

Answer to Anonymous Referee #1 comments

This manuscript provides detailed insights into the biogenic primary organic aerosol emission sources of the primary sugar compounds (SC), i.e., glucose, arabitol and mannitol. The study has been carefully designed and the results have been interpreted in detail. The study covers 16 nation-wide sites all over France and contains a very comprehensive data set. It is clearly shown that the main drivers of SC atmospheric concentrations are ambient air temperature, relative humidity and vegetation density.

We thank the reviewer for his/her review. We have studied the comments and we have made revisions point by point. The detailed responses to the comments are given below, point by point, *in blue color*, including changes directly made to the manuscript, *in red color*.

Specific comments:

1. Introduction: glucose is recognized as a tracer for plant pollen but also for biomass burning. I miss some discussion about this issue in the introduction.

The reviewer is right that glucose can also originate from the thermal degradation of the plant materials (e.g., cellulose, a polymer of glucose). However, as evidenced in Figure R1.1, the concentrations of levoglucosan (a well-established tracer of biomass burning source) and those of glucose clearly display very different annual atmospheric evolution cycles: higher concentrations of levoglucosan in France are observed in the coldest season (winter) due to the increased biomass burning while those of glucose are observed in warm seasons and coinciding with negligible ambient concentrations of levoglucosan. Such different temporal patterns indicate that the biomass burning is not an important source of atmospheric glucose.

Glucose can have a broad primary biogenic sources, e.g. from terrestrial plant pollen, fruits, and detritus, or from the degradation of the soil microorganisms (Xiao et al., 2018; Zhu et al., 2015) or even possibly from bubble bursting processes in remote oceans (Fu et al., 2013; Gao et al., 2011; Leck and Bigg, 2005).

For these reasons, we have removed the term “specific” in lines 54—59.

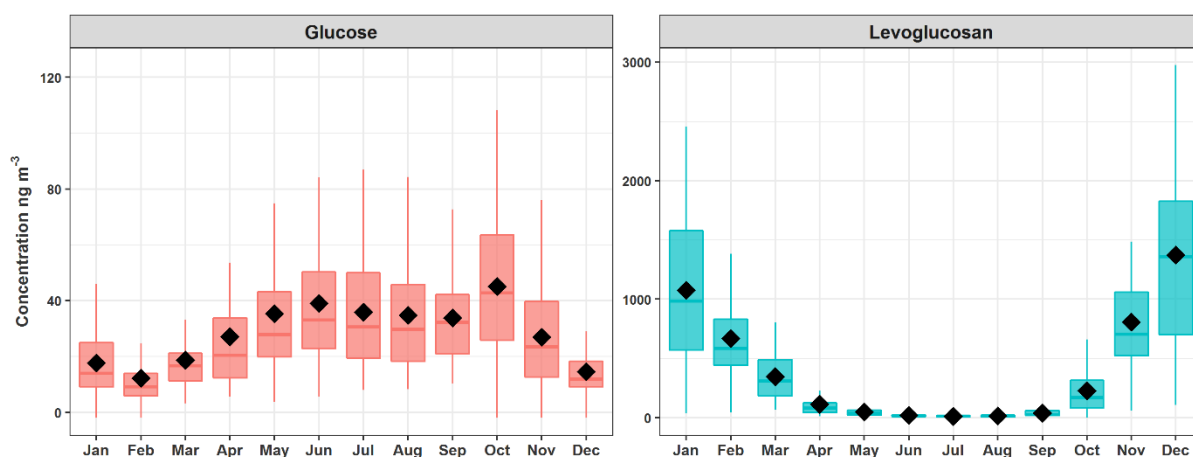


Figure R1.1: Annual evolution cycles of the glucose (left) and levoglucosan (right) concentrations in PM₁₀ measured at the urban site of Grenoble Les Frênes, from the years 2012 to 2018. The black marker inside each boxplot indicates the average value, while the top, middle and bottom of the box represent the 75th, median and 25th percentiles, respectively. The whiskers at the top and bottom of the box extend from the 95th to the 5th percentiles.

2. In several parts of the text, figures and tables, mention is made of "glucose" but in fact "free cellulose" is meant. In order to avoid confusion, I suggest to make this more clear and replace "glucose" by "free cellulose".

In fact, both glucose and free cellulose are measured and analyzed separately in the present work. We used glucose when the monosaccharide "glucose" is meant and free cellulose we considered the cellulose ambient cellulose.

Technical corrections: References: should be ordered chronologically.

The references are now ordered chronologically, as suggested by the reviewer.

References

Fu, P. Q., Kawamura, K., Chen, J., Charrière, B., and Sempéré, R.: Organic molecular composition of marine aerosols over the Arctic Ocean in summer: contributions of primary emission and secondary aerosol formation, *Biogeosciences*, 10(2), 653–667, doi:10.5194/bg-10-653-2013, 2013.

Gao, Q., Nilsson, U., Ilag, L. L., and Leck, C.: Monosaccharide compositional analysis of marine polysaccharides by hydrophilic interaction liquid chromatography-tandem mass spectrometry, *Anal. Bioanal. Chem.*, 399(7), 2517–2529, doi:10.1007/s00216-010-4638-z, 2011.

Leck, C. and Bigg, E. K.: Biogenic particles in the surface microlayer and overlaying atmosphere in the central Arctic Ocean during summer, *Tellus B*, 57(4), 305–316, doi:10.1111/j.1600-0889.2005.00148.x, 2005.

Xiao, M., Wang, Q., Qin, X., Yu, G., and Deng, C.: Composition, Sources, and Distribution of PM_{2.5} Saccharides in a Coastal Urban Site of China, *Atmosphere*, 9(7), 274, doi:10.3390/atmos9070274, 2018.

Zhu, C., Kawamura, K., and Kunwar, B.: Organic tracers of primary biological aerosol particles at subtropical Okinawa Island in the western North Pacific Rim: Organic biomarkers in the north pacific, *J. Geophys. Res. Atmospheres*, 120(11), 5504–5523, 2015.

Answer to Anonymous Referee #2 comments

The manuscript “Arabitol, mannitol and glucose as tracers of primary biogenic organic aerosol: influence of environmental factors on ambient air concentrations and spatial distribution over France” describes the primary sugar compounds (SC, defined as glucose, arabitol and mannitol) concentrations in PM₁₀ for 16 increasing space scale sites (local to nation-wide), distributed in several French geographic areas of different environmental conditions. This paper first time investigates the spatial behavior of these chemicals and evidencing their major effective environmental drivers.

We thank the reviewer for his/her attention to our manuscript that greatly contribute to improve the quality of this research paper. All comments have been considered and answered. The detailed responses to the comments are given below, point by point, *in blue color*, including changes directly made to the manuscript, *in red color*.

Major comments:

(1) Updating the references used in this manuscript to more current state is suggested.

We do agree with the reviewer and we have updated the references with several works recently published, including those in 2019. However, very few scientific papers have been published recently on the short term (daily) and the spatial characterization of polyols and glucose in PM₁₀. This is why older pioneering works are also cited in the present work.

(2) LOD (limit of detection) of the detected chemicals should be included in the experimental section. As suggested by reviewer, the information about the quantification limits have been included in the experimental section (lines 176—177).

(3) The regional transport is also very important for the spatial behavior and distribution of the chemical species in the ambient. In addition, only temporal variations and tracer ratios were shown and discussed in the results and discussion section. More deep analysis (i.e., the influences from nearby regions/sources, combine the chemical analysis results with synoptic data, . . .) are recommended to make this paper more interesting and innovative. At least, choose one or two cases to explain the contribution from regional transport by backward trajectory analysis.

We agree that regional transport may impact PM polyol concentrations. However, we do not think that it explains the main temporal signals observed in this work. Since the correlation matrix corresponds to averaged values of composite data, i.e. aggregate on consecutive three days or six days intervals, it already account for potential regional transport between sites, and a decreased of correlation with inter-site distance is observed and is so probably indicative of local source contribution rather than transportation.

However, as suggested by the reviewer, we achieved additional back trajectory analyses. , This was done for arabitol concentrations at the remote OPE-ANDRA site for the period 2012–2018, applying Potential Source Contribution Function (PSCF) to HYSPLIT data and using the pyPSCF python package¹. Results do not indicate clear source region(s) (Figure R2.2). Indeed, even if it seems that air-masses associated with high arabitol loading (>75th concentration percentile) never come from the East, it is in fact explained by the climatic wind condition in this region where no easterlies wind are observed during summer (anticyclonic condition). Finally, since no specific region is pointed out by the PSCF analysis, it may be explain either if the arabitol is emitted everywhere, or by a local (<few grid cells, within around 100 km from the station) source since all back-trajectories will then be associated with high concentration. Since correlations between sites decrease with the distance, the first hypothesis is

¹ <https://gricad-gitlab.univ-grenoble-alpes.fr/webersa/pyPSCF.git>
(documentation: <https://pypscf.readthedocs.io/en/latest>)

most probably not valid. These two arguments are in favor of local sources being predominant for the polyols, as opposed to regional (> 100 km) transport.

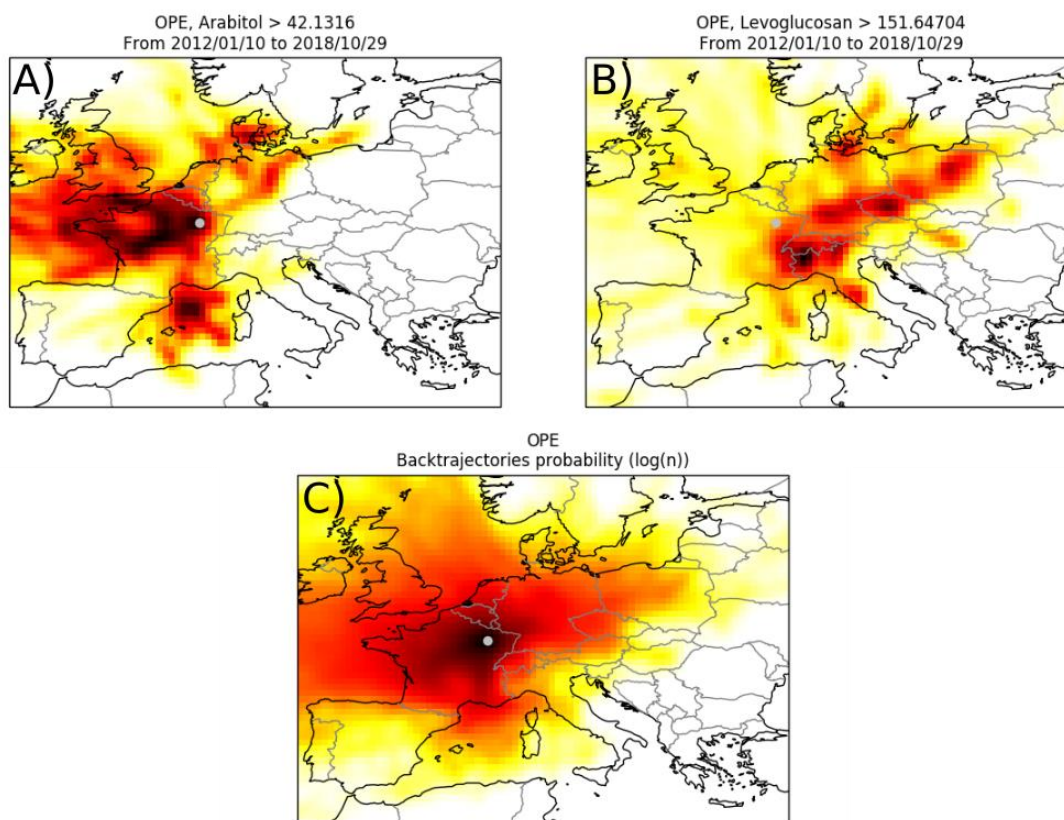


Figure R2.2:A) PSCF analysis for the OPE site (using pyPSCF and HYSPLIT). Back-trajectories associated with arabitol concentrations higher than the 75th percentile divided by the number of back-trajectories. B) displays all the back-trajectories. The colorscale indicated the probability that the specie comes from this cell (the darker color indicate higher probability).

(4) Page 6, Line 174-180. The normalized cross-correlation (NCC) test was chosen in this manuscript, and author mentioned a thorough discussion on the normalized cross-correlation method can be found elsewhere (Kaso, 2018; Yoo and Han, 2009). However, there was no related applied reference of NCC method was given, more field observation references used this methods are suggested to add.

The reviewer is right. The references related to the NCC method described only the concept and theory of NCC method. Sorry about it. Additional references (Bardal and Sætran, 2016; Dai and Zhou, 2017; Eisner et al., 2009; Lainer et al., 2016; Le Pichon et al., 2019) are now given to illustrate NCC applications in atmospheric sciences (lines 206—207).

(5) The lines in the figures are too thick to find the points, especially for Fig. (2a), Fig. (2b) and Fig. 5. It is difficult to separate the different color lines. Moreover, the thickness of the lines seems not consistent, i.e., the blue lines seem thicker than other color lines.

These figures have been modified accordingly. Thanks for suggestion.

(6) Figure S2 is suggested to add in Figure 3. Discuss the Normalized cross-correlation values for the daily evolution of particulate for glucose, polyols, calcium and sulfate together. It can exhibit the differences of NCC between these chemicals more directly. Moreover, how about NCC of other

inorganic ions, i.e., NH_4^+ , NO_3^- (similar as SO_4^{2-} , are the main components of secondary inorganic aerosols), K^+ (biomass burning tracer) and Cl^- .

The present work do not aim at discussing these species. Nevertheless, since they may act as a negative control for the local emission of polyols, we initially presented some of them in the submitted manuscript and SI. As suggested by the reviewer, former Fig S2 has also added in the main text together with fig 3. and we are now presenting some major secondary inorganics (ammonium) and biomass burning proxy (levoglucosan) as follows:

Lines 266-279 “Unlike SC, ambient air concentrations of sulfate (Fig. 3C) and ammonium (Fig. 3D), associated with long-range aerosol transport (Abdalmogith and Harrison, 2005; Amato et al., 2016; Coulibaly et al., 2015; Pindado and Perez, 2011; Waked et al., 2014) and levoglucosan ((Fig. 3E), associated with biomass burning in cold season (Weber et al., 2019; Xiao et al., 2018), display stronger positive correlations ($R > 0.72-0.98$, $p < 0.01$) at all pairs of sites considered in the present work. Moreover, ambient concentrations of calcium (Fig. 3F), associated with local fugitive dust sources or/and long-range aerosol transport (Ram et al., 2010; Wan et al., 2019) display random correlation patterns. These results are in agreement with Zhu et al. (2018) who also reported non-significant correlations between SC and sulfate in $\text{PM}_{2.5}$ aerosols measured at Shanghai, China. The distinct spatial behaviors between sulfate (or Ca^{2+}) and SC in the present work further suggest a dominant regional influence for atmospheric SC, as opposed to processes associated with either local sources for calcium or long-range transport for sulfate”.

For secondary species (sulfate and ammonium), potential long range transport (Ca^{2+}) and chemically stable species (levoglucosan, Figure R2.2B), the correlation are still high ($r > 0.7$) even after hundreds of kilometers. For these species, we can make the hypothesis that the regional transport play a major role in concentrations seen at a given site. However, an in-depth analysis of the sources and evolution of the concentrations of these species is beyond the scope of this study and would require a dedicated future work.

Specific comments:

(1) Line 190: The linear regression (lm) package in R was employed for multiple regression analyses. What does “lm” in the bracket means??

The linear model aka “lm” in the brackets is the name of the statistical package employed for multiple regression analyses. Definition of “lm” is now added in the main text (line 217).

(2) Line 320: these findings highlight that particulate SC PM_{10} and cellulose in both urban background and rural agricultural areas. . ., should be changed to “these findings highlight that SC in PM_{10} and cellulose in both urban background and rural agricultural areas

Thank you for your attentive review, this sentence has been corrected (lines 367—368).

Answer to Anonymous Referee #3 comments

This paper describes the evolutions of glucose, mannitol and arabitol in the aerosol covering 16 sites all over France. The study consists in a huge and precious dataset. For the first time, the distance-dependent correlation is demonstrated, investigating also the main drivers of atmospheric sugar concentrations.

We thank the reviewer for his/her attention to our manuscript that greatly contribute to improve the quality of this research paper. We have considered each of the comments and we have made revisions point by point. The detailed responses to the comments are given below, point by point, *in blue color*, including changes directly made to the manuscript, *in red color*.

General comment: please check all manuscript, including figures and tables, and modify the term “polyols” with “mannitol and arabitol”, as necessary, to avoid confusion., as suggested in the initial revision.

We do agree with the reviewer that we are not analyzing all the sugar alcohol species. However, we clearly specify that the term polyols is to refer to the sum of arabitol and mannitol concentrations (lines 178—179). This has been added to the main text:

“Hereafter, the term “Polyols” is used to refer uniquely to the sum of arabitol and mannitol concentrations”.

Line 41-43. The authors affirmed that “sugar alcohols . . . - including arabitol and mannitol. . . - have been recognized as tracers for airborne fungi”. One of the main objectives of my recent research is the source investigation of water soluble organic compounds, such as for example sugars, and I quite sure that some sugars alcohols have another source. For examples I saw that sorbitol have some correlation with biomass burning tracers, while arabitol and mannitol, mainly distributed in the coarse fraction of aerosol, plausibly originate from fungal spores. So, I suggest to focus your affirmation only on the arabitol and mannitol.

We do agree with the reviewer and we have focused our affirmation only on the arabitol and mannitol.

Line 45. The authors define glucose “a specific tracer for plant materials” but I think that the authors should remove “specific” because glucose can have different sources: plant materials, soil emissions (as suggested by the authors) and also marine biogenic material derived from degradation of polysaccharides present in the marine microlayer. I suggest to read some papers of Prof. Leck because she investigated the organic compounds (such as saccharides) in the marine aerosol. I know that the paper is focused on the aerosol samples collected in the areas far from the coast but in the introduction I think that the authors should consider all sources.

Indeed, glucose can have a broad of biogenic sources, e.g. from terrestrial plant pollen, fruits, and detritus, or from the degradation of the soil microorganisms (Kang et al., 2018; Li et al., 2018; Xiao et al., 2018) or even possibly from bubble bursting processes in remote oceans (Fu et al., 2013; Gao et al., 2011; Leck and Bigg, 2005); we have removed the term “specific” (see lines 54—59). This point is also further discussed in the response to the comment 1 of anonymous referee # 1.

Line 174. Can you specify some details about the dataset matrix using to perform the normalized cross correlation.

The raw data used in the present study consisted in to daily (24 hours) aerosols collected at 16 sites in different geographic regions in France. For pairwise normalized cross-correlation analyses, original daily series were first converted as follows: starting on identical days (for each pairs of sites), arrangement on the original daily data into consecutive 3-day intervals (or 6-day intervals in the case of

OPE-ANDRA) and calculation of the average concentration values for the middle-day were performed. We directly used this resultant data for the correlation analysis between site pairs (lines 207—212).

In this respect, the manuscript has been revised as follows: lines 207-212

“To achieve pair-wise correlation analysis between the sampling sites collected during the same periods, the original raw daily measurements were processed as follows: starting on identical days for each pairs of sites, arrangement on the original daily data into consecutive 3-day intervals (or 6-day intervals in the case of OPE-ANDRA) and calculation of the average concentration values for the middle-day were performed. The resultant data were used for correlation analysis between site pairs (Table S3)”.

Line 257. You correctly affirmed that mannitol-to arabitol ratio can suggest the temporal and spatial evolution of their emission processes, using this reference: Gosselin et al. 2016. This paper demonstrated also that, in some cases, mannitol and arabitol can have different sources: “mannitol is a common polyol in higher plants while arabitol is only found in fungal spores and lichen”. I suggest to insert this concept in the manuscript and to consider the R² between two polyols in the discussion because maybe either conclusion can be also obtained (this is just a suggestion).

Thank you for this interesting suggestion. We added discussion about R² between arabitol and mannitol (lines 288—292), as follows: lines 282-293.

“Based on parallel measurements of spore counts and PM₁₀ polyol concentrations at three sites within the area of Vienna (Austria), Bauer et al. (2008a) found an average arabitol and mannitol content per fungal spores of respectively 1.2 pg spore⁻¹ (range 0.8-1.8 pg spore⁻¹) and 1.7 pg spore⁻¹ (range 1.2-2.4 pg spore⁻¹). Mannitol and arabitol have also been often identified in the green algae and lower plants (Buiarelli et al., 2013; Di Filippo et al., 2013; Véléz et al., 2007; Xu et al., 2018; Zhang et al., 2010). Gosselin et al., 2016 observed a relatively low (R² = 0.31) to high (R² = 0.84) coefficient of determination between mannitol and arabitol for total suspended particles (TSP) collected at a pine-forested area during dry and rainy periods, respectively. High correlation in rainy periods possibly suggested that both chemical species in the TSP fraction in this pine-forested area could have been derived mainly from the same sources, i.e., actively wet-discharged ascospores and basidiospores, while the poor correlation in dry periods could have been likely due to more complex sources, i.e., dry discharged spores, plants, algae, etc.”

Note that the study by Gosselin et al., 2016 has been conducted on total suspended particles (TSP) and in a specific pine-forested area of North America. The high coefficient of determination reported by Gosselin et al., 2016 during the rainy periods possibly suggest that both chemical species in the TSP fraction in this pine-forested area could have been derived mainly from the same sources, i.e., actively wet-discharged ascospores and basidiospores. However, the relatively poor correlation in dry periods could have been likely due to more complex sources, i.e., a mixture of actively wet and/or dry discharged spores, or influence of additional biogenic sources such as plants, algae, etc. Indeed, active release of wet discharged ascospores and basidiospores occurs in most ascomycetes and in basidiomycetes (Ingold and Hudson, 1993; Zhang et al., 2010) which is influenced by ambient humidity and rainfall (Elbert et al., 2007; Zhang et al., 2015). In contrast, dry discharged spores are preferentially emitted under dry and warm conditions. Thus, these correlation patterns could be at least partially explained by the different fungal habitats and/or different emission processes during rainy and dry periods.

Section 3.2. The distance-dependent correlations and the SC evolution synchronous at an urban city scale and throughout the same geographical regions are the very interesting topics in the manuscript and I appreciate this work because it was a lack of the sugars knowledge. The distance-dependent correlations is very clear using your approach but I suggest to clarify the main reasons for the decrease of NCC when the distance was above 200 km. You report some explanations but I suggest to deeply discuss the reasons or the suggestion of this behavior.

Thank for this positive comment. We also believe that this point is quite innovative in the current literature and that it can considerably improve our knowledge about primary sugars in the atmosphere. We believe that the main reasons of such distance correlation patterns are most probably associated with different airborne microbial community assemblies that are shaped by different regional environmental factors (e.g. meteorological conditions, vegetation types and cover, etc.). Indeed, our recent interdisciplinary work (submitted for publication in “Science advances”) has shown that the atmospheric concentration dynamics of polyols and some major saccharides (trehalose, glucose) are driven by only a few specific airborne fungal and bacterial genera. Further analyses for sites located in different climatic regions of France have also shown that airborne microbial assemblies associated with these chemical species vary regionally (unpublished data). This makes sense since different biotopes (meteorological conditions, vegetation types and cover) harbor distinct microbial communities (Bowers et al., 2012; Liu et al., 2019).

Line 301. Please remove “s” from “corresponds”.

Thank you for your careful review, we have removed this “s”.

References:

Bowers, R. M., McCubbin, I. B., Hallar, A. G. and Fierer, N.: Seasonal variability in airborne bacterial communities at a high-elevation site, *Atmos. Environ.*, 50, 41–49, doi:10.1016/j.atmosenv.2012.01.005, 2012.

Elbert, W., Taylor, P. E., Andreae, M. O. and Pöschl, U.: Contribution of fungi to primary biogenic aerosols in the atmosphere: wet and dry discharged spores, carbohydrates, and inorganic ions, *Atmospheric Chem. Phys.*, 7(17), 4569–4588, doi:10.5194/acp-7-4569-2007, 2007.

Fu, P. Q., Kawamura, K., Chen, J., Charrière, B. and Sempéré, R.: Organic molecular composition of marine aerosols over the Arctic Ocean in summer: contributions of primary emission and secondary aerosol formation, *Biogeosciences*, 10(2), 653–667, doi:10.5194/bg-10-653-2013, 2013.

Gao, Q., Nilsson, U., Ilag, L. L. and Leck, C.: Monosaccharide compositional analysis of marine polysaccharides by hydrophilic interaction liquid chromatography-tandem mass spectrometry, *Anal. Bioanal. Chem.*, 399(7), 2517–2529, doi:10.1007/s00216-010-4638-z, 2011.

Gosselin, M. I., Rathnayake, C. M., Crawford, I., Pöhlker, C., Fröhlich-Nowoisky, J., Schmer, B., Després, V. R., Engling, G., Gallagher, M., Stone, E., Pöschl, U. and Huffman, J. A.: Fluorescent bioaerosol particle, molecular tracer, and fungal spore concentrations during dry and rainy periods in a semi-arid forest, *Atmospheric Chem. Phys.*, 16(23), 15165–15184, doi:10.5194/acp-16-15165-2016, 2016.

Ingold, T. C. and Hudson, H. J.: *The biology of fungi*, Springer Netherlands. [online] Available from: doi:10.1007/978-94-011-1496-7, 1993.

Kang, M., Fu, P., Kawamura, K., Yang, F., Zhang, H., Zang, Z., Ren, H., Ren, L., Zhao, Y., Sun, Y. and Wang, Z.: Characterization of biogenic primary and secondary organic aerosols in the marine atmosphere over the East China Sea, *Atmospheric Chem. Phys. Discuss.*, 1–45, doi:10.5194/acp-2018-318, 2018.

Leck, C. and Bigg, E. K.: Biogenic particles in the surface microlayer and overlaying atmosphere in the central Arctic Ocean during summer, *Tellus B*, 57(4), 305–316, doi:10.1111/j.1600-0889.2005.00148.x, 2005.

Li, Y.-C., Shu, M., Ho, S. S. H., Yu, J.-Z., Yuan, Z.-B., Wang, X.-X., Zhao, X.-Q. and Liu, Z.-F.: Effects of Chemical Composition of PM_{2.5} on Visibility in a Semi-Rural City of Sichuan Basin, *Aerosol Air Qual. Res.*, 18(4), 957–968, 2018.

