea where secondary organic compounds, including methanesulfonic acid and semivolatile amines abound in the aerosol composition; and 3) terrestrial coastal areas, traced by sugars such as sucrose, emitted by land vegetation. Finally, a new biogenic chemical marker, creatinine, was identified in the samples from the Weddell Sea, providing another confirmation of the importance of nitrogencontaining metabolites in Antarctic polar aerosols.

26

27 1. Introduction

28 The Antarctic continent is one of the last pristine areas of our planet but its natural ecosystems are now 29 threatened by an acceleration of the effects of global warming. Although at the beginning of the XXI 30 century the signals of climate change looked still weak in the region, the ice-sheet mass loss in Western 31 Antarctica has greatly accelerated in the last ten years along with an increasing warming of the Southern 32 Ocean (Shepherd et al., 2018). Climate change impacts on Antarctic maritime and coastal environments 33 may lead to stronger westerly winds, reduced summer sea ice extent, shifting geographical ranges of bird 34 communities, expanding terrestrial vegetation, increasing glacier melt and freshwater formation over land 35 (Rintoul et al.; 2018). As all these specific ecosystem impacts involve factors deemed to be important for 36 aerosol production in Antarctica (Davison et al., 2006; Schmale et al., 2013; Kyrö et al., 2013; Barbaro et al. 37 2017), significant climate change feedbacks on atmospheric concentrations of aerosols and cloud 38 condensation nuclei (CCN) are expected. The field studies performed in maritime and coastal areas around 39 Antarctica in the austral summer since the 90's (Davison et al., 1996) have provided precious information 40 on the contribution of the emissions from natural ecosystems to atmospheric composition. In summer, sea 41 ice recedes allowing wind stress over the oceanic surface and sea spray to occur closer to the continent, 42 hence increasing the primary marine aerosol production. At the same time, the thinning of sea ice in its 43 marginal zone and the increased intensity of solar radiation allow microalgae colonize the ice (Fryxell and 44 Kendrick, 1988; Roukaerts et al. 2016). The microbiota produce low-molecular weight metabolites as 45 cryoprotectors and osmoregulators, like dimethylsulfoniopropionate (DMSP) and quaternary nitrogen 46 compounds (Dallosto et al., 2017). Once released in seawater, such compounds become precursors of 47 atmospheric reactive volatile compounds, such as dimethylsulfide (DMS) and methylamines, which 48 eventually contribute to secondary aerosol formation. Davison et al. (1996) observed concentrations of 49 DMS south of 60°S more than four times higher than in the Atlantic Ocean. DMS and other reactive volatile 50 species are known precursors to secondary marine aerosol which contribute to the aerosol population in 51 the marine boundary layer together with primary sea-spray particles. Marine aerosols impact global climate 52 by reducing the amount of solar radiation reaching dark surface of the ocean, both directly (through 53 scattering) and indirectly (by modulating cloud formation and lifetime) (O'Dowd and de Leeuw 2007). In 54 polar regions, cloud seeding by marine aerosols transported over glaciated regions also affects the 55 longwave radiation budget (Willis et al. 2018).

56 During the 2015 PEGASO cruise (Dall'Osto et al., 2017, 2019; Fossum et al., 2018), we conducted 57 continuous atmospheric observations for over 42 days, providing one of the longest shipborne aerosol 58 measurement records in this area of the world. We contrasted the composition of seawater north and 59 south of the Southern Boundary of the Antarctic Circumpolar Current (SBACC), which represents the 60 approximate boundary between the Southern Ocean and the waters directly affected by sea ice formation 61 and melting around Antarctica. Dall'Osto et al. (2017) showed that not only DMSP and DMS occurred in 62 greater concentrations in sympagic waters (south of the SBACC), but so did quaternary nitrogen 63 compounds and methyl amines. By contrast, other biological parameters of seawater, like chlorophyll a, 64 total organic carbon (TOC) and transparent exopolymeric particles (TEP), showed higher concentrations in

the open Southern Ocean north of the SBACC. Results of bubble bursting experiments conducted on 65 66 nascent seawater as well as using melted sea ice showed that organic nitrogen and organic carbon were 67 more abundant in the aerosol in the latter case. Moreover, the production of organic-rich particles was 68 better traced by markers of the ice biota, such as mycosporines, than by macro-tracers of biological 69 productivity (chlorophyll). These results indicate that not only productivity per se but also the composition 70 and ecophysiological state of the microbiota affect the production of aerosol precursors in seawater. 71 Indeed, the observations of organic nitrogen in the aerosol – carried out by both online and offline 72 chemical methods - pointed to strong sources in the area of the Weddell Sea that, at the time of the field 73 campaign, was heavily covered by sea ice.

74 These findings contribute to the growing observational dataset of aerosol chemical compositions for 75 coastal Antarctic and sub-Antarctic marine areas. Past measurements relied on the analysis of filter and 76 impactor samples (Davison et al., 2006; Virkkula et al., 2006) and, more recently, on the application of 77 online aerosol mass spectrometric techniques (Zorn et al., 2008; Schmale et al., 2013; Giordano et al., 78 2017). All the chemical observations performed so far agree on showing a reduction of sea salt aerosol 79 concentrations from the Southern Ocean to the coasts of Antarctica, while secondary species including 80 non-sea salt sulfate and methanesulfonate (MSA) were found in relatively higher concentrations at higher 81 latitudes as a result of the DMS emissions from marginal ice zone waters. Open questions remain about a) 82 the amount of non-MSA organic matter in Antarctic air masses, and b) its origin (either primary or 83 secondary). Recent studies suggest that blowing snow at high wind speeds can be another important yet 84 hitherto underestimated source (Giordano et al, 2018; Frey et al., 2019), adding complexity onto the source 85 apportioning of organic aerosols. First observations of organic carbon (OC) in size-segregated aerosol 86 samples collected at a coastal site in the Weddell Sea (Virkkula et al., 2006) showed that MSA represented 87 only a few % of the total OC in the submicron fraction. In contrast with such findings, aerosol mass 88 spectrometric (AMS) measurements showed that the organic matter in submicron aerosols transported in 89 Antarctic air masses was almost totally accounted for by MSA, while non-MSA organic compounds were 90 associated to aerosols originating from highly productive waters in the Southern Ocean (Zorn et al., 2008). 91 Non-MSA OC was also associated to insular terrestrial biomass emissions (Schmale et al., 2013). In 92 particular, organic particles emitted from seabird colonies contain large amounts of nitrogen exhibiting MS 93 spectral fingerprints overlapping with those of natural aminoacids. In the paper by Liu et al. (2018), FTIR 94 spectroscopy was employed to probe the sources of particulate organic compounds at another coastal 95 Antarctic site, and the results point to a contribution of marine polysaccharides transported in sea spray 96 aerosols. Finally, detailed organic speciation using offline analytical techniques with high sensitivity and 97 selectivity suggest that OC concentrations are contributed by marine proteinaceous material, terrestrial 98 lipids and secondary organic compounds (Bendle et al 2007; Barbaro et al., 2015; 2017). It is unclear, 99 however, to what extent the concentrations of compounds occurring at pg m⁻³ relate to that of bulk organic 100 matter. We present here the organic characterization of Antarctic aerosol employing proton nuclear 101 magnetic resonance (¹H-NMR) spectroscopy. NMR spectroscopy has been used for decades in several fields 102 of biogeochemistry for its ability to fingerprint several classes of biomolecules and natural organic matter in 103 aquatic and terrestrial environments (e.g., Pautler et al., 2012; Hertkorn et al., 2013). In this study, which 104 focuses on the analysis of samples collected during the PEGASO 2015 cruise, we contrast the NMR 105 composition of submicron aerosol samples with that of seawater samples and bubble-bursting aerosols. 106 The results provide new hints on the origin of non-MSA aerosol organic matter in fine aerosol particles in 107 the Antarctic and sub-Antarctic marine environment.

108

109 2. Experimental

110 **2.1. Ambient aerosol sampling on filters.**

111 The PEGASO (Plankton-derived Emissions of trace Gases and Aerosols in the Southern Ocean) cruise was 112 conducted on board of RV Hesperides in the regions of Antarctic Peninsula, South Orkney and South 113 Georgia Islands from 2 January to 11 February 2015 (Dall'Osto et al. 2017). A high volume sampler (TECORA 114 ECO-HIVOL, equipped with Digitel PM₁ sampling inlet) collected ambient aerosol particles with Dp < 1 μ m on pre-washed and pre-baked quartz-fibre filters, at a controlled flow of 500 L min⁻¹. Sampling was allowed 115 116 only when the samplers were upwind the ship exhaust with a relative wind speed threshold of 5 m s⁻¹. Due 117 to the necessity of collecting sufficient amounts of samples for detailed chemical analyses, sampling time was of the order of ~50 h for each sample. A total of eight PM1 samples were collected during the cruise 118 119 (Figure 1). The samples were stored at -20 °C until extraction and NMR analysis. 120 For HPLC-MS analyses, aerosol samples were collected on PTFE fiber filters (70 mm diameter, Pallflex

T60A20, Pall Life Science) with flow rates of $2.31 - 2.41 \text{ m}^3/\text{h}$ through a PM_{2.5} inlet. Sampling times ranged from 12 - 24 h, resulting sampling volumes of $28.1 - 56.1 \text{ m}^3$ of air. As outlined above, sampling was only allowed when the sampler was upwind the ship exhaust.

