Polyols and glucose particulate species as tracers of primary biogenic organic aerosols at 28 French sites

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Abstract. A growing number of studies is using specific primary sugar species, such as sugar alcohols or primary saccharides, as marker compounds to characterize and apportion primary biogenic organic aerosols (PBOA) in the atmosphere. To better understand their annual cycles, as well as their spatio-temporal abundance in terms of concentrations and sources, we conducted a large study focusing on three major atmospheric primary sugar compounds (i.e., arabitol, mannitol and glucose) measured in various environmental conditions on about 5,300 filter samples collected at 28 sites in France. Our results show significant atmospheric concentrations of polyols (defined here as the sum of arabitol and mannitol) and glucose at each sampling location, highlighting their ubiquity. Results also confirm that polyols and glucose are mainly associated with the coarse rather than the fine aerosol mode. At nearly all sites, atmospheric concentrations of polyols and glucose display a well-marked seasonal pattern, with maximum concentrations from late spring to early autumn, followed by an abrupt decrease in late autumn, and a minimum concentration during wintertime. Such seasonal patterns support biogenic emissions associated with higher biological metabolic activities (e.g., sporulation, growth, etc.) during warmer periods. Results from a previous comprehensive study using Positive Matrix Factorization (PMF) based on an extended aerosol chemical composition dataset of up to 130 species for 16 of the same sample series has also been used in the present work. The Polyols-to-PM_{PBOA} ratio is 0.024 ± 0.010 on average for all sites, with no clear distinction between traffic, urban or rural typology. Overall, even if the exact origin of the PBOA source is still under investigation, it appears to be an important source of PM, especially during summertime. Results also show that PBOA are significant sources of total OM in PM_{10} (13 ± 4 % on a yearly average, and up to 40 % in some environments in summer) at most of the investigated sites. The mean PBOA chemical profile is clearly dominated by contribution from organic matter (OM) (78 ± 9 % of the mass of the PBOA PMF factor on average), and only a minor contribution from dust class (3 ± 4 %), suggesting that ambient polyols are most likely associated with biological particle emissions (e.g., active spore discharge) rather than soil dust resuspension.

1. Introduction

Airborne particles (or particulate matter, PM) are of major concern due to their multiple effects on climate and adverse human health impacts (Boucher et al., 2013; Cho et al., 2005; Ntziachristos et al., 2007). The diversity of PM impacts is closely linked to their complex and highly variable nature: size distribution, concentration and chemical composition, or specific surface properties. PM consists of inorganic and elemental substances, and a large fraction made of carbonaceous matter (organic carbon (OC) and elemental carbon (EC)) (Franke et al., 2017; Putaud et al., 2004a; Yttri et al., 2007a). Substantial amounts of atmospheric organic matter (OM) remain unidentified and uncharacterized at the molecular level. In most studies, a maximum of only 20 % of particulate OM mass can generally be speciated and quantified (Alfarra et al., 2007; Fortenberry et al., 2018; Liang et al., 2017; Nozière et al., 2015). This detailed composition of OM and its spatial and seasonal distribution can give important insights on the adverse effects of PM. So far, the majority of air pollution studies have focused on organic atmospheric particles associated with anthropogenic and secondary sources, whereas a significant fraction of OM can also be associated with primary emissions from biogenic sources (Bauer et al., 2008a; Jaenicke, 2005; Liang et al., 2016). Therefore, the characterization of primary OM biogenic sources at the molecular level is still limited (Fuzzi et al., 2006; Liang et al., 2017; Zhu et al., 2015), and should be further investigated for a better understanding of aerosol sources and formation processes.

71 Primary biogenic organic aerosols (PBOA) are emitted directly from the biosphere to the atmosphere where they

72 are ubiquitous and participate in many atmospheric processes (Elbert et al., 2007; Fröhlich-Nowoisky et al., 2016).

73 Additionally, their inhalation has long been associated with human respiratory impairments (e.g., asthma,

74 aspergillosis, etc.) (Després et al., 2012; Morris et al., 2011). PBOA comprise living and dead microorganisms

75 such as bacteria, fungi, viruses, bacterial and fungal spores, and microbial fragments, endotoxins, mycotoxins, or

76 pollens (Elbert et al., 2007; Jaenicke, 2005; Morris et al., 2011). In most semi-urban European sites, PBOA can

77 account for up to 25 % of the atmospheric aerosol mass, in the size range of 0.2 to 50 µm (Fröhlich-Nowoisky et

78 al., 2016; Jaenicke, 2005; Huffman et al., 2012; Manninen et al., 2014; Morris et al., 2011). However, their sources

79 and contribution to total airborne particles are still poorly documented, partly because of the difficulty to recognize

80 them by conventional microbiological methods (cell culture, microscopic examination, etc.) (Di Filippo et al., 2013;

81 Heald and Spracklen, 2009; Jia et al., 2010a).

82 Several specific chemical components, such as primary sugar compounds (i.e., primary saccharides and sugar

83 alcohols) emitted persistently from biogenic sources, have long been suggested as powerful and unique biomarkers

84 in tracing sources, and abundances of PBOA as well (Bauer et al., 2008a; Medeiros et al., 2006; Simoneit et al.,

85 2004b; Zhang et al., 2010; Zhu et al., 2016). For instance, ambient concentrations of glucose have been used as

86 markers for plant materials (such as pollen, leaves, and their fragments) or soil emissions from several areas in the

87 world (Fu et al., 2012; Jia et al., 2010a, 2010b; Pietrogrande et al., 2014; Rathnayake et al., 2017). Many studies

88 indicated that glucose is the most abundant monosaccharide in vascular plants, where it serves as the common

89 energy material, and an important source of carbon for soil active microorganisms (such as bacteria or fungi) (Jia

90 et al., 2010a; Medeiros et al., 2006; Pietrogrande et al., 2014; Zhu et al., 2015). Additionally, sugar alcohols (also

91 called polyols) including arabitol and mannitol have been proposed as markers for airborne fungi, and are widely

92 used to quantify their contributions to PBOA mass (Bauer et al., 2008a, 2008b; Golly et al., 2018; Srivastava et

93 al., 2018; Zhang et al., 2010). These sugar alcohols have also been found to correlate very well with fluorescent

94 PBOA in the ultraviolet aerodynamic particle sizer (UV-APS) and wideband integrated bioaerosol sensor (WIBS-

95 3) online studies, particularly in rainy periods (Gosselin et al., 2016), favoring microbial sporulation (such as fungi

96 belonging to Ascomycota and Basidiomycota phyla) (China et al., 2016; Elbert et al., 2007; Jones and Harrison,

97 2004). Polyols are produced in large amounts by many fungi and bacteria, and several functions have been

98 described for these compounds, such as common energy storage materials, intracellular protectants against

99 stressful conditions (e.g., heat or drought), storage or transport of carbohydrates, quencher of oxygenated reactive

100 species, or regulators of intracellular pH by acting as a sink or source of protons (Jennings et al., 1998; Medeiros

101 et al., 2006; Vélëz et al., 2007). Hence, polyols, especially arabitol and mannitol, may represent a significant

102 fraction of the dry weight of fungi, and mannitol can contribute between 20 to 50 % of the mycelium dry weight

103 (Ruijter et al., 2003; Vélëz et al., 2007). However, polyols are also often identified in the lower plants (leaves,

104 pollens) and green algal lichens (Medeiros et al., 2006; Vélëz et al., 2007; Yang et al., 2012). The primary sugar

105 compounds (defined as polyols and primary saccharide species) are thought to be relatively stable in the

106 atmosphere (Wang et al., 2018), although studies investigating their atmospheric lifetime are quite limited. One

107 previous laboratory study has been conducted by the US-EPA to evaluate the stability of these chemicals on filter

108 material exposed to gaseous oxidants as well as in aqueous solutions (simulating clouds and fog droplet chemistry).