Liu, H., Hu, Z., Zhou, M., Hu, J., Yao, X., Zhang, H., Li, Z., Lou, L., Xi, C., Qian, H., Li, C., Xu, X., Zheng, P. and Hu, B.: The distribution variance of airborne microorganisms in urban and rural environments, *Environ. Pollut.*, 247, 898–906, doi:10.1016/j.envpol.2019.01.090, 2019.

Xiao, M., Wang, Q., Qin, X., Yu, G. and Deng, C.: Composition, Sources, and Distribution of PM_{2.5} Saccharides in a Coastal Urban Site of China, *Atmosphere*, 9(7), 274, doi:10.3390/atmos9070274, 2018.

Zhang, T., Engling, G., Chan, C.-Y., Zhang, Y.-N., Zhang, Z.-S., Lin, M., Sang, X.-F., Li, Y. D. and Li, Y.-S.: Contribution of fungal spores to particulate matter in a tropical rainforest, *Environ. Res. Lett.*, 5(2), 024010, doi:10.1088/1748-9326/5/2/024010, 2010.

Zhang, Z., Engling, G., Zhang, L., Kawamura, K., Yang, Y., Tao, J., Zhang, R., Chan, C. and Li, Y.: Significant influence of fungi on coarse carbonaceous and potassium aerosols in a tropical rainforest, *Environ. Res. Lett.*, 10(3), 034015, doi:10.1088/1748-9326/10/3/034015, 2015.

Arabitol, mannitol and glucose as tracers of primary biogenic organic aerosol: influence of environmental factors on ambient air concentrations and spatial distribution over France

Abdoulaye Samaké¹, Jean-Luc Jaffrezo¹, Olivier Favez^{2,3}, Samuël Weber¹, Véronique Jacob¹, Trishalee Canete¹, Alexandre Albinet^{2,3}, Aurélie Charron^{1,4,16}, Véronique Riffault^{5,3}, Esperanza Perdrix^{5,3}, Antoine Waked¹, Benjamin Golly¹, Dalia Salameh^{1*}, Florie Chevrier^{1,6,13,4}, Diogo Miguel Oliveira^{2,5,3}, Jean-Luc Besombes^{4,6}, Jean M.F. Martins¹, Nicolas Bonnaire^{7,5}, Sébastien Conil^{8,6}, Géraldine Guillaud^{9,7}, Boualem Mesbah^{8,10}, Benoit Rocq^{11,9}, Pierre-Yves Robic^{10,2}, Agnès Hulin^{14,3}, Sébastien Le Meur^{12,4}, Maxence Descheemaeker^{15,3}, Eve Chretien^{16,4}, Nicolas Marchand^{17,5}, and Gaëlle Uzu¹.

¹University Grenoble Alpes, CNRS, IRD, INP-G, IGE (UMR 5001), 38000 Grenoble, France

²INERIS, Parc Technologique Alata, BP 2, F-60550 Verneuil-en-Halatte, ~~France~~France

³Laboratoire Central de Surveillance de la Qualité de l'Air (LCSQA), F-60550 Verneuil-en-Halatte, France

⁴IFSTTAR, F-69675 Bron, France

⁵IMT Lille Douai, University Lille, SAGE – Département Sciences de l'Atmosphère et Génie de l'Environnement, 59000 Lille, ~~France~~France

¹⁶IFSTTAR, F-69675 Bron, France

⁶University Savoie Mont-Blanc, LCME, 73000 Chambéry, France

⁷LSCE, UMR CNRS-CEA-UVSQ, 91191 Gif-sur Yvette, France

⁸ANDRA DRD/GES Observatoire Pérenne de l'Environnement, F-55290 Bure, France

⁹Atmo Auvergne-Rhône-Alpes, 38400 Grenoble, France

¹⁰Air PACA, 03040, France

¹¹Atmo Hauts de France, 59000, France

¹²Atmo Occitanie, 31330 Toulouse, France

¹³Atmo Nouvelle Aquitaine, 33000, France

¹⁴Atmo Normandie, 76000, France

¹⁵Lig' Air, 45590 Saint-Cyr-en-Val, France

¹⁶Atmo Grand Est, 16034 Strasbourg, France

¹⁷University Aix Marseille, LCE (UMR7376), Marseille, France

¹⁶IFSTTAR, F-69675 Bron, France

*Now at: Airport pollution control authority (ACNUSA), 75007 Paris, France

Corresponding author(s): A Samaké (abdoulaye.samake2@univ-grenoble-alpes.fr) and JL Jaffrezo (Jean-luc.Jaffrezo@univ-grenoble-alpes.fr)

1 **Abstract.** The primary sugar compounds (SC, defined as glucose, arabinol and mannitol) are widely recognized as
2 suitable molecular markers to characterize and apportion primary biogenic organic aerosol emission sources. This
3 work improves our understanding of the spatial behavior and distribution of these chemical species and evidences
4 their major effective environmental drivers. We conducted a large study focusing on the daily (24 h) PM₁₀ SC
5 concentrations for 16 increasing space scale sites (local to nation-wide), over at least one complete year. These
6 sites are distributed in several French geographic areas of different environmental conditions. Our analyses, mainly
7 based on the examination of the short-term evolutions of SC concentrations, clearly show distance-dependent
8 correlations. SC concentration evolutions are highly synchronous at an urban city-scale and remain well correlated
9 throughout the same geographic regions, even if the sites are situated in different cities. However, sampling sites
10 located in two distinct geographic areas are poorly correlated. Such pattern indicates that the processes responsible
11 for the evolution of the atmospheric SC concentrations present a spatial homogeneity over typical areas of at least
12 tens of kilometers. Local phenomena, such as resuspension of topsoil and associated microbiota, do not account for
13 the major emissions processes of SC in urban areas not directly influenced by agricultural activities. The
14 concentrations of SC and cellulose display remarkably synchronous temporal evolution cycles at an urban site in
15 Grenoble, indicating a common source ascribed to vegetation. Additionally, higher concentrations of SC at another
16 site located in a crop field region occur during each harvest periods, pointing out resuspension processes of plant
17 materials (crop detritus, leaf debris) and associated microbiota for agricultural and nearby urbanized areas. Finally,
18 ambient air temperature, relative humidity and vegetation density constitute the main effective drivers of SC
19 atmospheric concentrations.

20 **1. Introduction**

21 Primary biogenic organic aerosols (PBOA), which notably comprise bacterial and fungal cells or spores; viruses;
22 or microbial fragments such as endotoxins and mycotoxins; and pollens and plant debris, are ubiquitous particles
23 released from the biosphere to the atmosphere ([Amato et al., 2017](#); [Fang et al., 2018](#); [Martin et al., 2010](#); [Perrino](#)
24 [and Marcovecchio, 2016](#); [Wéry et al., 2017](#)) (~~Amato et al., 2017; Després et al., 2012; Elbert et al., 2007; Fang et~~
25 ~~al., 2018; Fröhlich-Nowoisky et al., 2016; Morris et al., 2011; Wéry et al., 2017~~). PBOA can contribute
26 significantly to the total coarse aerosol mass ([Amato et al., 2017](#); [Bozzetti et al., 2016](#); [Coz et al., 2010](#); [Fröhlich-](#)
27 [Nowoisky et al., 2016](#); [Jaenicke, 2005](#); [Manninen et al., 2014](#); [Morris et al., 2011](#); [Samaké et al., 2019](#); [Vlachou](#)
28 [et al., 2018](#); [Yue et al., 2017](#)). Besides their expected negative human health effects ([Fröhlich-Nowoisky et al.,](#)
29 [2009, 2016](#); [Humbal et al., 2018](#); [Lecours et al., 2017](#); [Zamfir et al., 2019](#)) (~~Fröhlich-Nowoisky et al., 2009, 2016;~~
30 ~~Humbal et al., 2018; Lecours et al., 2017~~), they substantially influence the carbon and water cycles at the global
31 scale, notably acting as cloud and ice nuclei ([Ariya et al., 2009](#); [Elbert et al., 2007](#); [Fröhlich-Nowoisky et al., 2016](#);
32 [Hill et al., 2017](#); [Humbal et al., 2018](#); [Morris et al., 2014](#); [Rajput et al., 2018](#)). While recent studies have revealed
33 highly relevant information on the abundance and size partitioning of PBOA ([Fröhlich-Nowoisky et al., 2017](#);
34 [Huffman and Santarpia, 2017](#)), their emission sources and contribution to total airborne particles are still poorly
35 documented, partly due to the analytical limitations to distinguish PBOA from other types of carbonaceous
36 particulate matter ([Bozzetti et al., 2016](#); [China et al., 2018](#); [Di Filippo et al., 2013](#); [Perrino and Marcovecchio,](#)
37 [2016](#); [Yan et al., 2019](#)) (~~Bozzetti et al., 2016; China et al., 2018; Di Filippo et al., 2013; Heald and Spracklen,~~
38 ~~2009; Jia et al., 2010~~). Notably, the global emissions of fungal spore emitted into the atmosphere are still poorly
39 constrained and range from 8 Tg.y⁻¹ to 186 Tg.y⁻¹ ([Després et al., 2012](#); [Elbert et al., 2007](#); [Jacobson and Streets,](#)
40 [2009](#); [Sesartic and Dallafior, 2011](#); [Tanarhte et al., 2019](#)) (~~Després et al., 2012; Elbert et al., 2007; Jacobson and~~
41 ~~Streets, 2009; Sesartic and Dallafior, 2011~~).

42 Recently, source-specific tracer methodologies have been introduced to estimate their contribution to aerosol
43 loadings ([Di Filippo et al., 2013](#); [Gosselin et al., 2016](#); [Li et al., 2018](#); [Medeiros et al., 2006b](#); [Verma et al., 2018](#);
44 [Wang et al., 2018](#)) (~~Bauer et al., 2008a; Di Filippo et al., 2013; Gosselin et al., 2016; Zhang et al., 2010,~~

45 ~~2015~~). Indeed, atmospheric organic aerosols (OA) contain specific chemical species that can be used as reliable
46 biomarkers in tracing the sources and abundance of PBOA ([Bauer et al., 2008](#); [Gosselin et al., 2016](#); [Holden et al.,](#)
47 [2011](#); [Jia et al., 2010](#); [Li et al., 2018](#); [Medeiros et al., 2006b](#); [Wang et al., 2018](#))~~([Bauer et al., 2008a](#); [Gosselin](#)~~
48 ~~[et al., 2016](#); [Holden et al., 2011](#); [Jia and Fraser, 2011](#); [Medeiros et al., 2006b](#))~~. For instance, among sugar
49 alcohols, (aka polyols) ~~including~~ arabitol and mannitol (two common storage soluble carbohydrates in fungi)—
50 have been recognized as tracers for airborne fungi, and their concentrations are widely used to estimate PBOA
51 contributions to OA mass ([Amato et al., 2017](#); [Bauer et al., 2008](#); [Buiarelli et al., 2013](#); [Golly et al., 2018](#); [Medeiros](#)
52 [et al., 2006b](#); [Samaké et al., 2019](#); [Srivastava et al., 2018](#); [Verma et al., 2018](#); [Weber et al., 2018, 2019](#))~~([Amato et](#)~~
53 ~~[al., 2017](#); [Bauer et al., 2008a, 2008b](#); [Golly et al., 2018](#); [Medeiros et al., 2006b](#); [Samaké et al., 2019](#); [Verma et al.,](#)~~
54 ~~[2018](#); [Weber et al., 2018](#); [Zhang et al., 2010](#); [Zhu et al., 2015, 2016](#))~~. Similarly, glucose has also been used as a
55 ~~specific~~ tracer for plant materials (such as pollen, leaves, and their fragments) or soil emissions within various
56 studies around the world ([Chen et al., 2013](#); [Medeiros et al., 2006b](#); [Pietrogrande et al., 2014](#); [Rathnayake et al.,](#)
57 [2017](#); [Rogge et al., 2007](#); [Wan et al., 2019](#); [Xiao et al., 2018](#); [Zhu et al., 2015](#))~~([Chen et al., 2013](#); [Fu et al., 2013](#);~~
58 ~~[Liang et al., 2016](#); [Medeiros et al., 2006b](#); [Pietrogrande et al., 2014](#); [Rathnayake et al., 2017](#); [Rogge et](#)~~
59 ~~[al., 2007](#); [Simoneit et al., 2004b](#); [Wan and Yu, 2007](#); [Wan et al., 2019](#))~~.

60 In this context, atmospheric concentrations of specific ~~polyols~~ sugar alcohols and/or primary monosaccharides
61 (including glucose) have been previously quantified at sites in several continental, agricultural, coastal or polar
62 regions ([Barbaro et al., 2015](#); [Chen et al., 2013](#); [Gladius et al., 2018](#); [Li et al., 2018](#); [Pietrogrande et al., 2014](#);
63 [Verma et al., 2018](#); [Wan et al., 2019](#); [Yan et al., 2019](#); [Yttri et al., 2007](#))~~([Barbaro et al., 2015](#); [Chen et al.,](#)~~
64 ~~[2013](#); [Fu et al., 2012](#); [Golly et al., 2018](#); [Graham et al., 2003](#); [Jia et al., 2010](#); [Liang et al., 2016](#);~~
65 ~~[Pietrogrande et al., 2014](#); [Rogge et al., 2007](#); [Simoneit et al., 2004a](#); [Verma et al., 2018](#); [Yttri et al.,](#)~~
66 ~~[2007](#); [Zhu et al., 2018](#))~~. However, large datasets investigating their (multi)annual cycles, seasonal and
67 simultaneous short-term variations at multiple spatial scale resolutions (i.e. from local to continental) are still
68 lacking ([Liang et al., 2013](#); [Nirmalkar et al., 2018](#); [Pietrogrande et al., 2014](#); [Yan et al., 2019](#)). Such records are
69 essential to better understand the spatial behavior of primary sugar compound (SC) concentrations (i.e., glucose,
70 arabitol and mannitol) and PBOA emission processes, and to isolate their potential key drivers (e.g., vegetation
71 type and density, topography, weather conditions, etc.), which are still unclear ([Bozzetti et al., 2016](#)). This
72 information would be essential for further implementation into chemical transport models ([Heald and Spracklen,](#)
73 [2009](#); [Myriokefalitakis et al., 2017](#); [Tanarhte et al., 2019](#))~~([Heald and Spracklen, 2009](#); [Tanarhte et al., 2019](#))~~.

74 It is commonly acknowledged that SC (particularly arabitol and mannitol) originate from primary biogenic derived
75 sources such as bacterial, fungal spores, and plant materials ([Di Filippo et al., 2013](#); [Golly et al., 2018](#); [Gosselin et](#)
76 [al., 2016](#); [Holden et al., 2011](#); [Kang et al., 2018](#); [Medeiros et al., 2006b](#); [Wan et al., 2019](#); [Yan et al., 2019](#); [Yttri et](#)
77 [al., 2007](#); [Zhu et al., 2018a](#))~~([Di Filippo et al., 2013](#); [Golly et al., 2018](#); [Gosselin et al., 2016](#); [Graham et al.,](#)~~
78 ~~[2003](#); [Holden et al., 2011](#); [Medeiros et al., 2006b](#); [Simoneit et al., 2004b](#); [Wan et al., 2019](#); [Yan et al.,](#)~~
79 ~~[2019](#); [Yttri et al., 2007, 2011a](#); [Zhu et al., 2015](#))~~. Some studies have characterized the composition of SC in
80 topsoil samples (for fractions larger than PM₁₀) from both, natural (i.e., uncultivated) and agricultural regions
81 ([Medeiros et al., 2006a](#); [Rogge et al., 2007](#); [Simoneit et al., 2004](#); [Wan and Yu, 2007](#))~~([Medeiros et al., 2006a](#);~~
82 ~~[Rogge et al., 2007](#); [Simoneit et al., 2004b](#); [Wan and Yu, 2007](#))~~. The authors suggested that the particulate

83 arabitol, mannitol and glucose are introduced into the atmosphere mainly through resuspended soils or dust
84 particles and associated biota derived from natural soil erosion, unpaved road dust or agricultural practices.
85 Conversely, [Jia and Fraser \(2011\)](#)~~Jia and Fraser (2011)~~ reported higher concentrations of SC relative to PBOA
86 in size-segregated aerosol samples collected at a suburban site (Higley, USA) compared to the local size-
87 fractionated soils (equivalent to atmospheric PM_{2.5} and PM₁₀). This suggested that direct emissions from biota
88 (microbiota, vascular plant materials) could also be a significant atmospheric input process for SC at this suburban
89 site.

90 A large database on SC concentrations was obtained over France in the last decade. It already allowed the
91 investigation of the size distribution and seasonal variabilities of SC concentrations in aerosols at 28 French sites,
92 notably showing that SC are ubiquitous primary aerosols, accounting for a significant proportion of PM₁₀ organic
93 matter (OM) mass (Samaké et al., 2019). Results confirmed that their ambient concentrations display a well-
94 marked seasonality, with maximum concentrations from late spring to early autumn, followed by an abrupt
95 decrease in late autumn, and a minimum concentration during wintertime in France. This study also showed that
96 the mean PBOA chemical profile is largely dominated by organic compounds, with only a minor contribution of
97 dust particle fraction. The latter result indicated that ambient polyols could most likely be associated with direct
98 biological particle emissions (e.g. active spore discharge, microbiota released from phylloplane or phyllosphere,
99 etc.) rather than with the microorganism-containing soil resuspension. These observations call for more
100 investigations of the predominant SC (and PBOA) emission sources.

101 Cellulose, a linear polymer composed of D-glucopyranose units linked by β -1,4 bonds, is the most frequent
102 polysaccharide occurring in terrestrial environments ([Ramoni and Seiboth, 2016](#))~~(Ramoni and Seiboth, 2016)~~.
103 Plant materials contain cellulose which has been reported as a suitable proxy to evaluate the vegetative debris
104 contribution to OM mass ([Bozzetti et al., 2016](#); [Daellenbach et al., 2017](#); [Glasius et al., 2018](#); [Hiranuma et al.,](#)
105 [2019](#); [Puxbaum and Tenze-Kunit, 2003](#); [Sánchez-Ochoa et al., 2007](#); [Yttri et al., 2011b](#))~~(Bozzetti et al., 2016;
106 ~~Glasius et al., 2018; Puxbaum and Tenze Kunit, 2003; Sánchez Ochoa et al., 2007; Yttri et al., 2011b)~~. The
107 ambient PM₁₀ cellulose has been shown to be abundant in the European semi-rural or background environments
108 (accounting for 2 to 10 % of OM mass) ([Glasius et al., 2018](#); [Sánchez-Ochoa et al., 2007](#))~~(Glasius et al., 2018;
109 ~~Sánchez Ochoa et al., 2007)~~ and Nordic rural environments in Norway (contributing to 12 to 18 % of total carbon
110 mass) ([Yttri et al., 2011b](#))~~(Yttri et al., 2011b)~~. Thus, simultaneous concentration measurements of cellulose and
111 SC can provide essential information into their emission source dynamics.~~~~

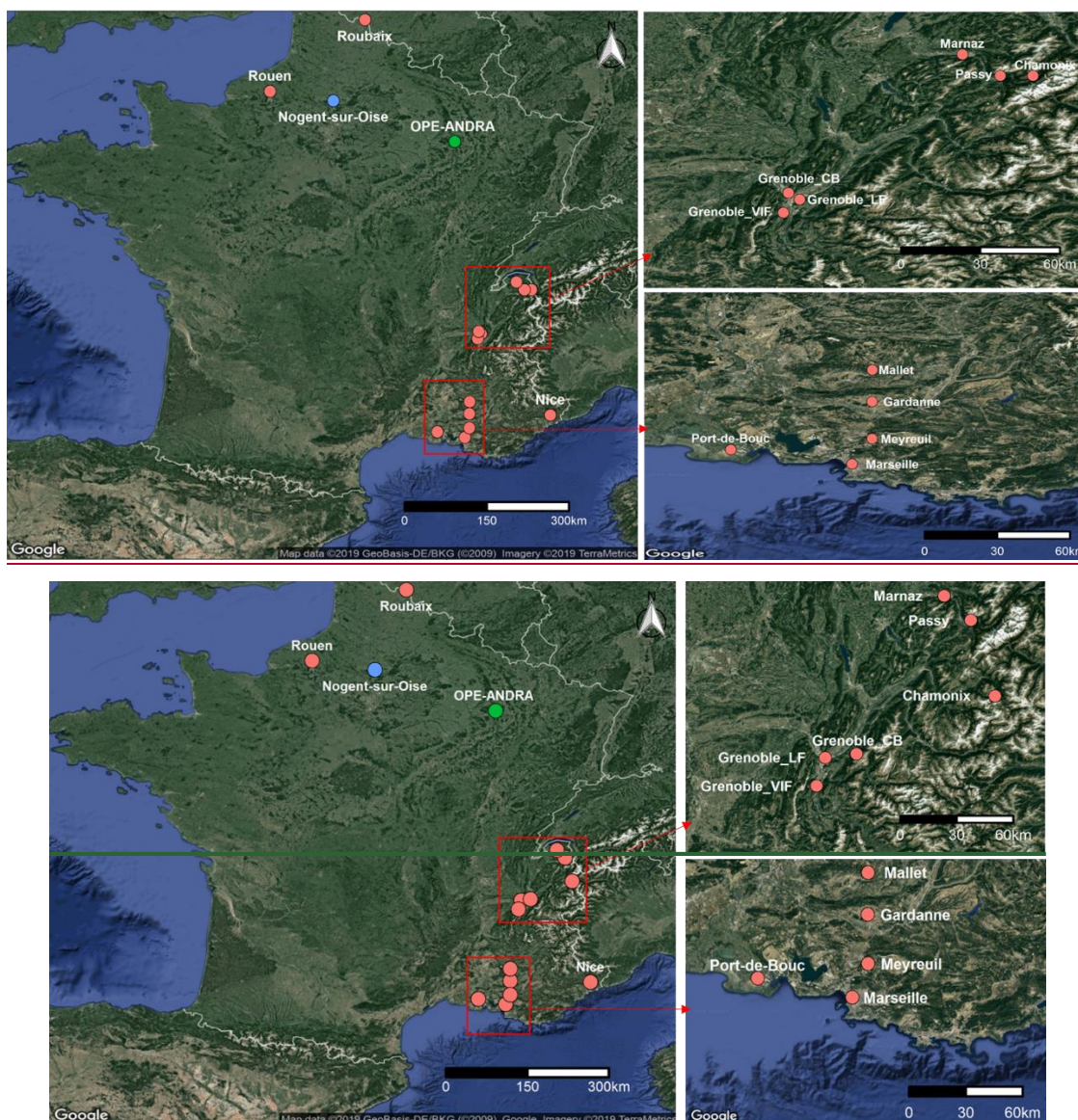
112 As the continuation of our previous work ([Samaké et al., 2019](#))~~(Samaké et al., 2019)~~, the present paper aims to
113 delineate the processes that drive the atmospheric concentrations of SC and then PBOA. This is achieved through
114 (i) the analysis of simultaneous annual short-term time series of particulate SC concentrations over pairs of sites
115 across multiple space ranges, including local, regional and nationwide sites, and (ii) the investigation of links
116 between concentrations and series key parameters such as meteorological and phenological ones. Simultaneous
117 annual short-term concentration measurements of SC and cellulose was performed to better understand of their
118 sources correlations.

119 **2. Material and methods**

120 **2.1 Sampling sites**

121 Daily PM₁₀ concentrations reported in the present work were obtained from different research and monitoring
122 programs conducted over the last six years in France. Within the framework of the present study, we carefully

123 selected sites sharing at least one complete year of concurrent monitoring with another one, to be representative
 124 of the annual variation cycles. The final dataset includes data from 16 sites, which are distributed in different
 125 regions of France (Figure 1) and cover several main types of environmental conditions in terms of site topography,
 126 local vegetation, and climate. The characteristics and data available at each sampling site are listed in Table S1 of
 127 the supplementary material (SM), together with the information on the annual average concentrations of aerosol
 128 chemical composition (Table S2). Detailed information on the sampling conditions can be found in Samaké et al.
 129 (2019), such as the campaign periods, number of collected PM samples, sampling flow rates, sample storage and
 130 handling, etc. Note that, the previous database (Samaké et al., 2019) (Samaké et al., 2019) has been updated here
 131 with arabitol and mannitol in PM₁₀ collected at the suburban site of Nogent-sur-Oise for a series covering the years
 132 2013 to 2017.