125 **2.2. Seawater sampling and tank experiments.**

124

126 Seawater samples were collected from a depth of 4 m using either the uppermost Niskin bottle of the CTD 127 rosette casts or the ship's flow-through underway pumping system. The samples were filtered with a 128 Millipore filtration apparatus on quartz-fiber filters (Whatman, Ø = 47mm) after a previous cut off at 10 μ m 129 performed with a polycarbonate filter (Millipore, Isopore, porosity=10 μ m, Ø= 47mm). In total 45 samples 130 were collected for subsequent quantification of the Particulate Organic Carbon (POC) and 20 mL of the 131 filtrates were stored for subsequent analysis of Dissolved Organic Carbon (DOC). All the samples were 132 stored at -20 °C until the chemical analyses. Three samples of sea ice from the marginal ice zone in the 133 northern Weddell Sea were also collected using the methodology described in Dall'Osto et al. (2017). The 134 samples, once melted, were filtered and treated similarly to the seawater samples.

135 For the bubble bursting experiments, seawater was pumped through the same ship's pumping system to fill 136 an airtight high grade stainless steel tank (200 L) designed for aerosol generation experiment. The sea ice 137 samples were also introduced and melted in the tank for dedicated experiments. Water was dropped from 138 the top of the tank as a plunging jet at a flow rate of 20 LPM. The entrained air formed bubbles that, upon 139 bursting, produced sea-spray aerosol, as reported in O'Dowd et al. (2015). Particle-free compressed air was 140 blown into the tank headspace (120 L min⁻¹), which had outlet ports leading to samplers for the collection 141 of filters and the subsequent off-line chemical characterization of the produced sea-spray. In particular, 142 nine sea-spray aerosol samples were collected for approximately 72 h by a PM₁ sampler (flow rate 40 L 143 min⁻¹) equipped with pre-washed and pre-baked quartz-fiber filters (PALL, Ø = 47mm). In six cases, bubble 144 bursting experiments were conducted in the tank continuously flushed with fresh seawater. In the three 145 sea-ice experiments, instead, bubble bursting was carried out in a closed loop system because of the 146 limited amount of water volume available from the melted sea ice samples. In this case, the bubble 147 bursting process could lead to chemical and biological modifications in the samples like a progressive 148 depletion of surfactants on the film. Quantification of such artefacts is unavailable. Nevertheless, past 149 studies carried out in different geographical region of North East Atlantic but with the same apparatus 150 showed no evidence of decreasing organic enrichment in the generated sea spray when operated in a 151 closed loop system (O'Dowd et al., 2015).

Parallel bubble-bursting aerosol generation experiments with the same seawater and sea ice samples were carried out using a smaller glass tank (10 L) continuously flushed with particle-free air (11 L min⁻¹) (Schwier et al. 2015) and were dedicated to seaspray aerosol characterization using online mass spectrometers (HR-ToF-AMS and ATOFMS). The results from the bubble bursting experiments in the small tank are discussed in Dall'Osto et al. (2017). 158

159 **2.3.** ¹H-NMR spectroscopy.

Quartz-fiber filters from both ambient, POC filter samples and sea-spray generation experiments were 160 extracted with deionized ultra-pure water (Milli-Q) in a mechanical shaker for 1 h and the water extract was 161 162 filtered on PTFE membranes (pore size: 0.45 µm) in order to remove suspended particles. The watersoluble organic carbon (WSOC) content was quantified using a TOC-TN thermal combustion analyser (Multi 163 164 N/C 2100 by Analytik Jena) (Rinaldi et al., 2007). Aliquots of the aerosol extract were dried under vacuum and re-dissolved in deuterium oxide (D₂O) for organic functional group characterization by ¹H-NMR 165 spectroscopy, as described in Decesari et al. (2000). The ¹H-NMR spectra were acquired at 600 MHz in a 5 166 167 mm probe using a Varian Unity INOVA spectrometer, at the NMR facility of the Department of Industrial 168 Chemistry (University of Bologna). Sodium 3-trimethylsilyl-(2,2,3,3-d₄) propionate (TSP-d₄) was used as an 169 internal standard by adding 50 µL of a 0.05% TSP-d₄ (by weight) in D₂O to the standard in the probe. To 170 avoid the shifting of pH-sensitive signals, the extracts were buffered to pH \sim 3 using a deuterated-171 formate/formic-acid (DCOO⁻/HCOOH) buffer prior to the analysis. The speciation of hydrogen atoms bound 172 to carbon atoms can be provided by ¹H-NMR spectroscopy in protic solvents. On the basis of the range of 173 frequency shifts, the signals can be attributed to H-C containing specific functionalities (Decesari et al., 174 2000, 2007). A total of eight HiVol PM1 ambient aerosol samples (+ one blank), four POC samples from 175 seawater and two POC samples from melted sea ice, and three samples from the tank experiments (from 176 aerosolization of one seawater sample and two melted sea ice samples) + one blank for the 47mm filters 177 were characterized by NMR spectroscopy. 178

179 **2.4. UHPLC-HESI-Orbitrap-MS.**

One half of each filter sample was extracted according to the following protocol: three times sonication in 180 181 1.5 mL, 1 mL, and 1 mL ACN/H2O (9:1, v/v) for 30 min. The extracts were filtered through PTFE membranes (pore size: 0.45 μ m), combined, dried at 50 °C under a gentle stream of N₂, resuspended in 200 μ L 182 ACN/H2O (1:4, v/v), and stored at -20 °C until analysis. Samples were analyzed in triplicate by UHPLC-HESI-183 184 HRMS using an Orbitrap mass analyzer (Q-Exactive hybrid quadrupole orbitrap mass spectrometer, Thermo 185 Scientific, Germany) equipped with an UHPLC-System (Dionex UltiMate 3000 UHPLC system, Thermo 186 Scientific, Germany) and a Hypersil Gold, C18, 50 x 2.0 mm column with 1.9 µm particle size (Thermo 187 Scientific, Germany). The injection volume was 20 µL and the eluents were ultrapure water with 2% 188 acetonitrile and 0.04% formic acid (eluent A), and acetonitrile with 2% water (eluent B). The gradient of the mobile phase with a flowrate of 0.5 mL min⁻¹ was as follows: starting with 2% B isocratic for 1 min, 189 190 increasing to 20% B in 0.5 min, isocratic for 2 min, increasing to 90% B in 2.5 min, isocratic for 4 min and 191 decreasing to 2% B in 0.5 min. Mass spectrometric analyses were performed using a ESI source under the 192 following conditions: 30°C ESI temperature, 4 kV spray voltage, 40 psi sheath gas flow, 20 psi auxiliary gas 193 flow and 350°C capillary temperature. Mass resolution was 70000 and the acquired mass range was m/z 194 80–550. Creatinine calibration results are shown in Table S2 of the supplementary information.

196 **2.5. Air mass back-trajectories.**

195

197 Five-day back trajectories arriving at the ship's position at 03:00, 09:00, 16:00 and 21:00 every day were 198 calculated using the HYSPLIT model (Draxler & Rolph, 2010) with GDAS data. In total, 140 air mass back 199 trajectories were obtained. A Polar Stereographic map was used to classify 24 x 24 km grid cells as land, sea 200 and ice. From this information we calculated the percentage of time spent by each trajectory over each 201 surface type, and particularly over sea ice. Daily maps of sea ice percentage concentration measured on a 202 12.5 km grid were used for this calculation. Sea ice abundance was derived from satellite microwave data 203 (Ezraty et al., 2007) available at IFREMER. This analysis allowed also assigning air mass trajectories (and 204 percentages of surface type overflown) to the aerosol samples collected on the filters (Figure 1).

205

206 **3. Results**

207 **3.1. Organic composition of seawater: POC samples.**

208 The composition of seawater in terms of pigments, metabolites, fluorescent organic matter and other 209 organic constituents from the PEGASO cruise has been characterized in great detail (Dall'Osto et al., 2017; 210 Nunes et al., 2019; Zamanillo et al., 2019). Marine organic substances occur in the ocean in dissolved and 211 particulate form. Particulate organic carbon (POC) is defined operationally by a filtration cutoff at 0.45 μ m, 212 and recovers phytoplankton cells, bacteria and of the large colloids, such as transparent exopolymeric 213 particles ("TEPs") (Passow et al., 2002). Dissolved organic carbon (DOC) is mostly contributed by the excreta 214 and metabolites of the marine biota but it also accounts for a pool of refractory compounds, resistant to 215 microbial degradation, and well mixed in the water column (Hertkorn et al., 2013). Past studies have 216 extensively characterized the NMR features of labile and refractory organic constituents of marine organic 217 matter (Repeta 2015). However, the NMR characterization of the dissolved organic substances was limited 218 to desalted fractions of DOC isolated by solid-phase extraction or ultrafiltration (Koprivnjak et al., 2009). 219 Therefore, the NMR analysis of low-molecular weight polar organic constituents of marine DOC remains 220 elusive. In our study, we screened the NMR features of POC in phytoplankton bloom areas. In addition, 221 samples of bubble bursting aerosols generated from seawater and melted sea ice were used to obtain 222 chemical fingerprints for primary marine aerosol (Dall'Osto et al., 2017). During the process of bubble 223 bursting performed in the tank experiments, aerosol particles became depleted in sea salt with respect to 224 seawater and enriched in surface-active DOC components and in buoyant POC substances. The chemical 225 characterization of the smallest POC component (0.45 – 10 μ m) aims to provide information about 226 composition of the buoyant particles, while the contribution from DOC to the surface film composition 227 could not be determined in this study.

