Technical Note: Preparation and purification of atmospherically relevant α -hydroxynitrate esters of monoterpenes

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Abstract. Organic nitrate esters are key products of terpene oxidation in the atmosphere. We report here the preparation and purification of nine nitrate esters derived from (+)-3-carene, limonene, α -pinene, β -pinene and perillic alcohol. The availability of these compounds will enable detailed investigations into the structure reactivity relationships of aerosol formation and processing and will allow individual investigations into aqueous phase reactions of organic nitrate esters.

1 Introduction

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Biogenic volatile organic compound (BVOC) emissions account for ~88% of non-methane VOC emissions. Of the total BVOC estimated by the Model of Emission of Gases and Aerosols from Nature version 2.1 (MEGAN2.1), isoprene is estimated to comprise half, and methanol, ethanol, acetaldehyde, acetone, α-pinene, β-pinene, limonene, ethene and propene together encompass another 30%. Of the terpenoids, α-pinene alone is estimated to generate ~66 Tg year ⁻¹ (Guenther et al., 2012). These monoterpenes can be oxidized by nitrate radicals that are projected to account for more than half of the monoterpene-derived secondary organic aerosol (SOA) in the US (Pye et al., 2010). Nitrate oxidation pathways have been shown to be important particularly during nighttime. A large portion (30-40%) of monoterpene emissions occur at night (Pye et al., 2010). These emissions can then react with NO₃ radicals, formed from the oxidation of NO₂ emissions by O₃, (Pye et al., 2010).

The full role of organic nitrates (ON) is complicated with many different sources and sinks (Perring et al., 2013). Fully deconvoluting the atmospheric processing of terpene-derived ON is difficult particularly due to partitioning into the aerosol phase in which hydrolysis and other reactivity can occur (Bleier and Elrod, 2013;

Rindelaub et al., 2014; 2015; Romonosky et al., 2015; Thomas et al., 2016). Hydrolysis reactions of nitrate esters of isoprene have been studied directly (Jacobs et al., 2014) and the hydrolysis of ON have been studied in bulk (Baker and Easty, 1950). These and other studies have shown that the hydrolysis of ON is dependent on structure (Darer et al., 2011). For example, primary and secondary ON are thought to be relatively stable (Hu et al., 2011). In contrast, tertiary nitrates have been shown to hydrolyze on the order of hours (Boyd et al., 2015; Liu et al., 2012) to minutes (Darer et al., 2011). To the best of our knowledge there is only one study of the hydrolysis of an isolated terpene-derived hydroxynitrate (2) (Rindelaub et al., 2016).

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Figure 1: Two hydroxynitrate esters with available spectral data. Relative stereochemistry is undefined.

Furthermore, fully understanding the atmospheric processes of organic molecules is restricted by the ability to identify these species (Nozière et al., 2015). Part of this challenge is, of course, related to the lack of available standards. While one certainly cannot synthesize all of the atmospherically relevant ON, having access to representative compounds from monoterpenes would enable key studies. With these molecules in hand, the atmospheric chemistry community could directly study the ON reactivity, such as hydrolysis, and deconvolute the structure-reactivity relationships. Additionally, novel method development would be enabled and validated (Rindelaub et al., 2016b). For example, Nozière and co-authors called attention to "the lack of NMR spectra libraries for atmospheric markers" as a barrier for utilizing NMR spectroscopy in atmospheric science. While the preparation of α -hydroxynitrates of terpenes has been alluded to in a handful of reports, we have only been able to identify NMR data for two species (Figure 1). Two main methods appear to have been attempted, treating an epoxide with either fuming nitric acid (Rollins et al., 2010) or bismuth nitrate (Rindelaub et al., 2016b; Romonosky et al., 2015). The utility of the former is limited by the extreme hazards involved with mixing fuming nitric acid with organic materials (Parker, 1995; Univ. of Calif., Berkeley, 2018) and only provides characterization data for a single compound. On the surface the latter appears to be a usable method but on closer inspection lacks spectral data, perhaps due to intractable and inseparable mixtures (Romonosky et al., 2015) and has been subject of a retracted study (Pöschl et al., 2011).

2 Experimental

2.1 Instrumentation and materials

All starting materials and reagents were purchased from commercial sources and used without further purification unless otherwise noted. Bismuth nitrate (98%) was purchased from Strem Chemicals and ground to a fine powder prior to use. Isoprene, MVK, (+)-3-carene, β-pinene, α-pinene oxide and 1,2-limonene oxide were purchased from Aldrich. Silica gel chromatography was performed on a TeldyneIsco CombiFlash+ Lumen using 25 μm SiliCycle spherical silica gel. ¹H and ¹³C NMR spectra were recorded on a Bruker AV400 spectrometer with chloroform (7.26 ppm) or benzene (7.28 ppm) as internal standards. IR spectra were recorded on a ThermoNicolet IR100 spectrometer using a Thunderdome Attenuated Total Reflectance (ATR) sample accessory. Melting points were collected on a DigiMelt 160 melting point apparatus. Exact masses were collected at the BioAnalytical Mass Spectrometry facility at Portland State University using a ThermoElectron LTQ-Orbitrap Discovery high resolution mass spectrometer with a dedicated Accela HPLC system. Low resolution masses were collected on a Varian Saturn 2000 GC-MS with a Restek Crossbind Rxi-5ms column. Abbreviations include: *meta*-chloroperoxybenzoic acid (m-CPBA), hexanes (Hex), ethyl acetate (EtOAc), column volume (CV), thin layer chromatography (TLC), retention factor (R_f), diastereomeric ratio (d.r.), broad (br), singlet (s), doublet (d), doublet of doublets (dd), doublet of doublets (dd), doublet of quartet (dq), triplet (t), multiplet (m), broad (b), weak (w), medium (m), strong (s), very strong (vs).

2.2 Preparation of epoxides

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2.2.1 trans-Carene oxide (3)

To a solution of (+)-3-carene (0.86 g, 1 mL, 7.34 mmol) in dichloromethane (25 mL) at 0 °C was added *meta*-chloroperoxybenzoic acid (m-CPBA) (1.85 g, 8.1 mmol, 1.1 equiv). The solution was warmed to 23 °C over 1 hour. The solution was poured onto sat. aq sodium bicarbonate (30 mL) and extracted with dichloromethane (3 x 30 mL). The combined organics were washed with sat. aq sodium bicarbonate (2 x 30 mL), dried (MgSO₄) and concentrated to yield a crude clear, colorless oil (1.17 g). The crude oil was purified by column chromatography (SiO₂; 0 – 25% EtOAc/Hex over 10 column volumes) to yield clear colorless oil (685 mg, 4.5 mmol; 61%

yield): ¹H NMR (400 MHz, chloroform-d) δ 2.82 (s, 1H), 2.28 (ddd, J = 16.4, 9.0, 1.9 Hz, 1H), 2.13 (dd, J = 16.2, 9.1 Hz, 1H), 1.63 (dt, J = 16.4, 2.3 Hz, 1H), 1.48 (dd, J = 16.1, 2.3 Hz, 1H), 1.24 (s, 3H), 1.00 (s, 3H), 0.72 (s, 3H), 0.52 (td, J = 9.1, 2.3 Hz, 1H), 0.44 (td, J = 9.1, 2.3 Hz, 1H) ppm. Spectral data is consistent with literature reports (Cabaj et al., 2009).