133

134

135 **Figure 3: Geographical location of the selected sampling sites. The red and blue dots indicate respectively urban and**
 136 **suburban sites while the green one corresponds to a rural site, surrounded by field crop areas.**

137 2.2 Chemical analyses

138 Daily (24 h) PM₁₀ samples were collected onto prebaked quartz fiber filter (Tissuquartz PALL QAT-UP 2500 150
 139 mm diameter) every third or sixth day, but not concurrently at all sites. They were then analyzed for various

140 chemical species using subsampled fractions of the collection filters and a large array of analytical methods. Details
141 of all the chemical analysis procedures are reported elsewhere ([Golly et al., 2018](#); [Samaké et al., 2019](#); [Waked et
142 al., 2014](#); [Weber et al., 2018](#))(~~[Golly et al., 2018](#); [Samaké et al., 2019](#); [Waked et al., 2014](#); [Weber et al.,
143 2018](#)~~). Briefly, primary sugar compounds were extracted from filter aliquots (punches typically about 10 cm²) into
144 ultrapure water. The extracts are then filtered using a 0.22 µm Acrodisc filter. Depending on the site, analyses
145 were conducted either by the IGE (Institut des Géosciences de l'Environnement) or by the LSCE (Laboratoire des
146 Sciences du Climat et de l'Environnement) ([Samaké et al., 2019](#))(~~[Samaké et al., 2019](#)~~). At the IGE, extraction
147 was performed during 20 min in a vortex shaker and analyses were achieved using high-performance liquid
148 chromatography with pulsed amperometric detection (HPLC-PAD). A first set of equipment was used until March
149 2016, consisting of a Dionex DX500 equipped with three columns Metrosep (Carb 1-Guard + A Supp 15-150 +
150 Carb 1-150), the analytical program was isocratic with 70 mM sodium hydroxide (NaOH) as eluent for 11 min,
151 followed by a gradient cleaning step with a 120 mM NaOH as eluent for 9 min. This procedure allows the analysis
152 of arabitol, mannitol and glucose (Waked et al., 2014). A second set of equipment was used after March 2016,
153 with a Thermo-Fisher ICS 5000+ HPLC equipped with 4 mm diameter Metrosep Carb 2 × 150 mm column and
154 50 mm pre-column. The analytical run was isocratic with 15 % of an eluent of sodium hydroxide (200 mM) and
155 sodium acetate (4 mM) and 85 % water, at 1 mL min⁻¹. At the LSCE, extraction was performed for 45 min by
156 sonication and analyses were achieved using ion chromatography instrument (IC, DX600, Dionex) with Pulsed
157 Amperometric Detection (ICS3000, Thermo- Fisher). In addition, a CarboPAC MA1 column has been used (4 ×
158 250 mm, Dionex) along with an isocratic analytical run with 480 mM sodium hydroxide eluent. This analytical
159 technique allows to quantify arabitol, mannitol and glucose ([Srivastava et al., 2018](#))(~~[Srivastava et al., 2018](#)~~).
160 For cellulose quantification, we used an optimized protocol based on that described by (Kunit and Puxbaum, 1996;
161 Puxbaum and Tenze-Kunit, 2003), in which the cellulose contained in the lignocellulosic material is enzymatically
162 hydrolyzed into glucose units before analysis. Since the alkaline peroxide pretreatment step used to remove lignin
163 in the original protocol results in a loss of sample material, it has been avoided in this study. Therefore, only the
164 “free cellulose” is reported in our samples. Note that Sánchez-Ochoa et al., (2007) consider that this free cellulose
165 could represent only about 70 % of the total cellulose in air samples and that the total cellulose could represent
166 only about 50 % of the “plant debris” content of atmospheric PM. Very few other results are available on this topic
167 ([Bozzetti et al., 2016](#); [Glasius et al., 2018](#); [Vlachou et al., 2018](#); [Yttri et al., 2011b](#))(~~[Bozzetti et al., 2016](#); [Glasius
168 et al., 2018](#); [Vlachou et al., 2018](#); [Yttri et al., 2011b](#)~~). The protocol has been improved to increase sensitivity
169 and accuracy, by reducing the contribution of glucose in the blanks and by using an HPLC-PAD as the analytical
170 method for the determination of glucose concentrations. Trichoderma reesei cellulase (>700 u g⁻¹, Sigma Aldrich)
171 and Aspergillus Niger glucosidase (>750 u g⁻¹, Sigma Aldrich) have been used as saccharification enzymes. The
172 protocol is detailed in Section 2 of the SM.
173 Field blank filters (about 10 % of samples) were handled as real samples for quality assurance. The present data
174 have been corrected from field blanks. The reproducibility of the analysis of primary sugar compounds (polyols,
175 glucose) and cellulose, estimated from the analysis of sample extracts from 10 punches of the same filters were in
176 the range of 10-15 %. [The quantification limits primary sugar compounds and cellulose ranged from 0.63 to 0.89
177 ng m⁻³](#). About 2 800 samples are considered in this work for the polyols and glucose series, while 290 samples

178 (from the sites of Grenoble_LF and OPE-ANDRA) are considered for the cellulose series. Hereafter, the term
179 “Polyols” is used to refer uniquely to the sum of arabitol and mannitol concentrations.

180 **2.3 Meteorological data and LAI measurements**

181 Ambient weather data were not available at all monitoring sites (see Table S1). In this study, data including daily
182 relative humidity (%), night-time temperature (°C), average and maximum temperatures (°C), wind speed (m s⁻¹),
183 solar radiation (W m⁻²), and rainfall level (mm) for the sites of Marnaz and OPE-ANDRA (Figure 1), representing
184 different climatic regions and environmental conditions, were obtained from the French meteorological data
185 sharing service system (Météo-France) and ANDRA (French national radioprotective agency, in charge of the
186 OPE-ANDRA site), respectively.

187 The leaf area index (LAI), which is defined as the projected area of leaves over a unit of land, is an important
188 measure of the local vegetation density variation (Heald and Spracklen, 2009; Yan et al., 2016a, 2016b) (Heald
189 and Spracklen, 2009; Yan et al., 2016a, 2016b). For this study, we used the MODIS Collection 6 LAI product
190 because it is considered to have the highest quality among all the MODIS LAI products (Yan et al., 2016a,
191 2016b) (Yan et al., 2016a, 2016b). The MCD15A3H product uses both Terra and Aqua reflectance observations
192 as inputs to estimate daily LAI at 500 m spatial resolution, and a 4-day composite is calculated to reduce the noise
193 from abiotic factors. Using a 2 × 2 km grid box around the monitoring site, the local vegetation density variation
194 was retrieved from LP DAAC (<https://lpdaac.usgs.gov/>, last accessed: 15 March 2019) for the sites of Marnaz,
195 OPE-ANDRA, and Grenoble_LF.

196 **2.4 Data analyses**

197 All the statistical analyses were carried out using the open-source R software (R studio interface, version 3.4.1).
198 Several statistical analyses were performed on the concentrations to identify the spatial patterns of emission
199 sources and the potential parameters of influence as explained below.

200 The normalized cross-correlation (NCC) test was chosen to examine the potential similarities among the
201 monitoring sites for particulate SC concentrations, in terms of short-term temporal trends (e.g. synchronized
202 periods of increase or decrease, simultaneous fluctuations during specific episodes). The main advantage of NCC
203 over the traditional correlation tests is that it is less sensitive to linear changes in the amplitudes of the two-time
204 series compared. Therefore, to reduce the possibility of spurious “anti-correlation” due to highly variable
205 concentration ranges, data were amplitude-normalized prior to correlation analysis. A thorough discussion on the
206 normalized cross-correlation method can be found elsewhere (Bardal and Sætran, 2016; Dai and Zhou, 2017;
207 Eisner et al., 2009; Kaso, 2018; Lainer et al., 2016; Le Pichon et al., 2019) (Kaso, 2018; Yoo and Han, 2009). To
208 achieve pair-wise correlation analysis between the sampling sites collected during the same periods, the original
209 raw daily measurements were processed as follows: starting on identical days for each pairs of sites, arrangement
210 on the original daily data into consecutive 3-day intervals (or 6-day intervals in the case of OPE-ANDRA) and
211 calculation of the average concentration values for the middle-day were performed. The resultant data were used
212 for correlation analysis between site pairs (Table S3).

213 Multiple linear regression (MLR) was used to assess the strength of the relationships between atmospheric
214 concentrations of particulate SC and local environmental factors including the daily mean relative humidity, night-
215 time temperature, average and maximum temperature, wind speed, solar radiation, rain levels and LAI. Because
216 the LAI is a 4-day composite, daily values of the other variables were re-scaled into consecutive 4-day averaged
217 values. The linear regression (linear model or lm) package in R was employed for multiple regression analyses.

218 The concentration data were log-transformed to obtain regression residual distributions as close as possible to the
219 normal Gaussian one (Fig. ~~ure~~ S1). Stepwise forward selection was used to select the predictors that explain well
220 the temporal variation of SC concentrations at the site of Marnaz.

221 It should be noted that due to the limited availability of external parameters, the environmental factors driving SC
222 atmospheric levels have been extensively investigated for only two monitoring sites with contrasted
223 characteristics: the urban background site of Marnaz located in an Alpine valley, and the rural OPE-ANDRA site
224 surrounded by field crop areas spreading over several tens of km.

225

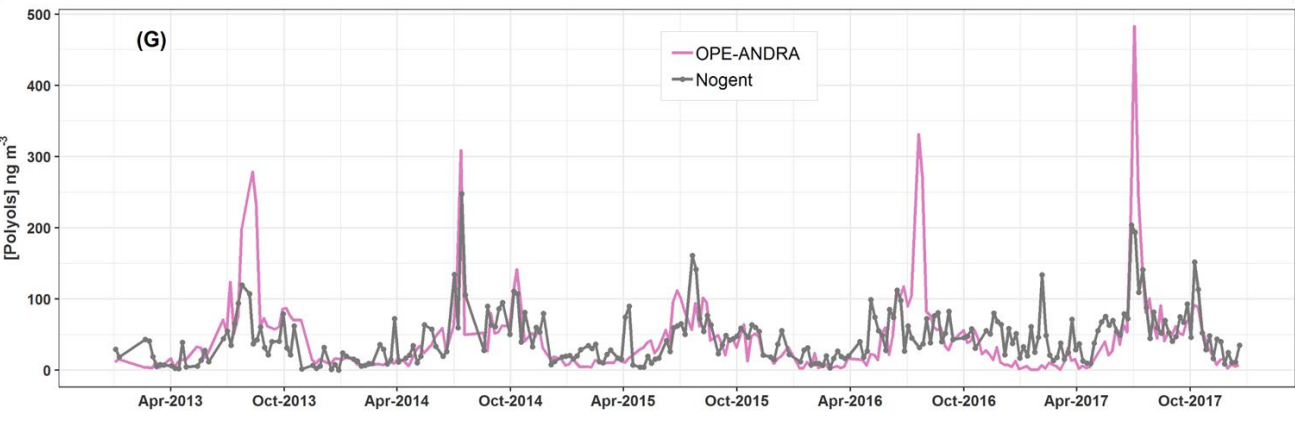
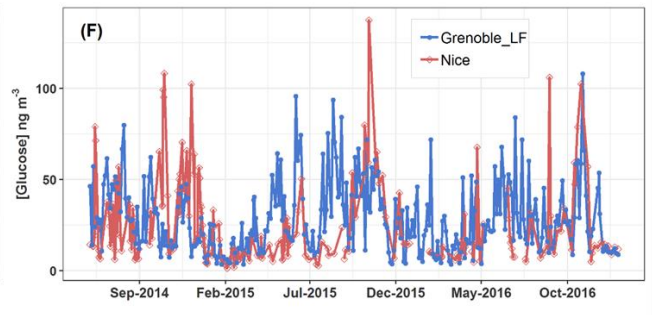
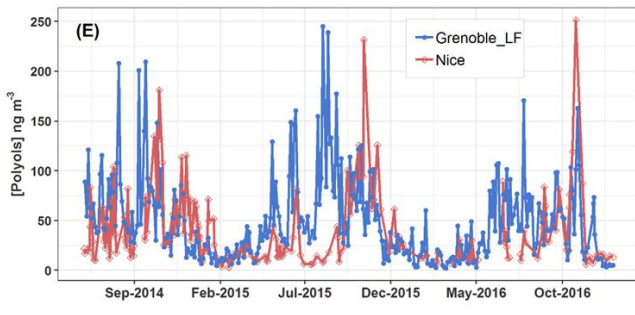
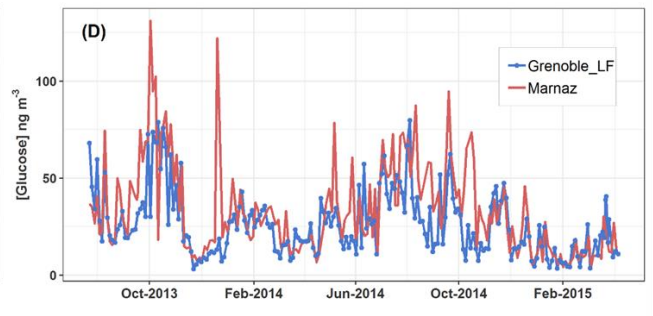
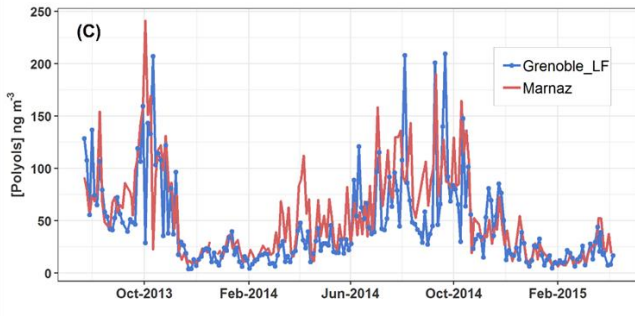
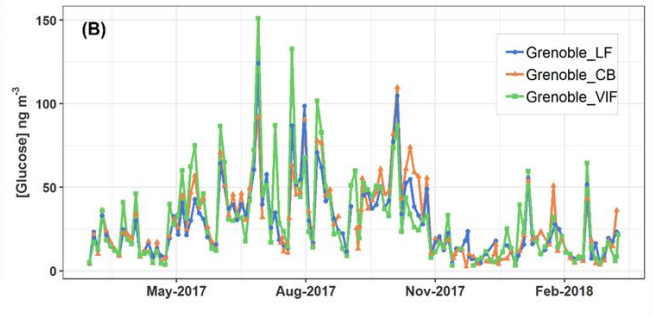
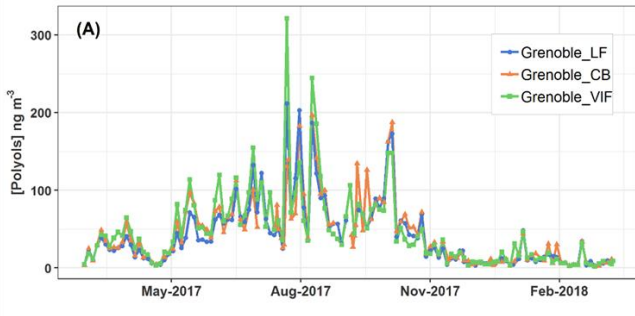
226 **3. Results and discussion**

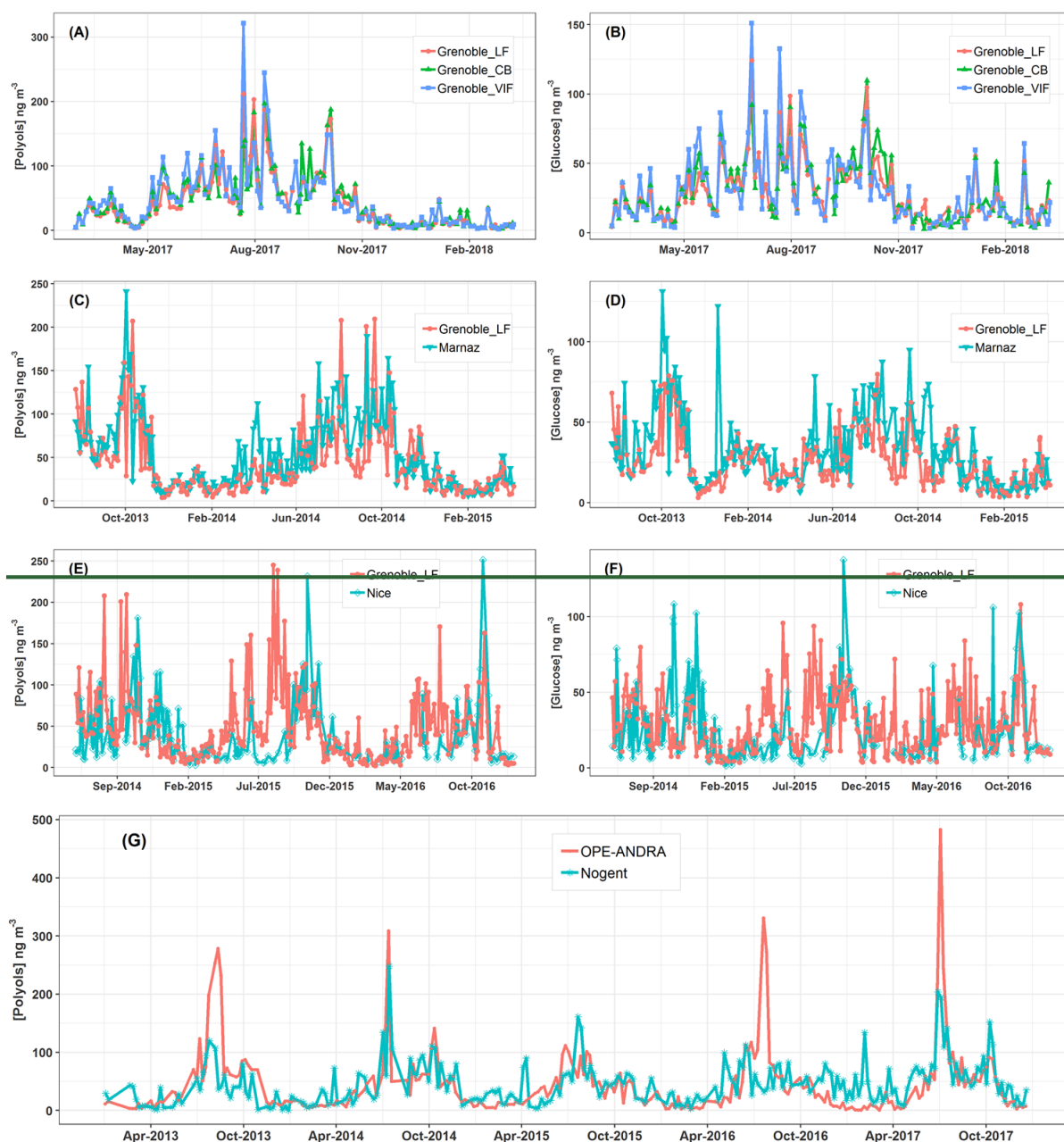
227 **3.1 Example of spatial coherence of the concentrations at different scales**

228 Our previous work ([Samaké et al., 2019](#))(~~Samaké et al., 2019~~) showed that particulate polyols and glucose are
229 ubiquitous primary compounds with non-random spatial and seasonal variation patterns over France. Here, an
230 inter-site comparison of their short-term concentration evolutions has been carried out at different space scales
231 (from local to national) for the pairs that can be investigated in our data base. Figure 2 presents some of these
232 comparisons for 3 spatial scales (15, 120, and 205 km).

233 The daily average concentrations of polyols (defined as sum of arabitol and mannitol) and glucose display highly
234 synchronous evolutionary trends (i.e., homogeneity in the concentrations, the timing of concentration peaks,
235 simultaneity of the daily specific episodes of increase/decrease of concentrations) over 3 neighboring monitoring
236 sites located 15 km apart in the Grenoble area (Figures 2A and B). Interestingly, remarkable synchronous patterns
237 both for short term (near-daily) and longer term (seasonal) still occur for sites located 120 km apart, as exemplified
238 for 2 sites in Alpine environments (Grenoble and Marnaz) (Figures 2C and D). However, as shown in Figures 2E
239 and F, the evolutions of concentrations become quite dissimilar and asynchronous in terms of seasonal and daily
240 fluctuations for more distant sites (Grenoble and Nice, 205 km apart), that are located in different climatic regions
241 (Alpine for Grenoble, Mediterranean for Nice). This is contrasting with results from the rural background site of
242 OPE-ANDRA and the suburban site of Nogent-sur-Oise, both located in a large field crop region of extensive
243 agriculture, and about 230 km apart from each other (Fig. ~~ure~~ 2G). Indeed, they present very similar variations of
244 daily concentrations for multi-year series, despite their distance apart, with concentration peaks generally more
245 pronounced at the rural site of OPE-ANDRA.

246 The following sections are dedicated to the investigation of the processes that can lead to these similarities and
247 differences according to these spatial scales.

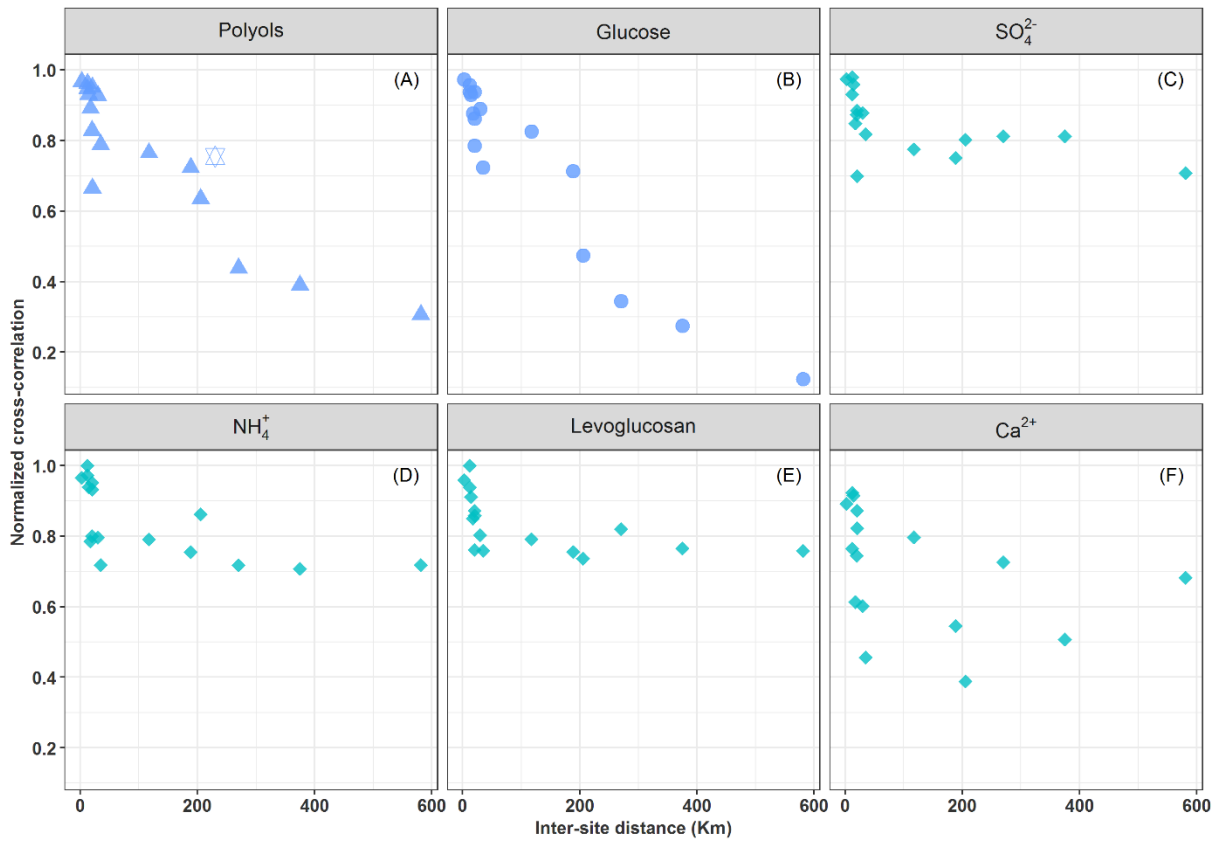




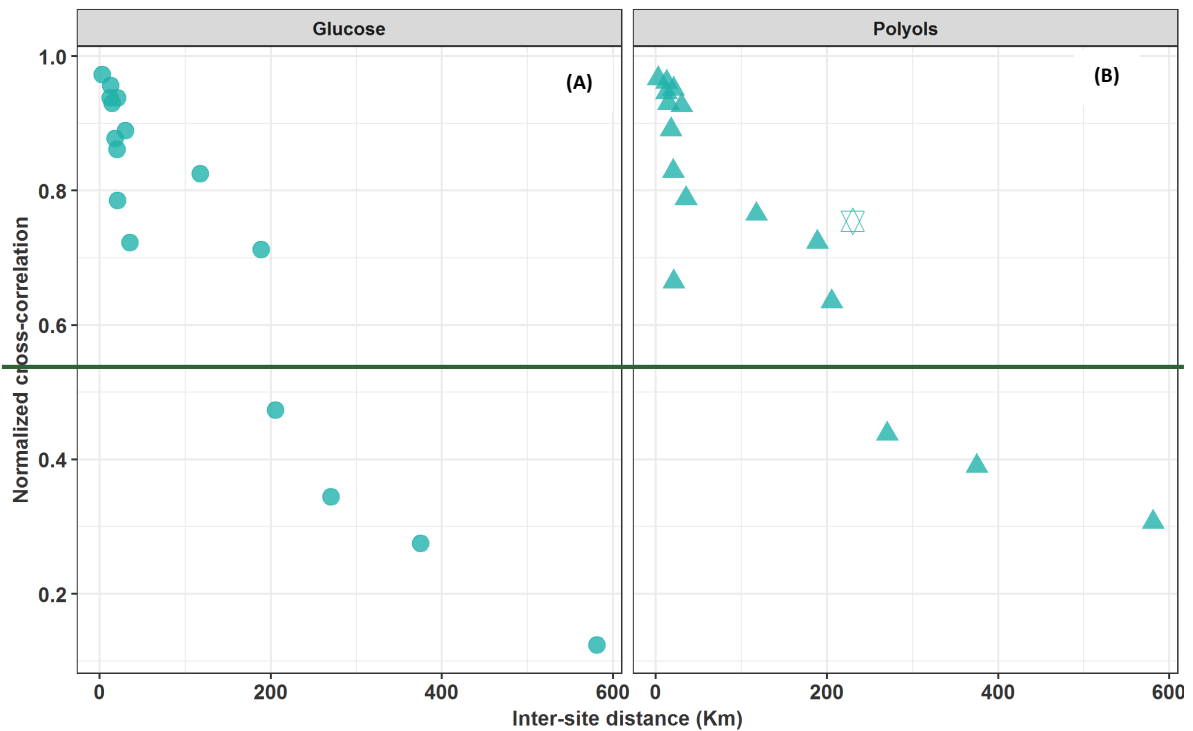
249
 250 **Figure 4: Concentrations (in ng m^{-3}) of (left) ambient particulate polyols (defined as the sum of arabitol and mannitol)**
 251 **and glucose (right) over different monitoring sites in France. Since PM_{10} were collected every 3-days at Nogent-sur-Oise**
 252 **and 6-days at OPE-ANDRA, the original data sets are averaged over consecutive 6-day intervals (bottom graph).**

253 3.2 Inter-site correlations and spatial scale variability

254 Figures 3A and 3B provide an overview of the cross-correlation coefficients for the daily evolution of
 255 concentrations (for polyols glucose and glucose polyols (SC)) between pairs of sites located at multiple increasing
 256 space scales across France (Table S3). Time series of concentrations for both SC show a clear distance-dependent
 257 correlation. The strength of the correlations is highly significant for distances up to 150-190 km ($R > 0.72$,
 258 $p < 0.01$) and gradually decreases with increasing inter-site distances. One exception is the pair OPE-ANDRA and
 259 Nogent-sur-Oise (high correlation for a distance above 230 km), both sites being located in highly-impacted
 260 agricultural areas. This overall pattern suggests that the processes responsible for the atmospheric concentrations
 261 of SC present a spatial homogeneity over typical areas of at least several tens of km.