228 Figure 2 shows the proton NMR spectra of three POC samples, one from seawater (POC W3101) and two 229 from melted sea ice (POC Sealce-1, and POC Sealce-3) as examples. It is worth noting that the samples were 230 pre-filtered through a polycarbonate membrane with 10 µm porosity, hence the analyzed POC fraction is 231 representative for only the small particles (with diameters between ~ 0.45 and 10 μ m). During PEGASO, the 232 concentration of the fine POC fraction (0.45 – 10 μ m) ranged between 8 and 12 μ molC L⁻¹ in bloom areas. 233 The sub-set of samples analysed by ¹H-NMR spectroscopy exhibited a concentration of 10.6 \pm 0.7 μ molC L⁻¹ 234 (n = 4). The sample POC W3101 whose spectrum is shown in Figure 2 originates from the bloom area west 235 of South Georgia island, while the two sea ice samples were collected in the marginal ice zone of the 236 Weddell Sea. The interpretation of the ¹H-NMR spectra was carried out by comparison with NMR datasets 237 provided by the literature on metabolomics (e.g., Bertram et al., 2009; Matulova et al., 2014; Li et al., 2015; 238 Upadhyay et al., 2016) as well as by the analysis of commercial standard compounds. Characteristic 239 patterns of NMR resonances for specific compounds (e.g., patterns in multiplicity) enabled an accurate 240 identification, while only a tentative attribution of the most simple NMR resonances (singlets) was 241 attempted when standards were not available because deviations with respect to published NMR data are 242 always possible when different experimental conditions (e.g., different pH) are used. Nevertheless, the ¹H-243 NMR spectra of the POC extracts (Figure 2) show several NMR features overlapping with the typical ones 244 for other biological matrices. In particular, the occurrence of most common aliphatic aminoacids was 245 observed in all three samples analysed and particularly in sample POC Sealce-1. Acidic aminoacids 246 dominated over the basic ones, while aromatic residuals were detected only in trace amounts (Figure S1). 247 The identification of modified aminoacids among the most typical natural products of the Antarctic 248 microbiota, such as mycosporines (Oyamada et al., 2007), could not be carried out in detail because of the 249 lack of suitable spectral libraries. The presence of metabolytes such as low-molecular weight nitrogen-250 containing compounds (choline, betaine, etc.) is confirmed by the singlets in the chemical shift range 3.1 - 251 3.3 ppm from methyls bound to nitrogen atoms (H_3C -N-). Resonances at higher chemical shift, between 3.4 252 and 4.2, recovered the -NCHRCO- groups of alpha-aminoacids and the H-C-O groups of sugars and polyols: 253 traces of glycerol were found in all three samples analyzed, while glucose was found in trace amounts in 254 sample POC W3101 and, as a major component, in sample POC Sealce-3 (Figure S2). These results confirm 255 the potential of ¹H-NMR spectroscopy for the characterization of marine metabolites and natural products. 256 The small set of POC samples analyzed in this study is, however, mainly aimed to provide spectral 257 fingerprints useful for the interpretation of the results of the aerosol sample analyses discussed in the 258 following sections.

259

260 **3.2. Organic composition of bubble bursting aerosols.**

261 The three primary marine aerosol samples collected in the 200 L tank and analysed by ¹H-NMR 262 spectroscopy included the following samples. One sample was collected from bubble bursting of nascent 263 seawater (BB W1101) obtained during almost four days of navigation west and north of the South Orkney 264 Islands with seawater continuously flushed onboard the RV maintaining continuous sea spray production in 265 the tank. The other two samples (BB Sealce-1 and BB Sealce-3) were obtained from two of the three sea ice 266 samples melted in the tank and run in a closed loop system. Sea ice was collected from the marginal ice 267 zone around 100 km south of the South Orkneys. The chemical information obtained for these bubble 268 bursting aerosols is, therefore, representative for primary marine particles in the northern sector of the 269 Weddell Sea. The natural process of sea spray – mimicked by the experiments carried out in the tank 270 onboard RV Hesperides- selectively transfers organic compounds from seawater into the aerosol 271 depending on the ability of the specific pools of organic substances to enrich in the surface microlayer 272 and/or to be scavenged by rising air bubbles. The selective nature of such process is witnessed by our NMR 273 data, showing that the seawater composition dominated by aminoacids, osmolytes and sugars/polyols 274 (Figure 2) differs quite substantially from that of bubble bursting aerosols from the tank experiments 275 (Figure 3, Figure S3). Bubble bursting aerosol was characterized by the occurrence of low-molecular weight 276 metabolites like lactic acid and amines (dimethylamine, DMA and traces of monomethyl- and trimethyl-277 amines) which likely originated from DOC components of seawater. The most characteristic feature of the 278 spectra is, however, the bands at 0.9 and 1.3 ppm of chemical shift. These correspond to aliphatic chains 279 with terminal methyl moyeties typical of lipids. Their occurrence in the aerosolized seawater and not in the 280 POC samples can be explained by an enrichment of surface-active compounds from DOC in the surface 281 microlayer. Lipid enrichment in the aerosol during past bubble bursting experiments was reported by Facchini et al. (2008) and Schmitt-Kopplin et al. (2012). Nevertheless, our findings clearly show that, beside 282 283 lipids, there are specific constituents of POC taking part in the formation of primary aerosol particles in the 284 tank experiments. In particular, the spectral region for sugars and polyols in bubble bursting aerosols is 285 completely consistent with the spectral features of POC (Figure S4), although the contribution of the -286 NCHRCO- groups of aminoacids in the same spectral window is clearly missing in the aerosol. The presence 287 of nitrogen-containing metabolytes (betaine) is confirmed in the aerosol samples from the tank. It is 288 plausible that betaine, glycerol and other sugars are chemically bond to lipids (glycolipids and 289 phospholipids) which can explain their preferential enrichment during the aerosolization process with 290 respect to other POC constituents like the aminoacids. It is a matter of fact that aminoacids could be 291 detected only in very trace amounts (the doublet of alanine at 1.45 ppm is barely visible) in the sea spray 292 samples. Other molecular tracers found in previous sea-spray experiments in other geographical regions, 293 such as acrylic acid (Schmitt-Kopplin et al., 2012), which is also product of DMSP degradation, were not 294 found in our experiment.

295

3.3. Organic composition of ambient submicron WSOC samples.

297 The eight ambient PM₁ HiVol samples analyzed for organic composition include six that were collected in 298 parallel to the impactor samples discussed in Dall'Osto et al. (2017). The proton NMR spectra of the eight 299 samples are reported in Figures S5-S7. Air mass origin varied largely during the cruise, with transport from 300 the Weddell sea prevalent during the first half of the cruise turning into open ocean prevailing air masses 301 during the second half (Fig. 1). Two samples (A-0701 and A-0102) of mixed origin had been omitted by 302 Dall'Osto et al (2017), who focused on the comparison between aerosols from the sympagic regions and 303 those from the open ocean. We applied hierarchical cluster analysis to investigate if a dual classification 304 also held with the ¹H-NMR spectra (Figure 4). The original spectra were normalized to their integrals and 305 binned to 354 points before clustering. Two main clusters were indeed identified: a first one recovering 306 three samples collected downwind the Weddell Sea during the first half of the cruise, and a second cluster 307 with samples representative of a greater diversity of conditions, from the Drake Channel, to the Antarctic 308 Peninsula and to the productive waters around South Georgia. This second cluster corresponds to the 309 samples characteristic for the open ocean conditions in Dall'Osto et al. (2017) plus samples A-0701 and A-310 0102. Unexpectedly, sample A-0701, whose air mass spent most of time over sympagic waters (Fig. 1) 311 clustered together with the samples from the open ocean according to NMR composition. It is noticeable, 312 however, that binned NMR spectra can only trace the distribution of the major organic functional groups 313 while the information carried by fine spectral features, which is critical to detect the presence of specific 314 molecular markers, is not accounted for by the cluster analysis. In the following sections, we will show that 315 sample A-0701 exhibits a peculiar NMR composition which must be put in relation to terrestrial sources of 316 organic compounds. On the basis of the back-trajectories (Figs. 1 and 8), the likely land sources were 317 located in the Antarctic Peninsula. In summary, the variability in the distribution of NMR functional groups 318 in ambient PM_1 samples (Table 1) was primarily driven by the air mass origin over sympagic (Weddell Sea) 319 or pelagic waters, in agreement with the results on inorganic compounds, WSOC and amines reported by 320 Dall'Osto et al. (2017; 2019). Nevertheless, the analysis of fine NMR spectral features supports the 321 existence of a third source area over land. In the following discussion, we will provide an in-depth 322 description of the NMR compositions for these three source sectors.

324 **3.3.1.** Ambient aerosols from the Weddell Sea.

