

Dear Dr. Hofzumahaus,

Thank you for your review of our paper. We have incorporated your suggestions into the revised manuscript. Below are our responses to your comments, highlighted in italics, with changes to the manuscript highlighted in red. The revised manuscript with track changes is attached.

Editor's Comments

The revised paper has been greatly improved and the comments of the reviewers and the editor have been adequately answered. Before the paper can be published, some minor changes are needed for clarification.

Minor changes

The abstract should be more specific and mention which chemical mechanisms have been tested and which version has given the best agreement with the interference corrected OH data. A quantitative statement should be made how much the modeled OH is increased by the LIM1 chemistry. Important boundary conditions (temperature, isoprene and NO concentrations) should be stated.

We have specified the chemical mechanisms tested in the abstract and have stated that the RACM2-LIM1 mechanism provides the best agreement with the corrected OH measurements. We have also quantified how much the base RACM2 and MCM 3.2 modeled OH concentrations increased with inclusion of versions of the LIM1 mechanism. We have also included the important boundary conditions as suggested:

This campaign took place in a forested area near the Indiana University, Bloomington campus characterized by high mixing ratios of isoprene (average daily maximum of approximately 4 ppb at 28°C) and low mixing ratios of NO (diurnal average of approximately 170 ppt). Supporting measurements of photolysis rates, VOCs, NO_x, and other species were used to constrain a zero-dimensional box model based on the Regional Atmospheric Chemistry Mechanism (RACM2) and the Master Chemical Mechanism (MCM 3.2), including versions of the Leuven isoprene mechanism (LIM1) for HO_x regeneration (RACM2-LIM1 and MCM 3.3.1).

The addition of versions of the LIM1 mechanism increased the base RACM2 and MCM 3.2 modeled OH concentrations by approximately 20% and 13% respectively, with the RACM2-LIM1 mechanism providing the best agreement with the measured concentrations, predicting maximum daily OH concentrations to within 30% of the measured concentrations.

Section 2.4: the difference between the two used isoprene mechanisms should be more precisely explained. The implementation in RACM2 as described in Tan et al. (2017) does not include the explicit LIM1 mechanism, but uses bulk reaction rates for the isoprene RO₂ isomerization via 1,6-H shift derived as parametrizations from LIM1 by Peters et al. (2014). Contrary, MCM

v3.3.1 contains the full LIM1 mechanism with a description of the equilibrium between different isoprene RO₂ isomers and isomerization by H shift reactions of specific isomers (Jenkin et al., 2015). Following a reviewer recommendation by Peeters et al. (2015), Jenkin et al. (2015) adjusted the rate coefficients for equilibration and H shift reactions to match preliminary experimental results by Crounse et al. (2014). Compared to the original LIM1, these changes mean a reduction of the effective bulk rate coefficient for the 1,6-H shift by about a factor of 5.

We have clarified the differences between the isoprene mechanisms in RACM2 and MCM as suggested on pages 9-10 of the revised manuscript:

The isoprene oxidation mechanism in RACM2 was updated as described in Tan et al. (2017) to include the Leuven Isoprene Mechanism (LIM1) originally proposed by Peeters, et al. (2009) involving peroxy radical isomerization reactions leading to additional HO_x radical production. This is a condensed version of the LIM1 mechanism and includes the updated bulk reaction rate constants for the isoprene peroxy radical 1,6-H shift isomerization reactions as parameterized in Peeters et al. (2014). These isomerization reactions lead to the formation of HO₂ and hydroperoxy aldehydes (HPALDs) (Crounse et al., 2011; Teng et al., 2017; Berndt et al., 2019) which can subsequently photolyze leading to OH production, as well as di-hydroperoxy carbonyl peroxy radicals (di-HPCARP-RO₂) which can rapidly decompose to produce additional OH radicals (Teng et al., 2017; Wennberg et al., 2018). The addition also includes a revision of the chemistry of first-generation isoprene oxidation products, including methyl vinyl ketone (MVK), methacrolein (MACR), and isoprene hydroperoxides (ISHP) (Tan et al., 2017).

In addition, the ambient measurements were also modeled with version 3.3.1 of the Master Chemical Mechanism. In comparison to version 3.2, MCM 3.3.1 incorporates the explicit LIM1 mechanism, including the equilibrium between different isoprene peroxy radical isomers and H-shift isomerization reactions of specific isomers, resulting in HO_x radical recycling through the production of HPALDs as well as di-HPCARP-RO₂ radicals (Jenkin et al., 2015). Based on the recommendation of Peeters (2015), the equilibrium rate coefficients between different peroxy radical isomers were increased and the 1,6 H-shift isomerization rate constants were decreased in order to match early experimental results of Crounse et al. (2014). These changes resulted in effective bulk 1,6-H shift peroxy radical isomerization rate constants in MCM 3.3.1 that are approximately a factor of 5 lower than the original LIM1 recommended rates (Novelli et al., 2020).

Page 12, line 22-23. Please quantify the difference between the model runs with and without LIM1 chemistry. The corresponding levels of NO from 9 to 17 EDT should be mentioned in this context.

We have quantified the increase in the modeled OH concentrations due to the LIM1 mechanism as suggested, and have included the average levels of NO from 9 to 17 EDT on page 12 of the revised manuscript:

Including versions of the LIM1 mechanism in both the MCM (3.3.1) and RACM2 (RACM2-LIM1) models increases the predicted daytime concentrations of OH by approximately 13% and 20%, respectively compared to the base mechanisms during the day (9:00-17:00 EDT), when mixing ratios of NO on these days decreased from an average morning maximum near 500 ppt to approximately 50 ppt in the afternoon.

Page 12, line 22-26. The comparison between the measured and modeled OH concentrations (Figure 5, upper right panel) needs to be done more careful. The statement "... with the RACM2-LIM1 results within 30% of the measured concentrations during the day (9:00-17:00 EDT)" is not true at 14h, where the RACM-LIM1 model and the measurements are numerically different by a factor of 1.8. Probably, this discrepancy is explainable by the total uncertainty of the measured data point (OH calibration error of 18%, plus statistical error bar, plus additional uncertainty from the interference correction mentioned on the same page) and the model uncertainty of 30%.

We have clarified the agreement between the RACM2-LIM1 model results with the measurements to include a statement that the measurements at 14:00 are still within the combined uncertainty of the measurements and the model on page 12 of the revised manuscript:

The RACM2-LIM1 results are generally within 30% of the measured concentrations during the day (Fig. 5, top right), with the difference at 14:00 EDT within the combined measurement precision, the accuracy of the calibration (18%, 1σ), and the model uncertainty (30%).

Page 12, line 27-29. The comparison with results by Novelli et al. is only meaningful, if the NO concentrations are comparable. To which NO concentration does the factor of 1.4 refer? The description of the improved mechanism in Novelli et al. is not entirely correct. The improvement is obtained by using the fast rate coefficients for the RO₂ equilibrium from MCM v3.3.1 and the fast 1,6-H-shift rate coefficients from the Caltech mechanism (Wennberg et al., 2018). This combination causes a faster production of HPALD and di-HPCARP-RO₂, both of which are likely important for the OH regeneration. The overall mechanism has a bulk rate coefficient for the 1,6-H shift which is close to the value in the original LIM1.

We have included the NO mixing ratio in the Novelli et al. study (less than 0.2 ppb), which is comparable to the NO mixing ratios in this study. We have also clarified the improved isoprene mechanism used in Novelli et al., on pages 12-13 of the revised manuscript as suggested:

These results are similar to that found by Novelli et al. (2020), who found that the MCM 3.3.1 underpredicted measurements of OH by a factor of approximately 1.4 during isoprene oxidation experiments in the SAPHIR chamber when mixing ratios of NO were less than 0.2 ppb, similar to the mixing ratios of NO measured in this study. The measured OH concentrations could be reproduced using a model that incorporated the larger equilibrium rate coefficients between the different peroxy radical isomers in the MCM 3.3.1 mechanism with the larger 1,6-H shift peroxy radical rate constants from the Caltech isoprene mechanism (Teng et al., 2017; Wennberg et al.,

2018). This combination increased production of HPALDs and di-HPCARP-RO₂ radicals in the oxidation mechanism, resulting in an effective bulk isoprene peroxy radical isomerization rate similar to that in the original LIM1 mechanism (Novelli et al., 2020). These larger bulk peroxy radical isomerization rates are similar to that incorporated into the RACM2-LIM1 mechanism used in the present study (Tan et al., 2017), leading to the higher modeled radical concentrations compared to the MCM 3.3.1 model results shown in Fig. 5.

Figure 6 and 8: which measurement days are averaged in the diurnal profiles?

For Figure 6, we have clarified that the model results are the diurnal average for the days which NO was measured simultaneously, and have specified the days in the main text (page 12).

We have also clarified that Figure 8 includes all the measurements made during the campaign.

Figure 9: what is the meaning of a median diurnal average for a single day?

We have corrected the caption to indicate that the figure represents the median total OH reactivity measurements for this day.

Figure 10 and 11: which components are meant by the label 'others' in the legends. Please give a short explanation in the figure captions.

We have clarified the components of the “Others + hv” categories in the caption of each figure as suggested, in addition to page 16 of the revised manuscript.

OH and HO₂ radical chemistry in a midlatitude forest: Measurements and model comparisons

Michelle M. Lew^{1*}, Pamela S. Rickly^{2**}, Brandon P. Bottorff¹, Emily Reidy¹, Sofia Sklaveniti^{2,3}, Thierry Léonardis³, Nadine Locoge³, Sebastien Dusanter³, Shuvashish Kundu^{4***}, Ezra Wood⁵, and Philip S. Stevens^{1,2}

¹ Department of Chemistry, Indiana University, Bloomington, IN 47405, USA

² O'Neill School of Public and Environmental Affairs, Indiana University, Bloomington, IN 47405, USA

³ IMT Lille Douai, Univ. Lille, SAGE – Département Sciences de l'Atmosphère et Génie de l'Environnement, 59000 Lille, France

⁴ Department of Chemistry, University of Massachusetts - Amherst, Amherst, MA 01003, USA

⁵ Department of Chemistry, Drexel University, Philadelphia, PA 19104, USA

* now at California Air Resources Board, Sacramento, CA 95812, USA

** now at Cooperative Institute for Research in Environmental Sciences, University of Colorado, Boulder, CO 80309, USA and Chemical Sciences Division, Earth System Research Laboratory, National Oceanic and Atmospheric Administration, Boulder, CO 80305, USA

*** now at Momentive Performance Materials, Inc., Tarrytown, NY 10591, United States

Correspondence: Philip S. Stevens (pstevens@indiana.edu)

Abstract. Reactions of the hydroxyl (OH) and peroxy radicals (HO₂ and RO₂) play a central role in the chemistry of the atmosphere. In addition to controlling the lifetimes of many trace gases important to issues of global climate change, OH radical reactions initiate the oxidation of volatile organic compounds (VOCs) which can lead to the production of ozone and secondary organic aerosols in the atmosphere. Previous measurements of these radicals in forest environments characterized by high mixing ratios of isoprene and low mixing ratios of nitrogen oxides (NO_x) (typically less than 1-2 ppb) have shown serious discrepancies with modeled concentrations. These results bring into question our understanding of the atmospheric chemistry of isoprene and other biogenic VOCs under low NO_x conditions.

During the summer of 2015, OH and HO₂ radical concentrations as well as total OH reactivity were measured using Laser-Induced Fluorescence - Fluorescence Assay by Gas Expansion (LIF-FAGE) techniques as part of the Indiana Radical, Reactivity and Ozone Production Intercomparison (IRRONIC). This campaign took place in a forested area near the Indiana University, Bloomington campus characterized by high mixing ratios of isoprene (average daily maximum of approximately 4 ppb at 28°C) and low mixing ratios of NO (diurnal average of approximately 170 ppt). Supporting measurements of photolysis rates, VOCs, NO_x, and other species were used to constrain a zero-dimensional box model based on the Regional Atmospheric Chemistry Mechanism (RACM2) and the Master Chemical Mechanism (MCM 3.2), including versions of the Leuven Isoprene Mechanism (LIM1) for HO_x regeneration (RACM2-LIM1 and MCM 3.3.1). Using an OH chemical scavenger technique, the study revealed the presence of an interference with the LIF-FAGE measurements of OH that increased with

both ambient concentrations of ozone and temperature, with an average daytime maximum equivalent OH concentration of approximately $5 \times 10^6 \text{ cm}^{-3}$. Subtraction of the interference resulted in measured OH concentrations of approximately $4 \times 10^6 \text{ cm}^{-3}$ (average daytime maximum) that were in better agreement with model predictions, although the models underestimated the measurements in the evening. The addition of versions of the LIM1 mechanism increased the base RACM2 and MCM 3.2 modeled OH concentrations by approximately 20% and 13% respectively, with the RACM2-LIM1 mechanism providing the best agreement with the measured concentrations, predicting maximum daily OH concentrations to within 30% of the measured concentrations. Measurements of HO_2 concentrations during the campaign (approximately $1 \times 10^9 \text{ cm}^{-3}$ average daytime maximum) included a fraction of isoprene-based peroxy radicals ($\text{HO}_2^* = \text{HO}_2 + \alpha \text{RO}_2$) and were found to agree with model predictions to within 10-30%. On average, the measured reactivity was consistent with that calculated from measured OH sinks to within 20%, with modeled oxidation products accounting for the missing reactivity, although significant missing reactivity (approximately 40% of the total measured reactivity) was observed on some days.