2.2.2 *cis*-Carene oxide (6)

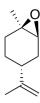
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According to the methods of Crocker and Grayson, 1969, a round bottom flask was charged with a solution of (+)-3-carene (2.7 g, 20 mmol) in dioxane (20 mL) and water (10 mL). Calcium carbonate (2 g, 20 mmol) and N-bromosuccinimide (7 g, 40 mmol) were added to the solution. The internal temperature rose to 50 °C after the initial addition. The mixture was stirred for 2 h then poured onto water (50 mL), filtered, and washed with diethyl ether. The filtrate was extracted with ether (2 x 100 mL). The combined extract was washed with water (3 x 100 mL), sodium thiosulfate (5% aq, 20 mL), dried (Na₂SO₄) and concentrated to yield a crude pale yellow oil. Purification by column chromatography (SiO₂; 0 - 50% EtOAc/Hex) yielded the bromohydrin as a white crystalline solid (4.6 g, 99% yield): ¹H NMR (400 MHz, chloroform-d) δ 4.07 (dd, J = 11.1, 7.6 Hz, 1H), 2.50 – $2.36 \text{ (m, 2H)}, 2.21 \text{ (dd, } J = 14.6, 10.1 \text{ Hz}, 1\text{H)}, 1.41 \text{ (dd, } J = 4.9, 1.2 \text{ Hz}, 1\text{H)}, 1.38 \text{ (s, 3H)}, 1.03 \text{ (s, 3H)}, 1.00 \text{ (s$ 3H), 0.91 – 0.80 (m, 1H), 0.70 (m, 1H). Spectral data is consistent with literature reports (Cocker & Grayson 1969). Bromohydrin (2.44 g, 10.5 mmol) was dissolved in 100 mL tBuOH (warmed with a water bath). Potassium *tert*-butoxide (2.44 g, 21.8 mmol, 2.08 equiv) was added and the solution was stirred at rt for 2 hours. The solution was poured onto water (50 mL), extracted with diethyl ether (3 x 75 mL), washed with water (3 x 50 mL), dried (MgSO₄) and concentrated to yield 2.5 g crude clear oil. Purification by column chromatography (40 g SiO₂; 0 – 25% EtOAc/Hex over 10 CVs) yielded a clear colorless oil (1.31 g, 63% yield): ¹H NMR (400 MHz, chloroform-d) δ 2.89 (d, J = 5.6 Hz, 1H), 2.30 (ddd, J = 16.7, 9.0, 5.6 Hz, 1H), 2.08 (dd, J = 16.4, 8.9 Hz, 1H), 1.82 (s, 1H), 1.78 (s, 1H), 1.32 (s, 3H), 0.98 (s, 3H), 0.94 (s, 3H), 0.63 – 0.49 (m, 2H). Spectral data is consistent with literature reports (Cocker and Grayson, 1969).

2.2.3 8,9-Limonene oxide (9)

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Methyl vinyl ketone (10 mL, 90 mmol), isoprene (10 mL, 75 mmol), and DCM (90 mL) were added to a round bottom flask with stir bar. The flask was purged with inert atmosphere and chilled to 0°C with stirring. AlCl₃ (1.2 g, 9 mmol) was added in three portions over 10 minutes. The ice bath was removed and the reaction mixture was stirred for 1.5 hours. The crude reaction mixture was filtered through a 1.5 inch silica gel pad (350 mL, 9 cm diameter) washed with 8% EtOAc:Hex (4 x 160 mL). The filtrate was concentrated to yield 1-(4methyl-3-cyclohexene) ethenone (8) as a clear yellow liquid (9.68 g, 88% yield): ¹H NMR (400 MHz, chloroform-d) δ 5.44 - 5.38 (m, 1H), 2.59 - 2.49 (m, 1H), 2.15 - 2.20 (s, 4H), 2.06 - 1.95 (m, 3H), 1.67 (s, 3H), 1.61 (m, 2H). Spectral data is consistent with literature reports (Buss et al., 1987). Sodium hydride (1.766 g, 44.16 mmol) was suspended in dry DMSO (25 mL). A solution of trimethylsulfonium iodide (10.356 g, 47.06 mmol) in DMSO (50 mL) was added via cannula. The solution was stirred at rt for 30 minutes then heated to 50 °C until gas evolution ceased (1 h). To this solution was added 1-(4-methyl-3-cyclohexen-1-yl) ethenone 8 (5 g, 36 mmol) and heated to 70 °C for 2 hours. The reaction mixture was cooled to room temperature and poured onto sat. aq. NH₄Cl (100 mL), extracted with MTBE (4 x 100 mL). The combined organics were washed with water (4 x 50 mL), dried (MgSO₄) and concentrated to yield 5.1 g crude oil. Purification by column chromatography (80 g SiO₂; 0-20% EtOAc/Hex over 10 CV) yielded 8,9-limonene oxide (1:1 d.r.) as a clear, colorless oil (3.76 g, 68% yield): ¹H NMR (400 MHz, chloroform-d) δ 5.43 – 5.33 (br s, 1H), 2.67 (t, J = 5.3Hz, 1H), 2.58 (dd, J = 10.9, 4.8 Hz, 1H), 2.17 - 1.73 (m, 5H), 1.66 (s, 3H), 1.44 (m, 2H), 1.29 (d, J = 4.5 Hz, 3H). Spectral data is consistent with literature reports (Almeida and Jr., 2005)



2.2.4 *cis*-1,2-Limonene oxide (12-*cis*) 12-cis

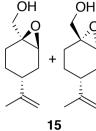
According to the methods of Steiner, et al., 2002, a 20 mL scintillation vial was charged with 1,2-limonene oxide, pyrrolidine & water. The vial was sealed with a Teflon lined cap and heated to 88 °C for 24 h. The vial was cooled to rt, transferred to a separatory funnel with 100 mL pentane, washed with sat. aq. ammonium chloride (3 x 30 mL) and water (1 x 50 mL). The organics were dried (MgSO₄) and concentrated to yield a light yellow oil. Purification by column chromatography (80 g spherical SiO₂; 0-15% EtOAc/Hex over 10 CV's) yielded *cis*-1,2-limonene oxide as a clear, colorless oil. (1.52 g, 64%): ¹H NMR (400 MHz, chloroform-*d*) δ 4.77 – 4.71 (m, 1H), 4.69 (s, 1H), 3.07 (s, 1H), 2.22 – 2.07 (m, 2H), 1.90 – 1.84 (m, 2H), 1.75 – 1.64 (m, 5H), 1.33 (s, 3H), 1.28 – 1.15 (m, 1H) ppm. Spectral data is consistent with literature reports (Steiner et al., 2002).