323

325 Sample A-0901 was collected in the marginal ice zone of the Weddell Sea. It spectrum exhibits no signals 326 from aromatic compounds or alkenes (Figure S7). The aliphatic region (Figures 5, S8) exhibits broad 327 similarity to that of the primary marine particles generated in the sea spray tank, but with a major 328 difference in the chemical shift range between 1.7 and 3.0 ppm where the background broad NMR bands 329 are much more intense in the ambient sample. This is also the region recovering the signals from acyl 330 groups (RCH-(C=O)-) in aliphatic carboxylic acids and ketoacids, which are formed by VOC oxidation in the 331 atmosphere (Barbaro et al., 2017). The most abundant individual compounds detected in these samples 332 were, however, MSA (Fossum et al., 2018) and the low-molecular methylamines (MMA, DMA, TMA). The 333 predominance of semivolatile C₁-C₃ alkyl-amines (Ge et al., 2011) indicates that amines become enriched in 334 the ambient aerosol thorough volatilization from the ocean surface and recondensation onto acidic aerosol 335 particles (Dall'Osto et al., 2019). The aliphatic bands at 0.9 and 1.3 ppm in sample A-0901 show a partial 336 overlap with the resonances of the lipids in the aerosolized seawater. However, the bands at 1.6 ppm and 337 2.2-2.3 ppm which, in lipids, correspond to methylenes in beta and alpha position to a C=O group, are much 338 more intense in the spectrum of A-0901 than in BB Sealce-3 (Figure S8), indicating that aliphatic chains are 339 shorter and more substituted in the ambient aerosol than in nascent primary aerosol particles. The pattern 340 of bands at 0.9, 1.3, 1.6, 2.2, 2.4 and 2.6 ppm follow the structure elucidated by Suzuki et al. (2001) and 341 attributed to C7-C9 aliphatic dicarboxylic acids and oxo-acids. This class of organic compounds, clearly 342 characterizing the aliphatic composition of the ambient samples in the Weddell Sea area, can originate 343 from degraded (oxidized) lipids (Kawamura et al., 1996) or from gas-to-particle conversion of carbonyls 344 produced by the photochemical oxidation of lipids at the air-sea interface (Bernard et al., 2016; Alpert et 345 al., 2017). Support to the latter hypothesis (secondary formation) is given by the fact that the N-osmolytes 346 (betaine, choline) present in the sea spray generated in the tanks were completely absent in the ambient 347 sample. Nevertheless, the resonances in the spectral window 3.5 - 3.8 ppm in sample A-0901 are 348 completely consistent with the occurrence of glycerol, indicating that in fact primary aerosol particles 349 contributed to the composition of the ambient aerosol in this region (Figure S9). There is another one 350 striking difference between the composition of the ambient aerosol and sea spray particles: the former 351 contains significant levels (1.65 ng m⁻³) of creatinine. This compound is responsible to the two singlets at 352 3.12 ppm and 4.27 ppm of chemical shift and was identified by the comparison with a standard under 353 identical NMR experimental conditions (Figure S11). The concentration of creatinine clearly follows that of 354 low-molecular weight amines (Figure 6) and shows a maximum in the three samples collecting most of the 355 air masses that travelled over the Weddell Sea. Creatinine was also determined by HPLC/-MS analysis in a 356 parallel set of filter samples collected onboard Hesperides during the PEGASO cruise (see Section 2.4). 357 Identification was based on MS/MS fragmentation patterns and retention time. Quantification was based 358 on chromatographic peak area. Figure 7 shows extracted ion chromatograms for m/z 114.0655-114.0667, 359 corresponding to creatinine, of the filter extract of sample 0119N obtained during the PEGASO campaign 360 and the neat creatinine standard. The HPLC/MS analysis indicate that creatinine occurred in concentrations 361 of $20 - 50 \text{ pg/m}^3$ in the samples from the Weddell Sea area (Table S1), much less than the concentrations determined by ¹H-NMR spectroscopy ($1.6 - 2.5 \text{ ng/m}^3$, Table 1). Such discrepancy can be due to the 362 363 different extraction protocols and to non-ideal chromatographic conditions in HPLC/MS for creatinine 364 quantification (elution close to the void volume). Nevertheless, our findings demonstrate that high-field 365 NMR methods can integrate HPLC/MS analysis for the identification of molecular markers in atmospheric 366 aerosol complex organic mixtures.

367

368 **3.3.2. Ambient aerosols in the open ocean.**

Sample A-2401 was collected during the northern transit of the cruise, RV Hesperides just west to South 369 370 Georgia (55° S) (Figure 1). During sampling, the air masses had a westerly component and can be 371 considered representative of Southern Ocean conditions. The ¹H-NMR spectrum of A-2401 shares 372 similarities with that of A-0901 described above: a) the resonances of MSA and methyl-amines are much 373 more intense than that of other low-molecular weight compounds (such as N-osmolytes); b) the spectral 374 region of acyls (1.8 - 3.0 ppm) accounting for unresolved carboxylic acids is clearly more intense than in the 375 spectrum of primary organic aerosols; c) the pattern of bands at 0.9, 1.3, 1.6 and 2.2-2.4 ppm highlights the 376 presence of linear aliphatic structures substituted with oxo- and carboxylic groups. Nevertheless, MSA and 377 the low molecular weight amines were less abundant in A-2401 than in the sample from the Weddell Sea 378 (Table 1). Also the ratio between acyl (C \underline{H} -C=O) and alkyl (C \underline{H} -CH) groups was smaller in A-2401 than in A-379 0901 (Figure S8). The linear aliphatic structures involved longer methylenic chains in A-2401 than in A-0901, 380 so that in the former case they were more similar to the aliphatic structures of the aerosolized melted sea 381 ice (Figure S8). Another difference between the two ambient aerosol samples is that the one from the 382 Southern Ocean contains much more alcoxy groups (HC-O, in the chemical shift range 3.4 – 4.2 ppm) of 383 polyols than the one from the Weddell Sea (Figure 5; Table 1). When comparing the functional group 384 distributions of the ambient aerosol samples to that of the aerosol generated during the tank experiments, 385 clearly the samples from the Southern Ocean show a better match than the samples from the Weddell Sea 386 do. Other similarities between the composition of A-2401 and the aerosol in the tank can be found in the 387 fine structures of the spectra, especially in the ranges of aromatics, acetals and polyols (Figure S12). A-2401 388 clearly contains traces of organic markers of primary aerosols and specifically glycerol, N-osmolytes (Figure 389 S10) and aminoacids (alanine). Finally, contrary to A-0901, sample A-2401 contains only trace amounts of 390 creatinine.

391

392 3.3.3. Ambient aerosols influenced by coastal land sources.

393 Sample A-0701 was collected in the western sector of the Weddell Sea. The air masses showed several overpasses on the Antarctic Peninsula. The ¹H-NMR spectrum shows unique features: isobutyric acid was 394 395 found in relatively high concentrations, together with an amine tentatively identified as cadaverine (Figure 396 5). The aliphatic chains occur in much lower amounts than in the samples described above, the band of 397 acyls is not as pronounced as in A-0901 (Figure S8), whereas alcoxyls are abundant, especially due to the 398 occurrence of sucrose at a remarkable concentration of 10 ng/m³. Finally, no creatinine was found in this 399 sample. Clearly, the composition of A-0701 is drastically different from that of the other samples collected in the Weddell Sea. The presence of sucrose (Figure S9) points to a contribution from primary biological 400 401 particles emitted from a terrestrial biota, not a marine one. Vegetation cover (scarce but existing) in the 402 Antarctic Peninsula can be responsible for such emissions. The NMR composition of A-0701 provides 403 evidence of the diversity of biogenic aerosol sources active in this area of the world.

404

405 4. Discussion

406 **4.1** Source apportioning of primary and secondary organic components in different regions

407 The comparison of the NMR compositions of the ambient aerosol samples collected onboard RV Hesperides 408 (Figure 8) supports the distinction of aerosol sources between the sympagic and pelagic environments 409 already introduced by Dall'Osto et al. (2017). The higher abundance of alkyl (C-H) and alcoxy (H-C-O) groups 410 detected in the second half of the cruise points to a larger fraction of primary organic compounds rich in 411 lipids and polyols in the aerosols of the open Southern Ocean. Analogous compositions were obtained 412 using FTIR spectroscopy at Ross Island (Liu et al. 2018). In our study, the attribution of compound classes 413 and molecular markers (such as glycerol and N-osmolytes) to primary marine particles was supported by 414 the comparison with the analysis of tank-generated sea-spray particles. According to our NMR datasets, 415 primary marine organics were ubiquitous in the region as witnessed by the presence of glycerol in all 416 samples. However, glycerol accounted for almost the entire polyol content in the three samples from the 417 eastern/north Weddell Sea, while the samples from the open ocean contained much larger and more 418 complex mixtures of polyols/sugars. Sub-ng/m³ levels of free aminoacids (alanine) and trace amounts of N-419 osmolytes along with the greater abundance of linear aliphatic structures similar to lipids in the samples 420 from the Southern Ocean point to a major contribution of primary organics to submicron organic aerosols 421 in this environment. These findings provide further confirmation to the importance of sea spray as a source 422 of marine organic particles in oceanic regions characterized by high productivity and strong wind stress.