262



263

264 **Figure 5: Normalized cross-correlation values for the daily evolution of particulate glucose-polyols (A), glucose-and
 265 polyols (B), sulfate (C), ammonium (D), levoglucosan (E) and calcium (F) concentrations over pairs of sites located at
 266 multiple increasing space scales across France. The hexagram corresponds to the correlation between the sites of OPE-
 267 ANDRA and Nogent-sur-Oise, both sites being surrounded by crop field areas.**

268 Unlike SC, ambient air concentrations of sulfate ([Fig. 3C](#)) and ammonium ([Fig. 3D](#)), associated with long-range
 269 aerosol transport ([Abdalmogith and Harrison, 2005](#); [Amato et al., 2016](#); [Coulibaly et al., 2015](#); [Pindado and Perez,](#)

270 [2011; Waked et al., 2014](#))~~(Abdalmogith and Harrison, 2005; Amato et al., 2016; Coulibaly et al., 2015;~~
271 ~~Pindado and Perez, 2011; Waked et al., 2014)~~ and levoglucosan ([Fig. 3E](#)), associated with biomass burning
272 in cold season ([Weber et al., 2019; Xiao et al., 2018](#)), display stronger positive correlations ($R > 0.72-0.98$,
273 $p < 0.01$) at all pairs of sites considered in the present work (~~Figure S2~~). Moreover, ambient concentrations of
274 calcium ([Fig. 3F](#)), associated with local fugitive dust sources or/and long-range aerosol transport ([Ram et al., 2010;](#)
275 [Wan et al., 2019](#)) display random correlation patterns (~~Figure S2~~). These results are in agreement with [Zhu et al.](#)
276 (2018) who also reported non-significant correlations between SC and sulfate in PM_{2.5} aerosols measured at
277 Shanghai, China. The distinct spatial behaviors between sulfate (or Ca²⁺) and SC in the present work further
278 suggest a dominant regional influence for atmospheric SC, as opposed to processes associated with either local
279 sources for calcium or long-range transport for sulfate.

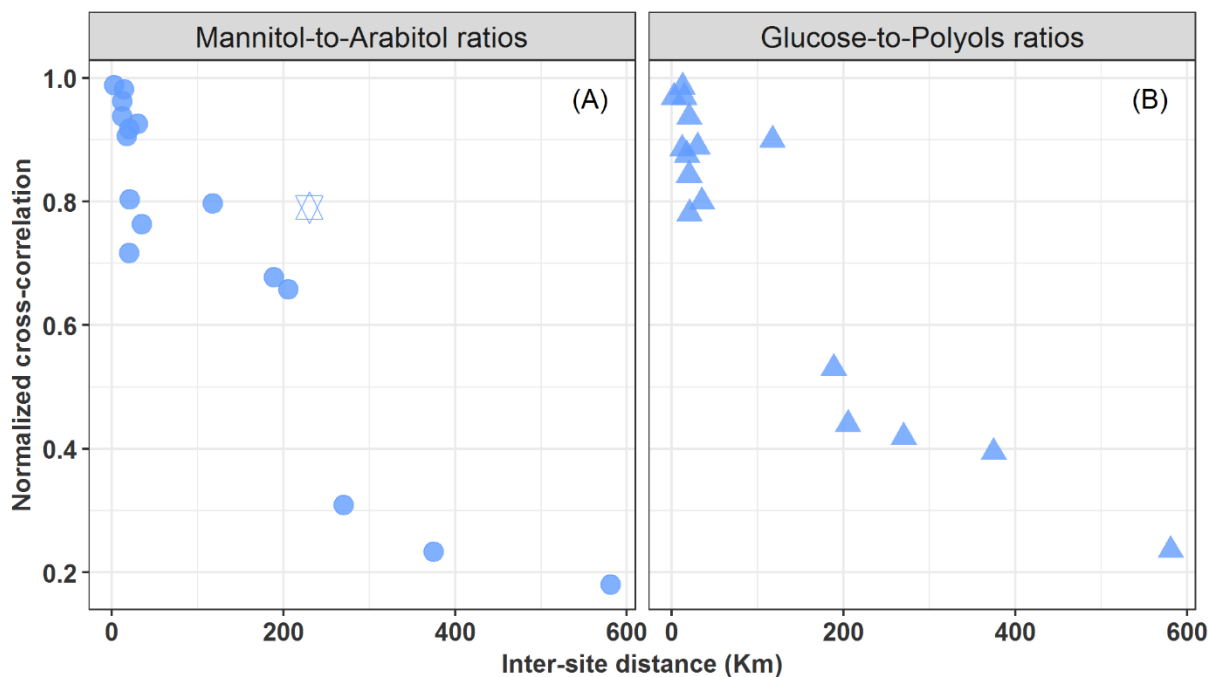
280 Mannitol and arabitol are well-known materials of fungal spores, serving as osmo-regulatory solutes ([Medeiros et](#)
281 ~~al., 2006b; Simoneit et al., 2004; Verma et al., 2018; Xiao et al., 2018; Zhang et al., 2015~~)~~(Medeiros et al., 2006b;~~
282 ~~Simoneit et al., 2004b; Verma et al., 2018; Zhang et al., 2010, 2015)~~. Based on parallel measurements of spore
283 counts and PM₁₀ polyol concentrations at three sites within the area of Vienna (Austria), [Bauer et al. \(2008a\)](#) found
284 an average arabitol and mannitol content per fungal spores of respectively 1.2 pg spore⁻¹ (range 0.8-1.8 pg spore⁻¹)
285 and 1.7 pg spore⁻¹ (range 1.2-2.4 pg spore⁻¹). Mannitol and arabitol have also been often identified in the green
286 algae and lower plants ([Buiarelli et al., 2013; Di Filippo et al., 2013; Gosselin et al., 2016; Véléz et al., 2007; Xu](#)
287 ~~et al., 2018; Zhang et al., 2010~~)~~(Buiarelli et al., 2013; Di Filippo et al., 2013; Véléz et al., 2007; Xu et al., 2018;~~
288 ~~Zhang et al., 2010)~~. [Gosselin et al., 2016](#) observed a relatively low ($R^2 = 0.31$) to high ($R^2 = 0.84$) coefficient of
289 determination between mannitol and arabitol for total suspended particles (TSP) collected at a pine-forested area
290 during dry and rainy periods, respectively. High correlation in rainy periods possibly suggested that both chemical
291 species in the TSP fraction in this pine-forested area could have been derived mainly from the same sources, i.e.,
292 actively wet-discharged ascospores and basidiospores, while the relatively poor correlation in dry periods could
293 have been likely due to more complex sources, i.e., dry discharged spores, plants, algae, etc. Being important
294 chemical species for the metabolism of these microorganisms ([Shcherbakova, 2007](#)), it may well be that the
295 concentration ratio of mannitol-to-arabitol could deliver some information on the spatial or temporal evolution of
296 their emission processes ([Gosselin et al., 2016](#))~~(Gosselin et al., 2016)~~. The annual average mannitol-to-arabitol
297 ratio at all sites is about 1.15 ± 0.59 , with ratios for the warm period (Jun-Sept) being 1 to 2 times higher than
298 those in the cold period (Dec-May) (Table S1). These ratios are within the range of those previously reported for
299 PM₁₀ aerosols collected at various urban and rural background sites in Europe ([Bauer et al., 2008; Yttri et al.,](#)
300 ~~2011b~~)~~(Bauer et al., 2008a; Yttri et al., 2011b)~~. Similarly, [Burshtein et al., \(2011\)](#) also reported comparable ratios
301 for PM₁₀ aerosols collected during autumn and winter from a Mediterranean region in Israel.

302 Similarly, the annual average glucose-to-polyols ratio at all sites is about 0.79 ± 0.77 . No literature data are
303 currently available for comparison. Further work is needed to relate these variations with microorganism
304 communities and plant growing stages.

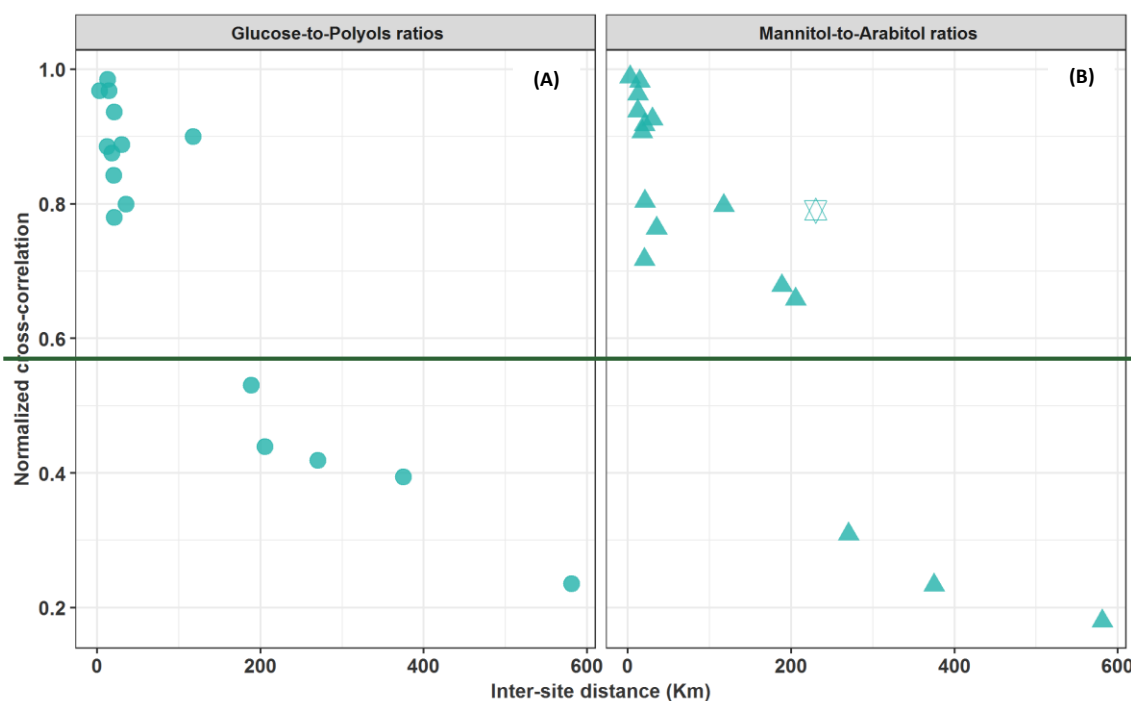
305 However, as evidenced in [Fig. 4](#), both mannitol-to-arabitol and glucose-to-polyols ratios show a clear distance-
306 dependent correlation, with higher correlations ($R = 0.64$ to 0.98 , $p < 0.01$) observed for pairs of sites within 150-
307 190 km distance. This spatial consistency highlights once again that the dominant emission processes should be
308 effective regionally, rather than being specific local input processes, and that atmospheric dynamics of the
309 concentration levels (i.e., driven by the interplay of emission and removal processes) are determined by quite

310 similar environmental factors (e.g. meteorological conditions, vegetation, land use, etc.) at such a regional scale.
311 This implies that local events and phenomena, such as the mechanical resuspension of topsoil and associated biota
312 (like bacteria, fungi, plant materials, etc.) might not be their major atmospheric input processes, particularly in
313 urban background areas typically characterized by less bare soil, and with a variable nature of the unpaved topsoil
314 at the regional scale ([Karimi et al., 2018](#))~~(Karimi et al., 2018)~~. Furthermore, [Karimi et al. \(2018\)](#)~~Karimi et al.~~
315 ~~(2018)~~ also recently reported heterogeneous topsoil microbial structure within patches of 43 to 260 km across
316 different regions of France. It follows that the hypotheses of emissions related to mechanical resuspension of
317 topsoil particles and associated biota, or microbiota emitted actively from surface soil into the air generally
318 assumed in most pioneering reports ([Medeiros et al., 2006b](#); [Rogge et al., 2007](#); [Simoneit et al., 2004](#); [Wan and](#)
319 [Yu, 2007](#))~~(Medeiros et al., 2006b; Rogge et al., 2007; Simoneit et al., 2004b; Wan and Yu, 2007)~~ are most
320 probably not valid.

321 Alternatively, the vegetation leaves have also been suggested as sources of atmospheric SC ([Bozzetti et al., 2016](#);
322 [Golly et al., 2018](#); [Jia et al., 2010](#); [Myriokefalitakis et al., 2017](#); [Pashynska et al., 2002](#); [Sullivan et al., 2011](#);
323 [Verma et al., 2018](#); [Wan et al., 2019](#))~~(Golly et al., 2018; Jia and Fraser, 2011; Pashynska et al., 2002; Sullivan~~
324 ~~et al., 2011; Verma et al., 2018; Wan et al., 2019)~~. In fact, vascular plant leaf surfaces is an important habitat
325 for endophytic and epiphytic microbial communities ([Kembel and Mueller, 2014](#); [Lindow and Brandl, 2003](#);
326 [Lymeropoulou et al., 2016](#); [Mhuireach et al., 2016](#); [Whipps et al., 2008](#))~~(Kembel and Mueller, 2014; Lindow~~
327 ~~and Brandl, 2003; Whipps et al., 2008)~~. Our results are more in agreement with a dominant atmosphere
328 entrance process closely linked to vegetation, which is more homogeneous than topsoil at the climatic regional
329 scale. Consistent with this, [Sullivan et al. \(2011\)](#)~~Sullivan et al. (2011)~~ also observed evident distinct regional
330 patterns for daily PM_{2.5} polyols and glucose concentrations at ten urban and rural sites located in the upper Midwest
331 (USA). The authors attributed such a spatial pattern to the differences in vegetation types and microbial diversity
332 over distinct geographical regions. Accordingly, the vegetation structure and composition have been previously
333 shown to play essential roles on airborne microbial variabilities in nearby areas ([Bowers et al., 2011](#); [Laforest-](#)
334 [Lapointe et al., 2017](#); [Lymeropoulou et al., 2016](#); [Mhuireach et al., 2016](#))~~(Bowers et al., 2011;~~
335 ~~Lymeropoulou et al., 2016; Mhuireach et al., 2016)~~.



336



337

338 **Figure 6: Normalized cross-correlation values for daily evolution of particulate mannitol-to-arabitol (A) glucose-to-**
 339 **polyols (B) and mannitol-to-arabitol (B)-ratios over pairs of sites located at multiple increasing space scales across**
 340 **France. The hexagram corresponds to the correlation between the sites of OPE-ANDRA and Nogent-sur-Oise, both**
 341 **sites being surrounded by crop field areas.**

342 **3.3 Influence of the vegetation on polyols and glucose concentrations**

343 The relationships between SC PM₁₀ concentrations and vegetation (plant materials) can be examined at the site of
 344 Grenoble Les Frênes (Grenoble_LF) by comparing the annual evolutions of SC and the free atmospheric cellulose
 345 concentrations, together with LAI ones.

346 The daily ambient concentration levels of SC and cellulose range respectively from 5.0 to 301.9 ng m⁻³ (with an
 347 average of 41.2 ± 39.9 ng m⁻³) and 0.7 to 207.2 ng m⁻³ (with an average of 52.9 ± 44.2 ng m⁻³), which corresponds

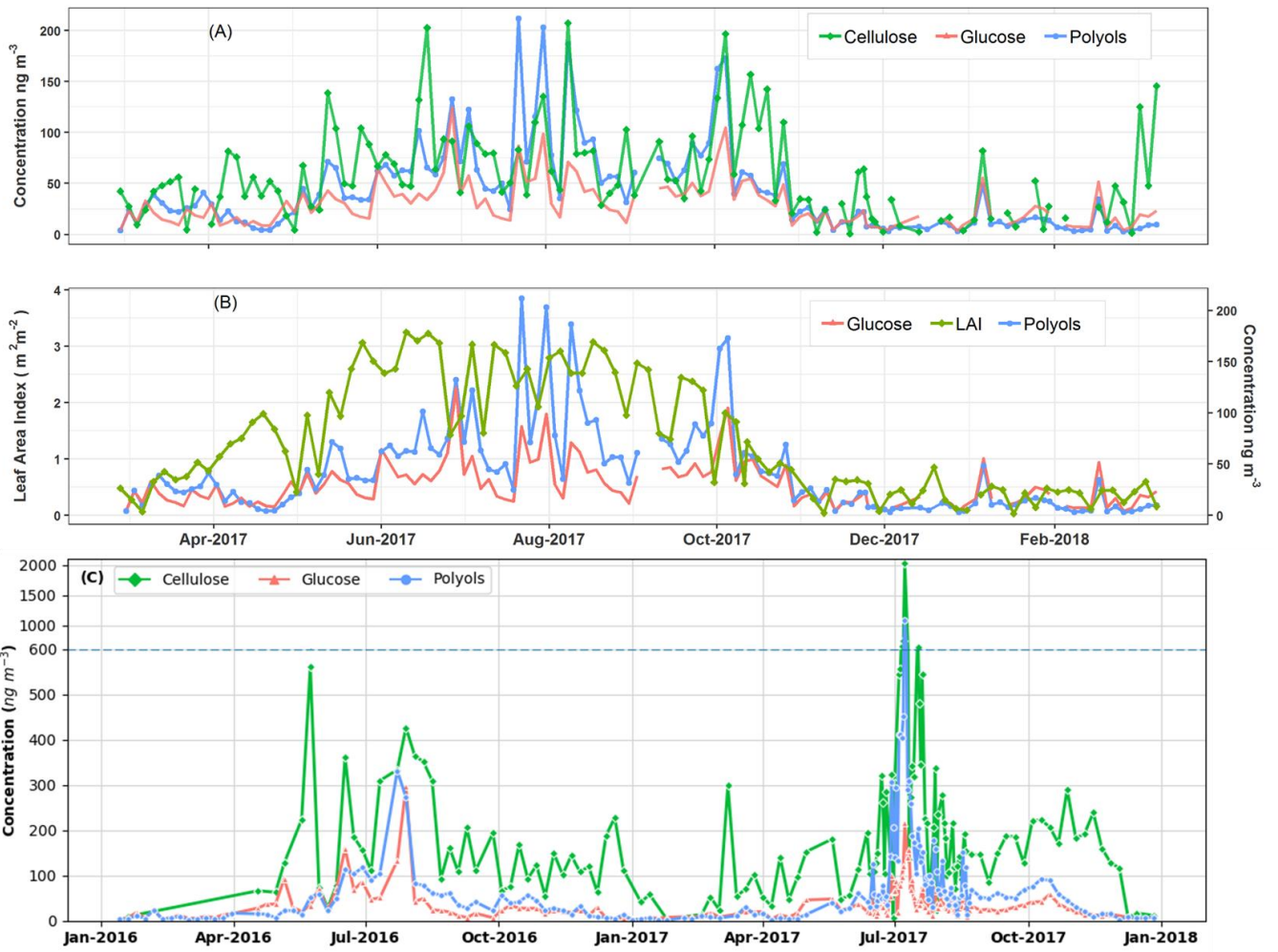
348 to respectively to 0.1 to 6.6 % and 0.01 to 5.3 % of total organic matter (OM) mass in PM₁₀. These values are
349 comparable to those previously reported for various sites in Europe ([Daellenbach et al., 2017](#); [Sánchez-Ochoa et
350 al., 2007](#); [Vlachou et al., 2018](#); [Yttri et al., 2011b](#)) (~~[Daellenbach et al., 2017](#); [Sánchez-Ochoa et al., 2007](#); [Vlachou
351 et al., 2018](#); [Yttri et al., 2011b](#)~~). Thus, a major part of PBOA could possibly be ascribed cellulose and SC derived
352 sources.

353 As evidenced in ~~Fig. 5A~~ 5A, ambient free cellulose concentrations vary seasonally, with maximum seasonal
354 average values observed in summer ($81.4 \pm 47.6 \text{ ng m}^{-3}$) and autumn ($64.2 \pm 49.2 \text{ ng m}^{-3}$), followed by spring
355 ($52.6 \pm 37.8 \text{ ng m}^{-3}$), and lower levels in winter ($23.0 \pm 19.9 \text{ ng m}^{-3}$). This is the same global pattern for polyols,
356 that are also more abundant in summer ($82.4 \pm 47.4 \text{ ng m}^{-3}$) and autumn ($48.7 \pm 41.6 \text{ ng m}^{-3}$), followed by spring
357 ($24.9 \pm 16.3 \text{ ng m}^{-3}$), and winter ($10.2 \pm 9.6 \text{ ng m}^{-3}$) in the Grenoble area. On a daily scale, the episodic increases
358 or decreases of polyols in PM₁₀ are very often well synchronized with that of cellulose (~~Fig. 5A~~ 5A). Moreover,
359 the maximum atmospheric concentrations of polyols also mainly occur when the vegetation density (LAI) is at its
360 highest in late summer (~~Fig. 5B~~ 5B). Similar global behaviors are also observed for atmospheric particulate glucose
361 and LAI (Figs. 5A and B). To further assess the relationships between SC PM₁₀ concentrations and vegetation at
362 a rural area, a two-year measurement of cellulose concentrations at the highly-impacted agricultural rural site of
363 OPE-ANDRA has been conducted. The average concentration of cellulose at OPE-ANDRA ($197.9 \pm 217.8 \text{ ng m}^{-3}$)
364 is 3.5 times higher than that measured in the urban area of Grenoble. In terms of temporal dynamics, the
365 evolution cycles (i.e., peaks and decreases) of both polyols and glucose are also very often well synchronized with
366 that of cellulose at OPE-ANDRA (Fig. 5C).

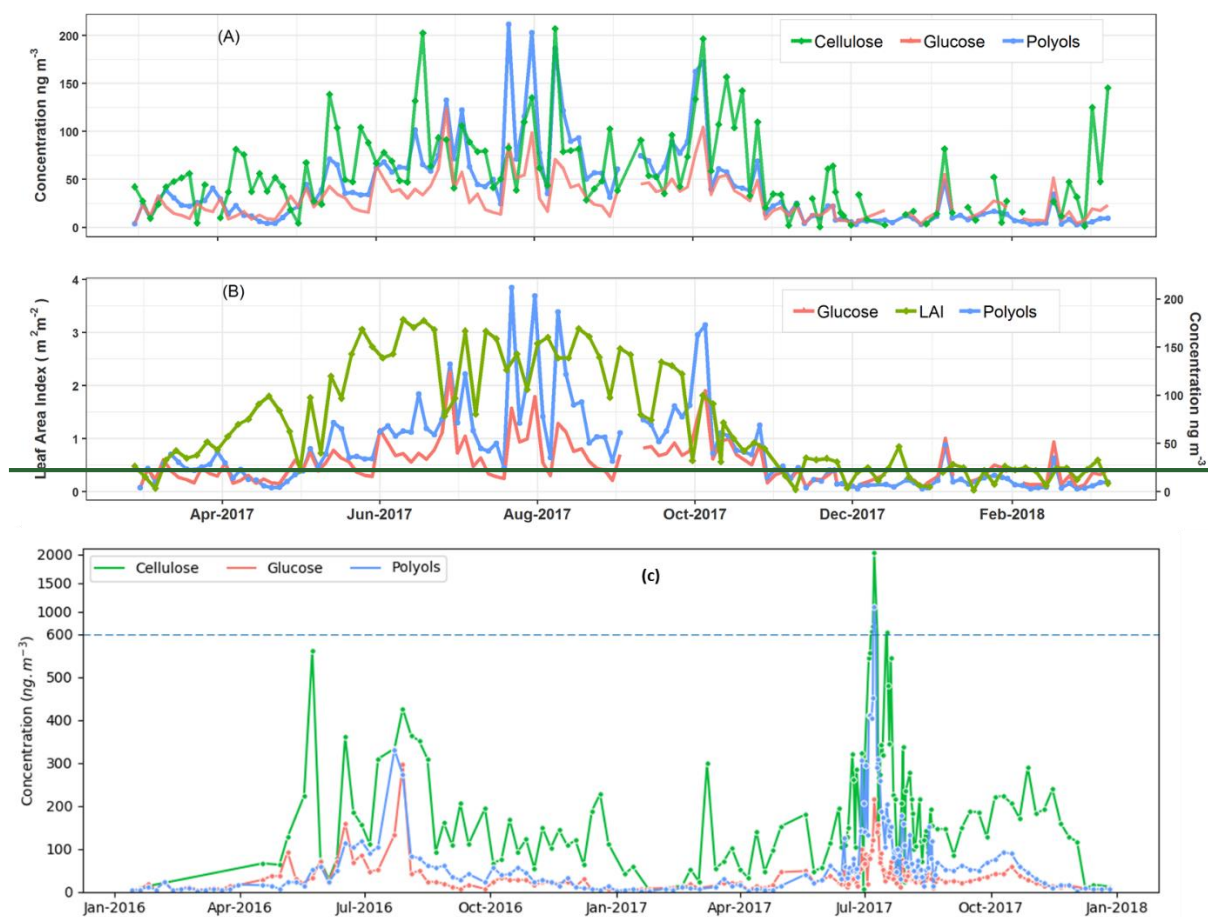
367 Altogether, ~~these findings highlight that particulate SC PM₁₀ and cellulose in both urban background and rural agricultural
368 areas~~ these findings highlight that SC in PM₁₀ and cellulose in both urban background and rural agricultural
369 ~~agricultural areas~~ most probably share a common source related to the vegetation. This is an additional evidence
370 in support of the hypothesis suggested in previous studies ([Bozzetti et al., 2016](#); [Burshtein et al., 2011](#); [Daellenbach
371 et al., 2017](#); [Pashynska et al., 2002](#); [Verma et al., 2018](#); [Vlachou et al., 2018](#); [Wan and Yu, 2007](#); [Yttri et al.,
372 2007](#)) (~~[Bozzetti et al., 2016](#); [Burshtein et al., 2011](#); [Daellenbach et al., 2017](#); [Pashynska et al., 2002](#);
373 [Verma et al., 2018](#); [Vlachou et al., 2018](#); [Yttri et al., 2007](#)~~). It is also in line with studies indicating that the
374 PBOA source profile identified using offline aerosol mass spectrometry (offline-AMS) correlates very well with
375 coarse cellulose concentrations ([Bozzetti et al., 2016](#); [Vlachou et al., 2018](#)). Noticeable contribution of cellulose
376 to PBOA mass (26 %) at the rural background site of Payerne (Switzerland), during summer 2012 and winter 2013,
377 was reported by [Bozzetti et al. \(2016\)](#) (~~[Bozzetti et al., 2016](#)~~).