109 Findings of this former study have shown that primary sugar compounds remain quite stable up to 7 days (the

110   extent of the testing period), pointing out their suitability for use as tracers of atmospheric transport (Fraser, 2010).
111   With all of this information, the use of primary sugar compounds (such as mannitol, arabitol, glucose, etc.) as
112   suitable tracers of PBOA is generally acknowledged (Jia and Fraser, 2011; Zhu et al., 2015, 2016).

113   Although atmospheric concentrations of polyols, including arabitol and mannitol, as well as that of some primary
114   monosaccharides (e.g., glucose), have been previously quantified as part of several studies in various environments
115   including urban/suburban, rural, rainforest, mountain, and marine areas (Fu et al., 2012; Graham et al., 2003; Jia
116   et al., 2010a, 2010b; Liang et al., 2016; Pietrogrande et al., 2014; Simoneit, 2004a, 2004b; Verma et al., 2018;
117   Yttri et al., 2007b; Zhu et al., 2015), large datasets investigating their annual cycles and spatial distributions are
118   still limited. Such information could give important insights on environmental factors influencing their
119   atmospheric levels such as climate and biotope, and therefore help to elucidate patterns regarding their major
120   sources and atmospheric emission pathways. Even if numerous sources and emission mechanisms have been
121   widely proposed, including among others, metabolic active microbial wet emissions, entrainment of farmland or
122   natural soils and associated microbiota (Elbert et al., 2007; Fu et al., 2013; Gosselin et al., 2016; Jia et al., 2010a,
123   2010b; Medeiros et al., 2006; Pietrogrande et al., 2014; Simoneit et al., 2004a, 2004b; Verma et al., 2018; Yttri
124   et al., 2007b), the dominant atmospheric input processes have not been yet sufficiently elucidated.

125   In this context, the present study was designed to provide a large overview of the spatial and seasonal variations
126   of polyols and glucose mass concentrations, as well as their contribution to the aerosol organic mass fraction in
127   France. To do so, data was collected at many sites in different environments (rural, traffic, urban), in order to
128   represent various sampling conditions in terms of site typologies and meteorological conditions. Thanks to the
129   availability of results from an extended Positive Matrix Factorization (PMF) analysis performed for the
130   corresponding datasets, the overall contributions of the main polyols and glucose emission sources could also be
131   investigated in light of their spatial patterns. To the best of our knowledge, this is the first study providing such an
132   extended phenomenology of these compounds over multiple sites with different typologies.

133   2. Material and methods

134   2.1 Aerosol sampling

135   Ambient aerosol samples considered in the present work come from different research and monitoring programs,
136   conducted over the last 5 years in France (Figure 1). Each program includes at least one-year of field sampling,
137   providing a total number of 5,343 daily filter samples available for the sake of the present study. These sites offer
138   diverse conditions in terms of typologies (i.e., rural, traffic, urban sites, Alpine valley environments, etc.), local
139   climate and vegetation types and were selected in order to cover the complex and variable national environmental
140   conditions. These sites are assumed to represent typical environmental conditions in France, and our
141   observations/and general tendency could therefore be extrapolated to neighboring western European countries
142   presenting quite homogeneous environmental conditions.

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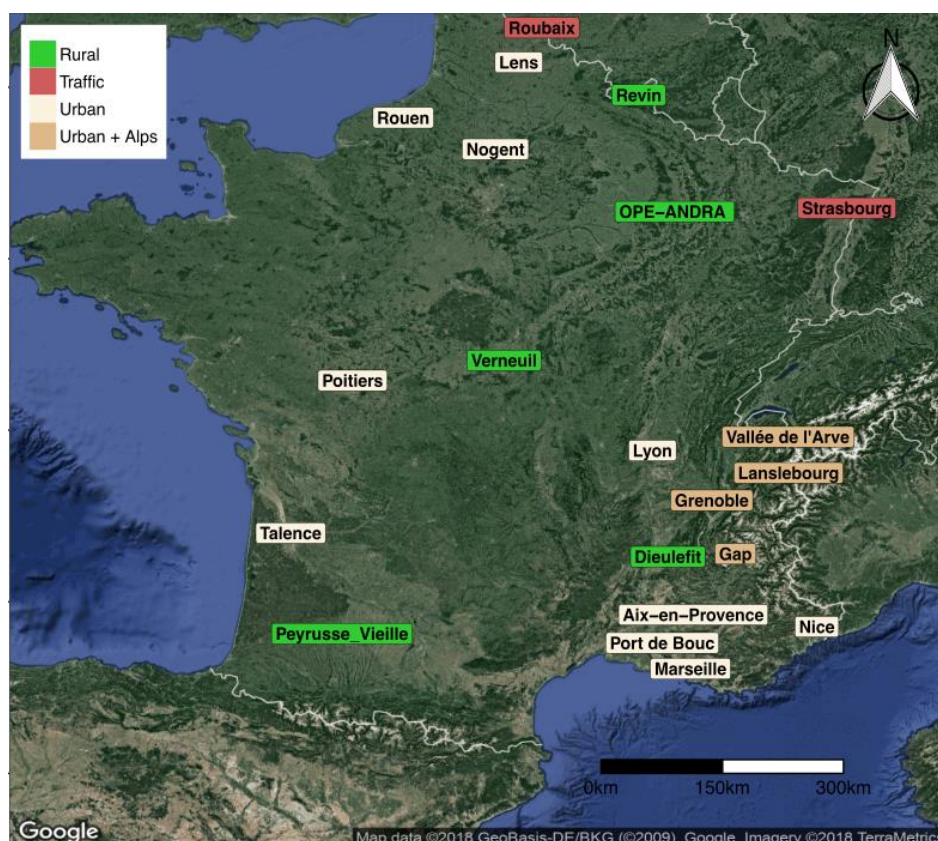
144 **Figure 1: Map of sampling site locations in France. Green: rural background, red: traffic, wheat: urban background**
145 **and dark wheat: urban background in Alpine valley sites. The areas of Grenoble (Grenoble_LF, Grenoble_CB and**
146 **Grenoble_VIF) and Vallée de L'Arve (Marnaz, Passy and Chamonix) include 3 sites each. The area of Marseille**
147 **includes four sites: Marseille, Mallet, Meyreuil and Gardanne.**

148 The site of OPE-ANDRA is a specific monitoring observatory in a rural environment, without any village or

149 industry within several kilometers (description available from: http://www.andra.fr). All other sites correspond to

150 stations of regional air quality monitoring networks (AASQA). The availability of filter samples was variable from

151 one site to the other one, depending on the sampling frequency (typically every third or sixth day). Filter collection

152 was conducted within the PM_{10} or the $PM_{2.5}$ aerosol size fraction, depending on the investigated site (Table SI-1).

153 Moreover, co-located and simultaneous PM_{10} and $PM_{2.5}$ samplings were conducted at OPE-ANDRA and Revin,

154 allowing to investigate the distribution of primary sugar compounds between the fine and the coarse aerosol size

155 fractions at these two sites.