1 Introduction

The hydroxyl radical (OH) is one of the primary oxidants in the atmosphere (Levy, 1972). The OH radical initiates the oxidation of volatile organic compounds (VOCs) that leads to the production of hydroperoxy radicals (HO_2) and organic peroxy radicals (RO_2). In the presence of nitrogen oxides ($\text{NO}_x = \text{NO} + \text{NO}_2$), reactions of these radicals can lead to the production of ozone and secondary organic aerosols in the atmosphere, the primary components of photochemical smog. Because of their short atmospheric lifetimes, measurements of OH and HO_2 (together HO_x) and total OH reactivity can provide a robust test of our understanding of this complex chemistry (Heard and Pilling, 2003).

Multiple field campaigns have been conducted over the years measuring OH and HO_2 radicals in both urban and forested environments. Measurements of OH in urban areas characterized by high mixing ratios of NO_x and anthropogenic VOCs have been generally consistent with model predictions (Ren et al., 2003; Shirley et al., 2006; Kanaya et al., 2007a; Dusanter et al., 2009b; Hofzumahaus et al., 2009; Griffith et al., 2016; Tan et al., 2017; 2018; 2019), while measurements in remote forested environments characterized by low mixing ratios of NO_x and high mixing ratios of biogenic VOCs have often been greater than model predictions (Tan et al., 2001; Lelieveld et al., 2008; Whalley et al., 2011; Rohrer et al., 2014).

However, recent measurements by Mao et al. (2012) in a northern California forest using a new chemical scavenging technique that removes ambient OH before air enters the detection cell revealed a significant interference associated with their Laser-Induced Fluorescence (LIF) measurements of OH. The unknown interference was a factor of 2 to 3 times higher than ambient OH concentrations (Mao et al., 2012). Similar results were observed in a boreal forest by Novelli et al. (2014), who observed an interference using a similar chemical scrubbing technique that was a factor of 3 to 4 times higher than ambient OH concentrations. One possible source of this observed interference may be the decomposition of Criegee intermediates produced from the ozonolysis of biogenic emissions in the low-pressure detection cells used by LIF instruments, although the ambient concentration of these intermediates in the atmosphere may be too low to explain all of the observed interference

(Novelli et al., 2017; Rickly and Stevens, 2018). Another proposed source of the interference is the decomposition of ROOOH molecules inside the FAGE detection cell formed from the reaction of OH with RO₂ radicals (Fittschen et al., 2019). Nevertheless, interferences associated with measurements of OH could explain part of the discrepancies between measured and modeled OH concentrations in forested environments. Monitoring potential interferences associated with OH measurements using LIF techniques may be crucial for understanding the discrepancies between measurements and models.

In contrast to measurements of OH, the agreement between measured and modeled HO₂ concentrations have been highly variable. In urban environments, measured HO₂ concentrations were sometimes found to agree with model predictions (Shirley et al., 2006; Emmerson et al., 2007; Dusanter et al., 2009b; Michoud et al., 2012; Lu et al., 2013; Ren et al., 2013; Griffith et al., 2016; Tan et al., 2017), while other times the measurements were found to be both lower (George et al., 1999; Konrad et al., 2003) and higher than model predictions (Martinez et al., 2003; Ren et al., 2003; Emmerson et al., 2005; Kanaya et al., 2007a; Chen et al., 2010; Sheehy et al., 2010; Czader et al., 2013; Griffith et al., 2016; Tan et al., 2018). In forested environments, measured HO₂ concentrations were sometimes found to agree with model predictions (Tan, D. et al., 2001; Ren et al., 2005; 2006), but were often found to be either lower (Carslaw et al., 2001; Kanaya et al., 2007b; Whalley et al., 2011; Kanaya et al., 2012; Mao et al., 2012; Griffith et al., 2013; Mallik et al., 2018), or higher than model predictions (Carslaw et al., 2001; Kubistin et al., 2010; Kim et al., 2013; Hens et al., 2014). Part of this variability may be due to interferences from alkene and aromatic based RO₂ radicals converting to HO₂ in systems that detect HO₂ through conversion to OH by addition of NO in the sample cell. The degree to which the RO₂ species can interfere with HO₂ measurements has been quantified through several laboratory experiments (Fuchs et al., 2011; Whalley et al., 2013; Lew et al., 2018) and estimated in some field studies (Hens et al., 2014; Crowley et al., 2018; Mallik et al., 2018). However, the extent of RO₂ radical contributions during HO₂ measurements in many of the campaigns mentioned above is unclear.

Total OH reactivity measurements can complement HO_x measurements by providing a constraint on the total loss of OH that can be compared to that calculated from co-located measurements of OH sinks. Several recent studies have identified discrepancies between measured and calculated OH reactivity in which the measured values are significantly greater than the calculated values (Di Carlo et al., 2004; Hansen et al., 2014; Nölscher et al., 2016; Zannoni et al., 2016; Bsaibes et al., 2020). This difference has been attributed to OH loss from unmeasured VOCs and their oxidation products. In general, significant missing OH reactivity has not been observed as often in urban environments as it has in forested areas, bringing into question our understanding of the chemistry of biogenic emissions and their oxidation products (Dusanter and Stevens, 2017).

This study reports measurements and model simulations of HO_x radical chemistry as well as OH reactivity for a forested site located in Bloomington, Indiana during the 2015 IRRONIC (Indiana Radical Reactivity and Ozone production InterComparison) field campaign. This work compares the measured HO_x radical concentrations to model predictions incorporating the Regional Atmospheric Chemistry Mechanism 2 (RACM2), in addition to a version updated to include the Leuven Isoprene Mechanism (RACM2-LIM1), as well as the Master Chemical Mechanism versions 3.2 and 3.3.1 in order to test the ability of each model to reproduce the observed radical concentrations and total OH reactivity.

2 Experimental section

2.1 IRRONIC location and supporting measurements

The IRRONIC campaign site was located within a mixed deciduous forest (sugar maple, sycamore, tulip polar, ash and hickory trees) at the Indiana University Research and Teaching Preserve (IU-RTP) field lab (39.1908° N, 86.502° W) located approximately 2.5 km northeast of the center of the Indiana University campus, and 1 km from the IN 45/46 bypass at the northern perimeter. The goals of the campaign included an informal intercomparison of peroxy radical measurements by two different techniques (Kundu et al., 2019), an analysis of ozone production sensitivity at this site (Sklaveniti et al., 2018), a comparison of measured OH radical reactivity with that calculated from measured VOCs, and a comparison of measured OH, HO₂, and RO₂ radicals with model predictions. The main biogenic emission within this area was isoprene, with an average daytime maximum mixing ratio of approximately 4 ppb during the campaign. This area exhibited low anthropogenic influence from the campus area, with an average daytime maximum mixing ratio of NO of approximately 315 ppt and an average daytime maximum NO₂ mixing ratio of approximately 2 ppb. Measurements were conducted on top of two scaffolding platforms adjacent to the field lab, approximately 1.8 m from the ground. At this site, solar noon occurred at approximately 13:52 EDT. Additional information regarding the field site and the IRRONIC campaign can be found in Sklaveniti et al. (2018) and Kundu et al. (2019).

Table 1 summarizes the major instrumentation employed during the campaign. NO was measured every 10 s using a chemiluminescence instrument (Thermo model 42i-TL, detection limit 50 ppt / 2 min). Periodic problems with the sensor's high voltage power supply that required an eventual replacement limited the coverage of the measurements. NO₂ was measured every 1 s by a Cavity Attenuated Phase Shift (CAPS) instrument (detection limit 40 ppt / 10 s), and ozone was measured every 10 sec using a 2B Technologies model 202 UV absorbance instrument (detection limit 3 ppb / 10 s). Further details on the calibration and baseline measurements for the NO, NO₂, and O₃ measurements are described in Kundu et al. (2019). Nonmethane hydrocarbons, including C2-C10 alkanes and alkenes, butadiene, C6-C9 aromatic compounds, isoprene, α -pinene, and β -pinene, were measured using a thermal desorption GC/FID instrument with a 1.5-h time resolution. Oxygenated VOCs (OVOCs), including C2-C10 aldehydes, C2-C6 ketones, and C2-C4 alcohols, were measured by thermal desorption GC/FID-MS with a 1.5-h time resolution. Offline sampling focused on measurements of oxygenated VOCs including formaldehyde and C2-C6 aldehydes, acetone, MEK, glyoxal and methylglyoxal using DNPH cartridges and HPLC-UV analysis. C6-C16 VOCs including α -pinene, β -pinene, limonene, camphene, heptane-hexadecane, methylpentene-pentadecene were measured using Sorbent cartridges and GC-MS analysis. Measurements of J(NO₂) were made by spectral radiometry courtesy of the University of Houston. HONO was measured using a newly developed Laser Photofragmentation/Laser-Induced Fluorescence instrument (Bottorff et al., 2015; Bottorff et al., manuscript in preparation).

2.2 HO_x radical measurements

The Indiana University LIF-FAGE instrument (IU-FAGE) has been described in detail previously and consists of a single axis for alternating measurements of OH and HO₂ or HO₂* (Dusanter et al., 2009a Griffith et al., 2013; 2016). In the LIF-FAGE technique, OH radicals are detected by laser-induced fluorescence after expansion of ambient air to low pressure. This extends the OH fluorescence lifetime, allowing temporal filtering of the fluorescence from laser scatter (Heard and Pilling, 2003). Ambient air is expanded through a 0.64 mm diameter orifice located at the top of a cylindrical nozzle (5 cm in diameter and 20 cm long), resulting in a flow rate of approximately 3 SLPM through the sampling nozzle. Two scroll pumps (Edwards XDS 35i) connected in parallel maintain a pressure inside the cell of 7.3 hPa.

The laser system used in this study consisted of a Spectra Physics Navigator II YHP40-532Q that produces approximately 8 W of radiation at 532 nm at a repetition rate of 10 kHz which is used to pump a Sirah Credo Dye laser (255 mg/L of Rhodamine 610 and 80 mg/L of Rhodamine 101 in ethanol), resulting in 40 to 100 mW of radiation at 308 nm. After exiting the dye laser, a fraction of the radiation is focused onto the entrance of a 12-m optical fiber to transmit the radiation to the sampling cell which was placed on top of the 1.8-m platform adjacent to the field lab. In the detection cell, the laser crosses the expanded air perpendicular to the flow in a White cell configuration with 24 passes. For this campaign, the laser power entering the sampling cell ranged from 0.5 to 4.4 mW and was monitored using a photodiode at the exit of the White cell. This does not reflect the laser power density inside the detection cell due to overlap of the beams in the multipass configuration.

OH radicals are excited and detected using the $A^2\Sigma^+ v' = 0 \leftarrow X^2\Pi v'' = 0$ transition near 308 nm (Stevens et al., 1994). The net signal is measured by spectral modulation by tuning the wavelength on- and off-resonance in successive modulation cycles. A reference cell where OH is produced by thermal dissociation of water vapor is used to ensure that the laser is tuned on and off the OH transition. The OH fluorescence is detected using a microchannel plate photomultiplier tube (MCP-PMT) detector (Hamamatsu R5946U-50), a preamplifier (Stanford Research System SR445) and a gated photon counter (Stanford Research Systems SR 400). The MCP-PMT is switched off during the laser pulse through the use of electronic gating allowing the OH fluorescence to be temporally filtered from laser scattered light. A Teflon injector located approximately 2.5 cm below the inlet and 17.5 cm above the detection axis allowed for the addition of NO (approximately 2 sccm, $1.4 \times 10^{13} \text{ cm}^{-3}$, Matheson Gas, 10% in N₂) to convert ambient HO₂ to OH through the fast $\text{HO}_2 + \text{NO} \rightarrow \text{OH} + \text{NO}_2$ reaction, allowing for indirect measurements of HO₂.