423

424 In sympagic waters, other mechanisms of aerosol formation take place. Sympagic waters are rich in S- and 425 N- osmolytes produced by the algal communities colonizing the sea ice. The osmolytes degrade to VOCs 426 which are then converted to SOA components, such as MSA (Davison et al., 1996) and low-molecular 427 weight methyl-amines (Facchini et al., 2008). Further differences between the two regimes of aerosol 428 formation are visible in the distribution of the oxygenated functional groups. If alcoxyl groups (H-C-O) from 429 polyols and sugars accounted for almost 50% of total alcoxyl (H-C-O) and acyls (H-C-C=O) in the samples 430 from the Southern Ocean, such fraction decreased to less than 30% in the three samples from the offshore 431 areas of the Weddell Sea (Fig. 8). The mixtures of organic compounds carrying acyls, like carboxylic and 432 oxocarboxylic acids, are not associated to primary marine aerosols and are likely components of SOA. 433 Carboxylic acids can form photochemically (Cui et al., 2019) during the austral summer. The nature of 434 parent VOCs for carboxylic acids in our samples is unknown, but the occurrence of linear aliphatic 435 compounds containing oxo- and carboxylic groups indicates that one of the possible sources stands in the 436 oxidative degradation of lipids - either in the aerosol or in the marine microlayer - as suggested by past studies in Antarctica (Kawamura et al., 1996) and consistent with recent AMS observations in the Arcticmarginal ice zone (Willis et al., 2017).

439 In the Weddell Sea, under the influence of air masses that had travelled over the Peninsula (sample A-440 0701), the contribution of the emissions from the land biota became evident, therefore supporting the 441 observations of Schmale et al. (2013) on the contribution of primary biological particles from the coastal 442 land ecosystems. Our data suggest that beside animal colonies, also the land vegetation (grasses, mosses, 443 lichens) of the Antarctic Peninsula can contribute to primary aerosols emission and to their sugar content. 444 Other biological compounds of primary origin, the aminoacids, were not found in the Weddell Sea in our 445 study. These results contrast with the previous findings that a significant fraction of the ambient PM₁ mass 446 was accounted for by proteinaceous material at an island site in the Southern Ocean (Schmale et al., 2013). 447 On the other hand, the observations of Schmale et al. (2013) were carried out under the direct influence of 448 the emissions of seabird colonies, while our observations were carried out offshore. More research is 449 needed to quantify the range and extent to which primary particles from the terrestrial biota impact the 450 marine aerosol composition in the Antarctic region.

451

452 **4.2 A new potential marker: creatinine.**

453 The sources of creatinine in the ambient aerosol is controversial. On the basis of its chemical structure, it is 454 water-soluble but clearly less volatile than the methyl-amines and, as a consequence, its Henry coefficient 455 must be much less favorable for transferring this amine out of seawater into the gas phase. A primary origin 456 via sea spray is also doubtful because creatinine is not a strong surfactant. On the other hand, Prather et al. 457 (2013) showed that sea-spray aerosols encompass several classes of organic particles, including some made 458 of biological material: POC particles and large colloids can be scavenged by rising bubbles and injected in 459 the atmosphere by jet drops. Jet drop emission represents a plausible mechanism to transfer primary 460 organic compounds which are not strong surfactants from seawater to the atmosphere. If this is the 461 relevant pathway for creatinine, it must have occurred in source areas other than the algal blooms where 462 we conducted the tank experiments, since we did not detect any creatinine in the aerosolized sea water 463 and sea ice. Creatinine is a common metabolite of mammals, therefore an alternative source via the 464 excreta of sea lions in Antarctic coastal areas can be postulated. However, a much more vast potential 465 source in seawater is also possible under the hypothesis that creatinine results from the enzymatic 466 conversion of creatine, which is a known metabolite of the urea cycle in marine animals (Whitledge and 467 Dugdale, 1972) and phytoplankton (Allen et al., 2011) that contributes to pelagic DOC across the world's 468 oceans (e.g., Wawrik et al., 2017).

- 469
- 470

471 **5. Conclusions**

472 Our results demonstrate that, beside MSA, a complex mixture of biogenic organic compounds contributes 473 to the composition of submicron aerosol particles in the Antarctic atmosphere. Although individual organic 474 markers encompassing sugars, aminoacids and carboxylic acids have already been identified in past studies, 475 our results indicate that non-MSA biogenic organic compounds impact the bulk composition of organic 476 aerosol in this environment (Figure 8). ¹H-NMR analysis provides evidence for both secondary (more 477 important in sympagic regions) and primary marine (more important in pelagic areas) sources. A third 478 contribution from the terrestrial biota in the Antarctic Peninsula was also identified. The emission of sea-479 spray organics in offshore areas was unambiguously demonstrated by the determination of molecular 480 tracers for lipids and polyols and by the comparison of the fine structures in the ¹H-NMR spectra of the 481 ambient samples and of the aerosol generated in the tank experiments. A new biogenic marker, creatinine, 482 was identified for the first time in the ambient aerosol, extending the list of reduced nitrogen containing 483 molecular tracers in the atmosphere. The discovery of creatinine also exemplifies the usefulness of

- 484 employing non-targeted analytical techniques like NMR spectroscopy for screening the organic composition
- of the aerosol in remote environments where the sources of atmospheric particulate matter are still poorly
- 486 known. The complexity of the organic composition illustrated in this study calls for more research on
- 487 suitable methodologies both online and offline and combinations of them to investigate the nature of
- 488 non-MSA marine organic particles in off-shore regions around the Antarctic continent.
- 489

490 Data Availability

- 491 The NMR data sets are available on request to the corresponding author.
- 492

493 Author Contribution

494 SD wrote the paper; MDO and RS coordinated the experimental activities in the field; MP, MDO and SG 495 collected the aerosol samples; MDO, MP and DC collected the sea ice samples; COD, JO and DC set up the 496 bubble bursting tank; MR, MP, MR, NZ and FV performed the sample extraction and preparation for WSOC 497 and NMR analysis; NZ and MP performed the NMR analyses; SG and CJK carried out the HPLC/MS analyses; 498 SD, MP and ET elaborated the NMR data; MDO, RS and ET contributed to the interpretation of the analyses 499 of the seawater samples; SD, MP, MR, MDO, TH, CJK and ET contributed to the interpretation of the 500 analyses of the aerosol samples; all authors contributed to the general discussion and to work out the main 501 conclusions of this study.

502

503 Competing interests

- 504 The authors declare that they have no competing interests.
- 505

506 Acknowledgments

507 The cruise was funded by the Spanish Ministry of Economy through projects PEGASO (CTM2012-37615) and 508 Bio-Nuc (CGL2013-49020-R). The research leading to these results has received funding from the European 509 Union's Seventh Framework Programme (FP7/2007-2013) Project BACCHUS under Grant Agreement 503445. The research activities of CNR were also supported by the project AirSEaLab: Progetto Laboratori 510 Congiunti. We would like to thank Prof. Andrea Mazzanti for his advice in performing the NMR experiments 512 at the NMR facility of the Dep. Industrial Chemistry, University of Bologna. We also thank Dr. David 513 Beddows (Uni. Birmingham) for help in drawing figures, in particular air mass back trajectories.

514

515 References

Allen A.E., Dupont, C.L., Obornik, M., Horak, A., Nunes-Nesi, A., McCrow, J.P., Zheng, H., Johnson, D.A., Hu,
H., Fernie, A.R., Bowler, C.: Evolution and metabolic significance of the urea cycle in photosynthetic
diatoms, Nature, 473, 203-209, 2011.

Alpert, P. A., Ciuraru, R., Rossignol, S., Passananti, M., Tinel, L., Perrier, S., Dupart, Y., Steimer, S. S.,
Ammann, M., Donaldson, D. J., George, C.: Fatty Acid Surfactant Photochemistry Results in New Particle
Formation, Scientific Reports, 7, 12693, 2017.