378 As also evidenced in ~~Fig. 5~~ 5, the cellulose concentration peaks are not systematically correlated to those of
379 polyols. The development stage of the plants (developing or mature leaves, flowering plants) in addition to the
380 metabolic activities of endophytic and epiphytic biota (growth, sporulation), all closely related to meteorological
381 conditions ([Bodenhausen et al., 2014](#); [Bringel and Couée, 2015](#); [Lindow and Brandl, 2003](#); [Pirttilä and Frank,
382 2011](#); [Reddy et al., 2017](#)) (~~[Bodenhausen et al., 2014](#); [Bringel and Couée, 2015](#); [Lindow and Brandl, 2003](#); [Morieca
383 and Ragazzi, 2011](#); [Reddy et al., 2017](#)~~), could explain such observations. The influence of local meteorological
384 conditions for an urban Alp valley site is discussed in Section 3.4. Consistent with our observations, previous
385 studies conducted at various urban background sites in Europe have suggested that particulate polyols are
386 associated to mature plant leaves and microorganisms (bacterial and fungal spores) while glucose, which is a
387 monomer of cellulose, would most likely be linked to the developing leaves ([Bozzetti et al., 2016](#); [Burshtein et al.,](#)

388 [2011; Pashynska et al., 2002; Yttri et al., 2007; Zhu et al., 2015](#) ([Bozzetti et al., 2016; Burshtein et al., 2011;](#)
389 [Pashynska et al., 2002; Yttri et al., 2007](#)).



390



391
 392 **Figure 7: Temporal covariation cycles of the daily particulate polyols and glucose concentrations along with vegetation**
 393 **indicators at the urban background site of Grenoble (A and B) and the rural agricultural background site of OPE-**
 394 **ANDRA (C), respectively. Note that PM₁₀ aerosols are intensively collected at OPE-ANDRA every day (24-h) from 12**
 395 **June 2017 to 22 August 2017, and that the concentration scale is changing above 600 ng m⁻³ in Figure C, due to extreme**
 396 **concentration peak in July 2017.**

397 **3.4 Influence of meteorological parameters on ambient concentrations of polyols and glucose**

398 We used here a multiple linear regression analysis (MLR) approach to gain further insight about the environmental
 399 factors influencing the annual and short time variation cycles of atmospheric SC concentrations. This tentative
 400 MLR analysis is focused on the urban background site of Marnaz only since meteorological and other data are
 401 readily available for this site and are not influenced too much by some large city effects. Several variables were
 402 tested, that are already mentioned in the literature as drivers of SC concentrations. It includes the ambient relative
 403 humidity, rainfall level, wind speed, solar radiation, night-time temperature, average (or maximum) temperature,
 404 and LAI. Night-time temperature was selected since the time series in Marnaz and Grenoble indicate that the major
 405 drop of concentrations in late fall (Fig. 4.10 2C) is related to the first night of the season with night-time temperature
 406 below 5°C. The use of the night-temperature is also consistent with the bi-modal distribution of polyols during
 407 night and day time found in previous studies (Claeys et al., 2004; Graham et al., 2003; Yan et al., 2019; Yttri et
 408 al., 2011a)(Claeys et al., 2004; Graham et al., 2003).

409 Overall, the environmental factors including the mean night-time temperature, relative humidity, wind speed and
 410 the leaf area index explain up to 82 % (adjusted R² = 0.82, see Table 1) of the annual temporal variation cycles of
 411 SC concentrations. The mean night-time temperature and LAI contribute respectively to 54 % and 37 % of the
 412 observed annual variabilities of SC concentrations. The atmospheric humidity is also a driver for these chemical

413 species (3 % of the explained variation). These results are consistent with previous studies showing that
 414 concentrations of mannitol (in both PM₁₀ and PM_{2.5} size fractions) linearly correlate best with the LAI, atmospheric
 415 water vapor and temperature (Heald and Spracklen, 2009; Hummel et al., 2015; Myriokefalitakis et al.,
 416 2017)(Heald and Spracklen, 2009; Hummel et al., 2015). All of these drivers have been previously shown to
 417 induce the initial release and influence the long-term airborne microbial (i.e. bacteria, fungi) concentrations (China
 418 et al., 2016; Elbert et al., 2007; Grinn-Gofroń et al., 2019; Jones and Harrison, 2004; Rathnayake et al., 2017;
 419 Zhang et al., 2015).

420 Besides, the wind speed (range of 0.2 to 5.6 m s⁻¹) seems an additional effective driver affecting the contribution
 421 of the local vegetation to SC concentrations in the atmosphere. Albeit enough air movement is required to passively
 422 release microorganisms along with plant debris into the atmosphere, strong air motions induce higher dispersion.
 423 These observations are in good agreement with those previously reported (Jones and Harrison, 2004; Liang et al.,
 424 2013; Zhang et al., 2010, 2015; Zhu et al., 2018b)(Jones and Harrison, 2004; Liang et al., 2013; Zhang et al.,
 425 2010, 2015; Zhu et al., 2018). For instance Liang et al. (2013)Liang et al. (2013) have found a negative
 426 correlation between wind speed and polyols concentrations, and the highest atmospheric fungal spores
 427 concentrations were observed for a wind speed range of 0.6 to 1.0 m s⁻¹.

428 **Table 1: Multiple linear regression for ambient polyols and glucose concentrations and their effective environmental**
 429 **factors at the Marnaz site. Contributions of predictor are normalized to sum 1. “Relaimpo package under R” was**
 430 **used to compute bootstrap confidence intervals for importance of effective predictors (n=1000) (Grömping, 2006).**

	<i>Dependent variable</i>	<i>Variability explained by effective predictors</i>
	log(Polyols + Glucose)	
Night-time temperature (°C)	0.112*** (0.090, 0.133)	0.538 (0.453, 0.604)
Relative Humidity (%)	0.017*** (0.005, 0.030)	0.030 (0.018, 0.067)
Leaf Area Index	0.386** (0.034, 0.737)	0.372 (0.286, 0.444)
Wind speed (m s ⁻¹)	0.226 (-0.203, 0.655)	0.021 (0.015, 0.058)
Leaf Area Index × Wind Speed ^a	-0.596*** (-1.001, -0.191)	0.039 (0.014, 0.085)
Constant	2.023*** (0.787, 3.260)	
Observations	87	
R ²	0.837	
Adjusted R ²	0.824	
Residual Std. Error	0.297 (df = 81)	
F Statistic	66.677*** (df = 5; 81)	
Note	**p < 0.01; ***p < 0.001	^a stands for interaction between predictors

431
 432 One of the limitations of this study is that 4-day averaged observations do not allow to evaluate the driver
 433 contributions that might explain some short term events for which the influence of meteorological parameters such
 434 as rainfall or solar radiation could also be significant (Grinn-Gofroń et al., 2019; Heald and Spracklen, 2009; Jones
 435 and Harrison, 2004)(Grinn-Gofroń et al., 2019; Heald and Spracklen, 2009; Jones and Harrison, 2004). However,
 436 such simple parameterizations could be a first step in considering SC concentrations in CTM models, and further
 437 work is required in this direction in order to generate a robust parametrization of the emissions.

438 3.5 Specific case of a highly-impacted agricultural area

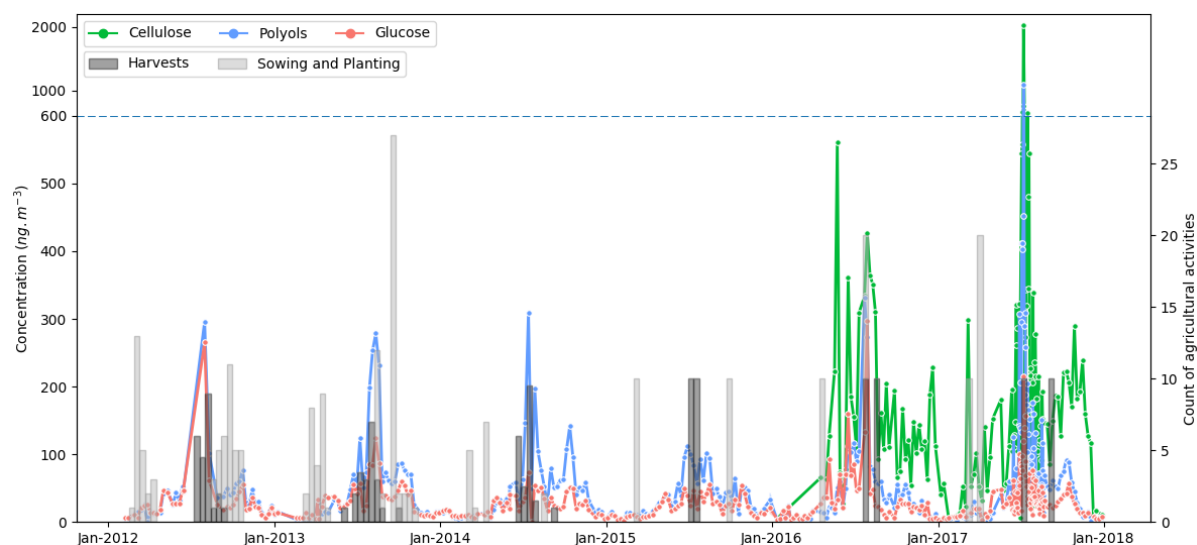
439 This section focuses on evidencing the environmental drivers of PM₁₀ SC concentrations specific to agricultural
 440 areas. To achieve this objective, the site of OPE-ANDRA has been selected because it is extensively impacted by
 441 agricultural activities, without being too prone to influences by other sources. OPE-ANDRA is a specific rural

442 background observatory located at about 230 km east of Paris at an altitude of ~~293~~²⁹³ m. It is characterized by a
 443 low population density (< 22 inhabitants km^{-2} within an area of 900 km^2), with no surrounding major transport
 444 road or industrial activities. The air monitoring site itself lies in a “reference sector” of 240 km^2 , in the middle of
 445 a field crop area (tens of kilometers in all directions). The daily agricultural practices within this reference sector
 446 are recorded and made available by ANDRA. The parcels within the agricultural area are submitted to a 3-year
 447 crop-rotation system. The major crops are wheat, barley, rape, pea and sunflower. Additionally, OPE-ANDRA is
 448 also characterized by a homogeneous type of soil, with a predominance of superficial clay-limestone.

449 Figure 6 shows the daily evolution of polyols concentrations in the PM_{10} fraction at OPE-ANDRA from 2012 to
 450 2018, together with the agricultural activities recorded daily and averaged over 12 days.

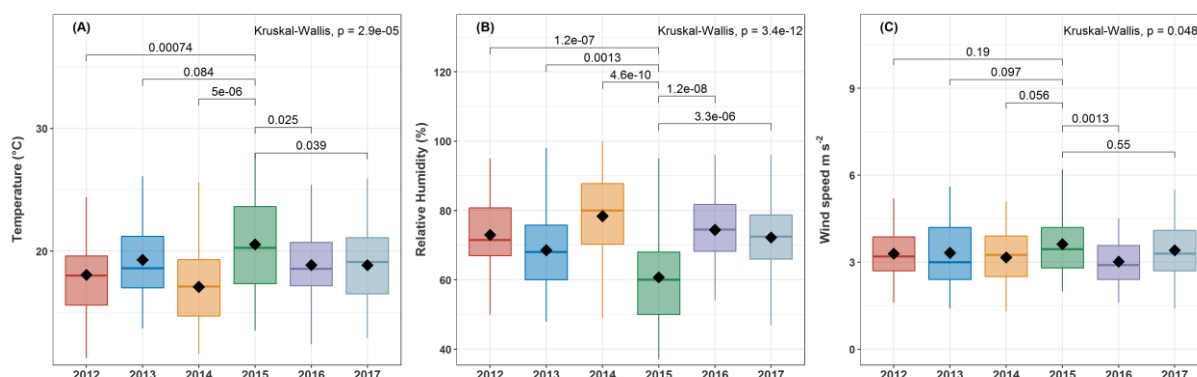
451 Although the concentration of polyols fluctuates from a year to another, they display clear annual variation cycles,
 452 with higher values in the warm periods (Jun. to Nov.) and lower concentration values in the cold periods (Oct. to
 453 May). Interestingly, the annual concentrations of polyols in 2015 ($4.2\text{-}111.7 \text{ ng m}^{-3}$; annual average:
 454 $37.0 \pm 29.1 \text{ ng m}^{-3}$) are significantly lower than those observed for the other years ($0.6\text{-}1084.6 \text{ ng m}^{-3}$; annual
 455 average: $62.9 \pm 96.8 \text{ ng m}^{-3}$). Similar inter-annual evolution trends, but with variable intensities, are also observed
 456 for glucose concentrations (Fig. ~~ure~~ 6). Year 2015 has been found to be particularly hot and dry at OPE-ANDRA
 457 (Fig. ~~ure~~ 7) whereas the local averaged wind conditions are quite stable over the years within the period of study,
 458 suggesting that the wind conditions are not the main driver of the observed inter-annual variability. These results
 459 highlight that ambient air temperature and humidity are key meteorological drivers of the annual variation cycles
 460 of polyols and glucose concentrations. Hot and dry ambient air conditions may decrease the metabolic activity of
 461 the microorganisms (e.g. microbial growth and sporulation) (Fang et al., 2018; Liang et al., 2013; Meisner et al.,
 462 2018)(Fang et al., 2018; Liang et al., 2013; Meisner et al., 2018).

463 Finally, maximum ambient concentration levels for both SC and cellulose are observed in excellent temporal
 464 agreement with the harvest periods (late summer) at the ~~OPE-ANDRA-OPE~~ site (Fig. ~~ure~~ 6). Harvesting activities
 465 have been previously reported as the major sources for particulate polyols and glucose to the atmosphere in
 466 agricultural and nearby urbanized areas (Golly et al., 2018; Rogge et al., 2007; Simoneit et al., 2004)(Golly et al.,
 467 2018; Rogge et al., 2007; Simoneit et al., 2004b). Hence, the resuspension of plant materials (crop detritus,
 468 leaves debris) and associated microbiota (e.g., bacteria, fungi) originating from cultivated lands are most-likely
 469 major input processes of PM_{10} polyols and glucose at field crop sites.



470

471 **Figure 8: Daily evolution cycles of polyols and glucose concentrations in aerosols collected from the OPE-ANDRA**
 472 **monitoring site, from 2012 to 2018. Cellulose concentrations have been measured from January 2016 to January 2018.**
 473 **Colored bars correspond to the sum of the various agricultural practices performed (data for 69 parcels are averaged**
 474 **over 12 days for better clarity). Records of agricultural activities after October 2014 were available for only two parcels**
 475 **within the immediate vicinity of the PM₁₀ sampler. Records are multiplied by 10 for this period.**



476
 477 **Figure 9: Boxplots of (A) maximum ambient temperature, (B) relative humidity and (C) wind speed at OPE-ANDRA**
 478 **from 2012 to 2017. Analyses are performed for warmer periods (June to November). Only statistically different**
 479 **meteorological factors are presented. The black marker inside each boxplot indicates the average value, while the top,**
 480 **middle and bottom of the box represent the 75th, median and 25th percentiles, respectively. The whiskers at the top and**
 481 **bottom of the box extend from the 95th to the 5th percentiles. Statistical differences between average values were assessed**
 482 **with the Kruskal-Wallis method ($p < 0.05$).**

483 4. Conclusions

484 The short-term temporal (daily) and spatial (local to nation-wide) evolutions of particulate polyols ([defined here](#)
 485 [as the sum of arabitol and mannitol](#)) and glucose concentrations are rarely discussed in the current literature. The
 486 present work aimed at investigating the spatial behavior of these chemicals and evidencing their major effective
 487 environmental drivers. The major results mainly showed that:

- 488 • The short-term evolution of ambient polyols and glucose concentrations is highly synchronous across an
 489 urban city-scale and remains very well correlated throughout the same geographic areas of France, even
 490 if the monitoring sites are situated in different cities at about 150-190 km. However, sampling sites
 491 located in two distinct geographic areas are poorly correlated. This indicates that emission sources of
 492 these chemicals are uniformly distributed, and their accumulation and removal processes are driven by
 493 quite similar environmental parameters at the regional scale. Therefore, local phenomena such as
 494 atmospheric resuspension of topsoil particles and associated microbiota, microbial direct emissions (e.g.
 495 sporulation), cannot be the main emission processes of particulate polyols and glucose in urban areas not
 496 directly influenced by agricultural activities.
- 497 • The atmospheric concentrations of polyols (or glucose) and cellulose display remarkably synchronous
 498 temporal evolution cycles at the background urban site of Grenoble, indicating a common source related
 499 to plant debris.
- 500 • Higher ambient concentrations of polyols and glucose at the rural site of OPE-ANDRA occur during each
 501 harvest period, pointing out resuspension processes of plant materials (crop detritus, leaves debris) and
 502 associated microbiota for agricultural and nearby urbanized areas. This is associated with higher PM₁₀
 503 cellulose concentration levels, as high as 0.4 to 2.0 $\mu\text{g}\cdot\text{m}^{-3}$ on a daily basis (accounting up to 7.5 to 32.4 %
 504 of the OM mass).

505 • Multiple linear regression analysis of the yearly series from the site of Marnaz gave insightful information
506 on which parameter controls the ambient concentrations of polyols and glucose. Ambient air night-time
507 temperature, relative humidity and vegetation density are the most important drivers, whilst wind speed
508 conditions tend to affect the contribution of local vegetation.

509 Altogether, these results improve our understanding of the spatial behavior tracers of PM₁₀ PBOA emission sources
510 in France, and in general, which is imperative for further implementation of this important mass fraction of OM
511 into chemical transport models. Further investigations of airborne microbial fingerprint (bacteria and fungi) are
512 ongoing, which may deepen our understanding of the PBOA source profile.

513 **Acknowledgements:** We would like to express special acknowledgements to Pierre Taberlet (LECA, Grenoble,
514 France) for fruitful discussions about the importance of endophytic and epiphytic biota for aerobiology. The PhD
515 of AS and SW are funded by the Government of Mali and ENS Paris, respectively. We gratefully acknowledge
516 the LEFE-CHAT and EC2CO programs of the CNRS for financial supports of the CAREMBIOS multidisciplinary
517 project, and the LEFE-CHAT program for the MECEA project for the development of the atmospheric cellulose
518 measurements. Samples were collected and analyzed in the frame of many different programs funded by ADEME,
519 Primequal, the French Ministry of Environment, the CARA program led by the French Reference Laboratory for
520 Air Quality Monitoring (LCSQA), ANDRA, and actions funded by many AASQA, IMT Lille Douai (especially
521 Labex CaPPA ANR-11-LABX-0005-01 and CPER CLIMIBIO projects). Analytical aspects were supported at
522 IGE by the Air-O-Sol platform within Labex OSUG@2020 (ANR10 LABX56). We acknowledge the work of
523 many engineers in the lab at IGE for the analyses (Aude Wack, Céline Charlet, Fany Donaz, Fany Masson, Sylvie
524 Ngo, Vincent Lucaire, Claire Vérin, and Anthony Vella). Finally, the authors would like to kindly thank the
525 dedicated efforts of many other people at the sampling sites and in the laboratories for collecting and analyzing
526 the samples.

527 **Author contributions:** JLJ was the (co-)supervisor for the PhD for AS, FC, SW, and for the post-doc of DS,
528 BG, and AW. He directed all the personnel who performed the analysis at IGE. He is the coordinator for the CNRS
529 LEFE-EC2CO CAREMBIOS program that is funding the work of AS. GU and JMF-M were the co-supervisor for
530 the PhD of AS or SW. EP, OF, and VR supervised the PhD of DMO who investigated the sites in northern France.
531 OF, JL-J, JL-B, AA and NM were coordinating and partners of the different initial programs for the collection and
532 chemical analysis of the samples. VJ developed the analytical techniques for polyols and cellulose measurements.
533 TC performed the cellulose measurements. Samples analyses at LSCE were performed by NB. AC gave advices
534 for the statistical aspects of the data processing. AS and JLJ processed the data and wrote up the manuscript. SW
535 participated to the visualization of the results. SC is supervising the OPE station and collected the agricultural
536 activities records. All authors from AASQA (author affiliation nos. 97 to 164) are representatives for each network
537 that conducted the sample collection and the general supervision of the sampling sites. All authors reviewed and
538 commented on the manuscript.

539 **Competing interests:** The authors declare that they have no conflict of interest.

540 References

541 [Abdalmogith, S. S. and Harrison, R. M.: The use of trajectory cluster analysis to examine the long-range transport](#)
542 [of secondary inorganic aerosol in the UK, Atmos. Environ., 39\(35\), 6686–6695,](#)
543 [doi:10.1016/j.atmosenv.2005.07.059, 2005.](#)

544 [Amato, F., Alastuey, A., Karanasiou, A., Lucarelli, F., Nava, S., Calzolari, G., Severi, M., Becagli, S., Gianelle, V.,](#)
545 [L., Colombi, C., Alves, C., Custódio, D., Nunes, T., Cerqueira, M., Pio, C., Eleftheriadis, K., Diapouli, E., Reche,](#)
546 [C., Minguillón, M. C., Manousakas, M.-I., Maggos, T., Vratolis, S., Harrison, R. M., and Querol, X.: Airuse-life+:](#)
547 [a harmonized PM speciation and source apportionment in five southern European cities, Atmos. Chem. Phys.,](#)
548 [16\(5\), 3289–3309, doi:10.5194/acp-16-3289-2016, 2016.](#)

549 [Amato, P., Brisebois, E., Draghi, M., Duchaine, C., Fröhlich-Nowoisky, J., Huffman, J. A., Mainelis, G., Robine,](#)
550 [E., and Thibaudon, M.: Main biological aerosols, specificities, abundance, and diversity, in Microbiology of](#)
551 [Aerosols, pp. 1–21, John Wiley & Sons, Ltd., doi:10.1002/9781119132318, 2017.](#)

552 [Ariya, P. A., Sun, J., Eltouny, N. A., Hudson, E. D., Hayes, C. T., and Kos, G.: Physical and chemical](#)
553 [characterization of bioaerosols – implications for nucleation processes, *Int. Rev. Phys. Chem.*, 28\(1\), 1–32,](#)
554 [doi:10.1080/01442350802597438, 2009.](#)

555 [Barbaro, E., Kirchgeorg, T., Zangrando, R., Vecchiato, M., Piazza, R., Barbante, C., and Gambaro, A.: Sugars in](#)
556 [Antarctic aerosol, *Atmos. Environ.*, 118, 135–144, doi:10.1016/j.atmosenv.2015.07.047, 2015.](#)

557 [Bardal, L. M. and Sætran, L. R.: Spatial correlation of atmospheric wind at scales relevant for large scale wind](#)
558 [turbines, *J. Phys. Conf. Ser.*, 753, 032033, doi:10.1088/1742-6596/753/3/032033, 2016.](#)

559 [Bauer, H., Claeys, M., Vermeylen, R., Schueller, E., Weinke, G., Berger, A., and Puxbaum, H.: Arabitol and](#)
560 [mannitol as tracers for the quantification of airborne fungal spores, *Atmos. Environ.*, 42\(3\), 588–593,](#)
561 [doi:10.1016/j.atmosenv.2007.10.013, 2008.](#)

562 [Bodenhausen, N., Bortfeld-Miller, M., Ackermann, M., and Vorholt, J. A.: A synthetic community approach](#)
563 [reveals plant genotypes affecting the phyllosphere microbiota, *PLoS Genet.*, 10\(4\), doi:](#)
564 [10.1371/journal.pgen.1004283, 2014.](#)

565 [Bowers, R. M., Sullivan, A. P., Costello, E. K., Collett, J. L., Knight, R., and Fiereri, N.: Sources of bacteria in](#)
566 [outdoor air across cities in the midwestern United States., *Appl. Environ. Microbiol.*, 77\(18\), 6350–6356,](#)
567 [doi:10.1128/AEM.05498-11, 2011.](#)

568 [Bozzetti, C., Daellenbach, K. R., Hueglin, C., Fermo, P., Sciare, J., Kasper-Giebl, A., Mazar, Y., Abbaszade, G.,](#)
569 [El Kazzi, M., Gonzalez, R., Shuster-Meiseles, T., Flasch, M., Wolf, R., Křepelová, A., Canonaco, F., Schnelle-](#)
570 [Kreis, J., Slowik, J. G., Zimmermann, R., Rudich, Y., Baltensperger, U., El Haddad, I., and Prévôt, A. S. H.: Size-](#)
571 [resolved identification, characterization, and quantification of primary biological organic aerosol at a European](#)
572 [rural site, *Environ. Sci. Technol.*, 50\(7\), 3425–3434, doi:10.1021/acs.est.5b05960, 2016.](#)

573 [Bringel, F. and Couée, I.: Pivotal roles of phyllosphere microorganisms at the interface between plant functioning](#)
574 [and atmospheric trace gas dynamics, *Front. Microbiol.*, 6, 486, doi:10.3389/fmicb.2015.00486, 2015.](#)

575 [Buiarelli, F., Canepari, S., Di Filippo, P., Perrino, C., Pomata, D., Riccardi, C., and Speziale, R.: Extraction and](#)
576 [analysis of fungal spore biomarkers in atmospheric bioaerosol by HPLC–MS–MS and GC–MS, *Talanta*, 105, 142–](#)
577 [151, doi:10.1016/j.talanta.2012.11.006, 2013.](#)

578 [Burshtein, N., Lang-Yona, N., and Rudich, Y.: Ergosterol, arabitol and mannitol as tracers for biogenic aerosols](#)
579 [in the eastern Mediterranean, *Atmospheric Chem. Phys.*, 11\(2\), 829–839, doi:10.5194/acp-11-829-2011, 2011.](#)

580 [Chen, J., Kawamura, K., Liu, C.-Q., and Fu, P.: Long-term observations of saccharides in remote marine aerosols](#)
581 [from the western North Pacific: A comparison between 1990–1993 and 2006–2009 periods, *Atmos. Environ.*, 67,](#)
582 [448–458, doi:10.1016/j.atmosenv.2012.11.014, 2013.](#)

583 [China, S., Wang, B., Weis, J., Rizzo, L., Brito, J., Cirino, G. G., Kovarik, L., Artaxo, P., Gilles, M. K., and Laskin,](#)
584 [A.: Rupturing of biological spores as a source of secondary particles in Amazonia, *Environ. Sci. Technol.*, 50\(22\),](#)
585 [12179–12186, 2016.](#)