The IU-FAGE instrument is calibrated by producing known quantities of OH and HO₂ from the photolysis of water vapor in air using a mercury penlamp within the calibration source as described previously (Dusanter et al., 2008). For these calibrations, zero air was sent through a humidifier and delivered at a flow rate of 38-50 L min⁻¹ to the calibration source. Uncertainties associated with the UV water photolysis calibration technique have been described previously (Dusanter et al., 2008) and are estimated to be 18% (1 σ) for both OH and HO₂.

2.2.1 Measurement of OH interferences

The LIF-FAGE measurements are subject to potential interferences where OH radicals are generated inside the detection cell. For example, ozone can be photolyzed by the laser and in the presence of water vapor can produce hydroxyl radicals (Davis et al., 1981a; 1981b) (reactions R1 and R2):



This interference in the IU-FAGE instrument is monitored through laboratory calibrations utilizing various concentrations of ozone, water vapor, and laser power. To characterize this and any other interference during ambient measurements, a chemical scrubbing technique is used to remove ambient OH prior to entering the detection cell (Griffith et al., 2016; Rickly and Stevens, 2018). This chemical modulation technique is used to monitor levels of the laser-generated ozone-water interference and any other processes that may produce OH radicals within the excitation axis.

Hexafluoropropylene (C_3F_6 , 95.5% in N_2 , Matheson) is added through a circular injector 1 cm above the nozzle with a flow rate of approximately 3.5 sccm to remove 95% of externally generated OH (Rickly and Stevens, 2018). During ambient measurements, chemical addition of C_3F_6 is modulated in between ambient OH measurements every 15 minutes for a duration of 10 minutes. The differences between the measured OH during C_3F_6 addition and OH measurements including the interference represents the net ambient OH concentration in the atmosphere. Taking the measurement of potential interferences into account results in a limit of detection for OH for this campaign of approximately $7.9 \times 10^5 \text{ cm}^{-3}$ for a 30 min average (S/N = 1).

2.2.2 Contribution of RO_2 interferences during HO_2 measurements

As discussed above, HO_2 radicals are measured indirectly after sampling ambient air at low pressure through chemical conversion to OH by addition of NO and subsequent detection of OH by LIF:



It was previously believed that the detection of HO_2 radicals using this technique was free from interferences from the reaction of RO_2 radicals with NO, as model simulations and measurements suggested that the rate of conversion of RO_2 radicals to HO_2 by reactions R4 and R5 and subsequent conversion to OH through reaction R3 were negligible. This was due to the slow rate of reaction R5 under the reduced oxygen concentration in the low pressure LIF-FAGE cell and the short reaction time between injection of NO and detection of OH (Heard and Pilling, 2003).



For example, RO_2 radicals produced from the OH-initiated oxidation of small alkanes were found to produce a negligible yield of HO_2 (Stevens et al., 1994; Kanaya et al., 2001; Tan, et al., 2001; Creasey et al., 2002; Holland et al., 2003).

1 However, recent laboratory studies have shown that there are interferences associated with measurements of HO₂ from the
2 conversion of RO₂ radicals derived from the OH-initiated oxidation of alkenes and aromatics to HO₂ (and subsequently OH)
3 by reaction with NO. The high conversion efficiency of alkene-based peroxy radicals to HO₂ is due to the ability of the β-
4 hydroxyalkoxy radicals produced from OH + VOC reactions to rapidly decompose, forming a hydroxyalkyl radical which then
5 reacts rapidly with O₂ leading to the production of a carbonyl compound and HO₂ (Fuchs et al., 2011; Whalley et al., 2013;
6 Lew et al., 2018). Because of this interference, measurements of peroxy radicals that are sensitive to this interference are
7 denoted as HO₂* ($[HO_2^*] = [HO_2] + \alpha [RO_2]$, $0 < \alpha < 1$). The conversion efficiency depends on the instrumental characteristics
8 and configurations employed as well as the amount of NO added. The RO₂-to-HO₂ conversion efficiencies for a number of
9 different peroxy radicals have been characterized for current and past configurations of the IU-FAGE instrument (Lew et al.,
10 2018). For the configuration of the IU-FAGE instrument used in this study, the conversion efficiency of isoprene-based peroxy
11 radicals was found to be approximately 83%, while the conversion efficiency of propane peroxy radicals was found to be
12 approximately 15%. A high concentration of NO leading to a high conversion efficiency of isoprene-based peroxy radicals to
13 HO₂ was used throughout the study to provide a useful intercomparison of the IU-FAGE HO₂* measurements with the
14 RO₂+HO₂ measurements by the Drexel University Ethane – Nitric Oxide Chemical Amplifier (ECHAMP) instrument (Kundu
15 et al., 2019), as HO₂ and isoprene-based peroxy radicals accounted for approximately 70% of the total peroxy radicals at this
16 site (see below). To maximize measurement overlap for the intercomparison, low concentrations of added NO were not used
17 and as a result no measurements of HO₂ with minimal interference from RO₂ radicals were obtained. The instrumental precision
18 for the HO₂* measurement based on the variability of the background signal due to laser scatter and detector noise results in a
19 limit of detection for HO₂* during this campaign of $7 \times 10^7 \text{ cm}^{-3}$ for a 30 second average (S/N = 1).
20

21 2.3 OH reactivity measurements

22 The IU Total OH Loss rate Method (TOHLM) instrument is based on the method of Kovacs and Brune (2001) and is described
23 in detail elsewhere (Hansen et al., 2014). Briefly, the instrument is comprised of a flow tube reactor measuring 5 cm in diameter
24 and 75 cm in length. Ambient air is introduced through an 8 cm diameter perfluoroalkoxy polymer film hose attached to the
25 flow tube at a flow rate of approximately 180 SLPM using a regenerative blower (Spencer VB001) to establish turbulent flow
26 conditions. Previous measurements have demonstrated that different lengths of this inlet tubing do not significantly impact the
27 measured OH reactivity (Hansen et al., 2014). A pitot-static tube (Dwyer Instruments) is positioned just before the exit of the
28 flow tube facing the turbulent core of the flow, approximately 1 cm from the flow tube wall. The pitot-static tube is connected
29 to a differential pressure gauge (MKS Instruments) to measure the total flow tube velocity.

30 OH radicals are produced in a movable injector that houses a mercury pen lamp (UV Pen-Ray) in which the top of
31 the pen lamp was positioned at the end of the injector, just before a spiral Teflon spray nozzle used to promote mixing within
32 the flow tube (McMaster Carr). In addition, a turbulizer is attached to the injector tube 24 cm before the spray nozzle consisting

of four 1 cm wide fins to promote turbulent flow conditions as well as to provide support of the injector throughout the flow tube. The injector is inserted along the main axis and is configured for automated movement acquiring continuous measurements in the forward and backward directions. A nitrogen flow of 10 standard liters per minute (SLPM) is bubbled through high-purity water (EMD Chemicals) producing water vapor which is directed through the injector and photolyzed by the penlamp to produce OH with typical concentrations on the order of 10^9 cm^{-3} . This method is known to also produce HO_2 radicals, which can lead to a regeneration of OH at NO mixing ratios greater than 1 ppbv (Kovacs and Brune, 2001). However, because the average NO mixing ratio measured over the course of the campaign was below this value, no correction to the measured reactivity was applied (Hansen et al., 2014).

OH radicals were measured using a similar FAGE detection cell described above. Ambient air was expanded through a 1 mm diameter orifice to a total pressure of approximately 8 hPa. OH radicals were excited by a portion of the 308 nm output of the dye laser, with the resulting fluorescence detected by a gated channel photomultiplier tube detector (Excelitas MP 1300) and monitored by a photon counter (Stanford Research SRS 400). A 2 meter long optical fiber was used to transmit the 308-nm laser beam to the OH reactivity detection cell which was located inside the field lab. The laser power was measured at the exit of the detection cell and monitored with a photodiode.

As ambient air entered the flow tube, the automated OH source injector allowed for varying reaction time with the ambient air over a distance of approximately 15 cm for a period of 2.5 minutes. This produced an OH decay over a reaction time of 0-0.15 s from which the OH reactivity was determined. Losses of OH on the walls of the flow tube were measured by flowing high-purity nitrogen (Indiana Oxygen) at 180 SLPM through the flow tube in addition to the OH production through the injector to measure the decay of OH in the absence of any VOCs. Several measurements of this wall loss (k_b) resulted in an average value of $10 \pm 2 \text{ s}^{-1}$ (1σ).

The calculated OH reactivity for a measured compound X (k_X), can be determined from the product of the concentration of X and its second-order rate constant with OH:

$$k_X = k_{OH+X}[X] \quad (1)$$

Summation of this value for each reacting species gives the total OH reactivity (k_{OH}):

$$k_{OH} = \sum_i k_{OH+X_i} [X_i] \quad (2)$$

Under pseudo-first order conditions ($[\text{OH}] \ll [\text{X}]$), the OH concentration within the flow tube can be expressed as a first-order exponential decay:

$$[\text{OH}]_t = [\text{OH}]_0 e^{-(k_{OH}+k_b)t} \quad (3)$$

Solving for k_{OH} , the OH reactivity, gives:

$$k_{OH} = -\frac{\Delta \ln[\text{OH}]}{\Delta t} - k_b \quad (4)$$

Measurements of the change in the concentration of OH over the reaction time produces the measured OH reactivity value. These measurements can be compared to the calculated total reactivity from measured OH sinks (Eq. 2) to determine whether the measured total OH reactivity can be accounted for by the measured sinks. The difference between the measured and calculated total OH reactivity is referred to as the “missing” OH reactivity.

Laboratory measurements of the reactivity of several VOCs with well-known rate constants, including butane, isoprene, and propane showed that the OH reactivity measurements for these compounds were on average 30% lower than calculated when the measured velocity of the turbulent core is used to determine the reaction time. This consistent underestimation of the OH reactivity is likely due to either incomplete mixing of the reactants or a systematic underestimation of the reaction time, and is similar to that measured previously by Hansen et al. (2014). As a result, the measured ambient OH reactivity values were scaled by a factor of 1.41. Measurements performed over a range of OH reactivity values suggest that the IU-TOHLM instrument can measure OH reactivity up to 45 s^{-1} with a precision (1σ) of $1.2 \text{ s}^{-1} + 4\%$ of the measured value for a 10 min average (Hansen et al., 2014).

2.4 Modeling HO_x concentrations and OH reactivity

Ambient measurements of OH, HO₂^{*}, and total OH reactivity were modeled with the Regional Atmospheric Chemistry Mechanism (RACM2) (Goliff et al., 2013) and the Master Chemical Mechanism (MCM) version 3.2 (Jenkin et al., 1997; Saunders et al., 2003). While the MCM model provides a near-explicit chemical mechanism and is expected to better represent complex chemical atmospheres, the lumped RACM mechanism is easier to use in radical budget calculations. The isoprene oxidation mechanism in RACM2 was updated as described in Tan et al. (2017) to include the Leuven Isoprene Mechanism (LIM1) originally proposed by Peeters, et al. (2009) involving peroxy radical isomerization reactions leading to additional HO_x radical production. This is a condensed version of the LIM1 mechanism and includes the updated bulk reaction rate constants for the isoprene peroxy radical 1,6-H shift isomerization reactions as parameterized in Peeters et al. (2014). These isomerization reactions lead to the formation of HO₂ and hydroxyperoxy aldehydes (HPALDs) (Crounse et al., 2011; Teng et al., 2017; Berndt et al., 2019) which can subsequently photolyze leading to OH production, as well as di-hydroperoxy carbonyl peroxy radicals (di-HPCARP-RO₂) which can rapidly decompose to produce additional OH radicals (Teng et al., 2017; Wennberg et al., 2018). The addition also includes a revision of the chemistry of first-generation isoprene oxidation products, including methyl vinyl ketone (MVK), methacrolein (MACR), and isoprene hydroperoxides (ISHP) (Tan et al., 2017).

In addition, the ambient measurements were also modeled with version 3.3.1 of the Master Chemical Mechanism. In comparison to version 3.2, MCM 3.3.1 incorporates the explicit LIM1 mechanism, including the equilibrium between different isoprene peroxy radical isomers and H-shift isomerization reactions of specific isomers, resulting in HO_x radical recycling through the production of HPALDs as well as di-HPCARP-RO₂ radicals (Jenkin et al., 2015). Based on the recommendation of Peeters (2015), the equilibrium rate coefficients between different peroxy radical isomers were increased and the 1,6 H-shift isomerization rate constants were decreased in order to match early experimental results of Crounse et al. (2014). These

changes resulted in effective bulk 1,6-H shift peroxy radical isomerization rate constants in MCM 3.3.1 that are approximately a factor of 5 lower than the original LIM1 recommended rates (Novelli et al., 2020).