- Barbaro E., Zangrando, R., Vecchiato, M., Piazza, R., Cairns, W. R. L., Capodaglio, G., Barbante, C., Gambaro,
 A.: Free amino acids in Antarctic aerosol: potential markers for the evolution and fate of marine aerosol,
 Atmospheric Chemistry and Physics, 15, 5457–5469, 2015.
- Barbaro, E., Zangrando, R., Padoan, S., Karroca, O., Toscano, G., Cairns, W.R.L., Barbante, C., Gambaro, A.:
 Aerosol and snow transfer processes: An investigation on the behavior of water-soluble organic compounds
 and ionic species, Chemosphere, 183, 132 138, 2017.
- 528 Bendle, J., Kawamura, K., Yamazaki, K., Niwai, T.: Latitudinal distribution of terrestrial lipid biomarkers and 529 n-alkane compound-specific stable carbon isotope ratios in the atmosphere over the western Pacific and 530 Southern Ocean, Geochimica Cosmochimica Acta, 71, 5934–5955, 2007.
- Bernard, F., Ciuraru, R., Boréave, A., George, C.: Photosensitized Formation of Secondary Organic Aerosols
 Above the Air/Water Interface. Environmental Science and Technology, 50, 8678–8686, 2016.
- Bertram, H. C., Duus, J.Ø., Petersen, B. O., Hoppe, C., Larnkjaer, A., Schack-Nielsen, L,, Mølgaard, C.,
 Michaelsen, K. F.: Nuclear magnetic resonance–based metabonomics reveals strong sex effect on plasma
 metabolism in 17-year–old Scandinavians and correlation to retrospective infant plasma parameters,
 Metabolism Clinical and Experimental, 58, 1039–1045, 2009.
- Cui, T., Green, H. S., Selleck, P. W., Zhang, Z., O'Brien, R. E., Gold, A., Keywood, M., Kroll, J. H., Surratt, J. D.:
 Chemical characterization of isoprene- and monoterpene-derived secondary organic aerosol tracers in
 remote marine aerosols over a quarter century, ACS Earth Space Chem., 3, 935–946, 2019.
- Dall'Osto, M., Ovadnevaite, J., Paglione, M., Beddows, D. C. S., Ceburnis, D., Cree, C., Cortés, P., Zamanillo,
 M., Nunes, S. O., Pérez, G. L., Ortega-Retuerta, E., Emelianov, M., Vaqué, D., Marrasé, C., Estrada, M.,
 Montserrat Sala, M., Vidal, M., Fitzsimons, M. F., Beale, R., Airs, R., Rinaldi, M., Decesari, S., Facchini, M. C.,
 Harrison, R. M., O'Dowd, C., Simó, R.: Antarctic sea ice region as a source of biogenic organic nitrogen in
 aerosols, Scientific Reports, 7, 6047, doi:10.1038/s41598-017-06188-x, 2017.
- Dall'Osto, M., Airs, R. L., Beale, R., Cree, C., Fitzsimons, M. F., Beddows, D., Harrison, R. M., Ceburnis, D.,
 O'Dowd, C., Rinaldi, M., Paglione, M., Nenes, A., Decesari, S., Simó, R.: Simultaneous detection of
 alkylamines in the surface ocean and atmosphere of the Antarctic sympagic environment, ACS Earth Space
 Chem., 3, 5, 854-862, 2019.
- Davison, B., Hewitt, C. N., D. O'Dowd, C., Lowe, J. A., Smith, M. H., Schwikowski, M., Baltensperger, U.,
 Harrison, R. M.: Dimethyl sulfide, methane sulfonic acid and physicochemical aerosol properties in Atlantic
 air from the United Kingdom to Halley Bay, J. Geophys. Res., 101, 22855 22867, 1996.
- 552 Decesari, S., Facchini, M. C., Fuzzi, S., Tagliavini, E.: Characterization of water-soluble organic compounds in 553 atmospheric aerosol: a new approach, J. Geophys. Res., 105, 1481 – 1489, 2000.
- Decesari, S., Mircea, M., Cavalli, F., Fuzzi, S., Moretti, F., Tagliavini, E., Facchini, M. C.: Source attribution of
 water-soluble organic aerosol by Nuclear Magnetic Resonance spectroscopy, Environ. Sci. Technol, 41, 2479
 2484, 2007.
- Draxler, R. R. & Rolph, G. D.: HYSPLIT (HYbrid Single-Particle Lagrangian Integrated Trajectory) Model access
 via NOAA ARL READY Website (<u>http://ready.arl.noaa.gov/HYSPLIT.php</u>), 2010.
- Ezraty, R., Girard-Ardhuin, F., Piollé, J.F., Kaleschke, L. & Heygster, G. Arctic and Antarctic sea ice
 concentration and Arctic sea ice drift estimated from Special Sensor Microwave data. Département
 d'Océanographie Physique et Spatiale, IFREMER, Brest, France and University of Bremen Germany, 2.1edn,
 2007.

Facchini, M. C., Decesari, S., Rinaldi, M., Carbone, C., Finessi, E., Mircea, M., Fuzzi, S., Moretti, F., Tagliavini,
E., Ceburnis, D., O'Dowd, C. D.: Important Source of Marine Secondary Organic Aerosol from Biogenic
Amines, Environmental Science and Technology, 42, 9116 – 9121, 2008.

Fossum, K. N., Ovadnevaite, J., Ceburnis, D., Dall'Osto, M., Marullo, S., Bellacicco, M., Simó, R., Liu, D.,
Flynn, M., Zuend, A., O'Dowd, C.: Summertime primary and secondary contributions to Southern Ocean
cloud condensation nuclei, Scientific Reports., 8, 13844, 2018.

- Frey, M. M., Norris, S. J., Brooks, I M., Anderson, P. S., Nishimura, K., Yang, X., Jones, A. E., Nerentorp
 Mastromonaco, M. G., Jones, D H., Wolff, E. W.: First direct observation of sea salt aerosol production from
 blowing snow above sea ice, Atmos. Chem. Phys. Discuss., doi: 10.5194/acp-2019-259, 2019.
- 572 Fryxell, G. A. and Kendrick, G. A.: Austral spring microalgae across the Weddell Sea ice edge: spatial 573 relationships found along a northward transect during AMERIEZ 83. Deep-sea Res. 35, 1-20., 1988.
- 574 Ge, X., Wexler, A. S., Clegg, S. L.: Atmospheric amines Part II. Thermodynamic properties and gas/particle 575 partitioning, Atmos. Environ., 45, 561-577, 2011.
- 576 Giordano, M. R., Kalnajs, L. A., Avery, A., Goetz, J. D., Davis, S. M., DeCarlo, P. F.:A missing source of 577 aerosols in Antarctica – beyond long-range transport, phytoplankton, and photochemistry, Atmospheric 578 Chemistry and Physics, 17, 1–20, 2017.
- Hertkorn, N., Harir, M., Koch, B.P., Michalke, B., Schmitt-Kopplin, P.: High-field NMR spectroscopy and
 FTICR mass spectrometry: powerful discovery tools for the molecular level characterization of marine
 dissolved organic matter, Biogeosciences, 10 (3), 1583–1624, 2013.
- 582 Kawamura, K., Seméré, R., Imai, Y., Fujii, Y., Hayashi, M.: Water soluble dicarboxylic acids and related 583 compounds in Antarctic aerosols, J. Geophys. Res., 101, 18721 – 18728, 1996.
- Koprivnjak, J. F., Pfromm, P. H., Ingall, E., Vetter, T. A., Schmitt-Kopplin, P., Hertkorn, N., Frommberger, M.,
 Knicker, H., Perdue, E. M.: Chemical and spectroscopic characterization of marine dissolved organic matter
 isolated using a coupled reverse osmosis-electrodialysis. Geochim. Cosmochim. Acta 73, 4215 4231, 2009.
- 587 Kyrö, E.-M., Kerminen, V.-M., Virkkula, A., Dal Maso, M., Parshintsev, J., Ruíz-Jimenez, J., Forsström, L., 588 Manninen, H. E., Riekkola, M.-L., Heinonen, P., Kulmala, M.: Antarctic new particle formation from 589 continental biogenic precursors, Atmosheric Chemistry and Physics, 13, 3527–3546, 2013.
- Li, S., Winters, H., Villacorte, L.O., Ekowati, Y., Emwas, A. -H. M., Kennedy, M.D., Amy, G. L.: Compositional
 similarities and differences between transparent exopolymer particles (TEPs) from two marine bacteria and
 two marine algae: Significance to surface biofouling, Marine Chemistry, 174, 131–140, 2015.
- Liu, J., Dedrick, J., Russell, L. M., Senum, G. I., Uin, J., Kuang, C., Springston, S. R., Leaitch, W. R., Aiken, A. C.,
 Lubin, D.: High summertime aerosol organic functional group concentrations from marine and seabird
 sources at Ross Island, Antarctica, during AWARE, Atmos. Chem. Phys., 18, 8571–8587, 2018.
- Matulová, M., Husárová, S., Capek, P., Sancelme, M., Delort, A. M.: Biotransformation of Various
 Saccharides and Production of Exopolymeric Substances by Cloud-Borne Bacillus sp. 3B6, Environ. Sci.
 Technol., 48, 14238–14247, 2014.
- Nunes, S., Latasa, M., Delgado, M., Emelianov, M., Simó, R., Etrada, M., Phytoplankton community
 structure in contrasting ecosystems of the Southern Ocean: South Georgia, South Orkneys and western
 Antarctic Peninsula, Deep-Sea Res. I, 151, 103059, doi: 10.1016/j.dsr.2019.06.005, 2019.