156 Ambient aerosols were collected onto quartz fiber filters (Tissu-quartz PALL QAT-UP 2500 150 mm diameter),

157 preheated at 500 °C for 4 h minimum before use. After collection, all filter samples were wrapped in aluminum

158 foils, sealed in zipper plastic bags and stored at <4°C until further chemical analysis. Field blank filters were also

159 collected, at least once a month, using the same handling procedure as for PM samples. More detailed information

160 on the sampling periods, air sampler, number of filters and nature of PM samples are provided in Table SI-1 and

161 Fig. SI-1.

162 **2.2 Chemical analyses**

163 PM samples were analyzed for various chemical species using sub-sampled fractions of the collection filters. In

164 the frame of the present study, the carbonaceous matter (organic carbon (OC) and elementary carbon (EC)) was

165 analyzed using a thermo-optical method on a Sunset Lab analyzer (Birch and Cary, 1996) as described by Aymoz

166   et al. (2007), using the EUSAAR2 temperature program (Cavalli et al., 2010), except for the five sites of Northern

167   France where the NIOSH870 protocol was employed (Birch and Cary, 1996). OM contents were then estimated

168   by multiplying the organic carbon mass concentrations by a fixed factor, with $OM = 1.8 \times OC$. This OM-to-OC

169   ratio value of 1.8 was chosen based on previous studies performed in France (Favez et al., 2010; Petit et al., 2015

170   and reference therein) and around the world. (e.g., Aiken et al., 2008; Li et al., 2018; Ruthenburg et al., 2014;

171   Vlachou et al., 2018), with a typical range of 1.2-2.4 values.

172   For the analysis of anhydrosugars, sugar alcohols, and primary saccharides, filter punches (typically of about 10

173   cm²) were first extracted into ultrapure water, then filtered using a 0.22 µm Acrodisc filter. Depending on the site,

174   analyses were conducted either by IGE (Institut des Géosciences de l'Environnement) or by LSCE (Laboratoire

175   des Sciences du Climat et de l'Environnement) (Table SI-1). At IGE, extraction was performed during 20 min in

176   a vortex shaker and analyses were achieved using High-Performance Liquid Chromatography (HPLC) with Pulsed

177   Amperometric Detection. A first set of equipment was used until March 2016, consisting of a Dionex DX500

178   equipped with three columns Metrosep (Carb 1-Guard + A Supp 15-150 + Carb 1-150), the analytical run being

179   isocratic with 70 mM sodium hydroxide eluent, followed by a gradient cleaning step with a 120 mM NaOH eluent.

180   This analytical technique enables to detect anhydrous saccharides (levoglucosan, mannosan, galactosan), polyols

181   (arabitol, sorbitol, mannitol), and glucose (Waked et al., 2014). A second set of equipment was used after this date,

182   with a Thermo-Fisher ICS 5000⁺ HPLC equipped with 4 mm diameter Metrosep Carb 2 × 150 mm column and

183   50 mm pre-column. The analytical run is isocratic with 15 % of an eluent of sodium hydroxide (200 mM) and

184   sodium acetate (4 mM) and 85 % water, at 1 mL min^{-1}. This method allows for additional separation and

185   quantification of erythritol, xylitol, and threalose. At LSCE, extraction was performed during 45 min by sonication

186   and analyses were achieved using an ion chromatography (IC) instrument (DX600, Dionex) with Pulsed

187   Amperometric Detection (ICS3000, Dionex). A CarboPAC MA1 column has be used (4 × 250 mm, Dionex), the

188   analytical run being isocratic with 480 mM sodium hydroxide eluent. This analytical technique enables to detect

189   anhydrous saccharides (levoglucosan, mannosan, galactosan), polyols (arabitol, mannitol), and glucose.

190   Field blank filters were handled as real samples for quality assurance. The present data were corrected with field

191   blanks. The reproducibility of the analysis of primary sugar species (polyols, glucose), estimated from the analysis

192   of extracts of 10 punches from the same filters is generally in the range of 10-15 %.

193   Additional chemical analyses were conducted for most of the sites, allowing to quantify up to 130 different

194   chemical species (Calas et al., 2018). 30- 35 chemical species were then selected in order to achieve PMF analyses

195   as discussed hereafter.

2.3 Statistical analysis

197   Species concentration measurements were first analyzed for normality using Shapiro-Wilk's method with the

198   statistical program *R studio interface* (version 3.4.1). Since data were generally not distributed normally, we used

199   non-parametric statistical methods. The strength of the relationship between species concentrations was

200   investigated using the non-parametric Spearman rank correlation method. Multiple mean comparison analyses

201   were performed with the Kruskall-Wallis test method. Statistical significance was set at $p < 0.05$.

202   Positive Matrix Factorization for the source apportionment of the PM was previously performed at several sites of

203   this study, as part of the SOURCES (Favez et al., 2016; Salameh et al., in prep.) and DECOMBIO (Chevrier, 2017)

204   projects. We used the US EPA PMF 5.0 software (US EPA, 2015), following the general recommendation

guidelines of the European Joint Research Centre (JRC) (Belis et al., 2014). Briefly, the SOURCES program aimed at performing source apportionment at 15 sites using a harmonized methodology, i.e., using the same chemical species, uncertainties, constraints, and criteria for factor identification. The PMF conducted within SOURCES project uses about 30 different species (Table SI-6), including carbonaceous fraction (OC, EC), ions (Cl$^-$, NO$_3^-$ SO$_4^{2-}$, NH$_4^+$ K$^+$, Mg^{2+}, Ca^{2+}), organic markers (polyols i.e., sum of arabitol, mannitol and sorbitol; levoglucosan; mannosan) and metals (Al, As, Ba, Cd, Co, Cs, Cu, Fe, La, Mn, Mo, Ni, Pb, Rb, Sb, Se, Sn, Sr, Ti, V, Zn). The PMF conducted within the DECOMBIO project, for the sites of Marnaz, Chamonix, and Passy, used aethalometer (AE 33) measurements instead of EC (Chevrier, 2016). This complementary measure gives the total black carbon (BC), thus enabling the deconvolution of BC concentrations into its two main constituents: wood-burning BC (BC$_{wb}$) and fossil-fuel BC (BC$_{ff}$) (Sandradewi et al., 2008). For graphical simplicity, BC$_{wb}$ and BC$_{ff}$ were summed up and labeled as EC in the following Figures. PMF modelling was performed separately for each site. Statistical significance was validated with bootstrap higher than 80 % for each factor. Detailed methodology and results about these studies are given in their respective papers (Chevrier, 2017; Favez et al., 2016; Salameh et al., in prep.). It should be noted that glucose was not included in the final solution for any of these PMF, since it generally produced statistical instability of the solutions (this point is further discussed in Sect. 3.2).

The PMF analysis took advantage of the ME-2 algorithm to add constraints to different chemical profiles (see Tables SI-3 and SI-4 for details). Mainly soft constraints were applied in order to add some prior knowledge about the emission sources and "clean" the different profiles without forcing the model toward an explicit solution. In particular, the polyol concentrations were "pulled up maximally", while levoglucosan and mannosan were set to zero, and EC was "pulled down maximally" in the PBOA factor. This was achieved to avoid mixing with the biomass burning factor as well as possible influences of unrealistic high contributions of EC to PBOA. Other constraints were added parsimoniously to other factors, targeting specific proxies of sources (Table SI-4).

As for the general results of this large PMF study, we identified some well-known sources for almost all the sites (biomass-burning, road traffic, secondary inorganics, dust and sea salt). Two other less-common factors were identified for all sites: secondary biogenic aerosols (probably from marine origin), traced mainly by the presence of MSA, and PBOA, traced by the presence of more than 90% of the polyols total mass in the factor. Table SI-5 and Fig. SI-4 present more detailed description of the chemical tracers in each factor, together with their yearly average contribution for each site, respectively. Hereafter, only the PBOA chemical profile will be extensively investigated. The uncertainties of this PBOA factor are discussed below and its stability is presented in Fig. SI-5. Bootstrap analysis based on 100 resampling runs evidenced the very high stability of this PBOA factor since the PBOA initial constrained factor was mapped to PBOA bootstrap factor (BF) more than 99% of the time.