The Framework for 0-D Atmospheric Modeling (F0AM) was used to calculate the radical concentrations and OH reactivity observed at the IRRONIC site (Wolfe et al., 2016). The model was constrained by the 30 minute average measured mixing ratios of ozone, NO_x, and VOCs and processed through a 5 day spin-up to generate unmeasured secondary oxidation products. Table S1 summarizes the measured compounds and includes their grouping into the condensed RACM2 model inputs. Because the VOC measurements occurred every 90 minutes, the measurements were interpolated into 30 min bins before input to the model. Due to the minimal overlap of the NO measurements with the HO_x measurements, the model was only run for the days when there was measurements of NO, limiting the number of days for comparison with the radical measurements. Zero-dimensional models cannot explicitly account for emissions, and NO is emitted both by vehicles on the nearby highway 1 km to the Southwest and by soil. Such local perturbations to the NO_x-O₃-radical chemistry necessitate using constrained measurements of NO, NO₂, and O₃. The measured J(NO₂) was used to scale the model calculated J(NO₂) and other photolysis rates. The model uncertainty is approximately 30% (1σ), estimated from uncertainties associated with the input parameters and the rate constants for each reaction (Griffith et al., 2013; Wolfe et al., 2016).

3 Results and discussion

Campaign diurnal average measurements of J(NO₂), temperature, isoprene, O₃, NO₂, and NO are summarized in Fig. 1. The maximum average mixing ratio of NO of approximately 315 ppt was observed at approximately 08:00 (EDT), while the average mixing ratio of NO₂ reached a maximum of 2 ppb around 10:00. Average mixing ratios of isoprene ranged from 0.4 to 4.4 ppb, reaching a maximum around 18:00. The relatively high nighttime mixing ratios often observed at this site are likely due to the fact that the measurements were made below the forest canopy and relatively close to the surface. As a result, vertical stratification likely resulted in higher concentrations of isoprene near the surface during several nights, similar to other measurements of biogenic VOCs below the forest canopy (Bsaiibes, et al., 2020). Anthropogenic VOCs were relatively low at this site, with maximum mixing ratios of benzene less than 80 ppt. Day-to-day profiles (July 10 to July 25) are illustrated in Fig. 2, showing measurements of O₃, temperature, isoprene, NO_x, HO₂^{*}, and OH. Unfortunately, instrumental problems limited the NO measurements prior to 19 July.

3.1 OH measurements and model comparison

OH concentrations were determined using the chemical modulation technique described above using external C₃F₆ addition to scavenge ambient OH and measure interferences producing OH inside the IU-FAGE detection cell, including laser generated OH. The measured interferences were subtracted from the total OH signal determined from spectral modulation, resulting in net ambient OH concentrations (Fig. 2). As can be seen from this figure, the measured interference was a significant fraction of the total OH signal on many days.

Figure 3 illustrates the total measured OH radical signal by spectral modulation (black circles), the measured interference (blue squares), and the expected laser-generated interference from reactions 3 and 4 calculated from laboratory calibrations (Griffith et al., 2016) (green points) during 14 July and 15 July. On 15 July, the measured interference was similar to the calculated interference suggesting that the majority of the measured interference was laser-generated. However, on 14 July, the measured interference was much larger than the calculated interference, suggesting that the majority of the measured interference was due to an unknown source. Subtraction of the calculated laser-generated interference from the measured interference on all days resulted in a measurement of the unknown interference that increased with both ozone and temperature during the campaign (Fig. 4).

This result is consistent with the observations from Mao et al. (2012) who found that the interference measured in their LIF-FAGE instrument using a similar chemical modulation technique increased with ozone and total OH reactivity. The observed increase in the magnitude of the unknown interference with ozone and temperature suggests that the interference may be related to the ozonolysis of biogenic VOCs, whose emissions increase with temperature. This result is also consistent with the measurements of Novelli et al. (2017), who found that their observed interference correlated with the product of ozone and biogenic VOC concentrations, although the correlation in the present study was weak ($R^2 = 0.15$). Previous measurements have shown that some LIF-FAGE instruments, including the IU-FAGE instrument, are susceptible to an interference under high concentrations of ozone and biogenic VOCs, perhaps due to the decomposition of Criegee intermediates inside the FAGE detection cell (Novelli et al., 2014; Fuchs et al., 2016; Novelli et al., 2017; Rickly and Stevens, 2018). However, estimated concentrations of Criegee intermediates in similar environments on the order of $5 \times 10^4 \text{ cm}^{-3}$ (Novelli et al., 2017) are too low to explain the observed interference during the IRRONIC campaign.

The observation of a significant interference during this campaign is in contrast to previous measurements of OH by the IU-FAGE instrument in a forested environment during the CABINEX 2009 campaign (Griffith et al., 2013). During this campaign, several tests were conducted where C_3F_6 or CO was added to remove ambient OH. These tests did not reveal any significant interference, and measurements of OH were found to be in good agreement with model predictions (Griffith et al., 2013). One possible explanation for this discrepancy with the measurements during IRRONIC is the lower levels of ozone and temperatures observed during CABINEX compared to IRRONIC. Average mixing ratios of ozone during CABINEX were near 30 ppb and average temperatures were near 20°C during the day, with average mixing ratios of isoprene less than 2 ppb in the afternoon. These levels of ozone and temperature are lower than that where the interference was observed during IRRONIC (Fig. 4), suggesting that a similar interference was likely undetectable during CABINEX.

Recent measurements have found that NO_3 radicals can lead to an interference in FAGE instruments (Fuchs et al., 2016), although the mechanism for production of this interference is not known. Such an interference in the IU-FAGE instrument could explain the observed interference during some nights (Fig. 3), but is unlikely the source of the interference during the daytime. Another possible source of the interference is the decomposition of ROOOH molecules inside the FAGE detection cell formed from the reaction of OH with RO_2 radicals (Fittschen et al., 2019). However, assuming a rate constant of $1 \times 10^{-10} \text{ cm}^3 \text{ s}^{-1}$ for the OH + RO_2 reaction, it is unlikely that a significant fraction of RO_2 radicals will react to form

ROOOH under the mixing ratios of NO observed at this site, as the estimated lifetime of RO₂ radicals with respect to reaction with NO was an order-of-magnitude shorter than that for reaction with OH. Additional measurements and laboratory tests will be needed to identify and minimize interferences associated with LIF-FAGE measurements of OH.

The day-to-day measurements of OH after the interference has been subtracted for 10-20 July and 24-25 July are also illustrated in Fig. 2. Because the interference was not measured simultaneously as the ambient OH measurements, subtraction of the measured interference often resulted in both apparent negative concentrations as well as large positive concentrations at night. These large positive and negative values reflect the fact that the nighttime measurements of the interference was much greater than the ambient OH signals and was highly variable between measurement cycles (Fig. 3), resulting in an ambient OH measurement uncertainty that was sometimes larger than the precision calculated from a quadratic propagation of the errors associated with the individual measurements of the ambient OH plus the interference and the interference alone. Measurements on 21-22 July focused on measurements of HO₂* as part of the peroxy radical informal instrumental intercomparison (Kundu et al., 2019), with NO added continuously to the detection cell to provide measurements with a higher time resolution. Thus OH measurements were not conducted on these days. This figure also illustrates the day-to-day model results for OH and HO₂* for the days when NO was also measured simultaneously, showing the MCM 3.3.1 and the RACM2-LIM1 models for simplicity.

Figure 5 (top) shows the average diurnal profile of the 15-min OH measurements, both with and without the measured interference, binned into 1 hour time periods for all the days illustrated in Fig. 2 (left panel), and binned into 2 hour time periods along with the MCM and RACM model results for the days when NO was measured simultaneously ([July 16, 18-20, 24, right panel](#)). The average ambient diurnal OH radical concentration measured during the entire campaign reached a maximum of approximately $4\text{--}5 \times 10^6 \text{ cm}^{-3}$ after the measured interference was subtracted (Fig. 5, top left), and was slightly lower during the period when NO measurements were available (Fig. 5, top right). If the measured interference was not subtracted from the total OH signal determined by spectral modulation, the resulting OH radical concentrations would be as high as $9 \times 10^6 \text{ cm}^{-3}$ (Fig. 5), much greater than the MCM and RACM2 modeled diurnal average maximum concentrations of approximately $2\text{--}3 \times 10^6 \text{ cm}^{-3}$. The daytime OH radical concentration measurements after the interference has been subtracted are in better agreement with the model results. Including versions of the LIM1 mechanism in both the MCM (3.3.1) and RACM2 (RACM2-LIM1) models [increases the predicted](#) daytime concentrations of OH [by approximately 13% and 20%, respectively](#) compared to the base mechanisms [during the day \(9:00-17:00 EDT\), when mixing ratios of NO on these days decreased from an average morning maximum near 500 ppt to approximately 50 ppt in the afternoon. The](#) RACM2-LIM1 results [are generally](#) within 30% of the measured concentrations during the day (Fig. 5, top right), [with the difference at 14:00 EDT within the combined measurement precision, the accuracy of the calibration \(18%, 1 \$\sigma\$ \), and the model uncertainty \(30%\).](#) [In contrast](#) the MCM 3.3.1 model underpredicted the measurements during this period by approximately a factor of 2. These results are similar to that found by Novelli et al. (2020), who found that the MCM 3.3.1 underpredicted measurements of OH by a factor of approximately 1.4 during isoprene oxidation experiments in the SAPHIR chamber [when mixing ratios of NO were less than 0.2 ppb, similar to the mixing ratios of NO measured in this study.](#) The measured OH concentrations could be

reproduced using a model that incorporated the larger equilibrium rate coefficients between the different peroxy radical isomers in the MCM 3.3.1 mechanism with the larger 1,6-H shift peroxy radical rate constants from the Caltech isoprene mechanism (Teng et al., 2017; Wennberg et al., 2018). This combination increased production of HPALDs and di-HPCARP-RO₂ radicals in the oxidation mechanism, resulting in an effective bulk isoprene peroxy radical isomerization rate similar to that in the original LIM1 mechanism (Novelli et al., 2020). These larger bulk peroxy radical isomerization rates are similar to that incorporated into the RACM2-LIM1 mechanism used in the present study (Tan et al., 2017), leading to the higher modeled radical concentrations compared to the MCM 3.3.1 model results shown in Fig. 5.

Although the RACM2-LIM1 mechanism appears to be able to reproduce the daytime OH radical measurements compared to the MCM 3.3.1 mechanism, all the models underestimate the measurements in the early evening and night. While there is uncertainty associated with these nighttime measurements due to the large interference that was subtracted, similar concentrations of OH were observed during the evening by both the IU-FAGE instrument and the University of Colorado Chemical Ionization Mass Spectrometry (CIMS) instrument at this site in 2017 during an informal instrument intercomparison (Rosales et al., 2018; Reidy et al., 2018). These results suggest that there may be a missing radical source during this period, such as the ozonolysis of unmeasured biogenic VOCs, and additional measurements will be required to resolve this discrepancy.

3.2 HO₂* measurements and model comparison

The day-to-day measurements of HO₂* are also illustrated in Fig. 2 with the MCM 3.3.1 and RACM2-LIM1 model results for the days when NO was also measured simultaneously. Figure 5 shows the average diurnal profile of the HO₂* measurements for all the days illustrated in Fig. 2 (left panel, bottom) which were similar to the measured HO₂* concentrations for the days when NO was measured simultaneously, along with the MCM and RACM model results (right panel, bottom). The contribution of modeled RO₂ radicals to the modeled HO₂* is based on laboratory calibrations of the RO₂-to-HO₂ conversion efficiencies for the sampling conditions used in this study (Lew et al., 2018) and are incorporated into both versions of the RACM2, and MCM peroxy radical categories. Under the instrumental conditions during the campaign, the conversion efficiency of isoprene-based peroxy radicals to HO₂ was determined to be approximately 83 ± 7%, while the conversion efficiency of methyl peroxy radicals was estimated to be approximately 5% (Lew et al., 2018). These two peroxy radicals accounted for the majority of RO₂ radicals predicted by the models (see below). The maximum measured HO₂* concentration each day during the campaign was generally between approximately 2 × 10⁸ and 2 × 10⁹ molecules cm⁻³ (Fig. 2), with an average daily maximum value of approximately 1 × 10⁹ cm⁻³ (Fig. 5). The RACM2-LIM1 and MCM 3.3.1 modeled diurnal averaged HO₂* reached a maximum of approximately 1.4 × 10⁹ cm⁻³ and 1.0 × 10⁹ cm⁻³, respectively, compared to values of 1.2 × 10⁹ cm⁻³ and 1.0 × 10⁹ cm⁻³, respectively, for the RACM2 and MCM 3.2 modeled HO₂* (Fig. 5).