0 2.2.5 trans-1,2-Limonene oxide (12-trans) 12-trans

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According to the methods of Steiner, et al., 2002, a 50 mL flask was charged with 1,2-limonene oxide (4.57 g, 30 mmol), pyrazole (0.34 g, 5 mmol) and water (16.2 mL). The reaction was heated to reflux for 5 hours. The reaction mixture was cooled to 80 °C and transferred to a separatory funnel. Warm water (80 °C; 60 mL) was added. The emulsion was extracted with pentane (3 x 50 mL). The combined organic layers were dried (MgSO₄) and concentrated to yield 1.481 g crude oil. Purification by column chromatography (80 g spherical SiO₂; 0 – 20% EtOAc/Hex over 13.5 CV) yielded *trans*-1,2-limonene oxide (485 mg, 20% yield): 1 H NMR (400 MHz, chloroform-*d*) δ 4.68 (s, 2H), 3.01 (d, J = 5.3 Hz, 1H), 2.11 – 2.00 (m, 2H), 1.96 – 1.82 (m, 1H), 1.75 – 1.64 (m, 5H), 1.45 – 1.36 (m, 2H), 1.34 (s, 3H) ppm. Spectral data is consistent with literature reports (Steiner et al., 2002).



2.2.6 Perillic alcohol epoxide (15)

Perillic alcohol (5 mL, 31.46 mmol) was combined with dichloromethane (62 mL) in a round bottom flask with stir bar and cooled to 0 °C under inert atmosphere. m-CPBA (5.97 g, 34.61 mmol) was added in 5 portions over the course of 10 minutes. After 30 minutes the reaction was warmed back up to room temperature and allowed to stir for an hour at room temperature. The reaction mixture was filtered through celite, washed with DCM (3 x 30 mL), then transferred to a separatory funnel where it was washed with sodium bicarbonate (50 mL) and brine (50 mL). The organic layer was dried (Na₂SO₄) and concentrated to yield a crude oil (5.1 g) Purification by column chromatography (80 g spherical SiO₂; 0 – 100% EtOAc/Hex over 15 CV's) yielded the epoxide **15** as a clear, colorless oil (3.363 g, 19.8 % yield): ¹H NMR (400 MHz, chloroform-d) δ 4.77 – 4.66 (m, 2H), 3.78 – 3.59 (m, 2H), 3.41 – 3.31 (m, 1H), 2.26 – 2.14 (m, 1H), 2.09 (dtd, J = 14.9, 5.6, 1.8 Hz, 1H), 1.88 – 1.83 (m, 1H), 1.80 – 1.59 (m, 7H), 1.56 – 1.38 (m, 1H), 1.32 – 1.16 (m, 1H). Spectral data is consistent with literature reports (Thomas et al., 2016).



2.2.7 B -pinene oxide

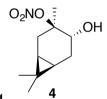
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Potassium peroxide monosulfate (4.750 g, 31.20 mmol) was dissolved in deionized water (60 mL). Sodium bicarbonate (4.019 g, 47.84 mmol) was placed into a 125 mL erlenmeyer flask and acetone (40 mL) was added, followed by β -pinene (1.59 mL, 9.974 mmol). The solution of potassium peroxide monosulfate mixture was slowly added into the β -pinene mixture over the course of three minutes via syringe while the mixture was stirring at 800 rpm. The reaction mixture was stirred for exactly 30 minutes while stirring at 800 rpm at room temperature. The reaction mixture was transferred to a separatory funnel and extracted with dichloromethane (2 x 40 mL), dried (MgSO₄) and concentrated to produce a clear oil (1.500 g, 96% yield). ¹H NMR (400 MHz, chloroform-d) δ 2.79 (d, J = 4.8 Hz, 1H), 2.62 (d, J = 4.8 Hz, 1H), 2.30 (dtd, J = 10.3, 5.9, 1.5 Hz, 1H), 2.19

(ddd, J = 15.1, 10.7, 8.1 Hz, 1H), 2.06 - 1.97 (m, 1H), 1.97 - 1.79 (m, 2H), 1.75 - 1.65 (m, 2H), 1.53 (t, J = 5.4 Hz, 1H), 1.27 (s, 3H), 0.94 (s, 3H). Spectral data is consistent with literature reports (Charbonneau et al., 2018).

2.3 General nitration method

A round bottom flask was charged with a solution of epoxide (2 mmol) in toluene, dioxane or dichloromethane (10 mL, 0.2 M). Bismuth nitrate (1.164 g, 2.4 mmol, 1.2 equiv) was added to the reaction mixture. The reaction mixture was stired for 30 – 60 minutes. When TLC indicated complete consumption of starting material the reaction was filtered through a 1 inch celite pad and washed with DCM (2 x 15 mL). The filtrate was transferred to a separatory funnel and washed with sodium bicarbonate (3 x 15 mL). The organic layer was dried with sodium sulfate, filtered and concentrated to yield a crude liquid. Desired products were isolated via column chromatography.



2.3.1 Nitrate ester 4

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Nitration of *trans*-3-carene oxide (3 mmol) was carried out according to the general method with dioxane. Purification of the crude oil by column chromatography(12 g SiO₂, 0-30% EtOAc/Hex) yielded nitrate ester **4** as a white crystalline solid (345 mg, 53% yield): mp = 52.2-54.0 °C; IR (ATR) cm⁻¹ 3412 (br w, OH), 2944 (w), 2360 (w), 1616 (vs, NO₂), 1291 (m, NO₂), 868 (m, NO₂); HRMS ESI+ m/z calc'd for [C₁₀H₁₈O₄N]⁺ (216.1230, observed; 216.1236, expected) ¹H NMR (400 MHz, chloroform-d) δ 3.70 (dd, J = 9.6, 7.6 Hz, 1H), 2.86 – 2.78 (m, 1H), 2.16 (dd, J = 14.8, 7.5 Hz, 1H), 1.83 (ddd, J = 14.8, 9.6, 7.9 Hz, 1H), 1.65 (s, 3H), 1.47 (dd, J = 14.3, 4.1 Hz, 1H), 1.03 (s, 3H), 1.01 (s, 3H), 0.83 – 0.73 (m, 2H); ¹³C NMR (101 MHz, chloroform-d) δ 95.68, 71.03, 28.37, 28.34, 27.02, 19.95, 18.98, 17.95, 15.70, 14.39 ppm.

Nitration of *cis*-3-carene oxide was attempted according to the general method with dioxane or dichloromethane. Purification of the crude oil by column chromatography (12 g SiO₂, 0-40% EtOAc/Hex) yielded *cis*-4-caranone as a clear, colorless oil (53% yield): R_f 0.51 (5% EtOAc:Hex; anisaldehyde); ¹H NMR (400 MHz, chloroform-*d*) δ 2.54 (ddd, J = 18.0, 8.4, 1.0 Hz, 1H), 2.44 – 2.25 (m, 3H), 1.33 – 1.20 (m, 1H), 1.14 – 0.99 (m, 5H), 0.97 (d, J = 6.4 Hz, 3H), 0.86 (s, 3H); ¹³C NMR (101 MHz, chloroform-*d*) δ 216.79, 41.99, 36.84, 29.77, 27.91, 22.83, 20.34, 19.47, 14.89, 14.11 ppm. Spectral data is consistent with literature reports (Kolehmainen et al., 1993).