3. Results and discussion

3.1 Relative distribution between sugar alcohols and glucose

Figure 2 presents an overview of the relative mass concentration distributions of individual chemical species quantified at two sites with very different characteristics, an urban site in Grenoble and the rural site of OPE-ANDRA. Data are presented for the warmer season (e.g., during summer and fall), when concentrations were at their maximum (see Sect. 3.4). Glucose is the most abundant species measured (average 37.6±26.4 ng m^{-3}), accounting on average for 25 % of primary sugar compounds (SC) total mass at both sites. Mannitol

243   (37.3±24.6 ng m^{-3}) and arabitol (32.0±22.2 ng m^{-3},) are the second and third most abundant species, accounting

244   respectively for 25 and 23 % of SC mass. Trehalose is relatively abundant in samples from these two sites

245   (20.1±16.2 ng m^{-3}), accounting for 14 % of SC mass, but in general its concentration is frequently below the limit

246   of quantification for samples from other sites in France. The other identified polyols (i.e., erythritol, inositol,

247   glycerol, sorbitol, and xylitol) present lower concentration levels (4.9±2.1 ng m^{-3}), corresponding altogether to

248   13 % of SC total mass.

249   Such ambient mass concentration distribution patterns are similar (but with variable intensities) to those previously

250   reported for aerosol samples collected at various locations around the world. For example, Verma et al. (2018)

251   found that glucose, and arabitol together with mannitol, contributed to 16.7 and 48.1 %, respectively, of total

252   primary sugar compounds in aerosols from Chichijima Island. Similarly, Yttri et al. (2007b) showed that glucose

253   and the pair arabitol-mannitol were the main contributors of total primary monosaccharides and sugar alcohols in

254   aerosols collected from four various background sites in Norway. In addition, Carvalho et al. (2003) reported that

255   arabitol, mannitol and glucose are the most dominant primary sugar compounds in aerosols from rural background

256   and boreal forest sites in Germany and Finland, respectively.

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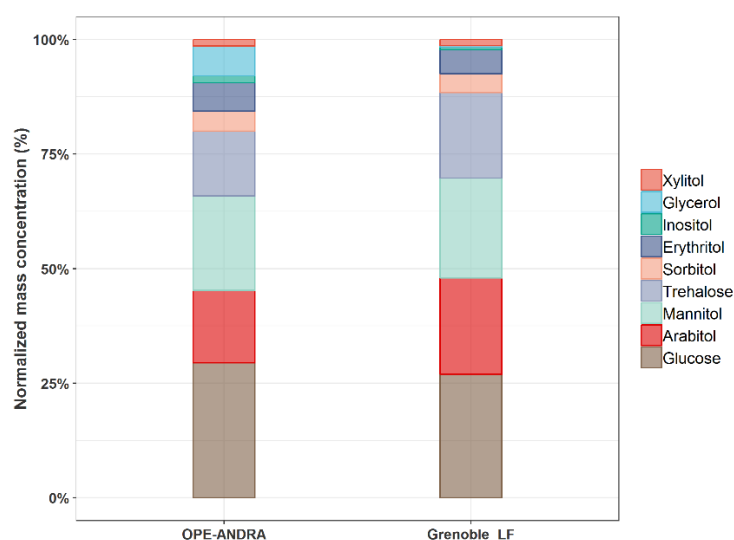
258 **Figure 2: Overview of relative mass distributions of individual primary sugar alcohols and saccharide compounds**
259 **quantified in PM$_{10}$ samples at two sites over summer and autumn periods (June to November) corresponding to**
260 **maximal atmospheric concentrations of sugar alcohols/saccharide compounds.**

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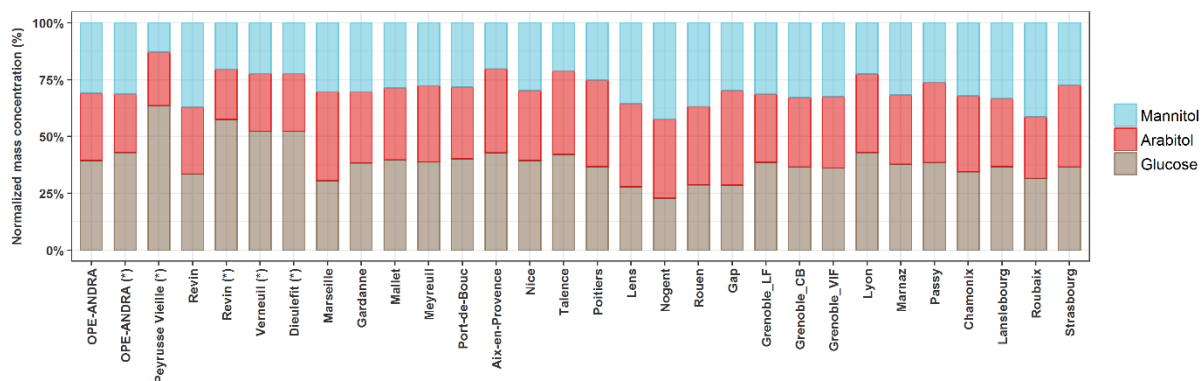
263 **Figure 3: Average mass concentration distributions of arabitol, mannitol, and glucose. Symbol (*) indicates PM$_{2.5}$**
264 **aerosol samples. Only data for warmer season (June to November), corresponding to maximal atmospheric**
265 **concentration of polyols and glucose are shown.**

266   Although various primary sugar alcohols and saccharides have been detected and quantified for most of the
267   investigated sites, the following study focuses only on the three major and ubiquitous species, namely arabitol,
268   mannitol and glucose. Figure 3 presents their average relative contributions at all sites, for the warmer period,
269   displaying very similar features at a first glance. However, discrepancies could be observed from site to site, as
270   discussed in following sections.

271   **3.2 Relationships between selected primary sugar compounds**

272   Figure 4 summarizes linear correlations obtained between arabitol and mannitol concentrations at each site during
273   the warmer period. Medium to very high coefficients of determination could be observed ($0.58 \leq R^2 \leq 0.93$; $30 \leq$
274    $n \leq 143$ or $45 \leq n \leq 341$ for $PM_{2.5}$ and PM_{10} series, respectively), with slopes in a rather narrow range (between
275   0.59 and 1.10), and quite low intercepts (always below 9 ng m^{-3}). Such covariations indicate that both species are
276   most probably co-emitted, by one or several type(s) of sources, at each site during the summer-autumn period.
277   These observations are in agreement with previous studies also showing strong covariations between arabitol and
278   mannitol (Kang et al., 2018; Verma et al., 2018; Zhu et al., 2015). Therefore, it seemed reasonable to consider both
279   species together, so that their concentrations are summed up and labelled as "polyols" in the following sections.
280   Conversely, linear correlations between glucose and polyols concentrations are generally weaker
281   ($0.10 < R^2 \leq 0.78$), with slopes varying over a much larger range (between 0.12 and 0.94), and variable intercepts
282   (between -5.6 and 16.4 ng m^{-3}). This suggests that glucose concentrations might follow a different pattern
283   compared to that of polyols, either due to different emission sources, or different chemical stability in the
284   atmosphere. It is therefore reasonable to keep glucose as a separate chemical species in the following discussion.
285   It should be emphasized that the variability in the slope of the regressions between the chemical concentrations is
286   most probably related to the emissions and atmospheric processing. Particularly in the case of mannitol and
287   arabitol, they may be influenced by biogenic or biotope characteristics. Nevertheless, no evident relationship
288   between the slope values and the typology or the geographical location of the sites could be observed (Figure 4).