- O'Dowd, C. D. & de Leeuw, G., Marine aerosol production: a review of the current knowledge. Philosophical
 Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences 365, 1753, 2007.
- O'Dowd, C., Ceburnis, D., Ovadnevaite, J., Bialek, J., Stengel, D. B., Zacharias, M., Nitschke, U., Connan, S.,
 Rinaldi, M., Fuzzi, S., Decesari, S., Facchini, M. C., Marullo, S., Santoleri, R., Dell'Anno, A., Corinaldesi, C.,
 Tangherlini, M., Danovaro, R., Connecting marine productivity to sea-spray via nanoscale biological
 processes: Phytoplankton Dance or Death Disco? Scientific Reports, 5, 14883, 2015.
- Oyamada, C., Kaneniwa, M., Ebitani, K., Murata, M., Ishihara, K.: Mycosporine-like amino acids extracted
 from scallop (Patinopecten yessoensis) ovaries: UV protection and growth stimulation activities on human
 cells, Marine Biotechnology, 10, 141–150, 2007.
- Pautler, B. G., Woods, G. C., Dubnick, A., Simpson, A. J., Sharp, M. J., Fitzsimons, S. J., Simpson, M. J.:
 Molecular characterization of dissolved organic matter in glacial ice: coupling natural abundance ¹H NMR
 and fluorescence spectroscopy, Environ. Sci. Technol., 46, 3753–3761, 2012.
- Passow, U., Transparent exopolymer particles (TEP) in aquatic environments, Prog. Oceanogr., 55, 287-333,
 2002.
- Prather, K. A., Bertram, T. H., Grassian, V. H., Deane, G. B., Stokes, M. D., DeMott, P. J., Aluwihare, L. I.,
 Palenik, B. P., Azam, F., Seinfeld, J. H., Moffet, R. C., Molina, M. J., Cappa, C. D., Geiger, F. M., Roberts, G. C.,
 Russell, L. M., Ault, A. P., Baltrusaitis, J., Collins, D. B., Corrigan, C. E., Cuadra-Rodriguez, L. A., Ebben, C. J.,
 Forestieri, S. D., Guasco, T. L., Hersey, S. P., Kim, M. J., Lambert, W. F., Modini, R. L., Mui, W., Pedler, B. E.,
 Ruppel, M. J., Ryder, O. S., Schoepp, N. G., Sullivan, R. C., Zhao, D.:Bringing the ocean into the laboratory to
 probe the chemical complexity of sea spray aerosol, Proceedings of the National Academy of Sciences of
 the Unites States of America, 7, 110, 7550-7555, 2013.
- Repeta, D. J., Chemical characterization and cycling of dissolved organic matter, in "Biogeochemistry of
 marine dissolved organic matter" second edition, ed. D. A. Hansell and C. A. Carlson, Elsevier 2015, ISBN:
 978-0-12-405940-5.
- Rintoul, S. R., Chown, S. L., DeConto, R. M., England, M. H., Fricker, H. A., Masson-Delmotte, V., Naish, T. R.,
 Siegert, M. J., Xavier, J. C.: Choosing the future of Antarctica, Nature, 558, 233 241, 2018.
- Roukaerts, A., Cavagna, A.-J., Fripiat, F., Lannuzel, D., Meiners, K. M., and Dehairs, F.: Sea-ice algal primary
 production and nitrogen uptake off East Antarctica, Deep-Sea Res. II, 131, 140–149,
 doi:10.1016/j.dsr2.2015.08.007, 2016.
- Schmale, J., Schneider, J., Nemitz, E., Tang, Y. S., Dragosits, U., Blackall, T. D., Trathan, P. N., Phillips, G. J.,
 Sutton, M., Braban, C. F.: Sub-Antarctic marine aerosol: dominant contributions from biogenic sources,
 Atmos. Chem. Phys., 13, 8669–8694, 2013.
- Schmitt-Kopplin, P., Liger-Belair, G., Koch, B. P., Flerus, R., Kattner, G., Harir, M., Kanawati, B., Lucio, M.,
 Tziotis, D., Hertkorn, N., Gebefügi, I.: Dissolved organic matter in sea spray: a transfer study from marine
 surface water to aerosols, Biogeosciences, 9, 1571–1582, 2012.
- Shepherd and the IMBIE team: Mass balance of the Antarctic ice sheet from 1992 to 2017, Nature, 558, 219
 222, 2018.
- Schwier, A. N., Rose, C., Asmi, E., Ebling, A. M., Landing, W. M., Marro, S., Pedrotti, M.-L., Sallon, A.,
 luculano, F., Agusti, S., Tsiola, A., Pitta, P., Louis, J., Guieu, C., Gazeau, F., Sellegri, K., Primary marine aerosol
 emissions from the Mediterranean Sea during pre-bloom and oligotrophic conditions: correlations to
 seawater chlorophyll a from a mesocosm study. Atmos. Chem. Phys. 15, 7961–7976 (2015).

- 543 Suzuki, Y., Kawakami, M., Akasaka, K.: ¹H NMR application for characterizing water-soluble organic 544 compounds in urban atmospheric particles, Environ. Sci. Technol., 35, 2656-2664, 2001.
- Upadhyay, D., Umaa, S., Govind, M., Naranamangalam, J.: Role of NMR metabonomics in Celiac Disease
 (CeD), Biomedical Spectroscopy and Imaging, 5, 27–40, 2016.
- Virkkula, A., Teinilä, K., Hillamo, R., Kerminen, V.-M., Saarikoski, S., Aurela, M., Viidanoja, J., Paatero, J.,
 Koponen, I. K., Kulmala, M.: Chemical composition of boundary layer aerosol over the Atlantic Ocean and at
 an Antarctic site, Atmos. Chem. Phys., 6, 3407–3421, 2006.
- Wawrik, B., Bronk, D. A., Baer, S. E., Chi, L., Sun, M., Cooper, J. T., Yang, Z.: Bacterial utilization of creatine in
 seawater. Aquat. Microb. Ecol., 80, 153-165, 2017.
- Whitledge, T. E., Dugdale, R. C.: Creatine in seawater, Limnol. Oceanogr. 17, 309-314, 1972.
- Willis, M. D., Köllner, F., Burkart, J., Bozem, H., Thomas, J. L., Schneider, J., Aliabadi, A. A., Hoor, P. M.,
 Schulz, H., Herber, A. B., Leaitch, R., Abbatt, J. P. D.:, Evidence for marine biogenic influence on
 summertime Arctic aerosol, Geophys. Res. Lett., 44, 10.1002/2017GL073359, 2017.
- Willis, M. D., Leaitch, W. R., Abbatt, J. P. D., Processes controlling the composition and abundance of Arctic
 aerosol. Reviews of Geophysics, 56, 621–671, 2018.
- 658 Zamanillo M., Ortega-Retuerta, E., Nunes, S., Estrada, M., Sala, M. M., Royer, S.-J., López-Sandoval, D. C., 659 Emelianov, M., Vaqué, D., Marrasé, C., Simó, R. : Distribution of transparent exopolymer particles (TEP) in 660 distinct regions of the Southern Ocean, Sci. Total Environ. 691: 736-748, doi: 661 10.1016/j.scitotenv.2019.06.524, 2019.

Zorn, S. R., Drewnick, F., Schott, M., Hoffmann, T., Borrmann, S.: Characterization of the South Atlantic
marine boundary layer aerosol using an aerodyne aerosol mass spectrometer, Atmos. Chem. Phys., 8,
4711–4728, 2008.

Table 1	L.
---------	----

sample ID:	A-0701	A-0901	A-1301	A-1801	A-2401	A-2801	A-0102	A-0602
sampling times:	07 Jan 20:00 – 09 Jan 09:00	09 Jan 14:50 – 13 jan 13:50	13 Jan 19:20 – 18 Jan 12:20	18 Jan 13:30 – 21 jan 23:55	24 Jan 15:00 – 28 jan 05:15	28 Jan 13:30 – 31 jan 13:50	01 Feb 14:50 – 06 Feb 03:15	06 Feb 22:00 – 10 Feb 11:00
average air mass type:	Weddell Sea / Antarctic Peninsula	Weddell Sea	Weddell Sea	Weddell Sea	Open Ocean	Open Ocean	Open Ocean / mixed	Open Ocean
Water-soluble organic carbon (μgC/m ³):								
WSOC	0.14	0.07	0.12	0.13	0.09	0.14	0.05	0.11
¹ H-NMR functional groups (nmolH/m ³):								
H-C	2.60	2.16	2.28	3.03	3.27	2.81	2.07	2.82
H-C-C=O	2.40	1.58	1.80	2.10	1.91	1.86	1.28	1.78
H-C-O	2.15	0.57	0.69	0.83	2.06	0.99	0.99	1.41
O-CH-O	0.20	0.07	0.05	0.04	0.08	0.09	0.07	0.09
Ar-H	0.12	0.05	0.00	0.10	0.09	0.10	0.11	0.07
MSA	2.13	1.95	2.63	4.54	1.72	2.90	2.22	1.53
Alkyl- Amines	0.30	0.79	0.53	1.32	0.34	0.49	0.13	0.15
Molecular marker	rs (ng/m³):							
MSA	68	62	84	145	55	93	71	49
methyl- amines	2.31	5.5	3.79	9.0	2.53	3.56	0.92	1.20
creati- nine	0.09	1.65	1.52	2.21	~0.05	1.00	0.29	0.41
glycerol	NA	1.1	0.7	0.7	3.0	0.8	0.7	1.3
sucrose	11							
alanine			traces1	traces ¹	0.6			0.7
betaine					traces ²			

¹ below the limit of quantification (0.3 ng/m³); ² below the limit of quantification (0.2 ng/m³)

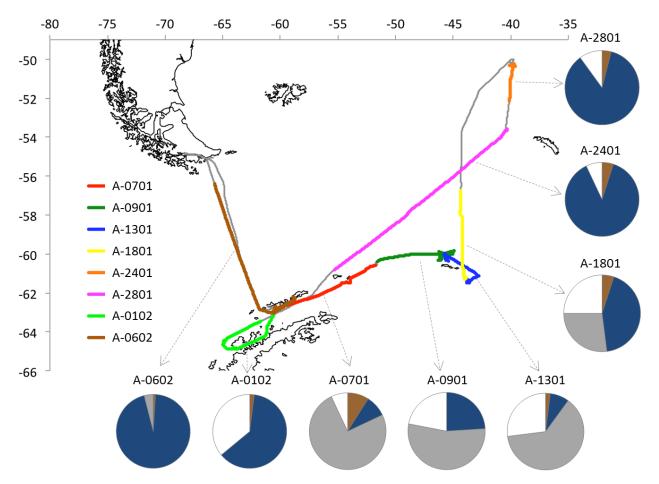


Figure 1. Cruise of RV Hesperides. The colors indicate the duration of the single aerosol samplings (short interruptions undertaken to avoid contamination from ship emissions are not indicated in the figure). The average time spent by air masses travelling over land (brown), marginal ice zone (1-99% surface coverage; grey), compact sea ice (100% coverage; white) and open ocean (dark blue) is indicated for each sample.