2.3.3 Reaction of 8,9-limonene oxide (9)

Nitration of 8,9-limonene oxide (9) was attempted according to the general method with acetonitrile or dioxane. Dioxane yielded a mixture of putative nitrate ester 10 and aldehyde 11 (1:0.25) as a crude oil. Nitrate 10 (1:1 d.r.): R_f 0.25 (15% EtOAc:Hex; anisaldehyde); IR (ATR) cm⁻¹ 3417 (br), 2913 (w), 1613 (vs, NO₂), 1439 (w), 1289 (m, NO₂), 865 (s, NO₂); ¹H NMR (400 MHz, chloroform-*d*) δ 5.39 – 5.35 (m, 1H), 4.06 – 3.86 (m, 2H), 2.41 – 2.26 (m, 1H), 2.09 – 1.82 (m, 5H), 1.66 (s, 3H), 1.49 (s, 1.5H), 1.48 (s, 1.5H), 1.43 – 1.31 (m, 2H) ppm; 13°C NMR (101 MHz, chloroform-*d*) δ 134.31, 134.01, 119.57, 119.31, 97.40, 97.17, 64.62, 64.27, 38.24, 38.19, 30.58, 30.47, 26.42, 25.87, 24.19, 23.32, 23.25, 23.22, 17.17, 16.78 ppm. Purification by column chromatography (12 g SiO₂ gold column, 0-50% EtOAc/Hex over 30 CV) yielded acetaldehyde 11 (1:1 d.r) as a clear, colorless oil (105 mg, 35% yield): ¹H NMR (400 MHz, chloroform-*d*) δ 9.69 (d, *J* = 2.4 Hz, 0.5H), 9.68 (d, *J* = 2.7 Hz, 0.5H), 5.42 – 5.35 (m, 1H), 2.35 – 2.25 (m, 1H), 2.10 – 1.73 (m, 6H), 1.68 – 1.62 (br s, 3H), 1.46 – 1.32 (m, 1H), 1.10 (s, 1.5H), 1.08 (s, 1.5H) ppm; ¹³C NMR (101 MHz, chloroform-*d*) δ 205.55, 205.47,

134.09, 134.05, 119.95, 119.88, 51.00, 50.68, 34.32, 34.25, 30.15, 29.99, 29.67, 28.05, 27.31, 25.43, 23.43, 10.34, 10.24 ppm. Spectral data is consistent with literature reports (Uehara et al., 2017; Reid & Watson, 2018).

Acetonitrile yielded oxazoline **10a** (1:1 d.r.) as a clear, colorless oil (76% yield): R_f 0.46 (75% EtOAc:Hex; anisaldehyde); IR (ATR) cm⁻¹ 3323 (br), 2963 (m), 2913 (m), 1674 (vs), 1437 (m), 1384 (m), 1265 (m), 991 (vs), 915 (m); 1 H NMR (400 MHz, chloroform-d) δ 5.38 (s, 1H), 4.10 (d, J = 6.2 Hz, 0.5H), 4.08 (d, J = 6.1 Hz, 0.5H), 3.82 (d, J = 8.3 Hz, 0.5H), 3.79 (d, J = 8.3 Hz, 0.5H), 2.16 – 1.91 (m, 6H), 1.89 – 1.75 (m, 1H), 1.73 – 1.56 (m, 5H), 1.29 – 1.14 (m, 4H); 13 C NMR (101 MHz, chloroform-d) 162.73, 162.64, 134.23, 133.89, 120.28, 120.23, 77.22, 76.08, 75.39, 72.75, 72.60, 42.55, 42.28, 30.65, 30.57, 26.75, 26.39, 25.03, 24.58, 23.79, 23.75, 23.37, 13.88, 13.87 ppm; GC-MS (100 – 275 °C; 15 °C/min): 6.691 min, m/z 193.8 (M+, 100%), 135.0 (M - [C₂H₃NO], 10%).

2.3.4 Nitrate ester 13

Nitration of *cis*-1,2-limonene oxide was carried out according to the general method with dioxane or dichloromethane. Purification by column chromatography (12 g SiO₂ gold column, 0-50% EtOAc/Hex over 30 CV) yielded nitrate ester **13** as a clear, colorless oil (53% yield in dioxane; 62% yield in DCM): IR (ATR) cm⁻¹ 3660 (w, alcoholic OH), 3340 (bd w, alcoholic OH), 2980 (m), 1618 (s, NO₂), 1291 (m, NO₂), 860 (vs, NO₂); HRMS ESI+ *m/z* calc'd for [C₁₀H₁₇O₄NNa]⁺ (238.1052, observed; 238.1055, expected); ¹H NMR (400 MHz,

chloroform-d) δ 4.76 (dd, J = 8.1, 1.5 Hz, 2H), 4.13 (s, 1H), 2.35 (ddt, J = 11.6, 8.4, 4.1 Hz, 1H), 2.24 (dtd, J = 14.8, 3.7, 1.4 Hz, 1H), 1.98 – 1.78 (m, 3H), 1.76 – 1.73 (m, 4H), 1.68 – 1.58 (m, 4H), 1.49 (tdd, J = 13.3, 11.6, 3.6 Hz, 1H); ¹³C NMR (101 MHz, chloroform-d) δ 148.49, 109.48, 91.34, 69.22, 36.84, 33.86, 29.90, 25.77, 20.98, 20.93 ppm.

5 **2.3.5** Nitrate ester 14

Nitration of *trans*-1,2-limonene oxide was carried out according to the general method with dioxane or dichloromethane. Purification by column chromatography (12 g SiO₂, 0-50% EtOAc/Hex over 30 CV) yielded nitrate ester **14** as a clear, colorless oil (63% in dioxane; 54% in DCM): R_f 0.39 (15% EtOAc:Hex; anisaldehyde); IR (ATR) cm⁻¹ 3414 (bd w, alcoholic OH), 2940 (w), 1627 (s, NO₂), 1279 (m, NO₂), 876(m, NO₂), 851 (m, NO₂); HRMS ESI+ m/z calc'd for $[C_{10}H_{17}O_4NNa]^+$ (238.1048, observed; 238.1055, expected); ¹H NMR (400 MHz, chloroform-d) δ 5.01 (s, 1H), 4.76 (d, J = 6.6 Hz, 2H), 2.19 (tt, J = 10.9, 3.1 Hz, 1H), 2.03 (ddd, J = 14.7, 12.1, 2.6 Hz, 1H), 1.92 (dq, J = 14.4, 3.5, 2.7 Hz, 1H), 1.74 (s, 3H), 1.69 – 1.56 (m, 4H), 1.32 (s, 3H); ¹³C NMR (101 MHz, chloroform-d) δ 148.27, 109.57, 84.43, 69.55, 37.96, 34.78, 29.91, 26.96, 20.89 ppm.