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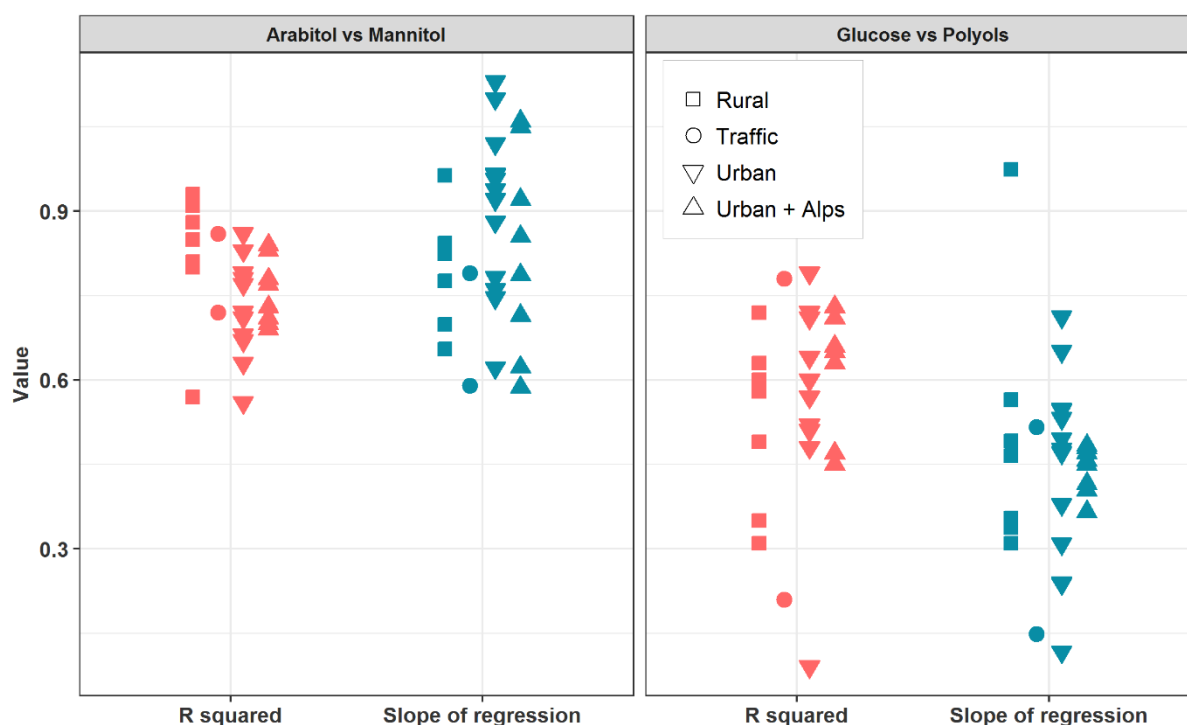
Figure 4: Linear regression analysis between selected primary sugar compounds mass concentrations (i.e., arabitol,
mannitol and glucose) during summer and autumn seasons (June to November), for all the sites considered in this study.

3.3 Relative distributions between PM$_{10}$ and PM$_{2.5}$

Figure 5 shows the average PM$_{10}$ and PM$_{2.5}$ concentrations of polyols and glucose at OPE-ANDRA and Revin during the summer and autumn seasons. The polyols mass concentrations ranged from 7.5±10.9 to 27.8±33.3 ng m^{-3} in PM$_{2.5}$, and from 48.9±38.2 to 73.5±61.8 ng m^{-3} in PM$_{10}$, in Revin and OPE-ANDRA sites, respectively. PM$_{10}$-to-PM$_{2.5}$ ratios were then on average of about 3 to 5. Similar size distribution patterns, with variable intensity, were observed for glucose (Fig. 5). These results indicate that polyols and glucose are mainly associated with the coarse PM fraction. This observation is in good agreement with several previous investigations where polyols (especially arabitol and mannitol), together with glucose, were prevalent in the coarse fraction (Fu et al., 2012; Fuzzi et al., 2007; Pio et al., 2008; Yttri et al., 2007b). However, Carvalho et al. (2003) reported different size distributions for polyols and glucose, with variable fine or coarse mode maxima depending upon sampling location. For instance, maximum atmospheric concentrations of mannitol were associated to fine and coarse aerosols from boreal forest (Finland) and rural background sites (Germany), respectively. The authors hypothesized that these observations are due to different assemblages of dominant fungal biota (with variable aerodynamic characteristics) at different sites. Some other previous studies showed aerodynamic diameters typically ranging from 2 to 10 µm, even though a few airborne bacterial/fungal spores could exceed that size (Bauer et al., 2008a; Elbert et al., 2007; Huffman et al., 2012; Zhang et al., 2015).

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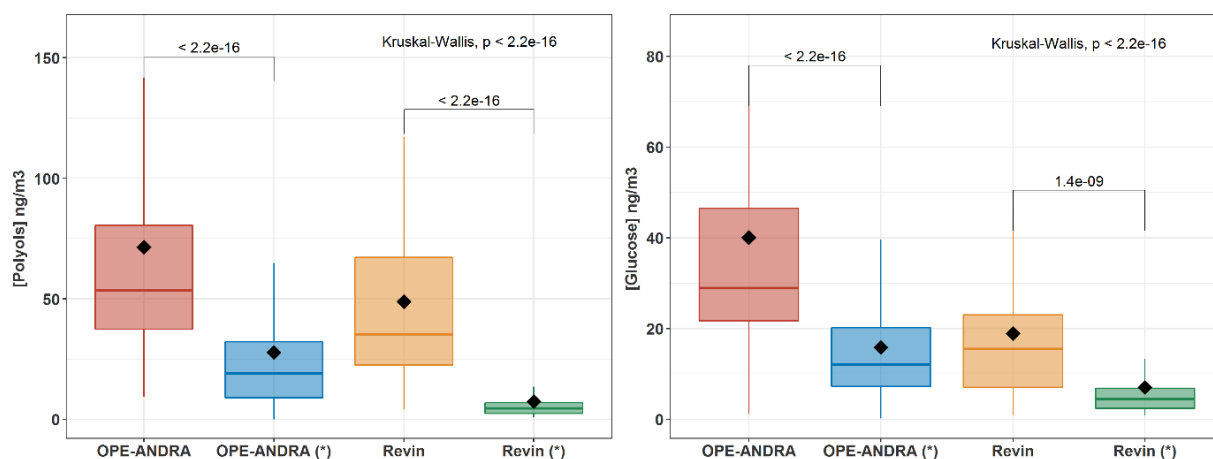
309 **Figure 5: Box plots of mass concentrations of polyols (left) and glucose (right) in PM$_{10}$ and PM$_{2.5}$ (with symbol (*)**
310 **samples). Black markers inside each boxplot indicate the mean concentration value, while the top, middle and bottom**
311 **lines of the box represent the 75th, median and 25th percentile, respectively. The whiskers at the top and bottom of the**
312 **box extend from the 95th to the 5th percentile. Number of samples were N = 123 for OPE-ANDRA and N = 87 for Revin,**
313 **respectively. Statistical differences between average mass concentrations were analyzed with the Kruskall-Wallis**
314 **methods (p < 0.05).**

315 Hence, although if the precise mechanisms of atmospheric emission of particulate polyols and glucose are not fully

316 resolved, our observations are in good agreement with ambient mass concentrations of polyols and glucose being

317 likely associated with biological particles, as already suggested elsewhere (Fu et al., 2012; Verma et al., 2018;

318 Zhang et al., 2015). These species could enter into the atmosphere through either natural or anthropogenic

319 resuspension of surface soils and associated bacterial/fungal spores (containing polyols and primary sugar

320 compounds), or via a direct input resulting from microbial activities (e.g., sporulation). Another hypothesis would

321 be the abrasion of leaves and the subsequent release of microbial organisms and plant debris (Fu et al., 2012;

322 Medeiros et al., 2006; Simoneit et al., 2004b).

323 **3.4 Spatial and seasonal distribution of atmospheric concentrations**

324 **3.4.1 Spatial and seasonal patterns of polyol concentrations**

325 As illustrated in Fig. 6, significant concentrations of polyols were measured at each investigated site, evidencing

326 the ubiquity of these organic compounds. The annual average concentration levels of polyols measured in PM$_{10}$

327 aerosols at all sites (33.2±33.5 ng m^{-3}; see Table SI-2) are within the range previously reported for urban and rural

328 sites across Europe (Burshtein et al., 2011; Di Filippo et al., 2013; Pietrogrande et al., 2014; Yttri et al., 2007b,

329 2011). Additionally, polyols mass concentrations clearly exhibit seasonal trends, with variable intensity according

330 to the sampling sites. On a seasonal average, polyols are more abundant in summer (46.8±43.6 ng m^{-3}) and autumn

331 (43.0±36.7 ng m^{-3}), followed by spring (19.0±13.6 ng m^{-3}) and winter (16.2±11.5 ng m^{-3}). The average

332 concentrations of polyols are at least 2 to 3 times higher during summer or autumn months than during the cold

333 months, with a ratio that can be as high as 8 to 10.