586 [China, S., Burrows, S. M., Wang, B., Harder, T. H., Weis, J., Tanarhte, M., Rizzo, L. V., Brito, J., Cirino, G. G.,](#)
587 [Ma, P.-L., Cliff, J., Artaxo, P., Gilles, M. K. and Laskin, A.: Fungal spores as a source of sodium salt particles in](#)
588 [the Amazon basin, *Nat. Commun.*, 9\(1\), doi:10.1038/s41467-018-07066-4, 2018.](#)

589 [Claeys, M., Graham, B., Vas, G., Wang, W., Vermeylen, R., Pashynska, V., Cafmeyer, J., Guyon, P., Andreae, M.](#)
590 [O., Artaxo, P., and Maenhaut, W.: Formation of secondary organic aerosols through photooxidation of isoprene,](#)
591 [*Science*, 303\(5661\), 1173, doi:10.1126/science.1092805, 2004.](#)

592 [Coulibaly, S., Minami, H., Abe, M., Hasei, T., Sera, N., Yamamoto, S., Funasaka, K., Asakawa, D., Watanabe,](#)
593 [M., Honda, N., Wakabayashi, K., and Watanabe, T.: Seasonal fluctuations in air pollution in Dzaifu, Japan, and](#)
594 [effect of long-range transport from mainland east Asia, *Biol. Pharm. Bull.*, 38\(9\), 1395–1403,](#)
595 [doi:10.1248/bpb.b15-00443, 2015.](#)

596 [Coz, E., Artíñano, B., Clark, L. M., Hernandez, M., Robinson, A. L., Casuccio, G. S., Lersch, T. L., and Pandis,](#)
597 [S. N.: Characterization of fine primary biogenic organic aerosol in an urban area in the northeastern United States,](#)
598 [Atmos. Environ., 44\(32\), 3952–3962, 2010.](#)

599 [Daellenbach, K. R., Stefanelli, G., Bozzetti, C., Vlachou, A., Fermo, P., Gonzalez, R., Piazzalunga, A., Colombi,](#)
600 [C., Canonaco, F., Hueglin, C., Kasper-Giebl, A., Jaffrezo, J.-L., Bianchi, F., Slowik, J. G., Baltensperger, U., El-](#)
601 [Haddad, I., and Prévôt, A. S. H.: Long-term chemical analysis and organic aerosol source apportionment at nine](#)
602 [sites in central Europe: source identification and uncertainty assessment, Atmos. Chem. Phys., 17\(21\), 13265–](#)
603 [13282, doi:10.5194/acp-17-13265-2017, 2017.](#)

604 [Dai, Y.-H. and Zhou, W.-X.: Temporal and spatial correlation patterns of air pollutants in Chinese cities, PLoS](#)
605 [ONE, 12\(8\), doi:10.1371/journal.pone.0182724, 2017.](#)

606 [Després, V. R., Alex Huffman, J., Burrows, S. M., Hoose, C., Safatov, A. S., Buryak, G., Fröhlich-Nowoisky, J.,](#)
607 [Elbert, W., Andreae, M. O., Pöschl, U., and Jaenicke, R.: Primary biological aerosol particles in the atmosphere:](#)
608 [a review, Tellus B., 64\(1\), 15598, doi:10.3402/tellusb.v64i0.15598, 2012.](#)

609 [Di Filippo, P., Pomata, D., Riccardi, C., Buiarelli, F., and Perrino, C.: Fungal contribution to size-segregated](#)
610 [aerosol measured through biomarkers, Atmos. Environ., 64, 132–140, 2013.](#)

611 [Eisner, A. D., Richmond-Bryant, J., Hahn, I., Drake-Richman, Z. E., Brixey, L. A., Wiener, R. W., and Ellenson,](#)
612 [W. D.: Analysis of indoor air pollution trends and characterization of infiltration delay time using a cross-](#)
613 [correlation method, J. Environ. Monit., 11\(12\), 2201, doi:10.1039/b907144j, 2009.](#)

614 [Elbert, W., Taylor, P. E., Andreae, M. O., and Pöschl, U.: Contribution of fungi to primary biogenic aerosols in](#)
615 [the atmosphere: wet and dry discharged spores, carbohydrates, and inorganic ions, Atmos. Chem. Phys., 7\(17\),](#)
616 [4569–4588, doi:10.5194/acp-7-4569-2007, 2007.](#)

617 [Fang, Z., Guo, W., Zhang, J., and Lou, X.: Influence of heat Events on the composition of airborne bacterial](#)
618 [communities in urban ecosystems, Int. J. Environ. Res. Public. Health, 15\(10\), 2295, doi:10.3390/ijerph15102295,](#)
619 [2018.](#)

620 [Fröhlich-Nowoisky, J., Pickersgill, D. A., Després, V. R. and Pöschl, U.: High diversity of fungi in air particulate](#)
621 [matter, Proc. Natl. Acad. Sci. U. S. A., 106\(31\), 12814–12819, doi:10.1073/pnas.0811003106, 2009.](#)

622 [Fröhlich-Nowoisky, J., Kampf, C. J., Weber, B., Huffman, J. A., Pöhlker, C., Andreae, M. O., Lang-Yona, N.,](#)
623 [Burrows, S. M., Gunthe, S. S., Elbert, W., Su, H., Hoor, P., Thines, E., Hoffmann, T., Després, V. R., and Pöschl,](#)
624 [U.: Bioaerosols in the earth system: climate, health, and ecosystem interactions, Atmos. Res., 182, 346–376,](#)
625 [doi:10.1016/j.atmosres.2016.07.018, 2016.](#)

626 [Fröhlich-Nowoisky, J., Amato, P., Renard, P., Brisebois, E., and Duchaine, C.: Quantification and characterization](#)
627 [of bioaerosols \(offline techniques\), in Microbiology of Aerosols, pp. 49–82, John Wiley & Sons, Ltd., doi:](#)
628 [10.1002/9781119132318, 2017.](#)

629 [Glasius, M., Hansen, A. M. K., Claeys, M., Henzing, J. S., Jedynska, A. D., Kasper-Giebl, A., Kistler, M.,](#)
630 [Kristensen, K., Martinsson, J., Maenhaut, W., Nøjgaard, J. K., Spindler, G., Stenström, K. E., Swietlicki, E., Szidat,](#)
631 [S., Simpson, D., and Yttri, K. E.: Composition and sources of carbonaceous aerosols in northern Europe during](#)
632 [winter, Atmos. Environ., 173, 127–141, doi:10.1016/j.atmosenv.2017.11.005, 2018.](#)

633 [Golly, B., Waked, A., Weber, S., Samaké, A., Jacob, V., Conil, S., Rangognio, J., Chrétien, E., Vagnot, M.-P.,](#)
634 [Robic, P.-Y., Besombes, J.-L., and Jaffrezo, J.-L.: Organic markers and OC source apportionment for seasonal](#)
635 [variations of PM_{2.5} at 5 rural sites in France, Atmos. Environ., 198, 142–157, doi:10.1016/j.atmosenv.2018.10.027,](#)
636 [2018.](#)

637 [Gosselin, M. I., Rathnayake, C. M., Crawford, I., Pöhlker, C., Fröhlich-Nowoisky, J., Schmer, B., Després, V. R.,](#)
638 [Engling, G., Gallagher, M., Stone, E., Pöschl, U., and Huffman, J. A.: Fluorescent bioaerosol particle, molecular](#)
639 [tracer, and fungal spore concentrations during dry and rainy periods in a semi-arid forest, Atmos. Chem. Phys.,](#)
640 [16\(23\), 15165–15184, doi:10.5194/acp-16-15165-2016, 2016.](#)

641 [Graham, B., Guyon, P., Taylor, P. E., Artaxo, P., Maenhaut, W., Glovsky, M. M., Flagan, R. C., and Andreae, M.](#)
642 [O.: Organic compounds present in the natural Amazonian aerosol: Characterization by gas chromatography-mass](#)
643 [spectrometry: organic compounds in Amazonian aerosols., J. Geophys. Res. Atmospheres, 108\(D24\), 4766,](#)
644 [doi:10.1029/2003JD003990, 2003.](#)

645 [Grinn-Gofroń, A., Nowosad, J., Bosiacka, B., Camacho, I., Pashley, C., Belmonte, J., De Linares, C., Ianovici, N.,](#)
646 [Manzano, J. M. M., Sadyś, M., Skjøth, C., Rodinkova, V., Tormo-Molina, R., Vokou, D., Fernández-Rodríguez,](#)
647 [S., and Damialis, A.: Airborne alternaria and Cladosporium fungal spores in Europe: forecasting possibilities and](#)
648 [relationships with meteorological parameters, Sci. Total Environ., 653, 938–946,](#)
649 [doi:10.1016/j.scitotenv.2018.10.419, 2019.](#)

650 [Heald, C. L. and Spracklen, D. V.: Atmospheric budget of primary biological aerosol particles from fungal spores,](#)
651 [Geophys. Res. Lett., 36\(9\), doi:10.1029/2009GL037493, 2009.](#)

652 [Hill, T. C. J., DeMott, P. J., Conen, F., and Möhler, O.: Impacts of bioaerosols on atmospheric ice nucleation](#)
653 [processes, in Microbiology of Aerosols, pp. 195–219, John Wiley & Sons, Ltd., doi:10.1002/9781119132318,](#)
654 [2017.](#)

655 [Hiranuma, N., Adachi, K., Bell, D. M., Belosi, F., Beydoun, H., Bhaduri, B., Bingemer, H., Budke, C., Clemen,](#)
656 [H.-C., Conen, F., Cory, K. M., Curtius, J., DeMott, P. J., Eppers, O., Grawe, S., Hartmann, S., Hoffmann, N.,](#)
657 [Höhler, K., Jantsch, E., Kiselev, A., Koop, T., Kulkarni, G., Mayer, A., Murakami, M., Murray, B. J., Nicosia, A.,](#)
658 [Petters, M. D., Piazza, M., Polen, M., Reicher, N., Rudich, Y., Saito, A., Santachiara, G., Schiebel, T., Schill, G.,](#)
659 [P., Schneider, J., Segev, L., Stopelli, E., Sullivan, R. C., Suski, K., Szakáll, M., Tajiri, T., Taylor, H., Tobo, Y.,](#)
660 [Ullrich, R., Weber, D., Wex, H., Whale, T. F., Whiteside, C. L., Yamashita, K., Zelenyuk, A., and Möhler, O.: A](#)
661 [comprehensive characterization of ice nucleation by three different types of cellulose particles immersed in water,](#)
662 [Atmos. Chem. Phys., 19\(7\), 4823–4849, doi:10.5194/acp-19-4823-2019, 2019.](#)

663 [Holden, A. S., Sullivan, A. P., Munchak, L. A., Kreidenweis, S. M., Schichtel, B. A., Malm, W. C., and Collett, J.](#)
664 [L.: Determining contributions of biomass burning and other sources to fine particle contemporary carbon in the](#)
665 [western United States, Atmos. Environ., 45\(11\), 1986–1993, doi:10.1016/j.atmosenv.2011.01.021, 2011.](#)

666 [Huffman, J. A. and Santarpia, J.: Online Techniques for quantification and characterization of biological aerosols,](#)
667 [in Microbiology of Aerosols, pp. 83–114, John Wiley & Sons, Ltd., doi:10.1002/9781119132318, 2017.](#)

668 [Humbal, C., Gautam, S. and Trivedi, U.: A review on recent progress in observations, and health effects of](#)
669 [bioaerosols, Environ. Int., 118, 189–193, doi:10.1016/j.envint.2018.05.053, 2018.](#)

670 [Hummel, M., Hoose, C., Gallagher, M., Healy, D. A., Huffman, J. A., O’Connor, D., Pöschl, U., Pöhlker, C.,](#)
671 [Robinson, N. H., Schnaiter, M., Sodeau, J. R., Stengel, M., Toprak, E., and Vogel, H.: Regional-scale simulations](#)
672 [of fungal spore aerosols using an emission parameterization adapted to local measurements of fluorescent](#)
673 [biological aerosol particles, Atmos. Chem. Phys., 15\(11\), 6127–6146, doi:10.5194/acp-15-6127-2015, 2015.](#)

674 [Jacobson, M. Z. and Streets, D. G.: Influence of future anthropogenic emissions on climate, natural emissions, and](#)
675 [air quality, J. Geophys. Res., 114\(D8\), D08118, doi:10.1029/2008JD011476, 2009.](#)

676 [Jaenicke, R.: Abundance of cellular material and proteins in the atmosphere, Science, 308\(5718\), 73–73,](#)
677 [doi:10.1126/science.1106335, 2005.](#)

678 [Jia, Y., Bhat, S., and Fraser, M. P.: Characterization of saccharides and other organic compounds in fine particles](#)
679 [and the use of saccharides to track primary biologically derived carbon sources, Atmos. Environ., 44\(5\), 724–732,](#)
680 [doi:10.1021/es103104e, 2010.](#)

681 [Jones, A. M. and Harrison, R. M.: The effects of meteorological factors on atmospheric bioaerosol](#)
682 [concentrations—a review, Sci. Total Environ., 326\(1\), 151–180, doi:10.1016/j.scitotenv.2003.11.021, 2004.](#)

683 [Kang, M., Ren, L., Ren, H., Zhao, Y., Kawamura, K., Zhang, H., Wei, L., Sun, Y., Wang, Z., and Fu, P.: Primary](#)
684 [biogenic and anthropogenic sources of organic aerosols in Beijing, China: Insights from saccharides and n-alkanes,](#)
685 [Environ. Pollut., doi:10.1016/j.envpol.2018.09.118, 2018.](#)

686 [Karimi, B., Terrat, S., Dequiedt, S., Saby, N. P. A., Horrigue, W., Lelièvre, M., Nowak, V., Jolivet, C., Arrouays,](#)
687 [D., Wincker, P., Cruaud, C., Bispo, A., Maron, P.-A., Bouré, N. C. P., and Ranjard, L.: Biogeography of soil](#)
688 [bacteria and archaea across France, *Sci. Adv.*, 4\(7\), eaat1808, doi:10.1126/sciadv.aat1808, 2018.](#)

689 [Kaso, A.: Computation of the normalized cross-correlation by fast Fourier transform, *PLOS ONE*, 13\(9\),](#)
690 [e0203434, doi:10.1371/journal.pone.0203434, 2018.](#)

691 [Kembel, S. W. and Mueller, R. C.: Plant traits and taxonomy drive host associations in tropical phyllosphere fungal](#)
692 [communities, *Botany*, 92\(4\), 303–311, doi:10.1139/cjb-2013-0194, 2014.](#)

693 [Kunit, M. and Puxbaum, H.: Enzymatic determination of the cellulose content of atmospheric aerosols, *Atmos.*](#)
694 [Environ., 30\(8\), 1233–1236, doi:10.1016/1352-2310\(95\)00429-7, 1996.](#)

695 [Laforest-Lapointe, I., Messier, C., and Kembel, S. W.: Tree leaf bacterial community structure and diversity differ](#)
696 [along a gradient of urban intensity, *mSystems*, 2\(6\), e00087-17, doi:10.1128/mSystems.00087-17, 2017.](#)

697 [Lainer, M., Hocke, K., and Kämpfer, N.: Variability of mesospheric water vapor above Bern in relation to the 27-](#)
698 [day solar rotation cycle, *J. Atmos. Sol.-Terr. Phys.*, 143–144, 71–87, doi:10.1016/j.jastp.2016.03.008, 2016.](#)

699 [Le Pichon, A., Blanc, E., and Hauchecorne, A.: Infrasound monitoring for atmospheric studies, 2nd ed., Springer](#)
700 [International Publishing, Switzerland., doi:10.1007/978-3-319-75140-5, 2019.](#)

701 [Lecours, P. B., Duchaine, C., Thibaudon, M., and Marsolais, D.: Health impacts of bioaerosol exposure, in](#)
702 [Microbiology of Aerosols, pp. 249–268, John Wiley & Sons, Ltd., doi:10.1002/9781119132318, 2017.](#)

703 [Li, L., Ren, L., Ren, H., Yue, S., Xie, Q., Zhao, W., Kang, M., Li, J., Wang, Z., Sun, Y., and Fu, P.: Molecular](#)
704 [characterization and seasonal variation in primary and secondary organic aerosols in Beijing, China, *J. Geophys.*](#)
705 [Res. Atmospheres, 123\(21\), 12,394–12,412, doi:10.1029/2018JD028527, 2018.](#)

706 [Liang, L., Engling, G., He, K., Du, Z., Cheng, Y., and Duan, F.: Evaluation of fungal spore characteristics in](#)
707 [Beijing, China, based on molecular tracer measurements, *Environ. Res. Lett.*, 8\(1\), 014005, doi:10.1088/1748-](#)
708 [9326/8/1/014005, 2013.](#)

709 [Lindow, S. E. and Brandl, M. T.: Microbiology of the Phyllosphere, *Appl. Environ. Microbiol.*, 69\(4\), 1875–1883,](#)
710 [doi:10.1128/AEM.69.4.1875-1883.2003, 2003.](#)

711 [Lymperopoulou, D. S., Adams, R. I., and Lindow, S. E.: Contribution of vegetation to the microbial composition](#)
712 [of nearby outdoor air, *Appl. Environ. Microbiol.*, 82\(13\), 3822–3833, doi:10.1128/AEM.00610-16, 2016.](#)

713 [Manninen, H. E., Bäck, J., Sihto-Nissilä, S.-L., Huffman, J. A., Pessi, A.-M., Hiltunen, V., Aalto, P. P., Hidalgo](#)
714 [Fernández, P. J., Hari, P., Saarto, A., Kulmala, M., and Petäjä, T.: Patterns in airborne pollen and other primary](#)
715 [biological aerosol particles \(PBAP\), and their contribution to aerosol mass and number in a boreal forest, *Boreal*](#)
716 [Environ. Res., 383–405, doi:hdl.handle.net/10138/165208, 2014.](#)

717 [Martin, S. T., Andreae, M. O., Artaxo, P., Baumgardner, D., Chen, Q., Goldstein, A. H., Guenther, A., Heald, C.](#)
718 [L., Mayol-Bracero, O. L., McMurry, P. H., Pauliquevis, T., Pöschl, U., Prather, K. A., Roberts, G. C., Saleska, S.](#)
719 [R., Silva Dias, M. A., Spracklen, D. V., Swietlicki, E., and Trebs, I.: Sources and properties of Amazonian aerosol](#)
720 [particles, *Rev. Geophys.*, 48\(2\), RG2002, doi:10.1029/2008RG000280, 2010.](#)

721 [Medeiros, P. M., Fernandes, M. F., Dick, R. P., and Simoneit, B. R. T.: Seasonal variations in sugar contents and](#)
722 [microbial community in a ryegrass soil, *Chemosphere*, 65\(5\), 832–839, doi:10.1016/j.chemosphere.2006.03.025,](#)
723 [2006a.](#)

724 [Medeiros, P. M., Conte, M. H., Weber, J. C. and Simoneit, B. R. T.: Sugars as source indicators of biogenic organic](#)
725 [carbon in aerosols collected above the Howland Experimental Forest, Maine, *Atmos. Environ.*, 40\(9\), 1694–1705,](#)
726 [2006b.](#)

727 [Meisner, A., Jacquiod, S., Snoek, B. L., ten Hooven, F. C., and van der Putten, W. H.: Drought legacy effects on](#)
728 [the composition of soil fungal and prokaryote communities, *Front. Microbiol.*, 9, doi:10.3389/fmicb.2018.00294,](#)
729 [2018.](#)

- 730 [Mhuireach, G., Johnson, B. R., Altrichter, A. E., Ladau, J., Meadow, J. F., Pollard, K. S., and Green, J. L.: Urban greenness influences airborne bacterial community composition, *Sci. Total Environ.*, 571, 680–687, doi:10.1016/j.scitotenv.2016.07.037, 2016.](#)
- 731
- 732
- 733 [Morris, C. E., Sands, D. C., Bardin, M., Jaenicke, R., Vogel, B., Leyronas, C., Ariya, P. A., and Psenner, R.: Microbiology and atmospheric processes: research challenges concerning the impact of airborne micro-organisms on the atmosphere and climate, *Biogeosciences*, 8\(1\), 17–25, doi:10.5194/bg-8-17-2011, 2011.](#)
- 734
- 735
- 736 [Morris, C. E., Conen, F., Alex Huffman, J., Phillips, V., Pöschl, U., and Sands, D. C.: Bioprecipitation: a feedback cycle linking Earth history, ecosystem dynamics and land use through biological ice nucleators in the atmosphere, *Glob. Change Biol.*, 20\(2\), 341–351, doi:10.1111/gcb.12447, 2014.](#)
- 737
- 738
- 739 [Myriokefalitakis, S., Fanourgakis, G., and Kanakidou, M.: The Contribution of bioaerosols to the organic carbon budget of the atmosphere, in *Perspectives on Atmospheric Sciences*, pp. 845–851, Springer International Publishing., doi:10.1007/978-3-319-35095-0, 2017.](#)
- 740
- 741
- 742 [Nirmalkar, J., Deshmukh, D. K., Deb, M. K., Tsai, Y. I., and Pervez, S.: Characteristics of aerosol during major biomass burning events over eastern central India in winter: A tracer-based approach, *Atmos. Pollut. Res.*, doi:10.1016/j.apr.2018.12.010, 2018.](#)
- 743
- 744
- 745 [Pashynska, V., Vermeylen, R., Vas, G., Maenhaut, W., and Claeys, M.: Development of a gas chromatographic/ion trap mass spectrometric method for the determination of levoglucosan and saccharidic compounds in atmospheric aerosols. Application to urban aerosols, *J. Mass Spectrom.*, 37\(12\), 1249–1257, doi:10.1002/jms.391, 2002.](#)
- 746
- 747
- 748 [Perrino, C. and Marcovecchio, F.: A new method for assessing the contribution of primary biological atmospheric particles to the mass concentration of the atmospheric aerosol, *Environ. Int.*, 87, 108–115, doi:10.1016/j.envint.2015.11.015, 2016.](#)
- 749
- 750
- 751 [Pietrogrande, M. C., Bacco, D., Visentin, M., Ferrari, S., and Casali, P.: Polar organic marker compounds in atmospheric aerosol in the Po valley during the supersito campaigns — part 2: seasonal variations of sugars, *Atmos. Environ.*, 97, 215–225, doi:0.1016/j.atmosenv.2014.07.056, 2014.](#)
- 752
- 753
- 754 [Pindado, O. and Perez, R. M.: Source apportionment of particulate organic compounds in a rural area of Spain by positive matrix factorization, *Atmos. Pollut. Res.*, 2\(4\), 492–505, doi:10.5094/APR.2011.056, 2011.](#)
- 755
- 756 [Pirttilä, A. M. and Frank, A. C., Eds.: Endophytes of forest trees, Springer Netherlands, Dordrecht., doi:10.1007/978-3-319-89833-9, 2011.](#)
- 757
- 758 [Puxbaum, H. and Tenze-Kunit, M.: Size distribution and seasonal variation of atmospheric cellulose, *Atmos. Environ.*, 37\(26\), 3693–3699, doi:10.1016/S1352-2310\(03\)00451-5, 2003.](#)
- 759
- 760 [Rajput, P., Chauhan, A. S., and Gupta, T.: Bioaerosols over the indo-gangetic plain: influence of biomass burning emission and ambient meteorology, in *Environmental Contaminants: measurement, modelling and control*, pp. 93–121, Springer Singapore., doi:10.1007/978-981-10-7332-8, 2018.](#)
- 761
- 762
- 763 [Ram, K., Sarin, M. M., and Hegde, P.: Long-term record of aerosol optical properties and chemical composition from a high-altitude site \(Manora Peak\) in central Himalaya, *Atmos. Chem. Phys.*, 13, doi:10.5194/acp-10-11791-2010, 2010.](#)
- 764
- 765
- 766 [Ramoni, J. and Seiboth, B.: Degradation of plant cell wall polymers by fungi, in *Environmental and microbial relationships*, vol. IV, pp. 127–148, Springer International Publishing, Cham., doi: 10.1007/978-3-540-71840-6 2016.](#)
- 767
- 768
- 769 [Rathnayake, C. M., Metwali, N., Jayarathne, T., Kettler, J., Huang, Y., Thorne, P. S., O’Shaughnessy, P. T., and Stone, E. A.: Influence of rain on the abundance of bioaerosols in fine and coarse particles, *Atmos. Chem. Phys.*, 17\(3\), 2459–2475, doi:10.5194/acp-17-2459-2017, 2017.](#)
- 770
- 771
- 772 [Reddy, S. M., Girisham, S., and Babu, G. N.: Applied Microbiology \(agriculture, environmental, food and industrial microbiology\), Scientific Publishers, doi:9789387307407, 2017.](#)
- 773

774 [Rogge, W. F., Medeiros, P. M., and Simoneit, B. R. T.: Organic marker compounds in surface soils of crop fields](#)
775 [from the San Joaquin Valley fugitive dust characterization study, *Atmos. Environ.*, 41\(37\), 8183–8204,](#)
776 [doi:10.1016/j.atmosenv.2007.06.030, 2007.](#)

777 [Samaké, A., Jaffrezo, J.-L., Favez, O., Weber, S., Jacob, V., Albinet, A., Riffault, V., Perdrix, E., Waked, A.,](#)
778 [Golly, B., Salameh, D., Chevrier, F., Oliveira, D. M., Bonnaire, N., Besombes, J.-L., Martins, J. M. F., Conil, S.,](#)
779 [Guillaud, G., Mesbah, B., Rocq, B., Robic, P.-Y., Hulin, A., Meur, S. L., Descheemaeker, M., Chretien, E.,](#)
780 [Marchand, N., and Uzu, G.: Polyols and glucose particulate species as tracers of primary biogenic organic aerosols](#)
781 [at 28 French sites, *Atmos. Chem. Phys.*, 19\(5\), 3357–3374, doi:10.5194/acp-19-3357-2019, 2019.](#)

782 [Sánchez-Ochoa, A., Kasper-Giebl, A., Puxbaum, H., Gelencser, A., Legrand, M., and Pio, C.: Concentration of](#)
783 [atmospheric cellulose: a proxy for plant debris across a west-east transect over Europe, *J. Geophys. Res.*,](#)
784 [112\(D23\), doi:10.1029/2006JD008180, 2007.](#)