2.3.6 Nitration of perillic alcohol epoxide (15)

Nitration of 4-(1-methylethenyl)-7-oxabicyclo[4.1.0]heptane-1-methanol (15) was carried out according to the general method with dichloromethane. Purification by column chromatography (12 g SiO₂, 0-40% EtOAc/Hex over 30 CV) yielded two regioisomers, nitrate esters 16 and 17 in 23% and 32% yield, respectively.

2.3.6.1 Nitrate ester **16**

Nitrate ester **16** was isolated as a white, crystalline solid (106 mg, 23% yield): mp = 72-75°C; R_f 0.31 (30% EtOAc:Hex; anisaldehyde); IR (ATR) cm⁻¹ 3285 (bd w, alcoholic OH), 2941 (w), 1627 (vs, NO₂), 1280 (s, NO₂), 880 (s, NO₂), 850 (s, NO₂); HRMS ESI+ *m/z* calc'd for [C₁₀H₁₇O₄NNa]⁺ (238.1054, observed; 238.1055, expected); ¹H NMR (400 MHz, chloroform-*d*) δ 5.23 (s, 1H), 4.80 – 4.73 (m, 2H), 3.74 (dd, *J* = 10.9, 5.0 Hz, 1H), 3.44 (dd, *J* = 11.0, 5.0 Hz, 1H), 2.59 (d, *J* = 1.0 Hz, 1H), 2.27 – 2.16 (m, 1H), 2.03 – 1.97 (m, 2H), 1.79 (t, *J* = 5.2 Hz, 1H), 1.71 – 1.61 (m, 3H), 1.58 (d, *J* = 0.8 Hz, 4H); ¹³C NMR (101 MHz, chloroform-*d*) δ 148.24, 109.58, 79.89, 70.98, 66.45, 38.47, 29.70, 29.55, 25.10, 20.77 ppm.

2.3.6.2 Nitrate ester 17 17

10 Nitrate ester **17** was isolated as a clear, colorless oil (149 mg, 32% yield): R_f 0.20 (30% EtOAc:Hex; anisaldehyde); IR (ATR) cm⁻¹ 3286 (bd w, alcoholic OH), 2941 (w), 1623 (vs, NO₂), 1288 (s, NO₂), 863 (vs, NO₂); HRMS ESI- *m/z* calc'd for [C₁₀H₁₆O₄N]⁻ (214.10813, observed; 214.10848, expected); ¹H NMR (400 MHz, chloroform-*d*) δ 4.76 (d, *J* = 8.5 Hz, 2H), 4.31 (s, 1H), 4.27 (d, *J* = 13.0 Hz, 1H), 3.99 (d, *J* = 13.0 Hz, 1H), 2.41 (tt, *J* = 12.1, 3.6 Hz, 1H), 2.27 – 2.19 (m, 1H), 1.98 – 1.78 (m, 3H), 1.75 (s, 3H), 1.72 – 1.65 (m, 1H), 1.51 – 1.38 (m, 1H); ¹³C NMR (101 MHz, chloroform-*d*) δ 148.58, 109.43, 92.51, 65.67, 62.91, 37.47, 33.53, 25.33, 25.26, 20.86 ppm.

2.3.7 Nitration of α-pinene oxide

Nitration of α -pinene oxide was carried out according to the general method. Crude oil from a 10 mmol reaction was purified by column chromatography (40 g SiO₂, 0-40% EtOAc/Hex over 30 CV) yielded the following isomerization products and nitrate esters.

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2.3.7.1 α-campholenic aldehyde (19)

α-Campholenic aldehyde **19** was isolated as a clear, colorless oil: R_f 0.59 (10% EtOAc:Hex; anisaldehyde); 1 H NMR (400 MHz, benzene- d_6) δ 9.51 (t, J = 2.1 Hz, 1H), 5.23 (s, 1H), 2.41 (dddt, J = 13.8, 6.0, 2.7, 1.6 Hz, 1H), 2.25 – 2.16 (m, 1H), 2.11 (ddd, J = 15.9, 4.4, 1.8 Hz, 1H), 2.00 (ddd, J = 15.9, 10.3, 2.3 Hz, 1H), 1.80 (ddp, J = 15.8, 9.2, 2.5 Hz, 1H), 1.58 (dt, J = 3.0, 1.6 Hz, 3H), 0.90 (s, 3H), 0.68 (s, 3H). Spectral data is consistent with literature reports (Thomas et al., 2016).

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2.3.7.2 5-isopropylidene-2-methyl-2-cyclohexen-1-ol (20)

Compound **20** was isolated as a white, crystalline solid: R_f 0.47 (20% EtOAc:Hex; anisaldehyde); ¹H NMR (400 MHz, chloroform-d) δ 5.49 (ddd, J = 4.4, 3.2, 1.5 Hz, 1H), 3.99 (t, J = 4.2 Hz, 1H), 2.86 (d, J = 19.6 Hz, 1H), 2.69 – 2.56 (m, 2H), 2.35 (d, J = 15.5 Hz, 1H), 1.78 – 1.75 (m, 3H), 1.74 (s, 3H), 1.68 (s, 3H); ¹³C NMR (101 MHz, chloroform-d) δ 135.94, 125.57, 124.23, 123.29, 70.34, 35.92, 29.83, 20.31, 20.14, 19.91 ppm. Spectral data is consistent with literature reports (Motherwell et al., 2004).

21

2.3.7.3 *trans*-Carveol (21)

trans-Carveol was isolated as a clear, colorless oil: R_f 0.37 (20% EtOAc:Hex; anisaldehyde); ¹H NMR (400 MHz, chloroform-*d*) δ 5.62 (dd, J = 5.2, 1.5 Hz, 1H), 4.78 – 4.73 (m, 2H), 4.06 – 4.03 (m, 1H), 2.34 (ddd, J = 13.7, 9.8, 2.9 Hz, 1H), 2.21 – 2.12 (m, 1H), 1.96 (dq, J = 13.6, 2.3 Hz, 1H), 1.93 – 1.85 (m, 1H), 1.85 – 1.81 (m, 3H), 1.77 (s, 3H), 1.64 (dd, J = 13.2, 4.0 Hz, 1H), 1.61 – 1.57 (m, 1H) ppm. Spectral data is consistent with literature reports). (Motherwell et al., 2004).

2.3.7.4 trans-Carveol nitrate ester 22

Nitrate ester **22** was isolated as a clear, colorless oil: IR (ATR) cm⁻¹ 3353 (bd w, alcoholic OH), 2917 (w), 1614 (vs, NO₂), 1293 (s, NO₂), 868 (m, NO₂); ¹H NMR (400 MHz, chloroform-d) δ 5.32 (ddd, J = 9.9, 3.5, 2.1 Hz, 1H), 4.12 (ddd, J = 10.6, 4.6, 1.9 Hz, 1H), 2.68 – 2.49 (m, 2H), 1.88 (t, J = 4.8 Hz, 1H), 1.43 (dd, J = 14.1, 3.7 Hz, 1H), 1.24 (dd, J = 13.8, 4.8 Hz, 1H), 1.11 (s, 3H), 0.97 (s, 3H), 0.94 (s, 3H) ppm. Spectral data is consistent with literature reports (Rindelaub et al., 2016b).