334 Previous studies also reported similar seasonal variation pattern for urban and rural aerosol samples collected at

335 various locations. For example, Pashynska et al. (2002) measured higher atmospheric polyol (arabitol, mannitol)

336 contents during late summer and autumn, in Belgium. Several other studies reported higher concentrations of

337 polyols in summer than spring and winter time, in aerosols collected from Texas, USA and Jeju Island, respectively

(Fu et al., 2012; Jia et al., 2010a, 2010b). More recently, Liang et al. (2016) and Verma et al. (2018) also reported similar seasonal distributions for aerosols sampled in Beijing, China and north-western Pacific, respectively.

The higher atmospheric polyols concentrations observed are likely due to the increased contribution from metabolically active microbial derived sources (fungi, bacteria, green algal lichens) as a result of external stressors such as heat, drought and relative moisture. Indeed, fungal and prokaryotic cells activities, including their growth and sporulation, are promoted by high temperature and humid conditions occurring in summer and autumn (China et al., 2016; Elbert et al., 2007b; Jones and Harrison, 2004; Rathnayake et al., 2017).

As also evidenced from Fig. 6, atmospheric polyols concentrations do not present any significant seasonal differences related to the site typology (rural, traffic, urban sites with/without Alpine influences), or latitude. There is some tendency toward higher concentrations in summer in Alpine environments, but some other sites (like the rural site of OPE-ANDRA, in the North-East of France) can reach the same levels of concentrations. We tested several types of hierarchical classifications, including variables like monthly or seasonal mean polyols concentrations, the ratio arabitol-to-mannitol, or linear regression parameters (slope, R square) but none of them led to a simple clustering of the sites that would explain the variability of the concentrations.
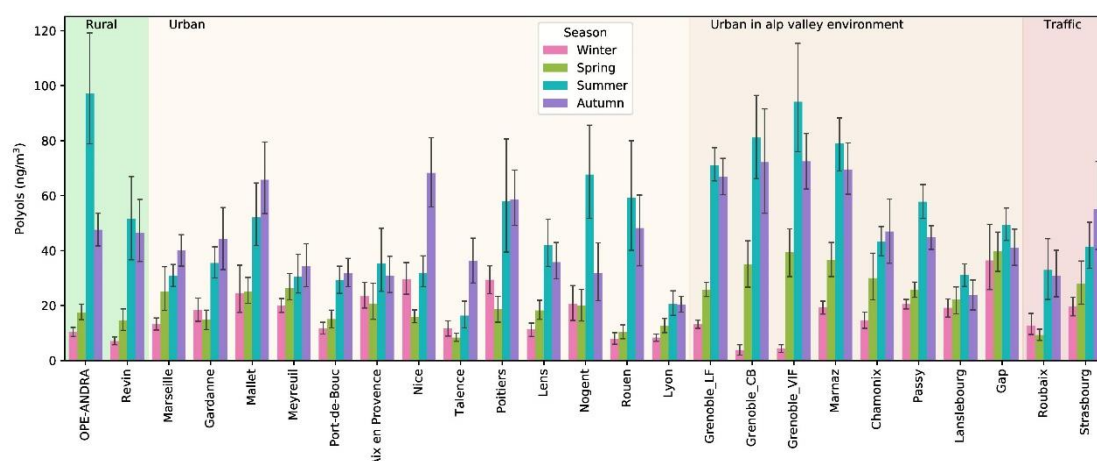
Figure 6: Spatial and seasonal distributions of atmospheric polyol average concentrations (ng m^{-3}) for various types of sites in France. Error bars correspond to standard deviations calculated with seasonal concentrations. Years of PM sampling campaigns are not concurrent at all sites (see Fig. SI-1). The seasons were defined as follows: Winter = December to February, Spring = March to May, Summer = June to August, and Autumn = September to November.

3.4.2 Spatial and seasonal patterns of glucose concentrations

The annual average concentrations of glucose measured in PM_{10} aerosols at all sites (20.4±15.6 ng m^{-3}; see Table SI-2) are comparable to those previously reported for various sites across Europe (Alves et al., 2006; Theodosi et al., 2018; Yttri et al., 2007b, 2011). Likewise polyols, the atmospheric concentrations of glucose also display seasonal and site-to-site variations (Fig. 7). The ambient seasonal mean concentrations (with standard deviations) of glucose are maximum in summer (25.0±24.2 ng m^{-3}) and autumn (24.6±19.8 ng m^{-3}), followed by spring (15.8±12.4 ng m^{-3}) and winter (12.6±10.2 ng m^{-3}). The summer / winter ratio for glucose seems generally lower than that of polyols, with higher ratios in the Alpine areas than in other parts of France. However, as for polyols, it remains difficult to classify the sites according to any criteria linked to site typology or latitude.

The seasonal trend of glucose concentrations in the present work is similar to that recently observed for aerosols (PM_{10} or total suspended particles) collected at various environmental background (suburban, urban and coastal)

sites around the world (Liang et al., 2016; Srithawirat and Brimblecombe, 2015; Verma et al., 2018). On average, a wide range of daily glucose concentrations (expressed as min-max, mean) in PM_{10} (0.1-297.2 ng m^{-3}, 20.4±15.6 ng m^{-3}) were observed in the present study. These values are comparable to those in PM_{10} (8.4-93.0, 47.0 ng m^{-3}) reported from an urban site in Norway (Yttri et al., 2007b). More recently, Liang et al. (2016) also reported similar concentrations in PM_{10} (3.1-343.6, 46.2±27.5 ng m^{-3}) from Beijing (China).