785 [Sesartic, A. and Dallafior, T. N.: Global fungal spore emissions, review and synthesis of literature data,](#)
786 [*Biogeosciences*, 8\(5\), 1181–1192, doi:10.5194/bg-8-1181-2011, 2011.](#)

787 [Shcherbakova, L. A.: Advanced methods of plant pathogen diagnostics, in *Comprehensive and molecular*](#)
788 [phytopathology, pp. 75–116, Elsevier, Amsterdam, doi:9780080469331, 2007.](#)

789 [Simoneit, B. R. T., Elias, V. O., Kobayashi, M., Kawamura, K., Rushdi, A. I., Medeiros, P. M., Rogge, W. F., and](#)
790 [Didyk, B. M.: Sugars dominant water-soluble organic compounds in soils and characterization as tracers in](#)
791 [atmospheric particulate matter, *Environ. Sci. Technol.*, 38\(22\), 5939–5949, doi:10.1029/2004JD004598, 2004.](#)

792 [Srivastava, D., Favez, O., Bonnaire, N., Lucarelli, F., Haeffelin, M., Perraudin, E., Gros, V., Villenave, E., and](#)
793 [Albinet, A.: Speciation of organic fractions does matter for aerosol source apportionment: part 2: intensive short-](#)
794 [term campaign in the Paris area \(France\), *Sci. Total Environ.*, 634, 267–278, doi:10.1016/j.scitotenv.2018.03.296,](#)
795 [2018.](#)

796 [Sullivan, A. P., Frank, N., Kenski, D. M., and Collett, J. L.: Application of high-performance anion-exchange](#)
797 [chromatography–pulsed amperometric detection for measuring carbohydrates in routine daily filter samples](#)
798 [collected by a national network 2: examination of sugar alcohols/polyols, sugars, and anhydrosugars in the upper](#)
799 [Midwest, *J. Geophys. Res. Atmospheres*, 116\(D8\), D08303, doi:10.1029/2010JD014169, 2011.](#)

800 [Tanarhte, M., Bacer, S., Burrows, S. M., Huffman, J. A., Pierce, K. M., Pozzer, A., Sarda-Estève, R., Savage, N.](#)
801 [J., and Lelieveld, J.: Global modeling of fungal spores with the EMAC chemistry climate model: uncertainties in](#)
802 [emission parametrizations and observations, *Atmospheric Chem. Phys. Discuss.*, 1–31, doi:10.5194/acp-2019-](#)
803 [251, 2019.](#)

804 [Véléz, H., Glassbrook, N. J., and Daub, M. E.: Mannitol metabolism in the phytopathogenic fungus *alternaria*](#)
805 [*alternata*, *Fungal Genet. Biol.*, 44\(4\), 258–268, doi:10.1016/j.fgb.2006.09.008, 2007.](#)

806 [Verma, S. K., Kawamura, K., Chen, J., and Fu, P.: Thirteen years of observations on primary sugars and sugar](#)
807 [alcohols over remote Chichijima Island in the western north Pacific, *Atmos. Chem. Phys.*, 18\(1\), 81–101,](#)
808 [doi:https://doi.org/10.5194/acp-18-81-2018, 2018.](#)

809 [Vlachou, A., Daellenbach, K. R., Bozzetti, C., Chazeau, B., Salazar, G. A., Szidat, S., Jaffrezo, J.-L., Hueglin, C.,](#)
810 [Baltensperger, U., Haddad, I. E., and Prévôt, A. S. H.: Advanced source apportionment of carbonaceous aerosols](#)
811 [by coupling offline AMS and radiocarbon size-segregated measurements over a nearly 2-year period, *Atmos.*](#)
812 [Chem. Phys., 18\(9\), 6187–6206, doi:10.5194/acp-18-6187-2018, 2018.](#)

813 [Waked, A., Favez, O., Alleman, L. Y., Piot, C., Petit, J.-E., Delaunay, T., Verlinden, E., Golly, B., Besombes, J.-](#)
814 [L., Jaffrezo, J.-L., and Leoz-Garziandia, E.: Source apportionment of PM₁₀ in a north-western Europe regional](#)
815 [urban background site \(Lens, France\) using positive matrix factorization and including primary biogenic](#)
816 [emissions, *Atmos. Chem. Phys.*, 14\(7\), 3325–3346, doi:10.5194/acp-14-3325-2014, 2014.](#)

817 [Wan, E. C. H. and Yu, J. Z.: Analysis of sugars and sugar polyols in atmospheric aerosols by chloride attachment](#)
818 [in liquid chromatography/negative ion electrospray mass spectrometry, *Environ. Sci. Technol.*, 41\(7\), 2459–2466,](#)
819 [doi:10.1021/es062390g, 2007.](#)

820 [Wang, X., Shen, Z., Liu, F., Lu, D., Tao, J., Lei, Y., Zhang, Q., Zeng, Y., Xu, H., Wu, Y., Zhang, R., and Cao, J.: saccharides in summer and winter PM_{2.5} over Xi'an, northwestern China: sources, and yearly variations of biomass](#)
821 [burning contribution to PM_{2.5}, Atmos. Res., 214, 410–417, doi:10.1016/j.atmosres.2018.08.024, 2018.](#)
822

823 [Wan, X., Kang, S., Rupakheti, M., Zhang, Q., Tripathee, L., Guo, J., Chen, P., Rupakheti, D., Panday, A. K.,](#)
824 [Lawrence, M. G., Kawamura, K., and Cong, Z.: Molecular characterization of organic aerosols in the Kathmandu](#)
825 [valley, Nepal: insights into primary and secondary sources, Atmos. Chem. Phys., 19\(5\), 2725–2747,](#)
826 [doi:10.5194/acp-19-2725-2019, 2019.](#)

827 [Weber, S., Uzu, G., Calas, A., Chevrier, F., Besombes, J.-L., Charron, A., Salameh, D., Ježek, I., Močnik, G., and](#)
828 [Jaffrezo, J.-L.: An apportionment method for the oxidative potential of atmospheric particulate matter sources:](#)
829 [application to a one-year study in Chamonix, France, Atmos. Chem. Phys., 18\(13\), 9617–9629, doi:10.5194/acp-](#)
830 [18-9617-2018, 2018.](#)

831 [Weber, S., Salameh, D., Albinet, A., Alleman, L. Y., Waked, A., Besombes, J.-L., Jacob, V., Guillaud, G.,](#)
832 [Meshbah, B., Rocq, B., Hulin, A., Dominik-Sègue, M., Chrétien, E., Jaffrezo, J.-L., and Favez, O.: Comparison](#)
833 [of PM₁₀ Sources profiles at 15 French sites using a harmonized constrained positive matrix factorization approach,](#)
834 [Atmosphere, 10\(6\), 310, doi:10.3390/atmos10060310, 2019.](#)

835 [Wéry, N., Galès, A., and Brunet, Y.: Bioaerosol sources, in Microbiology of Aerosols, pp. 115–135, John Wiley](#)
836 [& Sons, Ltd., doi:10.1002/9781119132318, 2017.](#)

837 [Whipps, J. M., Hand, P., Pink, D., and Bending, G. D.: Phyllosphere microbiology with special reference to](#)
838 [diversity and plant genotype, J. Appl. Microbiol., 105\(6\), 1744–1755, doi:10.1111/j.1365-2672.2008.03906.x,](#)
839 [2008.](#)

840 [Xiao, M., Wang, Q., Qin, X., Yu, G., and Deng, C.: Composition, sources, and distribution of PM_{2.5} saccharides](#)
841 [in a coastal urban site of China, Atmosphere, 9\(7\), 274, doi:10.3390/atmos9070274, 2018.](#)

842 [Xu, J., He, J., Xu, H., Ji, D., Snape, C., Yu, H., Jia, C., Wang, C., and Gao, J.: Simultaneous measurement of](#)
843 [multiple organic tracers in fine aerosols from biomass burning and fungal spores by HPLC-MS/MS, RSC. Adv.,](#)
844 [8\(59\), 34136–34150, doi:10.1039/C8RA04991B, 2018.](#)

845 [Yan, C., Sullivan, A. P., Cheng, Y., Zheng, M., Zhang, Y., Zhu, T., and Collett, J. L.: Characterization of](#)
846 [saccharides and associated usage in determining biogenic and biomass burning aerosols in atmospheric fine](#)
847 [particulate matter in the north China Plain, Sci. Total Environ., 650, 2939–2950,](#)
848 [doi:10.1016/j.scitotenv.2018.09.325, 2019.](#)

849 [Yan, K., Park, T., Yan, G., Chen, C., Yang, B., Liu, Z., Nemani, R., Knyazikhin, Y., and Myneni, R.: Evaluation](#)
850 [of MODIS LAI/FPAR product collection 6: part 1: consistency and improvements, Remote Sens., 8\(5\), 359,](#)
851 [doi:10.3390/rs8050359, 2016a.](#)

852 [Yan, K., Park, T., Yan, G., Liu, Z., Yang, B., Chen, C., Nemani, R., Knyazikhin, Y., and Myneni, R.: Evaluation](#)
853 [of MODIS LAI/FPAR product collection 6: part 2: validation and intercomparison, Remote Sens., 8\(6\), 460,](#)
854 [doi:10.3390/rs8060460, 2016b.](#)

855 [Yttri, K. E., Dye, C., and Kiss, G.: Ambient aerosol concentrations of sugars and sugar-alcohols at four different](#)
856 [sites in Norway, Atmos. Chem. Phys., 7\(16\), 4267–4279, doi:10.5194/acp-7-4267-2007, 2007.](#)

857 [Yttri, K. E., Simpson, D., Stenström, K., Puxbaum, H., and Svendby, T.: Source apportionment of the](#)
858 [carbonaceous aerosol in Norway – quantitative estimates based on 14C, thermal-optical and organic tracer](#)
859 [analysis, Atmos. Chem. Phys., 11, 9375-9394, doi:10.5194/acpd-11-7375-2011, 2011a.](#)

860 [Yttri, K. E., Simpson, D., Nøjgaard, J. K., Kristensen, K., Genberg, J., Stenström, K., Swietlicki, E., Hillamo, R.,](#)
861 [Aurela, M., Bauer, H., Offenberg, J. H., Jaoui, M., Dye, C., Eckhardt, S., Burkhardt, J. F., Stohl, A., and Glasius,](#)
862 [M.: Source apportionment of the summer time carbonaceous aerosol at Nordic rural background sites, Atmos.](#)
863 [Chem. Phys., 11\(24\), 13339–13357, doi:10.5194/acp-11-13339-2011, 2011b.](#)

864 [Yue, S., Ren, H., Fan, S., Wei, L., Zhao, J., Bao, M., Hou, S., Zhan, J., Zhao, W., Ren, L., Kang, M., Li, L., Zhang,](#)
865 [Y., Sun, Y., Wang, Z., and Fu, P.: High abundance of fluorescent biological aerosol particles in winter in Beijing,](#)
866 [China, ACS Earth Space Chem., 1\(8\), 493–502, doi:10.1021/acsearthspacechem.7b00062, 2017.](#)

867 [Zamfir, M., Gerstner, D. G., Walsler, S. M., Bünger, J., Eikmann, T., Heinze, S., Kolk, A., Nowak, D., Raulf, M.,](#)
868 [Sagunski, H., Sedlmaier, N., Suchenwirth, R., Wiesmüller, G. A., Wollin, K.-M., Tesseraux, I., and Herr, C. E.](#)
869 [W.: A systematic review of experimental animal studies on microbial bioaerosols: Dose-response data for the](#)
870 [derivation of exposure limits, Int. J. Hyg. Environ. Health, 222\(2\), 249–259, doi:10.1016/j.ijheh.2018.11.004,](#)
871 [2019.](#)

872 [Zhang, T., Engling, G., Chan, C.-Y., Zhang, Y.-N., Zhang, Z.-S., Lin, M., Sang, X.-F., Li, Y. D., and Li, Y.-S.:](#)
873 [Contribution of fungal spores to particulate matter in a tropical rainforest, Environ. Res. Lett., 5\(2\), 024010,](#)
874 [doi:10.1088/1748-9326/5/2/024010, 2010.](#)

875 [Zhang, Z., Engling, G., Zhang, L., Kawamura, K., Yang, Y., Tao, J., Zhang, R., Chan, C., and Li, Y.: Significant](#)
876 [influence of fungi on coarse carbonaceous and potassium aerosols in a tropical rainforest, Environ. Res. Lett.,](#)
877 [10\(3\), 034015, doi:10.1088/1748-9326/10/3/034015, 2015.](#)

878 [Zhu, C., Kawamura, K., and Kunwar, B.: Organic tracers of primary biological aerosol particles at subtropical](#)
879 [Okinawa Island in the western North Pacific Rim: organic biomarkers in the north pacific, J. Geophys. Res.](#)
880 [Atmospheres, 120\(11\), 5504–5523, 2015.](#)

881 [Zhu, W., Luo, L., Cheng, Z., Yan, N., Lou, S., and Ma, Y.: Characteristics and contributions of biogenic secondary](#)
882 [organic aerosol tracers to PM_{2.5} in Shanghai, China, Atmospheric Pollut. Res., 9\(2\), 179–188,](#)
883 [doi:10.1016/j.apr.2017.09.001, 2018a.](#)

884 [Zhu, W., Cheng, Z., Luo, L., Lou, S., Ma, Y., and Yan, N.: Investigation of fungal spore characteristics in PM_{2.5}](#)
885 [through organic tracers in Shanghai, China, Atmospheric Pollut. Res., 9\(5\), 894–900,](#)
886 [doi:10.1016/j.apr.2018.01.009, 2018b.](#)

887 [Abdalmogith, S. S. and Harrison, R. M.: The use of trajectory cluster analysis to examine the long-range transport](#)
888 [of secondary inorganic aerosol in the UK, Atmos. Environ., 39\(35\), 6686–6695,](#)
889 [doi:10.1016/j.atmosenv.2005.07.059, 2005.](#)

890 [Amato, F., Alastuey, A., Karanasiou, A., Lucarelli, F., Nava, S., Calzolari, G., Severi, M., Becagli, S., Gianelle, V.,](#)
891 [L., Colombi, C., Alves, C., Custódio, D., Nunes, T., Cerqueira, M., Pio, C., Eleftheriadis, K., Diapouli, E., Reche,](#)
892 [C., Minguillón, M. C., Manousakas, M. I., Maggos, T., Vratolis, S., Harrison, R. M., and Querol, X.: Airuse life+:](#)
893 [a harmonized PM speciation and source apportionment in five southern European cities, Atmos. Chem. Phys.,](#)
894 [16\(5\), 3289–3309, doi:10.5194/acp-16-3289-2016, 2016.](#)

895 [Amato, P., Brisebois, E., Draghi, M., Duchaine, C., Fröhlich-Nowoisky, J., Huffman, J. A., Mainelis, G., Robine,](#)
896 [E., and Thibaudon, M.: Main biological aerosols, specificities, abundance, and diversity, in Microbiology of](#)
897 [Aerosols, pp. 1–21, John Wiley & Sons, Ltd., doi:10.1002/9781119132318, 2017.](#)

898 [Ariya, P. A., Sun, J., Eltouny, N. A., Hudson, E. D., Hayes, C. T., and Kos, G.: Physical and chemical](#)
899 [characterization of bioaerosols—implications for nucleation processes, Int. Rev. Phys. Chem., 28\(1\), 1–32,](#)
900 [doi:10.1080/01442350802597438, 2009.](#)

901 [Barbaro, E., Kirchgeorg, T., Zangrando, R., Vecchiato, M., Piazza, R., Barbante, C., and Gambaro, A.: Sugars in](#)
902 [Antarctic aerosol, Atmos. Environ., 118, 135–144, doi:10.1016/j.atmosenv.2015.07.047, 2015.](#)

903 [Bauer, H., Claeys, M., Vermeylen, R., Schueller, E., Weinke, G., Berger, A., and Puxbaum, H.: Arabitol and](#)
904 [mannitol as tracers for the quantification of airborne fungal spores, Atmos. Environ., 42\(3\), 588–593,](#)
905 [doi:10.1016/j.atmosenv.2007.10.013, 2008a.](#)

906 [Bauer, H., Schueller, E., Weinke, G., Berger, A., Hitznerberger, R., Marr, I. L., and Puxbaum, H.: Significant](#)
907 [contributions of fungal spores to the organic carbon and to the aerosol mass balance of the urban atmospheric](#)
908 [aerosol, Atmos. Environ., 42\(22\), 5542–5549, doi:10.1016/j.atmosenv.2008.03.019, 2008b.](#)

909 Bodenhausen, N., Bortfeld Miller, M., Ackermann, M., and Vorholt, J. A.: A synthetic community approach
910 reveals plant genotypes affecting the phyllosphere microbiota, *PLoS Genet.*, 10(4), doi:
911 10.1371/journal.pgen.1004283, 2014.

912 Bowers, R. M., Sullivan, A. P., Costello, E. K., Collett, J. L., Knight, R., and Fierer, N.: Sources of bacteria in
913 outdoor air across cities in the Midwestern United States., *Appl. Environ. Microbiol.*, 77(18), 6350–6356,
914 doi:10.1128/AEM.05498-11, 2011.

915 Bozzetti, C., Daellenbach, K. R., Hueglin, C., Fermo, P., Sciare, J., Kasper Giebl, A., Mazar, Y., Abbaszade, G.,
916 El Kazzi, M., Gonzalez, R., Shuster Meiseles, T., Flasch, M., Wolf, R., Křepelová, A., Canonaco, F., Schnelle-
917 Kreis, J., Slowik, J. G., Zimmermann, R., Rudich, Y., Baltensperger, U., El Haddad, I., and Prévôt, A. S. H.: Size-
918 resolved identification, characterization, and quantification of primary biological organic aerosol at a European
919 rural site, *Environ. Sci. Technol.*, 50(7), 3425–3434, doi:10.1021/acs.est.5b05960, 2016.

920 Bringel, F. and Couée, I.: Pivotal roles of phyllosphere microorganisms at the interface between plant functioning
921 and atmospheric trace gas dynamics, *Front. Microbiol.*, 6, 486, doi:10.3389/fmicb.2015.00486, 2015.

922 Buiarelli, F., Canepari, S., Di Filippo, P., Perrino, C., Pomata, D., Riccardi, C., and Speziale, R.: Extraction and
923 analysis of fungal spore biomarkers in atmospheric bioaerosol by HPLC-MS-MS and GC-MS, *Talanta*, 105, 142–
924 151, doi:10.1016/j.talanta.2012.11.006, 2013.

925 Burshtein, N., Lang Yona, N., and Rudich, Y.: Ergosterol, arabinol and mannitol as tracers for biogenic aerosols
926 in the eastern Mediterranean, *Atmos. Chem. Phys.*, 11(2), 829–839, doi:10.5194/acp-11-829-2011, 2011.

927 Chen, J., Kawamura, K., Liu, C. Q., and Fu, P.: Long-term observations of saccharides in remote marine aerosols
928 from the western north Pacific: A comparison between 1990–1993 and 2006–2009 periods, *Atmos. Environ.*, 67,
929 448–458, doi:10.1016/j.atmosenv.2012.11.014, 2013.

930 China, S., Wang, B., Weis, J., Rizzo, L., Brito, J., Cirino, G. G., Kovarik, L., Artaxo, P., Gilles, M. K., and Laskin,
931 A.: Rupturing of biological spores as a source of secondary particles in Amazonia, *Environ. Sci. Technol.*, 50(22),
932 12179–12186, 2016.

933 China, S., Burrows, S. M., Wang, B., Harder, T. H., Weis, J., Tanarhte, M., Rizzo, L. V., Brito, J., Cirino, G. G.,
934 Ma, P. L., Cliff, J., Artaxo, P., Gilles, M. K., and Laskin, A.: Fungal spores as a source of sodium salt particles in
935 the Amazon basin, *Nat. Commun.*, 9(1), doi:10.1038/s41467-018-07066-4, 2018.

936 Claeys, M., Graham, B., Vas, G., Wang, W., Vermeylen, R., Pashynska, V., Cafmeyer, J., Guyon, P., Andreae, M.
937 O., Artaxo, P., and Maenhaut, W.: Formation of secondary organic aerosols through photooxidation of isoprene,
938 *Science*, 303(5661), 1173, doi:10.1126/science.1092805, 2004.

939 Coulibaly, S., Minami, H., Abe, M., Hasei, T., Sera, N., Yamamoto, S., Funasaka, K., Asakawa, D., Watanabe,
940 M., Honda, N., Wakabayashi, K., and Watanabe, T.: Seasonal fluctuations in air pollution in Dzaifu, Japan, and
941 effect of long range transport from mainland east Asia, *Biol. Pharm. Bull.*, 38(9), 1395–1403,
942 doi:10.1248/bpb.b15-00443, 2015.

943 Coz, E., Artúñano, B., Clark, L. M., Hernandez, M., Robinson, A. L., Casuccio, G. S., Lersch, T. L., and Pandis,
944 S. N.: Characterization of fine primary biogenic organic aerosol in an urban area in the northeastern United States,
945 *Atmos. Environ.*, 44(32), 3952–3962, 2010.

946 Daellenbach, K. R., Stefanelli, G., Bozzetti, C., Vlachou, A., Fermo, P., Gonzalez, R., Piazzalunga, A., Colombi,
947 C., Canonaco, F., Hueglin, C., Kasper Giebl, A., Jaffrezo, J. L., Bianchi, F., Slowik, J. G., Baltensperger, U., El-
948 Haddad, I., and Prévôt, A. S. H.: Long-term chemical analysis and organic aerosol source apportionment at nine
949 sites in central Europe: source identification and uncertainty assessment, *Atmos. Chem. Phys.*, 17(21), 13265–
950 13282, doi:10.5194/acp-17-13265-2017, 2017.

951 Després, V. R., Alex Huffman, J., Burrows, S. M., Hoose, C., Safatov, A. S., Buryak, G., Fröhlich-Nowoisky, J.,
952 Elbert, W., Andreae, M. O., Pöschl, U., and Jaenicke, R.: Primary biological aerosol particles in the atmosphere:
953 a review, *Tellus B.*, 64(1), 15598, doi:10.3402/tellusb.v64i0.15598, 2012.

954 Di Filippo, P., Pomata, D., Riccardi, C., Buiarelli, F., and Perrino, C.: Fungal contribution to size-segregated
955 aerosol measured through biomarkers, *Atmos. Environ.*, 64, 132–140, doi: 10.1016/j.atmosenv.2012.10.010, 2013.

956 Elbert, W., Taylor, P. E., Andreae, M. O., and Pöschl, U.: Contribution of fungi to primary biogenic aerosols in
957 the atmosphere: wet and dry discharged spores, carbohydrates, and inorganic ions, *Atmos. Chem. Phys.*, 7(17),
958 4569–4588, doi:10.5194/acp-7-4569-2007, 2007.

959 Fang, Z., Guo, W., Zhang, J., and Lou, X.: Influence of heat events on the composition of airborne bacterial
960 communities in urban ecosystems, *Int. J. Environ. Res. Public Health*, 15(10), 2295, doi:10.3390/ijerph15102295,
961 2018.

962 Fröhlich-Nowoisky, J., Pickersgill, D. A., Després, V. R., and Pöschl, U.: High diversity of fungi in air particulate
963 matter, *Proc. Natl. Acad. Sci. U. S. A.*, 106(31), 12814–12819, doi: 10.1073/pnas.0811003106, 2009.

964 Fröhlich-Nowoisky, J., Kampf, C. J., Weber, B., Huffman, J. A., Pöhlker, C., Andreae, M. O., Lang-Yona, N.,
965 Burrows, S. M., Gunthe, S. S., Elbert, W., Su, H., Hoor, P., Thines, E., Hoffmann, T., Després, V. R., and Pöschl,
966 U.: Bioaerosols in the earth system: climate, health, and ecosystem interactions, *Atmos. Res.*, 182, 346–376,
967 doi:10.1016/j.atmosres.2016.07.018, 2016.

968 Fu, P., Kawamura, K., Kobayashi, M., and Simoneit, B. R.: Seasonal variations of sugars in atmospheric particulate
969 matter from Gosan, Jeju Island: significant contributions of airborne pollen and Asian dust in spring, *Atmos.*
970 *Environ.*, 55, 234–239, doi: 10.1029/2003JD003697, 2012.

971 Fu, P. Q., Kawamura, K., Chen, J., Charrière, B., and Sempéré, R.: Organic molecular composition of marine
972 aerosols over the Arctic ocean in summer: contributions of primary emission and secondary aerosol formation,
973 *Biogeosciences*, 10(2), 653–667, doi:10.5194/bg-10-653-2013, 2013.

974 Glasius, M., Hansen, A. M. K., Claeys, M., Henzing, J. S., Jedynska, A. D., Kasper-Giebl, A., Kistler, M.,
975 Kristensen, K., Martinsson, J., Maenhaut, W., Nøjgaard, J. K., Spindler, G., Stenström, K. E., Swietlicki, E., Szidat,
976 S., Simpson, D., and Yttri, K. E.: Composition and sources of carbonaceous aerosols in northern Europe during
977 winter, *Atmos. Environ.*, 173, 127–141, doi:10.1016/j.atmosenv.2017.11.005, 2018.

978 Golly, B., Waked, A., Weber, S., Samaké, A., Jacob, V., Conil, S., Rangognio, J., Chrétien, E., Vagnot, M. P.,
979 Robic, P. Y., Besombes, J. L., and Jaffrezo, J. L.: Organic markers and OC source apportionment for seasonal
980 variations of PM_{2.5} at 5 rural sites in France, *Atmos. Environ.*, 198, 142–157,
981 doi:10.1016/j.atmosenv.2018.10.027, 2018.

982 Gosselin, M. I., Rathnayake, C. M., Crawford, I., Pöhlker, C., Fröhlich-Nowoisky, J., Schmer, B., Després, V. R.,
983 Engling, G., Gallagher, M., Stone, E., Pöschl, U., and Huffman, J. A.: Fluorescent bioaerosol particle, molecular
984 tracer, and fungal spore concentrations during dry and rainy periods in a semi-arid forest, *Atmos. Chem. Phys.*,
985 16(23), 15165–15184, doi: 10.5194/acp-16-15165-2016, 2016.