2.3.7.5 Nitrate ester 23

Nitrate ester 23 was isolated as a white, crystalline solid: mp = 132-134 °C; R_f 0.29 (25% EtOAc:Hex; anisaldehyde); IR (ATR) cm⁻¹ 3294 (bd w, alcoholic OH), 2961 (w), 2360 (w), 1624 (m, NO₂), 1282 (m, NO₂),

862 (m, NO₂); HRMS ESI- m/z calc'd for [C₁₀H₁₆O₄N]⁻ (214.10797, observed; 214.10848, expected); ¹H NMR (400 MHz, chloroform-d) δ 4.64 (s, 1H), 4.00 (ddd, J = 7.1, 3.3, 0.9 Hz, 1H), 2.37 (ddd, J = 13.8, 7.1, 2.5 Hz, 1H), 1.81 (d, J = 4.3 Hz, 1H), 1.60 – 1.51 (m, 3H), 1.32 (dt, J = 13.8, 4.0 Hz, 1H), 1.23 (s, 3H), 1.22 (s, 3H), 0.89 (s, 3H); ¹³C NMR (101 MHz, chloroform-d) δ 94.13, 68.52, 53.01, 47.34, 40.49, 37.76, 37.10, 29.24, 19.12, 14.75 ppm.

2.3.7.6 Nitrate ester 24

Nitrate ester **24** was isolated as a white, crystalline solid: mp = 62-64 °C; R_f 0.24 (15% EtOAc:Hex; anisaldehyde); HRMS ESI+ m/z calc'd for [C₁₀H₁₇O₄NNa]⁺ (238.10459, observed; 238.10498, expected); IR (ATR) cm⁻¹ 3434 (w, alcoholic OH), 2959 (w), 1622 (s, NO₂), 1284 (vs, NO₂), 856 (m, NO₂); ¹H NMR (400 MHz, chloroform-d) δ 5.31 (ddd, J = 9.9, 3.8, 2.1 Hz, 1H), 4.12 (m, 1H), 2.69 – 2.48 (m, 2H), 1.87 (t, J = 4.9 Hz, 1H), 1.43 (dd, J = 14.0, 3.7 Hz, 1H), 1.24 (dd, J = 13.7, 4.7 Hz, 1H), 1.11 (s, 3H), 0.97 (s, 3H), 0.94 (s, 3H); ¹³C NMR (101 MHz, chloroform-d) δ 91.04, 78.09, 52.58, 49.53, 42.73, 39.05, 36.71, 20.20, 19.70, 11.79 ppm.

2.3.8 Nitration of β-pinene oxide

Nitration of β-pinene oxide (11.1 mmol) was carried out according to the general procedure in toluene. Purification by column chromatography (80 g SiO₂, 0-30% EtOAc/Hex over 20 CV) yielded a 1:1 mix of myrtenol (26) and nitrate 29 (344 mg, 9 % combined yield), perillic alcohol (28, 600 mg, 36% yield) and nitrate 27 (71 mg, 3% yield). The combined myrtenol and nitrate 29 were separated by column chromatography (40 g SiO₂; 5-20% EtOAc/Hex over 25 CV).

20 Myrtenol was isolated as a clear, colorless oil: R_f 0.25 (15% EtOAc:Hex; anisaldehyde); ¹H NMR (400 MHz, chloroform-*d*) δ 5.49 (dt, J = 3.0, 1.5 Hz, 1H), 4.00 (t, J = 1.8 Hz, 2H), 2.42 (dt, J = 8.6, 5.6 Hz, 1H), 2.37 –

2.20 (m, 2H), 2.19 - 2.09 (m, 2H), 1.61 - 1.45 (m, 1H), 1.31 (s, 3H), 1.19 (d, J = 8.6 Hz, 1H), 0.85 (s, 3H) ppm.Spectral data is consistent with literature reports (Motherwell et al., 2004)

2.3.8.2 Nitrate ester 27

Nitrate ester **27** was isolated as a clear colorless oil: IR (ATR) cm⁻¹ 3362 (bd w, alcoholic OH), 2947 (w), 1624 (s, NO₂), 1281 (vs, NO₂), 864 (m, NO₂); HRMS ESI- m/z calc'd for [C₁₀H₁₆O₄N]⁻ (214.10813, observed; 214.10848, expected); ¹H NMR (400 MHz, chloroform-d) δ 5.57 – 5.49 (m, 1H), 3.71 (q, J = 11.5, 8.5 Hz, 2H), 2.55 (ddt, J = 14.1, 9.8, 3.9 Hz, 1H), 1.95 – 1.83 (m, 2H), 1.76 (t, J = 4.5 Hz, 1H), 1.52 – 1.22 (m, 5H), 1.10 (s, 3H), 1.01 (s, 3H); ¹³C NMR (101 MHz, chloroform-d) δ 85.30, 62.08, 53.45, 48.62, 45.59, 35.78, 27.53, 22.88, 20.20, 20.02 ppm.

10 **2.3.8.3** Nitrate ester **29**

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Nitrate ester **29** was isolated as a white, crystalline solid: mp = 59-61 °C; R_f 0.55 (15% EtOAc:Hex; anisaldehyde); IR (ATR) cm⁻¹ 3374 (bd w, alcoholic OH), 2915 (w), 1625 (s, NO₂), 1281 (vs, NO₂), 863 (m, NO₂); HRMS ESI+ m/z calc'd for [C₁₀H₁₇O₄NNa]⁺ (238.10474, observed; 238.10498, expected); ¹H NMR (400 MHz, chloroform-d) δ 4.92 (d, J = 1.6 Hz, 1H), 3.75 (d, J = 11.3 Hz, 1H), 3.65 (d, J = 11.3 Hz, 1H), 1.90 – 1.80 (m, 3H), 1.72 – 1.63 (m, 1H), 1.56 – 1.50 (m, 1H), 1.24 – 1.20 (m, 4H), 0.96 (s, 3H); ¹³C NMR (101 MHz, chloroform-d) δ 89.49, 63.82, 54.38, 48.25, 40.28, 37.03, 29.26, 24.74, 22.11, 19.46 ppm.

3 Results and discussion

3.1 Synthesis

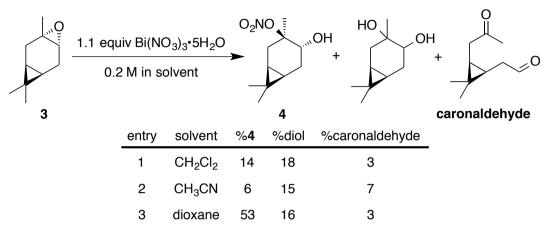


Figure 2: Effect of solvent on nitration of trans-3-carene oxide.