The sources and formation processes of glucose in the atmosphere are not currently well known and are rarely discussed. Glucose is an important carbon source for soil metabolic active microbiota, and it is commonly present in vascular plants. Additionally, cellulose (a linear polymer made of glucose subunits linked by β-1,4 bonds) is one of the most important form of organic compounds in terrestrial ecosystems and a major plant structural polymer (Boex-Fontvieille et al., 2014). It can also be quite abundant in the atmosphere (Puxbaum and Tenze-Kunit, 2003). Hence, it is hypothesized that ambient glucose could be formed through active microbial (i.e., bacteria, fungi, etc.) enzymatic hydrolysis of cellulose in plant debris. Consistent with these observations, glucose could be released into the atmosphere from both vascular plant materials (e.g., leaves, fruits, pollens, etc.) growing in spring and decomposing in autumn/summer, and soil microbiota, as already suggested elsewhere (Di Filippo et al., 2013; Jia et al., 2010a; Medeiros et al., 2006; Verma et al., 2018; Zhu et al., 2015).
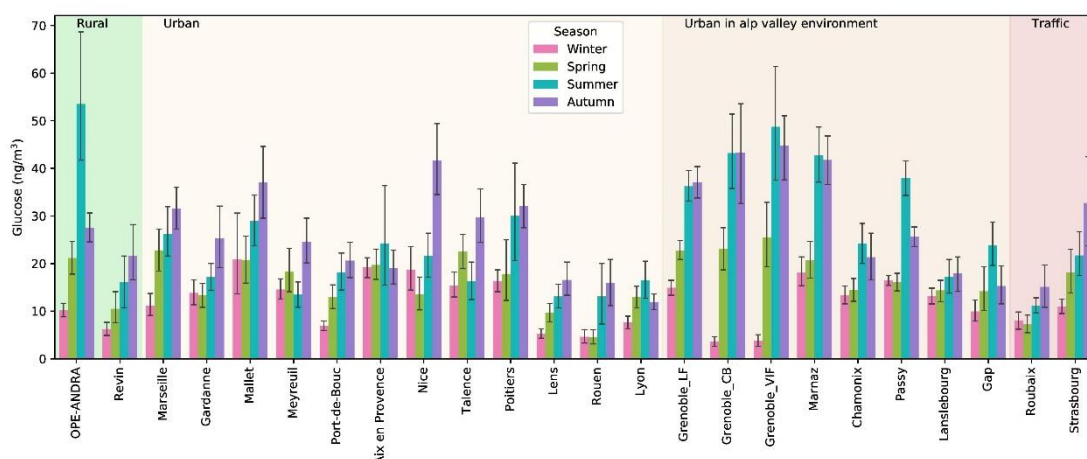
Figure 7: Spatial and seasonal distributions of atmospheric glucose levels (ng m^{-3}) for various types of sites in France (except the site of Nogent, which presented too many missing values. Error bars correspond to standard deviations calculated with seasonal concentrations.

3.4.3 Relative contributions to aerosol organic matter concentrations

The average contribution of polyols to the OM content of PM clearly displayed a seasonal behavior, as shown in Fig. 8. Here again, contributions are 2 to 10 times higher during summer and autumn compared to winter and spring, consistent with the assumption of higher emissions during these periods. The seasonal mean contribution of polyols to OM fluctuates from site to site, and accounts for 0.1 to 2.1 % of overall OM for these French sites (Fig. 8). Similarly, the seasonal mean concentrations of polyols together with glucose represent between 0.2 to 3.1 % of total OM at these sites (Fig. SI-2). However, on a daily basis (Samake et al., in prep.), atmospheric polyols mass concentrations can represent up to 6.3 % of total OM in PM_{10}, indicating that polyols can be amongst the major molecular species identified in aerosol organic matter (Fig. SI-3). Again, we could not find any simple way to group the sites according to their characteristics (typology or latitude, or climatic region), in order to better

understand the drivers behind the variability of this mass fraction. Further studies are currently conducted using
multi criterion examinations.

The seasonal polyols-to-OM distribution patterns in this study are comparable to those found for different urban or rural sites in Europe (around 0.2 to 2.5 % of OM) (Pashynska et al., 2002; Yttri et al., 2007b). Zhu et al. (2015) also reported similar seasonal polyols-to-OM contribution trend for aerosols sampled at Cape Hedo (coastal site, Japan).
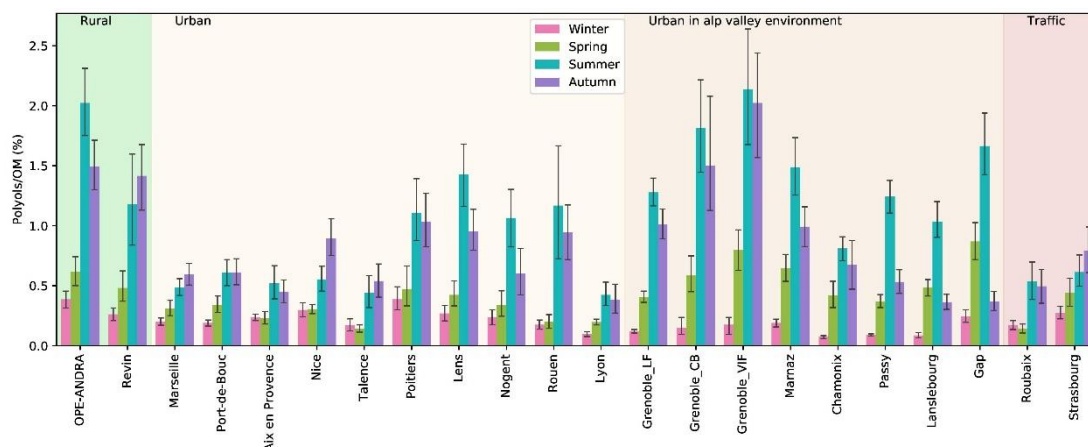
Figure 8: Spatial and seasonal distributions of mean contributions (in %) of polyols to the organic matter content of PM for various types of sites in France. Daily time-series of organic carbon (OC) were not available for the following sites: Gardanne, Mallet, and Meyreuil. Error bars correspond to standard deviations calculated with seasonal concentrations.

3.5 Primary biogenic factor in PMF studies

The sum of polyols (arabitol + mannitol) represents only a small fraction of the total OM. However, as proxies of PBOA, they are most probably emitted with other chemical species. Emission from biological particles is a complex topic since it may include a wide variety of compounds, both organic and inorganic (Elbert et al., 2007; Zhang et al., 2015). Moreover, it is not clear if polyols are mainly emitted directly in the atmosphere or are linked to other materials, for example with soil dust during resuspension processes. To investigate the relationship between polyols and other molecular tracers of emission sources, it would have been possible to perform simple correlation analysis with individual chemical species. This approach has the disadvantage of being a one-to-one relation and thus highly sensitive to the dynamics of all PM emission sources, not only the one we are interested in. Alternatively, another way is to use a PMF approach, also based on correlations but including much more information on the temporal variations of the different sources influencing the PM chemistry at a given receptor site.

As mentioned in Sect. 2.3, the PMF results used in this study include sites of different typologies (rural, traffic, urban sites in Alpine valley environments, etc.) for 16 different locations spread over France and part of the current dataset. At each site, the PMF studies allowed to identify a PBOA factor, characterized by the presence of more than 90 % of the total polyols content (sum of arabitol, mannitol and sorbitol), as presented in table SI-5 and Fig. SI-6. Moreover, the sensitivity of this factor to random noise in the data was investigated thanks to randomly re-sampling the input matrix of observation. In PMF analysis, this is done via the bootstrap method (Paatero et al., 2014) in the constrained run. The PBOA factor was always mapped to itself for 13 of the sites and quasi-always (97%) for the last three ones. It means that the PBOA factor does have a very high statistical stability since it never

428   swaps with another factor (see Fig. SI-5). Hence, the chemical composition of this factor may be informative to

429   investigate the PBOA source components (Table SI-6), and to evaluate the importance of PBOA emissions in

430   terms of OM mass apportionment.

431   **3.5.1. Contributions of PBOA to OM and polyols to PBOA**

432   Altogether, the results from the16 sites highlight the importance of the PBOA source contribution to total OM. As

433   shown in Fig. 9, the OM apportioned by the PBOA factor represents a significant fraction of the total OM mass

434   on a yearly average (range 6–28 %; average 13±6 %). When considering only the summer period (June-July-

435   August), this contribution is even larger and can exceed 40 % of the total OM at sites in the Alpine area (Marnaz,

436   Passy, Chamonix, Grenoble_LF) which are partially protected from large regional influences due to the local

437   topography. This result may be nuanced, in particular during summer, since some extent of mixing between PBOA

438   and Biogenic Secondary Organic Aerosols (BSOA) cannot be entirely excluded. However, several evidences tend

439   in favor of a non-significant mixing between BSOA and PBOA. First, the ratio of polyols-to-OC_{PBOA} shows a low

440   variability from site to site, while it is unlikely that such a secondary process led to the same amount of OC for all

441   sites since they present different meteorology, sunshine duration, etc. Second, the bootstrap analysis does not show

442   any "swap" between factors for the PBOA profile for all sites, indicative of a well-defined factor (see Fig. SI-5).