The predicted HO₂* concentrations by the base RACM2 model are in good agreement with the measured concentrations when NO was measured simultaneously, with the model results within approximately ±10% of the measurements on average and within the calibration uncertainty of the measurements (38%, 2σ). Including the LIM1

mechanism in the RACM2 mechanism increases the modeled HO_2^* by approximately 15-20% due to the modeled increase in HO_x radical production from the isomerization of isoprene-based peroxy radicals, overpredicting the measurements by approximately 10-30%. These results are in contrast to that observed during the CABINEX campaign, where a RACM-based model overpredicted the measured HO_2^* by as much as a factor of 2 (Griffith et al., 2013), likely related to the higher concentrations of NO observed during IRRONIC compared to CABINEX increasing the importance of the $\text{HO}_2 + \text{NO}$ and $\text{RO}_2 + \text{NO}$ reactions in determining the fate of these radicals. The MCM-based model results are also in good agreement with the measured HO_2^* (Fig. 2 and 5). The MCM 3.3.1 and 3.2 mechanisms predicted HO_2^* concentrations are approximately 5-30% less than the measurements between 12:00-18:00 EDT but within the calibration uncertainty of the measurements, with the MCM 3.3.1 mechanism predicting slightly greater concentrations due to the inclusion of HO_x production from the isomerization of isoprene-based peroxy radicals.

The RACM2-LIM1 and MCM 3.3.1 diurnal average modeled HO_2^* concentrations and the model contribution of peroxy radicals to HO_2^* are shown in Fig. 6 for the days when NO was measured simultaneously (left panels). The diurnal profile of the HO_2^* radical concentration predicted by these models includes contributions primarily from isoprene peroxy radicals and HO_2 radicals, with smaller contributions from methyl peroxy and acetyl peroxy radicals (Fig. 6). The RACM2 and MCM 3.2 models produced similar results, with HO_2 and isoprene peroxy radicals contributing to the majority of the modeled HO_2^* concentrations (Fig. S1). The total modeled RO_x ($\text{RO}_2 + \text{HO}_2$) concentrations by the different mechanisms for these days are also shown in Fig. 6 (right panels). The RACM2-LIM1 model predicted that the diurnal average total RO_x concentration consisted primarily of HO_2 (42%), isoprene peroxy radicals (30%), methyl peroxy (CH_3O_2 , 14%), and acetyl peroxy (CH_3CO_3 , 4%), with daytime (08:00 – 20:00) contributions of 42%, 32%, 13%, and 4% for HO_2 , isoprene peroxy, CH_3O_2 , and CH_3CO_3 , respectively. The MCM 3.3.1 model predicted that HO_2 (43%), isoprene peroxy (18%), methyl peroxy (14%), acetyl peroxy (4%) were the major contributors to the modeled diurnal average total RO_x concentration, with daytime contributions of 43%, 20%, 13%, and 4% (Fig. 6). Similar results were obtained from the MCM 3.2 and RACM2 models (Fig. S1). As discussed above, the configuration of the IU-FAGE instrument used in this study converted approximately 83% of isoprene peroxy radicals to HO_2 upon addition of NO and minimally converts methyl peroxy radicals to HO_2 (<5%) (Lew et al., 2018). Thus, the majority of the contributing species to the measured HO_2^* are HO_2 and isoprene peroxy radicals which together account for approximately 70% of the total peroxy radical concentration predicted by these models. Measurements of the total $\text{HO}_2 + \text{RO}_2$ radical concentrations using an Ethane – Nitric Oxide Chemical Amplifier (ECHAMP) were found to be in good agreement with the HO_2^* measurements reported here and are summarized in Kundu et al. (2019).

3.3 Total OH reactivity measurements and model comparison

The measured total OH reactivity and that calculated from measured OH sinks using both the RACM and MCM mechanisms are shown in Fig. 7, where the measured OH reactivity is averaged into 2 hour bins. As illustrated in this figure, the calculated OH reactivity was in relatively good agreement with the measured OH reactivity on some days and nights, specifically 15-16 July, with missing reactivity observed later in the campaign. Overall, the averaged measured OH reactivity

varied between the instrumental limit of detection of 1 s^{-1} to a maximum of approximately 31 s^{-1} with an overall diurnal average value of approximately 13 s^{-1} .

The campaign diurnal averaged measured OH reactivity is shown in Fig. 8 along with the calculated total OH reactivity from the measured OH sinks. As expected for this deciduous forest environment, isoprene was the dominant contributor making up 37% of the diurnally averaged total reactivity, followed by OVOCs (28%), inorganics (10%), alkanes and alkenes (5%), anthropogenic non-methane hydrocarbons (NMHC) (1%), and monoterpenes (<1%) with missing reactivity accounting for the remaining 18% (Fig. S2). During the daytime (08:00 and 20:00) the contributions are similar, with isoprene being the largest contributor at 47% followed by OVOCs (24%), inorganics (8%), alkanes and alkenes (4%), anthropogenic NMHC (1%), and monoterpenes (<1%) with missing reactivity accounting for the remaining 14%. During the nighttime, (20:00 to 08:00), OVOCs were the dominant contributor to the modeled OH reactivity at 32% followed by isoprene (24%), inorganics (11%), alkanes and alkenes (6%), anthropogenic NMHC (2%), and monoterpenes (<1%) with missing reactivity of 24% (Fig. S2).

The campaign diurnal average (Fig. 8) shows a correlation with temperature, with the maximum average OH reactivity of approximately 20 s^{-1} occurring around 13:30. The calculated reactivity was consistent with the measured reactivity for temperatures less than 294 K, while the observed reactivity is greater than that calculated from the measured sinks for higher temperatures, although at temperatures above 302 K the measured reactivity appears to be less than calculated (Fig. S3). These results are similar to several previous studies in which the measured missing reactivity appeared to increase with temperature (Di Carlo et al., 2004; Hansen et al., 2014; Bsaibes, et al., 2020).

Figure 8 also shows the campaign average OH reactivity including the reactivity of unmeasured oxidation products predicted by the MCM 3.3.1 model. On average, including the contribution of unmeasured oxidation products can account for the majority of the missing reactivity. While the model tends to overpredict the average measured reactivity in the afternoon and evening, the model results agree to within the combined uncertainty of the model and the precision of the measurement (Hansen et al., 2014). Similar results were obtained by the RACM2 models, although the predicted reactivity of unmeasured oxidation products by the RACM2 models are approximately a factor of two smaller than that predicted by the MCM models (Fig. S4). These results suggest that the models are generally able to reproduce the measured OH reactivity at this site, and that the missing reactivity observed during IRRONIC may be due to unmeasured oxidation products, with isoprene nitrates and isoprene epoxides within the RACM2 and MCM mechanisms being the primary contributors to the missing reactivity.

While the campaign averaged OH reactivity measurements appear to be in reasonable agreement with the calculated reactivity based on measured compounds, there were several days that displayed large missing reactivity similar to that observed by Hansen et al. (2014). The MCM 3.3.1 model results for a day with the largest missing reactivity (17 July) is shown in Fig. 9, indicating that the modeled reactivity including unmeasured oxidation products cannot explain the observed reactivity on this day. The reason for this discrepancy is unclear, as the missing reactivity on this day did not appear to correlate with changes in wind speed, direction, trajectory, or meteorological conditions, but may indicate the presence of additional

unmeasured emissions or oxidation products not accounted for by the model. Additional measurements and analyses will be necessary to determine the source of the missing reactivity.

3.4 Radical budgets

The analysis of the rates of radical initiation, propagation, and termination can provide insight to the importance of individual radical sources and sinks. For the IRRONIC campaign, the OH radical budget is illustrated in Fig. 10, where OH radical production reactions are represented in shades of blue and loss reactions are represented in shades of red. Daytime production includes reactions with both initiation and propagation that produces OH radicals (positive rates), while daytime OH loss reactions are represented by propagation and termination reactions that remove OH (negative rates). For simplicity only the RACM2 and RACM2-LIM1 radical budgets are shown.

The maximum rates for the OH radical budget of approximately $5 \times 10^7 \text{ cm}^{-3} \text{ s}^{-1}$ from the RACM2-LIM1 model were higher than the maximum value of approximately $4 \times 10^7 \text{ cm}^{-3} \text{ s}^{-1}$ in RACM2. The addition of the LIM1 mechanism increases the OH radical production rate mostly from photolysis of hydroxyperoxy aldehydes (HPALD) produced from the isomerization of isoprene-based peroxy radicals and their subsequent chemistry (Peeters et al., 2014; Tan et al., 2017). In the RACM2-LIM1 model, the daytime OH radical production is dominated by the $\text{HO}_2 + \text{NO}$ reaction from 10:00 to 14:00 (66%) and drops to 47% from 14:00 to 18:00. Ozone photolysis and the LIM1 mechanism contribute up to 18-24% and 8-15% of the total OH radical production during the morning and afternoon, with ozonolysis ($\text{VOC} + \text{O}_3$) and photolysis of HONO, H_2O_2 , organic peroxides (OP1, OP2), and HNO_3 all contributing to 4-7% and 4-6% of the total OH radical production (Fig. 10). A majority of the OH radical loss is due to OH reactions with VOCs (65-72%) and OVOCs (27-23%) during the morning and afternoon. As described above, the measured total OH reactivity was in reasonable agreement with the modeled OH reactivity; therefore, it is likely that the total OH loss is well represented in the model. An experimental radical budget for the period when the measurements were complete suggests that the total measured OH production rate during the day is nearly balanced by the total OH loss rate based on the measured total OH reactivity (Fig. S5), consistent with the agreement between the measured and modeled OH on these days as discussed above and within the combined uncertainties of all the measurements (38% (2σ) for OH and for HO_2^* for example), similar to that observed previously (Tan et al., 2019). However, the experimental budget is not balanced in the evening, suggesting that there may be a missing radical source during this period, consistent with the model underprediction of the OH measurements discussed above.

The total radical (RO_x) budget from the RACM2 mechanisms of OH, HO_2 , and RO_2 radicals is illustrated in Fig. 11. Overall, total radical initiation in the RACM2-LIM1 mechanism was larger, with a maximum value of approaching $3 \times 10^7 \text{ cm}^{-3} \text{ s}^{-1}$ compared to RACM2 maximum value of approximately $2 \times 10^7 \text{ cm}^{-3} \text{ s}^{-1}$. The increase in total radical initiation in the RACM2-LIM1 model is due to both the added radical initiation from the photolysis of HPALDs as well as increased radical initiation from other aldehydes produced in the LIM1 mechanism. Overall, radical initiation from the photolysis of HPALDs and the subsequent chemistry from the LIM1 mechanism contributed 16-22% of total radical initiation during the morning and afternoon, while photolysis of formaldehyde and others (HONO , H_2O_2 , aldehydes, organic peroxides, HNO_3) together

contributed to approximately 42% of total radical initiation, with ozone photolysis contributing to 29-33% of radical initiation (Fig. 11). In contrast, ozone photolysis contributes to approximately 44-46% of radical initiation in the RACM2 mechanism, with formaldehyde and other aldehydes together contributing 43% (Fig. 11). Radical termination for both mechanisms is dominated by peroxy radical self-reactions, such as the $\text{HO}_2 + \text{HO}_2$ reaction, as well as the reaction of HO_2 with isoprene-based peroxy radicals (ISOP) and other peroxy radicals (RO_2). These reactions account for approximately 67-95% of radical termination due to the low levels of NO_x used in the models, with reaction of $\text{OH} + \text{NO}_2$ and other NO_x radical reactions accounting for approximately 5-33% of radical termination in these models during the day (Fig. 11).