Previous reports have described the opening of epoxides and aziridines using bismuth nitrate in acetonitrile (Das et al., 2007). Others report optimal conditions in dichloromethane (Rindelaub et al., 2016b), or 1,4-dioxane with undesired side reactions in acetonitrile (Pinto et al., 2007). Thus, we began our investigations into the nitration of *trans*-carene oxide (3) using bismuth nitrate in various solvents (Figure 2). In our hands, dioxane clearly outperformed dichloromethane, yielding 53% of the desired product after 45 minutes (entry 3). A diol was isolated as a major (16%) byproduct. Attempts to mitigate hydrolysis by the use of base or molecular sieves were ineffective. Acetonitrile produced a complex intractable mixture with a high amount of diol and caronaldehyde (entry 2). Other solvents, such as THF and nitromethane, also produced complex mixtures with no trace of desired nitrate. Methanol, interestingly, only produced an undesired methyl ether. It is unclear if this product was generated through methanolysis of the nitrate ester or direct substitution of the epoxide. The regiochemistry of the nitration was easily elucidated with gHSQC NMR data. A doublet of doublets at 3.69 ppm correlated with a carbon shift at 71.1 ppm and was thus assigned to the alcohol methyne carbon. A tetrasubstituted carbon, with no correlations in the gHSQC, at 95.6 ppm was consistent with the tertiary nitrate ester. Finally, the IR of compound 4 displayed the expected strong absorbances at 1616, 1291, and 868 cm⁻¹.

Figure 3: Preparation of cis-3-carene oxide and attempted nitration.

Next, we aimed to investigate the impact of relative stereochemistry on the nitration. We hypothesized that the secondary nitrate would predominate due to classical stereoelectronic effects. (+)-3-Carene was treated with N-bromosuccinimide and calcium carbonate to produce a bromohydrin which subsequently cyclized under basic conditions to produce *cis*-carene oxide (Cocker and Grayson, 1969). Interestingly, we never observed the desired nitrate ester. Instead a 1,2-hydrogen shift generated *cis*-4-caranone (7) in 53% yield.

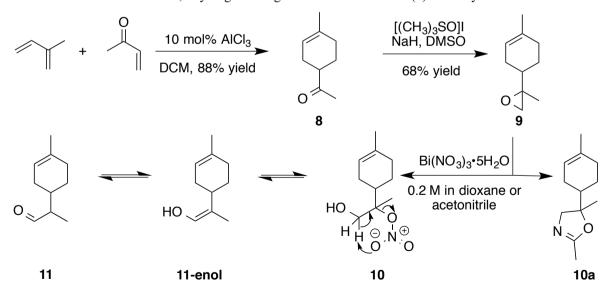


Figure 4: Preparation of 8.9-limonene oxide and attempted nitration.

The preparation of 8,9-limonene oxide (9) began with a Diels-Alder reaction to form 1-(4-methyl-3-cyclohexene) ethenone (8) in 88% yield. Corey-Chaykovsky addition of a sulfur ylide cleanly produced 8,9-limonene oxide (9). Nitration of 9 was attempted in a variety of solvents (DCM, benzene, dioxane, acetonitrile, nitromethane). In acetonitrile, oxazoline 10a (1:1 d.r.) was cleanly produced presumably via nucleophilic addition of acetonitrile and subsequent cyclization to the oxazoline. Pinto, et al. 2007 observed a similar bismuth nitrate mediated epoxide opening with acetonitrile. In dioxane, aldehyde 11 (1:1 d.r.) was observed along with a product consistent with a tertiary nitrate ester 10 (1:1 d.r.) in a 0.25:1 mixture. Formation of 11-enol presumably proceeds via a facile intramolecular elimination of nitric acid. The IR of the crude mixture showed the expected strong absorbances at 1613, 1288 and 865 cm⁻¹. A set of signals in the ¹³C NMR at 97.40 & 97.17 ppm, with no gHSQC correlations, are consistent with a tertiary nitrate. Attempts to purify this mixture on silica gel gave small amounts of 11 and unidentifiable decomposition products. Furthermore, the fast hydrolysis rates of tertiary nitrate esters hinder the ability to isolate these products (Boyd et al., 2015; Darer et al., 2011; Liu et al., 2012). No reaction was observed when non-polar solvents such as DCM and benzene were employed. We hypothesize that the steric hindrance at the β-carbon severely limits reactivity.

Figure 5: Nitration of cis- and trans-1,2-limonene oxide.

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Commercially available 1,2-limonene oxide was resolved into pure samples of *cis*- and *trans*-1,2-limonene oxide by treating with cyclic amine bases (Steiner et al., 2002). The nitration of the *cis* isomer was predicted to produce the tertiary nitrate ester via stereoelectronic effects. The dominant conformation of *cis*-1,2-limonene oxide is expected to be a pseudo-half chair (Figure 5). Nucelophilic substitution at the less substituted position would proceed through a lower energy chair-like transition state, whereas attack at the more substituted carbon leads to a very unstable twist-boat. The nitration of *cis*-1,2-limonene oxide (*cis*-12) proceeded smoothly in both dioxane (53%) and dichloromethane (63%). Respectively, the *trans* isomer was expected to produce the secondary nitrate ester. The nitration of *trans*-1,2-limonene oxide (*trans*-12) also proceeded smoothly in both dioxane (63%) and dichloromethane (61%). Contrary to previous reports (Romonosky et al., 2015), we did not observe the desired nitrate esters in acetonitrile. gHSQC NMR data was used to verify nitrate ester 13 was

tertiary and **14** was secondary. For **13**, a broad singlet at 4.12 ppm correlated with a carbon shift at 69.2 ppm and was thus assigned to the methyne adjacent to the alcohol. A tetrasubstituted carbon at 91.4 ppm was consistent with the tetrasubstituted nitrate ester. Similarly for **14**, a broad triplet at 5.00 ppm correlated with a carbon shift at 84.45 ppm and was thus assigned to the methyne adjacent to the nitrate ester. A tetrasubstituted carbon at 69.5 ppm was consistent with the tertiary alcohol.

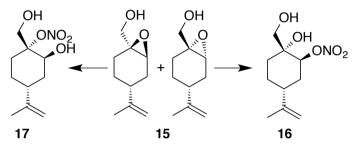


Figure 6: Nitration of perillic alcohol oxide.

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Under acidic conditions, β-pinene oxide undergoes facile rearrangement to form perillic alcohol. Accordingly, we prepared the epoxide **15** by treating perillic alcohol with 1 equivalent of m-CPBA. The inseparable mixture of diastereomers were treated with bismuth nitrate under the standard reaction conditions. Nitrate esters **16** and **17** were easily separated by column chromatography and assigned to the secondary and tertiary nitrate esters, respectively. These isomers were formed from the *cis*- and *trans*-epoxide diastereomers in an analogous fashion to the limonene isomers. For **16**, a broad singlet at 5.21 ppm correlated with a carbon shift at 80.1 ppm and was thus assigned to the methyne adjacent to the nitrate ester. A tetrasubstituted carbon at 70.9 ppm was consistent with the tertiary alcohol. Similarly for **15**, a broad singlet at 4.22 ppm correlated with a carbon shift at 65.6 ppm and was thus assigned to the methyne adjacent to the alcohol. A tetrasubstituted carbon at 92.8 ppm was consistent with the tertiary nitrate ester.