986 Graham, B., Guyon, P., Taylor, P. E., Artaxo, P., Maenhaut, W., Glovsky, M. M., Flagan, R. C., and Andreae, M.
987 O.: Organic compounds present in the natural Amazonian aerosol: Characterization by gas chromatography-mass
988 spectrometry: Organic compounds in Amazonian aerosols., *J. Geophys. Res. Atmos.*, 108(D24), 4766,
989 doi:10.1029/2003JD003990, 2003.

990 Grinn-Gofroń, A., Nowosad, J., Bosiacka, B., Camacho, I., Pashley, C., Belmonte, J., De Linares, C., Ianovici, N.,
991 Manzano, J. M. M., Sadyś, M., Skjoth, C., Rodinkova, V., Tormo Molina, R., Vokou, D., Fernández-Rodríguez,
992 S., and Damialis, A.: Airborne alternaria and cladosporium fungal spores in Europe: forecasting possibilities and
993 relationships with meteorological parameters, *Sci. Total Environ.*, 653, 938–946,
994 doi:10.1016/j.scitotenv.2018.10.419, 2019.

995 Grömping, U.: Relative importance for linear regression in R: the package relaimpo, *J. Stat. Softw.*, 17(1),
996 doi:10.18637/jss.v017.i01, 2006.

997 Heald, C. L. and Spracklen, D. V.: Atmospheric budget of primary biological aerosol particles from fungal spores,
998 *Geophys. Res. Lett.*, 36(9), doi:10.1029/2009GL037493, 2009.

- 999 Hill, T. C. J., DeMott, P. J., Conen, F., and Möhler, O.: Impacts of bioaerosols on atmospheric ice nucleation
1000 processes, in *Microbiology of Aerosols*, pp. 195–219, John Wiley & Sons, Ltd., doi:10.1002/9781119132318,
1001 2017.
- 1002 Holden, A. S., Sullivan, A. P., Munchak, L. A., Kreidenweis, S. M., Schichtel, B. A., Malm, W. C., and Collett, J.
1003 L.: Determining contributions of biomass burning and other sources to fine particle contemporary carbon in the
1004 western United States, *Atmos. Environ.*, 45(11), 1986–1993, doi:10.1016/j.atmosenv.2011.01.021, 2011.
- 1005 Humbal, C., Gautam, S., and Trivedi, U.: A review on recent progress in observations, and health effects of
1006 bioaerosols, *Environ. Int.*, 118, 189–193, doi:10.1016/j.envint.2018.05.053, 2018.
- 1007 Hummel, M., Hoose, C., Gallagher, M., Healy, D. A., Huffman, J. A., O'Connor, D., Pöschl, U., Pöhlker, C.,
1008 Robinson, N. H., Schnaiter, M., Sodeau, J. R., Stengel, M., Toprak, E., and Vogel, H.: Regional scale simulations
1009 of fungal spore aerosols using an emission parameterization adapted to local measurements of fluorescent
1010 biological aerosol particles, *Atmos. Chem. Phys.*, 15(11), 6127–6146, doi:10.5194/acp-15-6127-2015, 2015.
- 1011 Jacobson, M. Z. and Streets, D. G.: Influence of future anthropogenic emissions on climate, natural emissions, and
1012 air quality, *J. Geophys. Res.*, 114(D8), D08118, doi:10.1029/2008JD011476, 2009.
- 1013 Jaenicke, R.: Abundance of cellular material and proteins in the atmosphere, *Science*, 308(5718), 73–73,
1014 doi:10.1126/science.1106335, 2005.
- 1015 Jia, Y. and Fraser, M.: Characterization of saccharides in size fractionated ambient particulate matter and aerosol
1016 sources: the contribution of primary biological aerosol particles (PBAPs) and soil to ambient particulate matter,
1017 *Environ. Sci. Technol.*, 45(3), 930–936, doi:10.1021/es103104e, 2011.
- 1018 Jia, Y., Bhat, S., and Fraser, M. P.: Characterization of saccharides and other organic compounds in fine particles
1019 and the use of saccharides to track primary biologically derived carbon sources, *Atmos. Environ.*, 44(5), 724–732,
1020 doi: 10.1021/es103104e, 2010.
- 1021 Jones, A. M. and Harrison, R. M.: The effects of meteorological factors on atmospheric bioaerosol
1022 concentrations—a review, *Sci. Total Environ.*, 326(1), 151–180, doi: 10.1016/j.scitotenv.2003.11.021, 2004.
- 1023 Karimi, B., Terrat, S., Dequiedt, S., Saby, N. P. A., Horrigue, W., Lelièvre, M., Nowak, V., Jolivet, C., Arrouays,
1024 D., Wincker, P., Cruaud, C., Bispo, A., Maron, P. A., Bouré, N. C. P., and Ranjard, L.: Biogeography of soil
1025 bacteria and archaea across France, *Sci. Adv.*, 4(7), eaat1808, doi:10.1126/sciadv.aat1808, 2018.
- 1026 Kaso, A.: Computation of the normalized cross correlation by fast Fourier transform, *PLOS ONE*, 13(9),
1027 e0203434, doi:10.1371/journal.pone.0203434, 2018.
- 1028 Kembel, S. W. and Mueller, R. C.: Plant traits and taxonomy drive host associations in tropical phyllosphere fungal
1029 communities, *Botany*, 92(4), 303–311, doi:10.1139/cjb-2013-0194, 2014.
- 1030 Kunit, M. and Puxbaum, H.: Enzymatic determination of the cellulose content of atmospheric aerosols, *Atmos.*
1031 *Environ.*, 30(8), 1233–1236, doi:10.1016/1352-2310(95)00429-7, 1996.
- 1032 Lecours, P. B., Duchaine, C., Thibaudon, M., and Marsolais, D.: Health impacts of bioaerosol exposure, in
1033 *Microbiology of Aerosols*, pp. 249–268, John Wiley & Sons, Ltd., doi:10.1002/9781119132318, 2017.
- 1034 Liang, L., Engling, G., He, K., Du, Z., Cheng, Y., and Duan, F.: Evaluation of fungal spore characteristics in
1035 Beijing, China, based on molecular tracer measurements, *Environ. Res. Lett.*, 8(1), 014005, doi:10.1088/1748-
1036 9326/8/1/014005, 2013.
- 1037 Liang, L., Engling, G., Du, Z., Cheng, Y., Duan, F., Liu, X., and He, K.: Seasonal variations and source estimation
1038 of saccharides in atmospheric particulate matter in Beijing, China, *Chemosphere*, 150, 365–377,
1039 doi:10.1016/j.chemosphere.2016.02.002, 2016.
- 1040 Lindow, S. E. and Brandl, M. T.: Microbiology of the phyllosphere, *Appl. Environ. Microbiol.*, 69(4), 1875–1883,
1041 doi:10.1128/AEM.69.4.1875-1883.2003, 2003.

- 1042 Lymeropoulou, D. S., Adams, R. I., and Lindow, S. E.: Contribution of vegetation to the microbial composition
 1043 of nearby outdoor air, edited by F. E. Löffler, *Appl. Environ. Microbiol.*, 82(13), 3822–3833,
 1044 doi:10.1128/AEM.00610-16, 2016.
- 1045 Manninen, H. E., Bäck, J., Sihto Nissilä, S. L., Huffman, J. A., Pessi, A. M., Hiltunen, V., Aalto, P. P., Hidalgo
 1046 Fernández, P. J., Hari, P., Saarto, A., Kulmala, M., and Petäjä, T.: Patterns in airborne pollen and other primary
 1047 biological aerosol particles (PBAP), and their contribution to aerosol mass and number in a boreal forest, *Boreal
 1048 Environ. Res.*, 383–405, doi:hdl.handle.net/10138/165208, 2014.
- 1049 Medeiros, P. M., Fernandes, M. F., Dick, R. P., and Simoneit, B. R. T.: Seasonal variations in sugar contents and
 1050 microbial community in a ryegrass soil, *Chemosphere*, 65(5), 832–839, doi:10.1016/j.chemosphere.2006.03.025,
 1051 2006a.
- 1052 Medeiros, P. M., Conte, M. H., Weber, J. C., and Simoneit, B. R. T.: Sugars as source indicators of biogenic
 1053 organic carbon in aerosols collected above the howland experimental forest, Maine, *Atmos. Environ.*, 40(9), 1694–
 1054 1705, 2006b.
- 1055 Meisner, A., Jacquiod, S., Snoek, B. L., ten Hooven, F. C., and van der Putten, W. H.: Drought legacy effects on
 1056 the composition of soil fungal and prokaryote communities, *Front. Microbiol.*, 9, doi:10.3389/fmicb.2018.00294,
 1057 2018.
- 1058 Mhuireach, G., Johnson, B. R., Altrichter, A. E., Ladau, J., Meadow, J. F., Pollard, K. S., and Green, J. L.: Urban
 1059 greenness influences airborne bacterial community composition, *Sci. Total Environ.*, 571, 680–687,
 1060 doi:10.1016/j.scitotenv.2016.07.037, 2016.
- 1061 Moricca, S. and Ragazzi, A.: The holomorph *apiognomonina quercina*/*Diseula quercina* as a pathogen/endophyte
 1062 in oak, in *Endophytes of forest trees: biology and applications*, edited by A. M. Pirttilä and A. C. Frank, pp. 47–
 1063 66, Springer Netherlands, Dordrecht., doi:10.1007/978-94-007-1599-8, 2011.
- 1064 Morris, C. E., Sands, D. C., Bardin, M., Jaenicke, R., Vogel, B., Leyronas, C., Ariya, P. A., and Psenner, R.:
 1065 Microbiology and atmospheric processes: research challenges concerning the impact of airborne micro-organisms
 1066 on the atmosphere and climate, *Biogeosciences*, 8(1), 17–25, doi:10.5194/bg-8-17-2011, 2011.
- 1067 Morris, C. E., Conen, F., Alex Huffman, J., Phillips, V., Pöschl, U., and Sands, D. C.: Bioprecipitation: a feedback
 1068 cycle linking Earth history, ecosystem dynamics and land use through biological ice nucleators in the atmosphere,
 1069 *Glob. Change Biol.*, 20(2), 341–351, doi:10.1111/geb.12447, 2014.
- 1070 Nirmalkar, J., Deshmukh, D. K., Deb, M. K., Tsai, Y. I., and Pervez, S.: Characteristics of aerosol during major
 1071 biomass burning events over eastern central India in winter: a tracer based approach, *Atmos. Pollut. Res.*,
 1072 doi:10.1016/j.apr.2018.12.010, 2018.
- 1073 Pashynska, V., Vermeylen, R., Vas, G., Maenhaut, W., and Claeys, M.: Development of a gas chromatographic/ion
 1074 trap mass spectrometric method for the determination of levoglucosan and saccharidic compounds in atmospheric
 1075 aerosols. Application to urban aerosols, *J. Mass Spectrom.*, 37(12), 1249–1257, doi:10.1002/jms.391, 2002.
- 1076 Pietrogrande, M. C., Bacco, D., Visentin, M., Ferrari, S., and Casali, P.: Polar organic marker compounds in
 1077 atmospheric aerosol in the Po valley during the supersito campaigns — part 2: seasonal variations of sugars,
 1078 *Atmos. Environ.*, 97, 215–225, doi:10.1016/j.atmosenv.2014.07.056, 2014.
- 1079 Pindado, O. and Perez, R. M.: Source apportionment of particulate organic compounds in a rural area of Spain by
 1080 positive matrix factorization, *Atmos. Pollut. Res.*, 2(4), 492–505, doi:10.5094/APR.2011.056, 2011.
- 1081 Puxbaum, H. and Tenze Kunit, M.: Size distribution and seasonal variation of atmospheric cellulose, *Atmos.
 1082 Environ.*, 37(26), 3693–3699, doi:10.1016/S1352-2310(03)00451-5, 2003.
- 1083 Rajput, P., Chauhan, A. S., and Gupta, T.: Bioaerosols over the indo-gangetic plain: influence of biomass burning
 1084 emission and ambient meteorology, in *Environmental Contaminants: measurement, modelling and control*, edited
 1085 by T. Gupta, A. K. Agarwal, R. A. Agarwal, and N. K. Labhsetwar, pp. 93–121, Springer Singapore, Singapore.,
 1086 doi:10.1007/978-981-10-7332-8-2018.

- 1087 Ram, K., Sarin, M. M., and Hegde, P.: Long term record of aerosol optical properties and chemical composition
1088 from a high altitude site (Manora Peak) in central Himalaya, *Atmos. Chem. Phys.*, 13, doi:10.5194/acp-10-11791-
1089 2010, 2010.
- 1090 Ramoni, J. and Seiboth, B.: Degradation of plant cell wall polymers by fungi, in *Environmental and Microbial*
1091 *Relationships*, vol. IV, edited by I. S. Druzhinina and C. P. Kubicek, pp. 127–148, Springer International
1092 Publishing, Cham., doi: 10.1007/978-3-540-71840-6, 2016.
- 1093 Rathnayake, C. M., Metwali, N., Jayarathne, T., Kettler, J., Huang, Y., Thorne, P. S., O’Shaughnessy, P. T., and
1094 Stone, E. A.: Influence of rain on the abundance of bioaerosols in fine and coarse particles, *Atmos. Chem. Phys.*,
1095 17(3), 2459–2475, doi: 10.5194/acp-17-2459-2017, 2017.
- 1096 Reddy, S. M., Girisham, S., and Babu, G. N.: *Applied Microbiology (agriculture, environmental, food and*
1097 *industrial microbiology)*, Scientific Publishers, doi:9789387307407, 2017.
- 1098 Rogge, W. F., Medeiros, P. M., and Simoneit, B. R. T.: Organic marker compounds in surface soils of crop fields
1099 from the San Joaquin Valley fugitive dust characterization study, *Atmos. Environ.*, 41(37), 8183–8204,
1100 doi:10.1016/j.atmosenv.2007.06.030, 2007.
- 1101 Samaké, A., Jaffrezo, J. L., Favez, O., Weber, S., Jacob, V., Albinet, A., Riffault, V., Perdrix, E., Waked, A.,
1102 Golly, B., Salameh, D., Chevrier, F., Oliveira, D. M., Bonnaire, N., Besombes, J. L., Martins, J. M. F., Conil, S.,
1103 Guillaud, G., Mesbah, B., Rocq, B., Robic, P. Y., Hulin, A., Meur, S. L., Descheemaeker, M., Chretien, E.,
1104 Marehand, N., and Uzu, G.: Polyols and glucose particulate species as tracers of primary biogenic organic aerosols
1105 at 28 French sites, *Atmos. Chem. Phys.*, 19(5), 3357–3374, doi:10.5194/acp-19-3357-2019, 2019.
- 1106 Sánchez-Ochoa, A., Kasper-Giebl, A., Puxbaum, H., Gelencser, A., Legrand, M., and Pio, C.: Concentration of
1107 atmospheric cellulose: A proxy for plant debris across a west-east transect over Europe, *J. Geophys. Res.*,
1108 112(D23), doi:10.1029/2006JD008180, 2007.
- 1109 Sesartic, A. and Dallafior, T. N.: Global fungal spore emissions, review and synthesis of literature data,
1110 *Biogeosciences*, 8(5), 1181–1192, doi:10.5194/bg-8-1181-2011, 2011.
- 1111 Sheherbakova, L. A.: Advanced methods of plant pathogen diagnostics, in *Comprehensive and molecular*
1112 *phytopathology*, edited by Yu. T. Dyakov, V. G. Dzhavakhiya, and T. Korpela, pp. 75–116, Elsevier, Amsterdam,
1113 doi:9780080469331, 2007.
- 1114 Simoneit, B. R. T., Kobayashi, M., Mochida, M., Kawamura, K., Lee, M., Lim, H. J., Turpin, B. J., and Komazaki,
1115 Y.: Composition and major sources of organic compounds of aerosol particulate matter sampled during the ACE-
1116 Asia campaign, *J. Geophys. Res.*, 109(D19S10), doi:10.1029/2004JD004598, 2004a.
- 1117 Simoneit, B. R. T., Elias, V. O., Kobayashi, M., Kawamura, K., Rushdi, A. I., Medeiros, P. M., Rogge, W. F., and
1118 Didyk, B. M.: Sugars dominant water soluble organic compounds in soils and characterization as tracers in
1119 atmospheric particulate matter, *Environ. Sci. Technol.*, 38(22), 5939–5949, 2004b.
- 1120 Srivastava, D., Favez, O., Bonnaire, N., Lucarelli, F., Haefelin, M., Perraudin, E., Gros, V., Villenave, E., and
1121 Albinet, A.: Speciation of organic fractions does matter for aerosol source apportionment — part 2: intensive short-
1122 term campaign in the Paris area (France), *Sci. Total Environ.*, 634, 267–278, doi:10.1016/j.scitotenv.2018.03.296,
1123 2018.
- 1124 Sullivan, A. P., Frank, N., Kenski, D. M., and Collett, J. L.: Application of high performance anion exchange
1125 chromatography pulsed amperometric detection for measuring carbohydrates in routine daily filter samples
1126 collected by a national network 2: examination of sugar alcohols/polyols, sugars, and anhydrosugars in the upper
1127 Midwest, *J. Geophys. Res. Atmospheres*, 116(D8), D08303, doi:10.1029/2010JD014169, 2011.
- 1128 Tanarhte, M., Bacer, S., Burrows, S. M., Huffman, J. A., Pierce, K. M., Pozzer, A., Sarda-Estève, R., Savage, N.
1129 J., and Lelieveld, J.: Global modeling of fungal spores with the EMAC chemistry-climate model: uncertainties in
1130 emission parametrizations and observations, *Atmos. Chem. Phys. Discuss.*, 1–31, doi:10.5194/acp-2019-251,
1131 2019.

- 1132 Véléz, H., Glassbrook, N. J., and Daub, M. E.: Mannitol metabolism in the phytopathogenic fungus *alternaria*
 1133 *alternata*, *Fung. Genet. Biol.*, 44(4), 258–268, doi:10.1016/j.fgb.2006.09.008, 2007.
- 1134 Verma, S. K., Kawamura, K., Chen, J., and Fu, P.: Thirteen years of observations on primary sugars and sugar
 1135 alcohols over remote Chichijima Island in the western north Pacific, *Atmos. Chem. Phys.*, 18(1), 81–101,
 1136 doi:10.5194/acp-18-81-2018, 2018.
- 1137 Vlachou, A., Daellenbach, K. R., Bozzetti, C., Chazeau, B., Salazar, G. A., Szidat, S., Jaffrezo, J. L., Hueglin, C.,
 1138 Baltensperger, U., Haddad, I. E., and Prévôt, A. S. H.: Advanced source apportionment of carbonaceous aerosols
 1139 by coupling offline AMS and radiocarbon size segregated measurements over a nearly 2-year period, *Atmos.*
 1140 *Chem. Phys.*, 18(9), 6187–6206, doi:10.5194/acp-18-6187-2018, 2018.
- 1141 Waked, A., Favez, O., Alleman, L. Y., Piot, C., Petit, J. E., Delaunay, T., Verlinden, E., Golly, B., Besombes, J.-
 1142 L., Jaffrezo, J. L., and Leoz-Garziandia, E.: Source apportionment of PM₁₀ in a north-western Europe regional
 1143 urban background site (Lens, France) using positive matrix factorization and including primary biogenic
 1144 emissions, *Atmos. Chem. Phys.*, 14(7), 3325–3346, doi:10.5194/acp-14-3325-2014, 2014.
- 1145 Wan, E. C. H. and Yu, J. Z.: Analysis of sugars and sugar polyols in atmospheric aerosols by chloride attachment
 1146 in liquid chromatography/negative ion electrospray mass spectrometry, *Environ. Sci. Technol.*, 41(7), 2459–2466,
 1147 doi:10.1021/es062390g, 2007.
- 1148 Wan, X., Kang, S., Rupakheti, M., Zhang, Q., Tripathee, L., Guo, J., Chen, P., Rupakheti, D., Panday, A. K.,
 1149 Lawrence, M. G., Kawamura, K., and Cong, Z.: Molecular characterization of organic aerosols in the Kathmandu
 1150 Valley, Nepal: insights into primary and secondary sources, *Atmos. Chem. Phys.*, 19(5), 2725–2747,
 1151 doi:10.5194/acp-19-2725-2019, 2019.
- 1152 Weber, S., Uzu, G., Calas, A., Chevrier, F., Besombes, J. L., Charron, A., Salameh, D., Ježek, I., Močnik, G., and
 1153 Jaffrezo, J. L.: An apportionment method for the oxidative potential of atmospheric particulate matter sources:
 1154 application to a one-year study in Chamonix, France, *Atmos. Chem. Phys.*, 18(13), 9617–9629, doi:10.5194/acp-
 1155 18-9617-2018, 2018.
- 1156 Wéry, N., Galès, A., and Brunet, Y.: Bioaerosol sources, in *Microbiology of Aerosols*, pp. 115–135, John Wiley
 1157 & Sons, Ltd., doi:10.1002/9781119132318, 2017.
- 1158 Whipps, J. M., Hand, P., Pink, D., and Bending, G. D.: Phyllosphere microbiology with special reference to
 1159 diversity and plant genotype, *J. Appl. Microbiol.*, 105(6), 1744–1755, doi:10.1111/j.1365-2672.2008.03906.x,
 1160 2008.
- 1161 Xu, J., He, J., Xu, H., Ji, D., Snape, C., Yu, H., Jia, C., Wang, C., and Gao, J.: Simultaneous measurement of
 1162 multiple organic tracers in fine aerosols from biomass burning and fungal spores by HPLC-MS/MS, *RSC Adv.*,
 1163 8(59), 34136–34150, doi:10.1039/C8RA04991B, 2018.
- 1164 Yan, C., Sullivan, A. P., Cheng, Y., Zheng, M., Zhang, Y., Zhu, T., and Collett, J. L.: Characterization of
 1165 saccharides and associated usage in determining biogenic and biomass burning aerosols in atmospheric fine
 1166 particulate matter in the North China Plain, *Sci. Total Environ.*, 650, 2939–2950,
 1167 doi:10.1016/j.scitotenv.2018.09.325, 2019.
- 1168 Yan, K., Park, T., Yan, G., Chen, C., Yang, B., Liu, Z., Nemani, R., Knyazikhin, Y., and Myneni, R.: Evaluation
 1169 of MODIS LAI/FPAR product collection 6. part 1: consistency and improvements, *Remote Sens.*, 8(5), 359,
 1170 doi:10.3390/rs8050359, 2016a.
- 1171 Yan, K., Park, T., Yan, G., Liu, Z., Yang, B., Chen, C., Nemani, R., Knyazikhin, Y., and Myneni, R.: Evaluation
 1172 of MODIS LAI/FPAR product collection 6. part 2: validation and intercomparison, *Remote Sens.*, 8(6), 460,
 1173 doi:10.3390/rs8060460, 2016b.
- 1174 Yoo, J. C. and Han, T. H.: Fast normalized cross-correlation, *Circuits Syst. Signal Process.*, 28(6), 819–843,
 1175 doi:10.1007/s00034-009-9130-7, 2009.
- 1176 Yttri, K. E., Dye, C., and Kiss, G.: Ambient aerosol concentrations of sugars and sugar alcohols at four different
 1177 sites in Norway, *Atmos. Chem. Phys.*, 7(16), 4267–4279, doi:10.5194/acp-7-4267-2007, 2007.

1178 Yttri, K. E., Simpson, D., Stenström, K., Puxbaum, H., and Svendby, T.: Source apportionment of the
1179 carbonaceous aerosol in Norway—quantitative estimates based on ^{14}C , thermal optical and organic tracer
1180 analysis, *Atmos. Chem. Phys.*, 11(3), 7375–7422, doi:10.5194/acpd-11-7375-2011, 2011a.

1181 Yttri, K. E., Simpson, D., Nøjgaard, J. K., Kristensen, K., Genberg, J., Stenström, K., Swietlicki, E., Hillamo, R.,
1182 Aurela, M., Bauer, H., Offenberg, J. H., Jaoui, M., Dye, C., Eckhardt, S., Burkhardt, J. F., Stohl, A., and Glasius,
1183 M.: Source apportionment of the summer time carbonaceous aerosol at Nordic rural background sites, *Atmos.*
1184 *Chem. Phys.*, 11(24), 13339–13357, doi:10.5194/acp-11-13339-2011, 2011b.

1185 Yue, S., Ren, H., Fan, S., Wei, L., Zhao, J., Bao, M., Hou, S., Zhan, J., Zhao, W., Ren, L., Kang, M., Li, L., Zhang,
1186 Y., Sun, Y., Wang, Z., and Fu, P.: High abundance of fluorescent biological aerosol particles in winter in Beijing,
1187 China, *ACS Earth Space Chem.*, 1(8), 493–502, doi:10.1021/acsearthspacechem.7b00062, 2017.

1188 Zhang, T., Engling, G., Chan, C. Y., Zhang, Y. N., Zhang, Z. S., Lin, M., Sang, X. F., Li, Y. D., and Li, Y. S.:
1189 Contribution of fungal spores to particulate matter in a tropical rainforest, *Environ. Res. Lett.*, 5(2), 024010,
1190 doi:10.1088/1748-9326/5/2/024010, 2010.

1191 Zhang, Z., Engling, G., Zhang, L., Kawamura, K., Yang, Y., Tao, J., Zhang, R., Chan, C., and Li, Y.: Significant
1192 influence of fungi on coarse carbonaceous and potassium aerosols in a tropical rainforest, *Environ. Res. Lett.*,
1193 10(3), 034015, doi:10.1088/1748-9326/10/3/034015, 2015.

1194 Zhu, C., Kawamura, K., and Kunwar, B.: Organic tracers of primary biological aerosol particles at subtropical
1195 Okinawa Island in the western north pacific Rim: organic biomarkers in the north pacific, *J. Geophys. Res. Atmos.*,
1196 120(11), 5504–5523, 2015.

1197 Zhu, C., Kawamura, K., Fukuda, Y., Mochida, M., and Iwamoto, Y.: Fungal spores overwhelm biogenic organic
1198 aerosols in a midlatitudinal forest, *Atmos. Chem. Phys.*, 16(11), 7497–7506, doi:10.5194/acp-16-7497-2016, 2016.

1199 Zhu, W., Cheng, Z., Luo, L., Lou, S., Ma, Y., and Yan, N.: Investigation of fungal spore characteristics in $\text{PM}_{2.5}$
1200 through organic tracers in Shanghai, China, *Atmos. Pollut. Res.*, 9(5), 894–900, doi:10.1016/j.apr.2018.01.009,
1201 2018.

1202