The partitioning of the total radical budget production for IRRONIC is similar to the modeled budget observed during PROPHET 2008 and CABINEX 2009 (Griffith et al., 2013). The updated RACM model used during these campaigns predicted that radical termination was dominated by $\text{HO}_2 + \text{RO}_2$ reactions (including the $\text{HO}_2 + \text{ISOP}$ reaction), contributing to approximately 80% of total radical termination, similar to the 67-84% for the $\text{HO}_2 + \text{ISOP}$, $\text{HO}_2 + \text{RO}_2$, and $\text{HO}_2 + \text{HO}_2$ reactions predicted here by the RACM2 model. The photolysis of ozone accounted for approximately 20-30% of total radical initiation during these campaigns based on an updated version of the RACM model (Griffith et al., 2013) compared to approximately 45% predicted by the RACM2 mechanism during IRRONIC due to higher concentrations observed during this campaign. Ozonolysis reactions contributed to approximately 20-30% of total radical initiation during PROPHET and CABINEX compared to approximately 12% during IRRONIC. Photolysis of HCHO contributed to approximately 22-24% of the total rate of radical initiation during IRRONIC compared to 23% and 5% during PROPHET 2008 and CABINEX 2009, respectively, with the low contribution during CABINEX primarily due to the lower mixing ratios of HCHO observed during this campaign (Griffith et al., 2013). In contrast, photolysis of HONO was a significant radical source during PROPHET and CABINEX, contributing 14-17% of radical initiation compared to approximately 3% of total radical production during IRRONIC due to the lower mixing ratios of HONO observed during IRRONIC. On average, mixing ratios of HONO during IRRONIC were approximately 40 ppt at night decreasing to approximately 10 ppt during the day (Fig. S6) compared to daytime mixing ratios between 50 and 75 ppt during PROPHET and CABINEX (Griffith et al., 2013). The reason for the difference in the measured HONO values between these two sites is unclear, but may be related to increased production from photolysis of nitric acid on the forest canopy surfaces at the PROPHET site (Zhou et al., 2011).

4 Summary

Measurements of OH radical concentrations using the IU-FAGE instrument during the IRRONIC campaign revealed a significant unknown interference that appeared to correlate with both temperature and ozone. The average measured OH radical concentration after the interference was subtracted reached an average daytime maximum of approximately $4\text{--}5 \times 10^6 \text{ cm}^{-3}$. This is in contrast to the measurements including the interference which reached an average daytime maximum of approximately $9 \times 10^6 \text{ cm}^{-3}$. Similar concentrations of OH were observed after any measured interference was subtracted at

1 this site in 2017 during an informal intercomparison between the IU-FAGE instrument and the University of Colorado
2 Chemical Ionization Mass Spectrometry (CIMS) instrument (Rosales et al., 2018; Reidy et al., 2018).

3 After subtracting the interference, the OH measurements were in better agreement with model simulations utilizing
4 the Regional Atmospheric Chemical Mechanism 2 (RACM2) with an updated Leuven Isoprene Mechanism (LIM1) as well as
5 the Master Chemical Mechanism versions 3.2 and 3.3.1. Both the RACM2-LIM1 and MCM 3.3.1 mechanisms add radical
6 recycling reactions for isoprene oxidation that increase the modeled OH and peroxy radical concentrations. Similar to the
7 results of Novelli et al. (2020), the RACM2-LIM1 model results were in better agreement compared to the MCM 3.3.1
8 mechanism, predicting maximum daytime OH concentrations that were within 30% of the measurements, likely due to a larger
9 bulk isoprene peroxy radical isomerization rate leading to a greater rate of radical production. However, the models tend to
10 underpredict the measured concentrations during the evening, suggesting that a significant radical source may be missing from
11 the models. Additional measurements are needed in order to resolve this discrepancy. Nevertheless, it is clear that if the
12 measured interference was not taken into account, the apparent OH concentrations would have been a factor of 2-4 greater
13 than predicted by the model mechanisms, comparable to previous measurements under low NO_x and high isoprene conditions
14 (Rohrer et al., 2014). These results are similar to that reported by Mao et al. (2012) and Mallik et al. (2017) who found good
15 agreement between their OH measurements and model predictions when measured interferences were taken into account.
16 Because of differences in instrument design (geometry, cell pressure, flow, etc.) these interferences may not significantly
17 impact other LIF-FAGE instruments. However, future OH measurements using the LIF-FAGE technique should include
18 methods to quantify potential instrumental artifacts even if they are insignificant, to demonstrate that the measurements are
19 free from interferences.

20 Measurements of total OH reactivity were in reasonable agreement with that calculated from measured OH sinks,
21 with isoprene contributing approximately 37% and OVOCs 28% of the diurnally averaged measured reactivity, with 18% of
22 the measured reactivity missing. However, on average the missing reactivity fraction can be explained by unmeasured
23 oxidation products, specifically from isoprene nitrates and isoprene epoxides within the RACM2 and MCM mechanisms. This
24 indicates that these mechanisms are accurately representing the total OH loss at this site.

25 Measurements of HO₂ radicals by the IU-FAGE instrument using chemical conversion to OH by addition of NO has
26 been shown to be sensitive to alkene-based peroxy radicals (Lew et al., 2018). As a result, the measurements represent a sum
27 of HO₂ and a fraction of RO₂ radicals in the atmosphere (HO₂*). During the IRRONIC campaign, the measured HO₂*
28 concentration primarily reflected the sum of HO₂ and isoprene-based peroxy radicals, which contributed to approximately 70%
29 of the total modeled peroxy radicals. The average daytime ambient HO₂* measurements reached maximum concentrations of
30 approximately $1 \times 10^9 \text{ cm}^{-3}$. Both MCM models predicted HO₂* concentrations that were within 10-30% but generally lower
31 than the measurements, while the RACM mechanisms resulting in predicted concentrations that were within 10-30% but
32 generally greater than the measurements. However, all models predicted concentrations that were within the combined
33 uncertainty of both the model and the measurement. These results are in contrast to some previous measurements in forest
34 environments where model predictions were found to be significantly greater than measured HO₂* concentrations (Griffith et

1 al., 2013), perhaps as a result of the lower mixing ratios of NO observed at these sites. Additional measurements are needed
2 in order to resolve this discrepancy, which may be related to a gap in our understanding of peroxy radical chemistry under low
3 NO conditions.

Data availability. Data are available upon request from the corresponding author (pstevens@indiana.edu).

Competing interests. The authors declare that they have no conflicts of interest.

Author contributions. PS, SD and EW designed the research project. ML, PR, BB, and PS were responsible for the LIF-FAGE OH, HO₂*, OH reactivity, and HONO measurements. SK and EW were responsible for the supporting measurements of NO, NO₂, and O₃. SD, SS, TL, and NL were responsible for the measurements of VOCs and OVOCs. ML, PR, ER, and PS conducted the analysis and photochemical modelling and wrote the paper with feedback from all co-authors. ML and PR contributed equally to the paper.

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Table 1: Measurements conducted during the IRRONIC field campaign.

Measurement	Instrument	Technique	LOD	Reference
OH	LIF-FAGE	Laser-induced fluorescence –	$8 \times 10^5 \text{ cm}^{-3} / 30 \text{ min}$	Dusanter et al., 2009a;
HO ₂ *		fluorescence assay by gas expansion	$7 \times 10^7 \text{ cm}^{-3} / 20 \text{ s}$	Lew et al., 2018 ^a
NO	Thermo 42i-TL	Chemiluminescence	50 ppt / 2 min	
NO ₂	Aerodyne CAPS	Cavity attenuated phase shift spectroscopy	40 ppt / 10 s	
Ozone	2B Technologies Model 202	UV absorbance	3 ppb / 10 s	
OH reactivity	LIF-TOHLM	Total OH Loss Measurement	1 s ⁻¹ (10 min)	Hansen et al., 2013
HONO	LP LIF-FAGE	Laser-photofragmentation laser-induced fluorescence	20 ppt (30 min)	Bottorff et al., in prep
NMHCs	Online GC/FID	Gas chromatography with flame ionization detection	10-100 ppt (1.5 hr)	Badol et al., 2004
OVOCs	Online GC/FID-MS	Gas chromatography with mass spectrometer and FID	5-100 ppt (1.5 hr)	Roukos et al. (2009)
	Off-line Sorbent GC-MS	Sorbent cartridges analyzed by GC-MS		Detournay et al. (2011); Ait-Helal et al. (2014)
	Off-line DNPH HPLC-UV	Dinitrophenylhydrazine cartridges analyzed by high-performance liquid chromatography with UV detection		
J(NO ₂)		Spectral Radiometry	$0.3 \times 10^{-4} \text{ s}^{-1}$	Shetter and Muller (1999)

^aHO₂* = HO₂ plus a fraction of certain RO₂ radicals (see text)

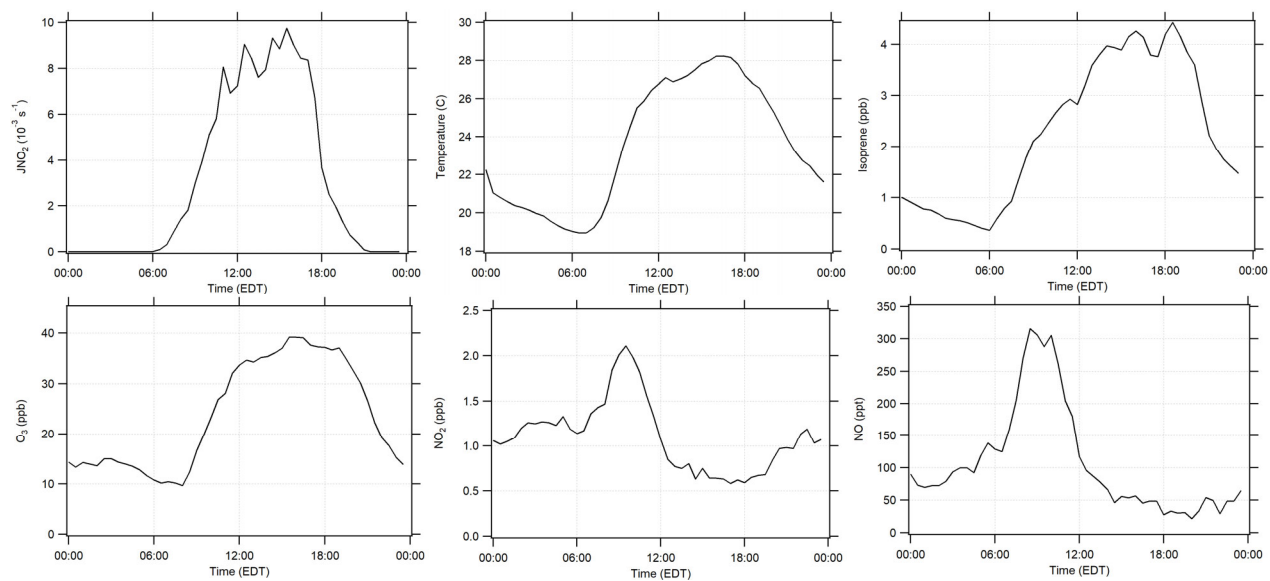


Figure 1. Diurnal campaign average profiles of J(NO₂), temperature, isoprene, O₃, NO₂, and NO.