1 equiv TBAN

Figure 7: Nitration of α-pinene oxide.

Toluene

Toluene

The facile rearrangements of α-pinene with both Brønsted and Lewis Acids (Kaminska et al., 1992) can be thought to proceed via a non-classical isobornyl cation (18) (Kong et al., 2010). We expected the reaction with bismuth nitrate to generate a complex mixture of products due to the many reactive sites. As shown in Figure 7, three rearrangement products (campholenic aldehyde 19, diene 20, and *trans*-carveol 21) and three nitrate esters (22, 23, 24) were observed. The structure of nitrate ester 23 was identified by a clear singlet at 4.64 ppm (s, 1H). 2D-NMR data (gCOSY, gHSQC and gHMBC) along with comparison to 6-exo-hydroxyfenchol, the analogous diol confirmed the structural assignment (Miyazawa and Miyamoto, 2004). Correspondingly, nitrate ester 24 displayed a distinct doublet of doublets of doublets at 5.31 ppm that correlated to the methyne adjacent to the nitrate ester. Again, 2D NMR data (gCOSY, gHSQC and gHMBC) was used in conjunction with the literature spectra for platydiol and its *trans* diastereomer to verify the assignment (Kuo et al., 1989).

Under all conditions campholenic aldehyde was the major product (20-28% yield). First generation nitrate ester 25 was not found under all conditions. All six components were isolated in small amounts when the reaction was run in dichloromethane (Figure 7, entry 1). Cooling the reaction to -78 °C completely shut down all

reactivity (entry 2). Interestingly, adding one equivalent of tetrabutylammonium nitrate (TBAN) as an external nitrate source in addition to bismuth nitrate at -78 °C generated mostly nitrate 22 (entry 3). More polar solvents, like dioxane (entry 4), generated slightly higher amounts of nitrates at room temperature. Aromatic solvents, such as benzene and toluene, provided the best yield of 23. Interestingly, adding TBAN to the nitration in toluene dramatically increased the amounts of diene 20 and *trans*-carveol 21. Using acetonitrile as a solvent generated a complex mixture that appeared to be mainly diols. Zirconium nitrate produced a similar distribution of products (Das et al., 2006), but other metal nitrate complexes (Y(NO₃)₃•6H₂O, Co(NO₃)₂•6H₂O) resulted in no observable reaction.

10 Figure 8: Nitration of β-pinene oxide.

The nitration of β -pinene was similarly complicated by isomerization pathways through the corresponding nonclassical carbocation. Nitrate **27** initially co-eluted with myrtenol (**26**) but was separable upon a second purification by column chromatography. The methyne proton of **27** at 5.51 ppm is correlated with a carbon at 85.3 ppm in the gHSQC NMR. This data is consistent with the secondary nitrate ester. Similarly, the methyne proton in nitrate **29** is a doublet at 4.91 ppm ($J^4 = 1.6$ Hz; W-coupling). This signal correlates with a carbon at 89.5 ppm. Interestingly, nitrates **30** and **31** were not observed.

3.2 Spectral data

Table 1. Collated ¹H and ¹³C NMR data with IR absorbances for nitrate esters.

compound	CHONO ₂ (ppm)	CHONO ₂ (ppm)	IR (cm ⁻¹)		
O ₂ NOOH		95.6	1616	1291	868

ONO ₂ OH		91.4	1618	1291	860
OH ONO ₂	5.00 (s, 1H)	84.4	1627	1279	876
OH OH ONO ₂	5.21 (s, 1H)	80.1	1627	1280	880
OH ONO ₂ OH		92.5	1623	1288	863
OH 22 ONO ₂		94.2	1614	1293	868
O ₂ NO , OH	4.64 (s, 1H)	94.13	1624	1282	862
O ₂ NO , OH	5.31 (ddd, <i>J</i> = 9.9, 3.8, 2.1 Hz, 1H)	91.04	1622	1284	856
O ₂ NO , 27	5.51 (m, 1H)	85.3	1624	1281	864
O ₂ NO , 29	4.91 (s, 1H)	89.5	1625	1281	863

As shown in table 1, all nitrate esters displayed the expected strong nitrate ester absorbances at ~1600, 1300, and 900 cm⁻¹. Methyne protons next to the secondary nitrate esters appeared between 4.64 ppm and 5.51 ppm. Carbon chemical shifts were found over a broader range than anticipated from 80.1 to 95.6 ppm. Finally, we observed masses consistent with methanolysis products (185.1 *m/z* M+; 223.1 *m/z* M+K) when nitrate esters were analysed (GC-MS) as solutions in methanol. Further experimentation is necessary to evaluate the implications of methanolysis.

3.3 Stability and storage

We were surprised to find that five of the nitrate esters (4, 16, 23, 24, and 29) are solids. We have stored these compounds at 0 °C for up to 9 months with no noticeable decline in purity. The remaining nitrate esters are stable at 0 °C for 2-4 weeks. Interestingly, some of these compounds appear to deviate from the expected stability patterns (*vide supra*). For example, tertiary nitrate ester 4 is particularly stable stored as a solid at 0 °C. In contrast, when secondary nitrate ester 14 was stored as a neat oil at 0 °C, it decomposed to a complex mixture within a few weeks. Freezing the samples as a solution in benzene is recommended for longer term (~6 month) storage.

4 Conclusions

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We have clearly delineated successful methods required to synthesize and purify nine nitrate esters derived from mono-terpenes. Seven of these compounds are undescribed in the literature and the remaining two had gaps in their characterization. Using our methods it is possible to cleanly isolate 50-100 mg even of the least prevalent isomers from the nitration reactions of α -pinene and β -pinene. Interestingly, we did not observe the formation of α -pinene oxide and β -pinene oxide products that retained their bicyclic ring structures. This is consistent with the solution phase synthetic literature (Kaminska et al., 1992) but in contrast to many atmospheric reports (For example, see: Rindelaub, 2016b; Duporte, G., et al., 2016). We believe that the availability of these compounds will enable further study of the structure-reactivity relationships. For example, comparing the specific hydrolysis rates for tertiary versus secondary nitrate esters in 13 and 14 as well as 17 and 16 could help deconvolute the fates of each terpene in the atmosphere. We also believe that these compounds will assist in confirming the identities of organic nitrates that have previously been limited to detection by MS based methods (Rindelaub, 2016b). A forthcoming report will describe our investigation into the behavior of these authentic compounds in mass spectrometers. In particular, the detailed MS-MS data, impact of various ionization conditions (ESI, CI, etc) and sample preparation will be described. Finally, the availability of these compounds is important for further studies into the influence of terpene structure on the fate and roles of organic nitrates in SOA formation.

25 Supplementary material related to this article is available online at:

Author contributions. RL designed the experiments and prepared the manuscript with contributions from all coauthors. All co-authors carried out experiments and contributed to reviewing and editing the manuscript.

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