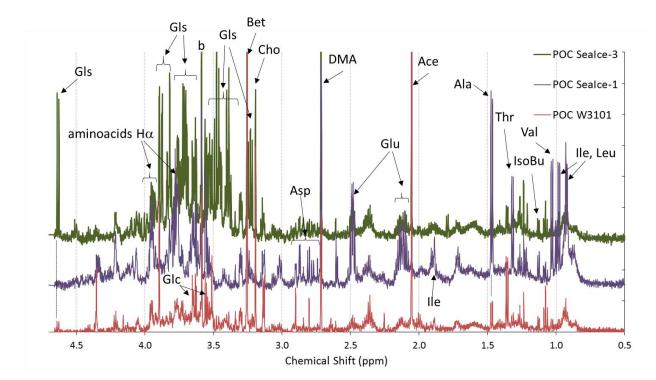


Figure 2. Aliphatic region of the ¹H-NMR spectra of three POC sample extracts: one for the seawater sample (POC W3201) and two from melted sea ice (POC Sealce-1 and POC Sealce-3). Specific NMR resonances were assigned to: the residuals of aminoacids (Ala, Thr, Val, Ile, Leu, Glu and Asp) and their alpha hydrogen atoms, isobutyric acid (IsoBu), acetic acid (Ace), dimethylamine (DMA), N-osmolytes (Bet: betaine; Cho: choline), glycerol (Glc) and to glucose (Gls).

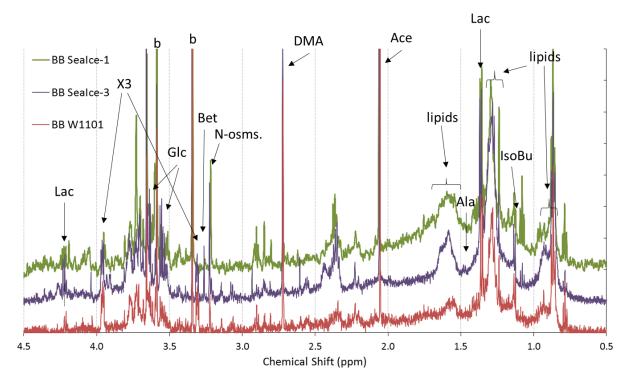
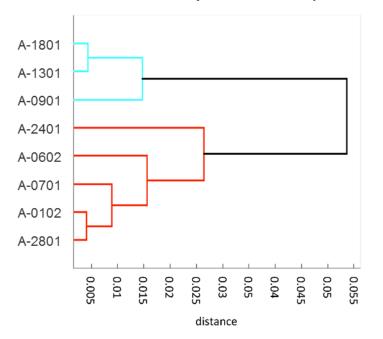


Figure 3. The same as Figure 2 but for the three bubble bursting aerosols: from seawater sample W1101 (BB W1101) and melted sea ice #1 and #3 (BB Sealce-1 and BB Sealce-3). Specific resonances were assigned to lactic acid (Lac), acetic acid (Ace), isobutyric acid (IsoBu), alanine (Ala), dimethylamine (DMA), glycerol (Glc), N-osmolytes (Bet: betaine;

"N-osms": unidentified, possibly phosphocholine) and to blank contaminations (b). Unresolved mixtures of aliphatic compounds were identified as lipids.



Hierarchical Cluster analysis of the NMR spectra

Figure 4. Cluster analysis of the ¹H-NMR spectra of the PM1 HiVol samples of ambient aerosol.

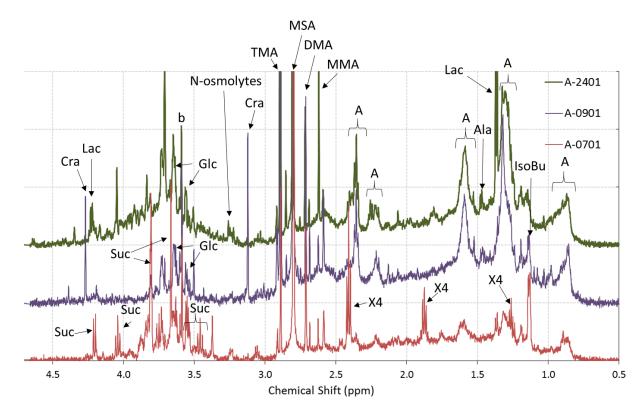


Figure 5. The same as Figure 2 but for the three ambient submicrometer aerosol samples. Specific resonances were assigned to lactic acid (Lac), isobutyric acid (IsoBu), alanine (Ala), monomethylamine (MMA), dimethylamine (DMA), trimethylamine (TMA), glycerol (Glc), sucrose (Suc), creatinine (Cra) and to blank contaminations (b). Unresolved

mixtures of linear aliphatic compounds (A), including possible contributions from lipids, are indicated in the spectra. Other NMR signals were only tentatively attributed to cadaverine (X4).

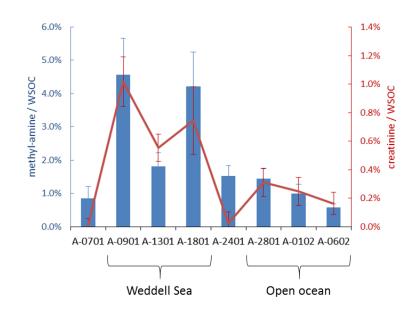


Figure 6. Concentrations of creatinine and methylamines in the PM1 samples. The concentrations are expressed as contributions to WSOC (mol% of carbon). "Weddell Sea" and "Open ocean" labels indicate the sampling periods identified by Dall'Osto et al. (2017) to characterize the aerosol composition in air masses travelling over sea ice and in the Southern Ocean, respectively.

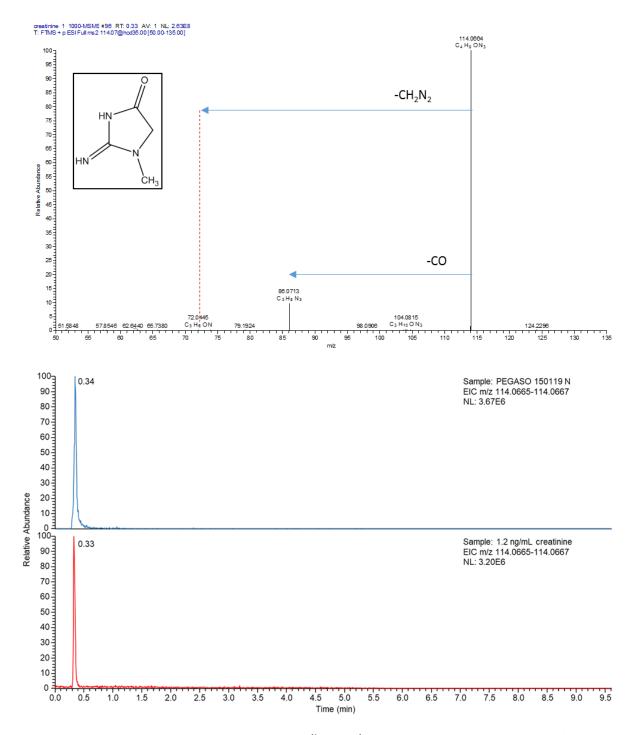


Figure 7. (top) MS spectrum of a creatinine standard. (bottom) Extracted ion chromatograms for m/z 114.0655-114.0667, corresponding to creatinine, of the filter extract of sample 0119n obtained during the PEGASO campaign and the neat creatinine standard. The retention time of creatinine was found to be 0.33 min using the conditions outlined in Section 2.4.

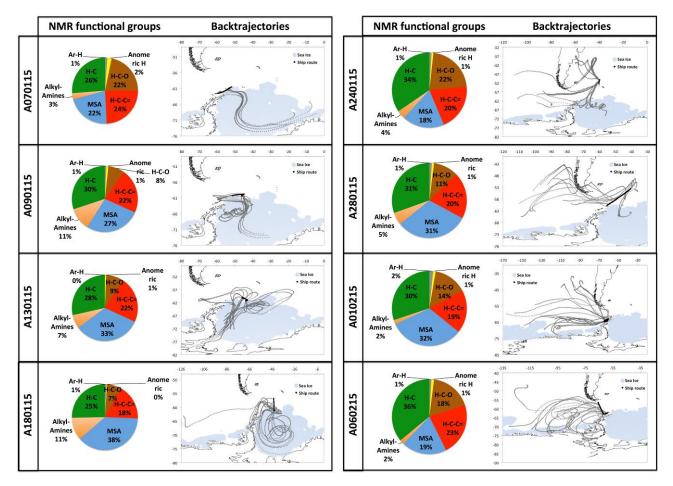


Figure 8. NMR functional group compositions of WSOC in the PM1 HiVol samples. Functionalities: H-C (alkyls), H-C-(C=) (acyls), H-C-O (alcoxyl), MSA, amines, anomeric, Ar-H (aromatic).