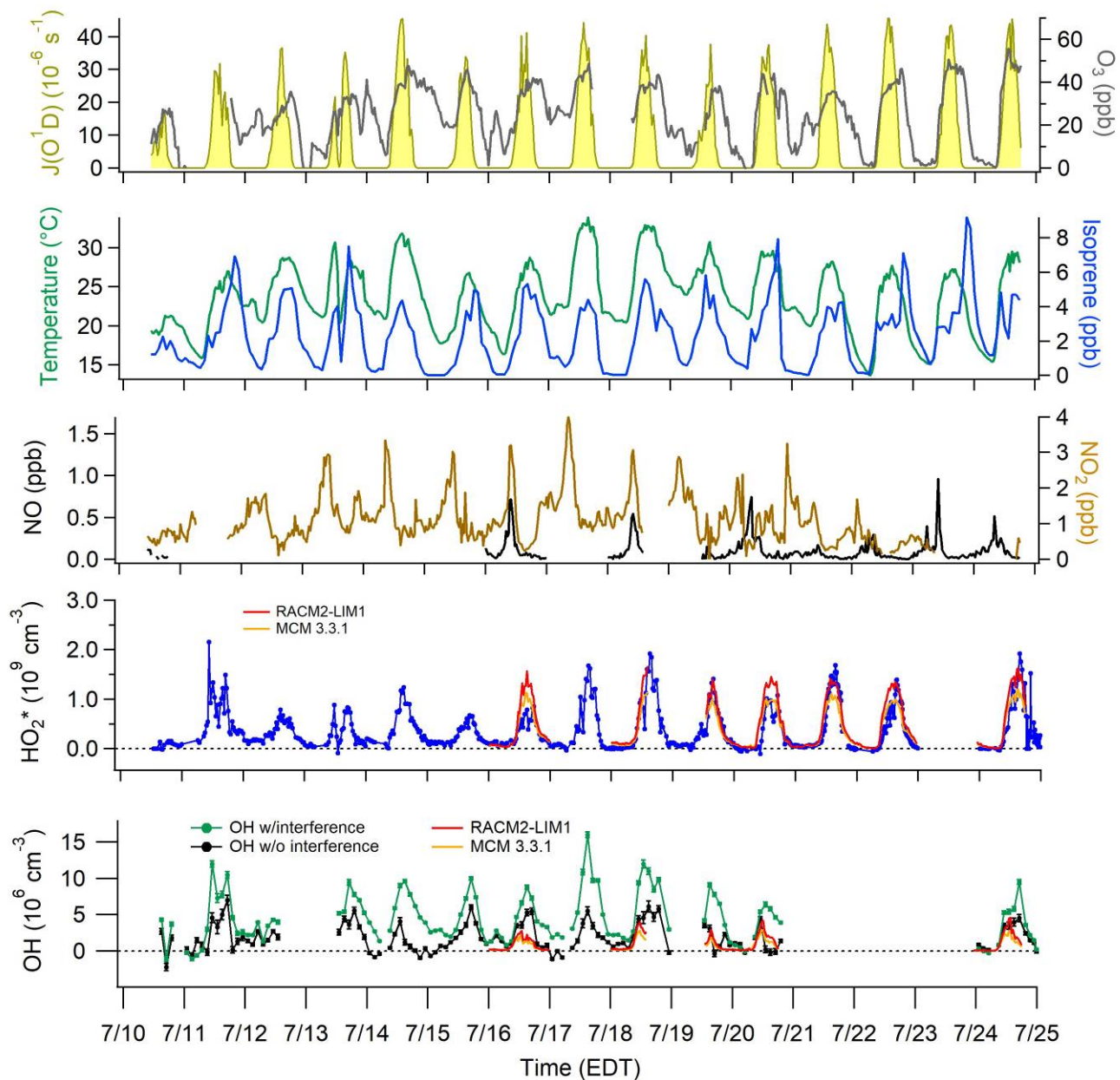


Figure 2. Time series of OH and HO_2^* from July 10 to July 25 with model calculated $J(O^1D)$ scaled to the measured $J(NO_2)$, and measured ozone, temperature, isoprene, and NO_x . OH measurements with interference ($\pm 1\sigma$) represented by the green line and measurements without interference ($\pm 1\sigma$) represented by the black line. For clarity, OH data shown are 2 hour averages. HO_2^* data are 30 s averages every 30 minutes. The daily RACM2-LIM1 and MCM 3.3.1 model results for the periods when NO was measured simultaneously are also shown.

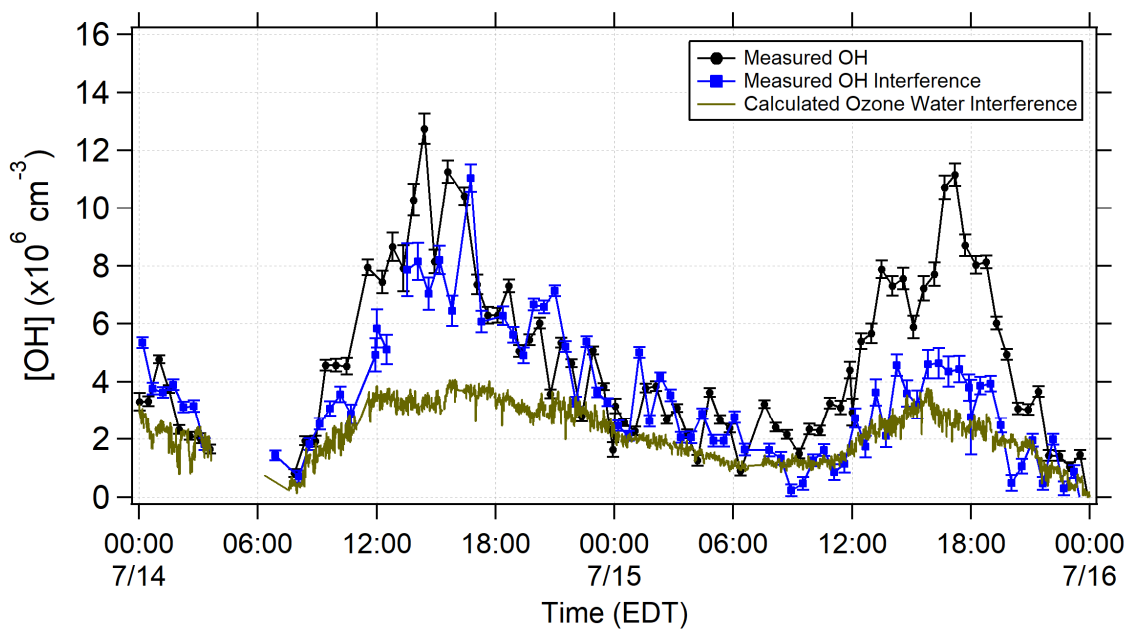


Figure 3. Averaged measured total OH signal using spectral modulation (black), and the measured interference using chemical modulation (blue) during July 14 and July 15. The calculated laser-generated interference from ozone photolysis for these days (reactions 1 and 2, green points) is also shown.

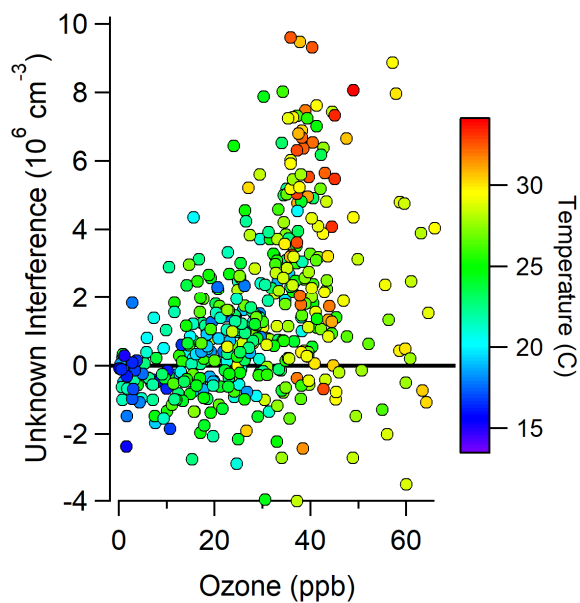


Figure 4. Measurements of the unknown interference as a function of ozone and temperature during the campaign.

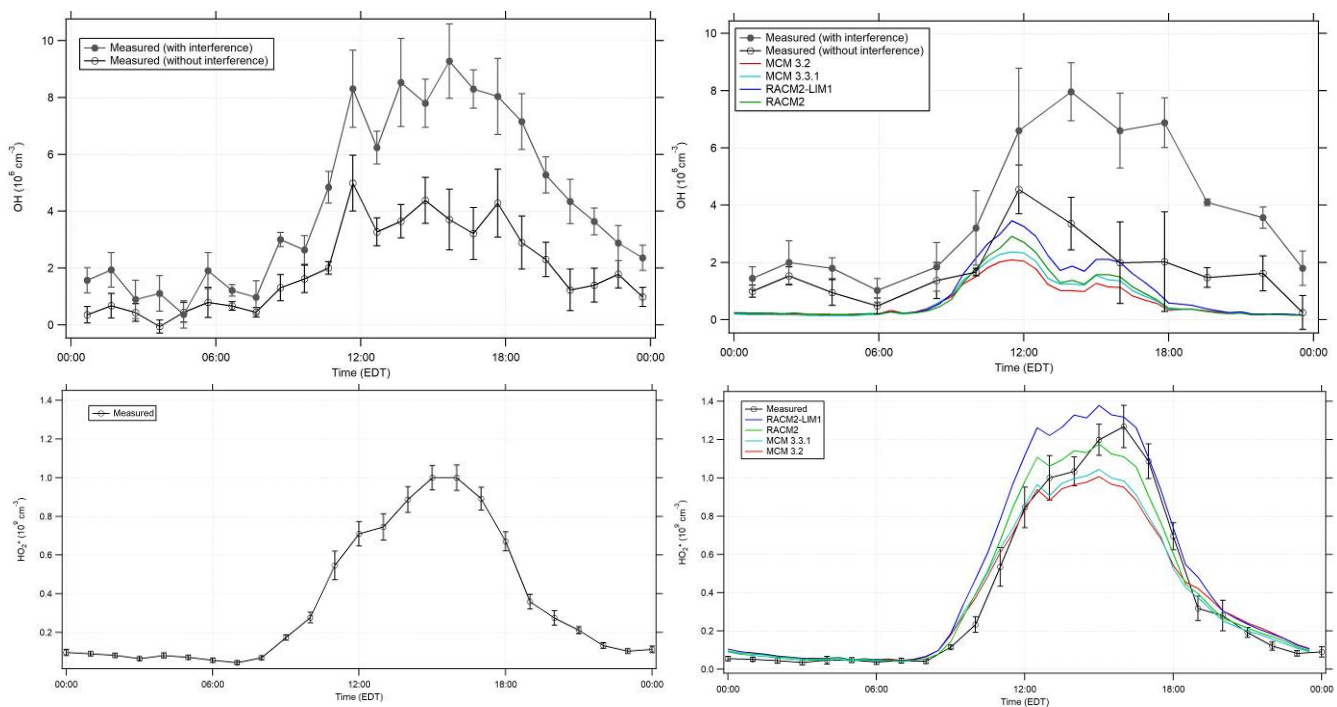


Figure 5. Diurnal profiles of OH (top) and HO₂* (bottom) with the RACM2, RACM2-LIM1, MCM 3.2, and MCM 3.3.1 model results. Left panels illustrate the average of all measurements during the campaign, while the right panels illustrate the average measurements when NO was measured simultaneously (see text). The open circles represent the 1 or 2 hour mean $\pm 1\sigma$ standard error of OH and HO₂* measurements. The filled circles represent the 1 or 2 hour mean $\pm 1\sigma$ standard error of the OH measurements with the interference.

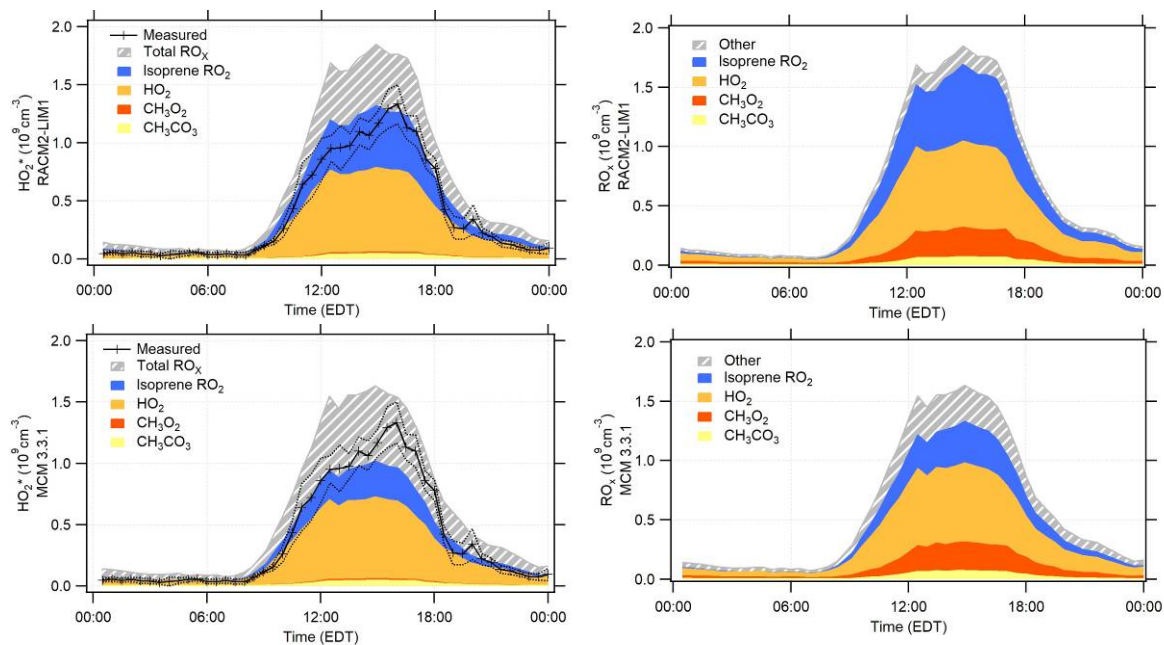


Figure 6. The RACM2-LIM1, and MCM 3.3.1 diurnal average modeled peroxy radical concentration and composition for the days when NO was measured simultaneously (see text). Left panels show the modeled contribution to the HO_2^* concentrations. The measured 30-min mean HO_2^* concentrations are shown by the black line with $\pm 1\sigma$ standard error of the measurements shown by the dotted lines. The calibration uncertainty of the measurements (not shown) is 38% (2σ). Right panels show the total RO_x ($\text{RO}_2 + \text{HO}_2$) composition predicted by each model.

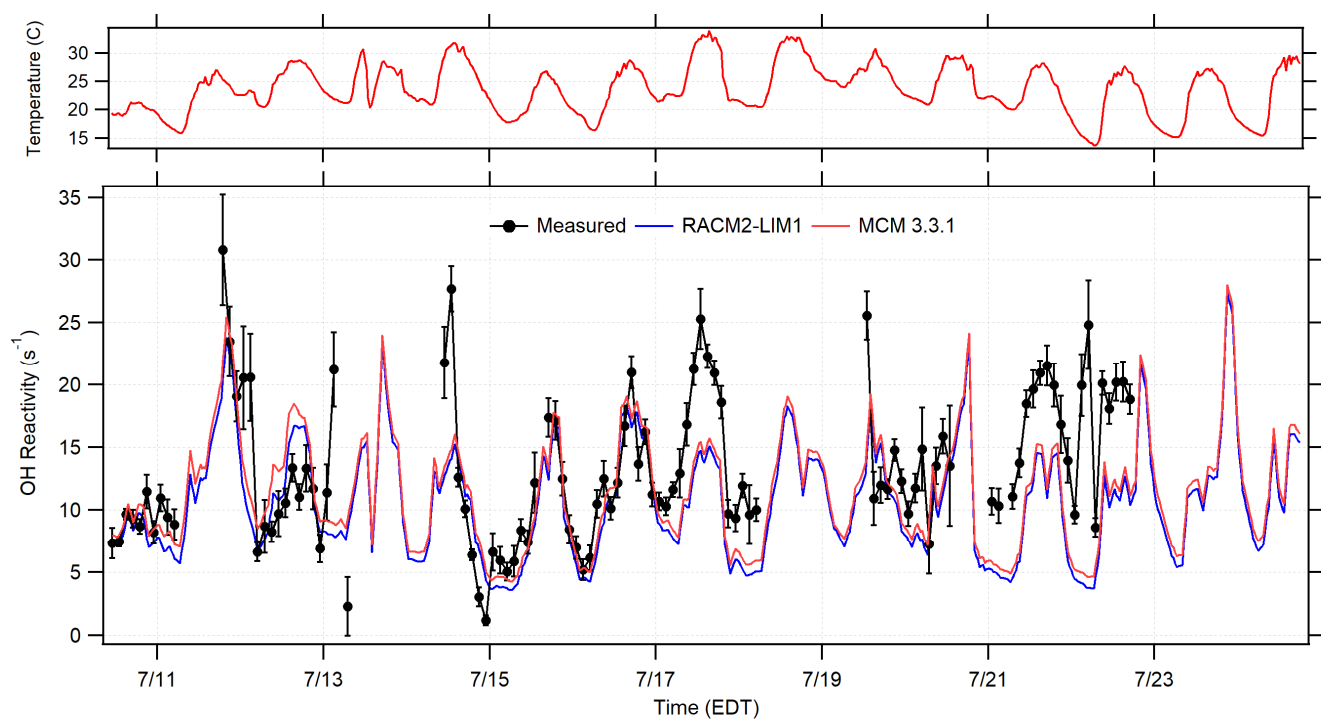


Figure 7. Time series of the 2 hour averaged OH reactivity measurements (black circles) in comparison to the RACM2-LIM1 and MCM 3.3.1 calculated OH reactivity based on measured OH sinks along with ambient temperature (top). Error bars represent the standard error of the average measurement.

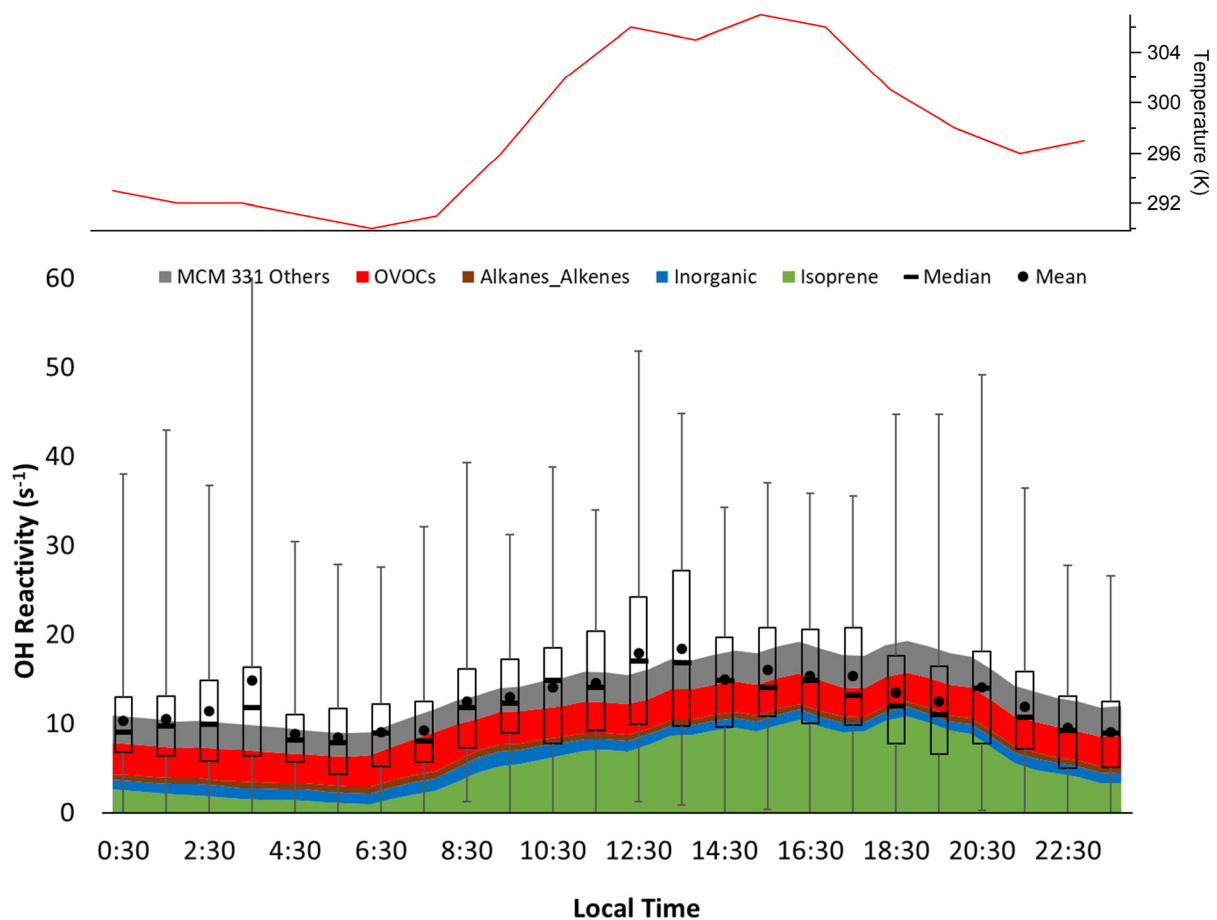


Figure 8. Diurnal temperature (top) and box and whiskers plot of observed total OH reactivity measured during the entire campaign showing the mean and median values for each hour, with the mean calculated values from the measured OH sinks as well as the unmeasured oxidation products from the MCM 3.3.1 model results (Others). Error bars show the range of individual 5-min measurements and bars show Q1 and Q3 for the measured OH reactivity.

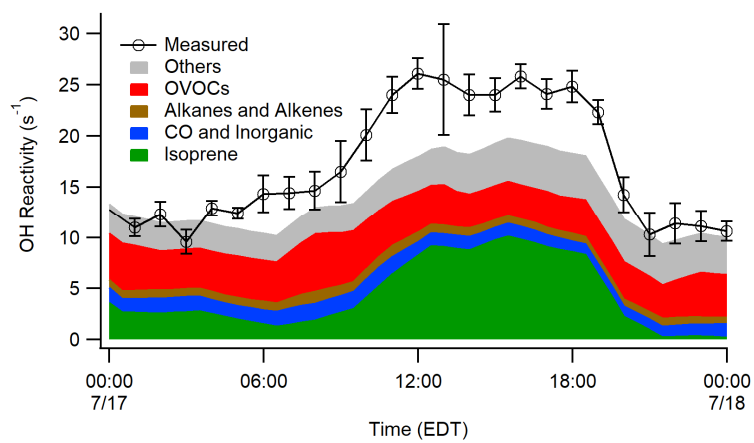


Figure 9. Median OH reactivity measurements from July 17 in comparison to modeled reactivity on this day from the MCM 3.3.1 mechanism.

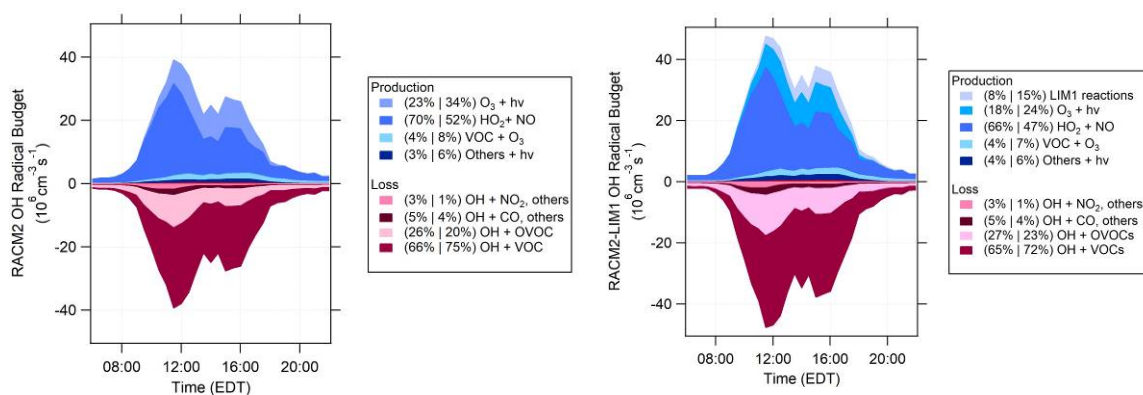


Figure 10. RACM2 (left) and RACM2-LIM1 (right) OH radical budgets for the days with NO measurements. Shades of blue represent production reactions and the shades of red represent loss rates. The percent contribution of each reaction to total production/loss are divided into two periods (10:00 to 14:00 and 14:00 to 18:00). The “Others + hv” category includes the photolysis of HONO, H₂O₂, organic peroxides, and HNO₃ (see text).

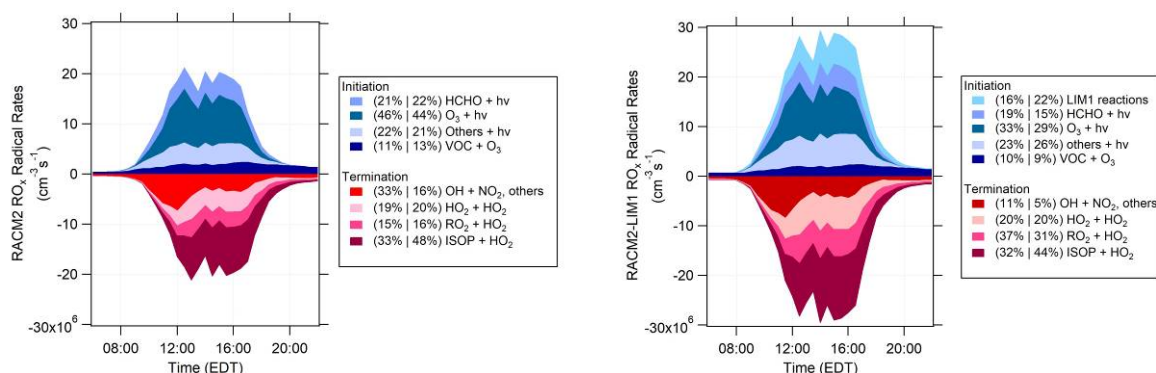


Figure 11. RACM2 (left) and RACM2-LIM1 (right) total RO_x radical budgets for the days with NO measurements. Shades of blue represent initiation rates and the shades of red represent termination rates. The percent contribution of each reaction to total initiation/termination are divided into two periods (10:00 to 14:00 and 14:00 to 18:00). The “Others + hv” category includes the photolysis of HONO, H₂O₂, aldehydes, organic peroxides, and HNO₃ (see text).