443   Finally, the OC_{PBOA}-to-polyols ratio in this work (about 16) is in the range of ratio expected for fungal spores (12

444   -27, when arabitol and mannitol are considered together) (Bauer et al., 2008a; Yttri et al., 2011).

445   Interestingly, some previous work using the same samples from the sites in the Arve valley (Passy, Chamonix)

446   showed that about 90 % of the OM is from modern origin (using ^{14}C measurements) during summer, with no

447   apparent correlation between this modern carbon and polyols concentrations (Bonvalot et al., 2016). Hence, despite

448   being an important contributing source, PBOA is not the major biogenic source in this type of environment.

449   Interestingly, opposite to the case of the Alpine valleys where this proportion is the highest, the ratios OM_{PBOA}-to-

450   OM_{total} are amongst the lowest for coastal environments (Talence, Marseille, Nice), a possible indication that the

451   marine environment is not a large emitter for these species. Recently, much lower concentrations of polyols in

452   aerosols from marine environments than those in terrestrially influenced sites were also reported off the coast of

453   Japan, also suggesting a higher contribution from terrestrial sources (Kang et al., 2018).

454   As illustrated in Fig. 9, polyols represent only a small fraction of the OM apportioned in the PBOA factor (1.2 %–

455   6.0 %; average 3.0±1.5 %) for the 16 studied sites. This variability is indeed rather small, considering the wide

456   range of sites and the diversity of other potential sources (on average 8 to 10 PMF factors were obtained for the

457   different sites). Indeed, this narrow range of the polyols fraction to the OM_{PBOA} highlights the stability of the

458   chemical profile of this source over a large regional scale. It indicates also that, if polyols are good proxies of the

459   PBOA sources, a large amount of other organic species are co-emitted, that still remain unknown.

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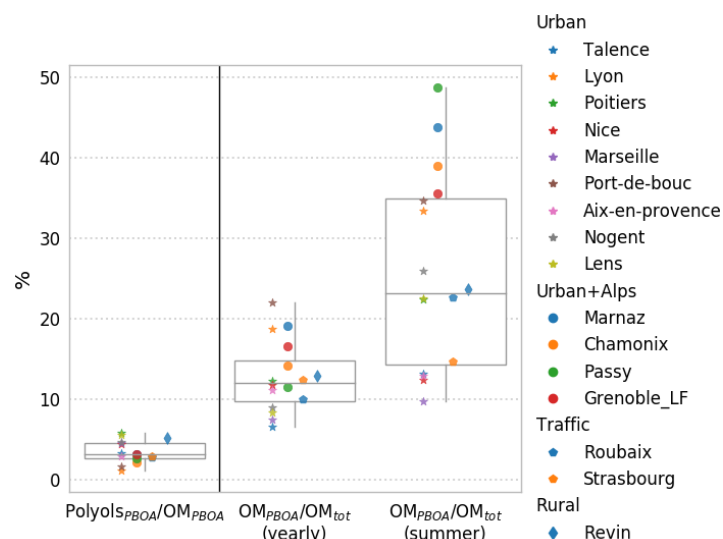

461 **Figure 9: Mass contribution of polyols to OM in the PBOA factor, and relative contributions of the OM$_{PBOA}$ factor to**
462 **the total OM in PM for the 16 studied sites where PMF model was run, over the year and summertime only. Stars and**
463 **circle refer to urban sites without/and with Alpine valley influence, respectively. Pentagon corresponds to traffic sites**
464 **and diamond to rural sites.**

465 **3.5.2 PBOA profile constituents and emission process**

466 Figure 10 shows the contribution (in µg of species per µg of PM in the PBOA factor profile) of each chemical

467 species included in the averaged PBOA factor from the 16 PMF studies. The principal contributors are OC and

468 EC, and significant fractions of crustal material also appear (Na$^+$, K$^+$, Ca^{2+}, Al, Ba, Cu, Fe, Mn, Ti, Zn) as well as

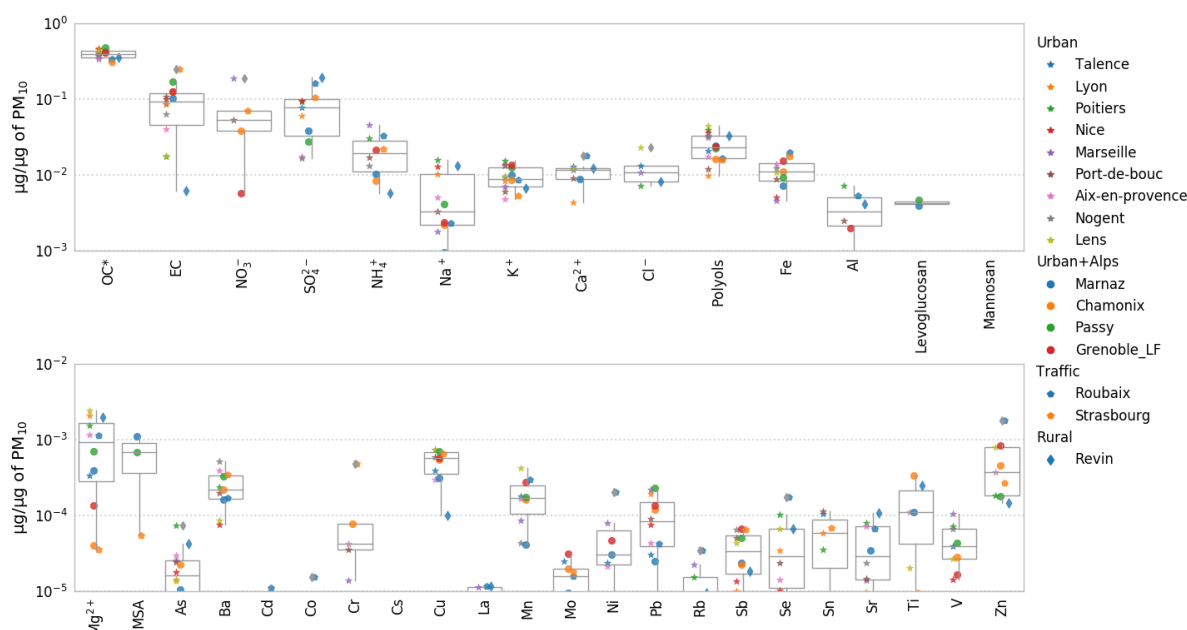
469 secondary elements such as nitrate and sulfate. However, EC appears to be highly variable both within and between

470 sites under consideration. The reader may refer to figure SI-7 for an estimation of the EC mass uncertainties in the

471 different sites. On average, the PBOA factor does not comprise a large fraction of metals and trace elements, most

472 of them being below 1 pg µg^{-1}. Here again, the low variability of the PBOA chemical profile encountered across

473 a large array of sites is remarkable.

474

475 **Figure 10: PMF chemical profile of the PBOA factor in the DECOMBIO and SOURCES programs expressed as a**
476 **fraction of the PM mass. Values lower than a few pg µg^{-1} are not displayed on purpose. For each boxplot, the top, middle**

480 The contribution from some crustal material could agree with the coarse mode distributions of polyols (Sect. 3.3)
481 and could be indicative of an emission process with the entrainment of spores with soil dust resuspension. To
482 investigate the importance of mineral dust in the PBOA factor, we clustered the chemical components of PM from
483 PBOA into 7 classes: OM (= $1.8 \times$ OC), EC, NO_3^-, NH_4^+, non-sea-salt sulfate (nss-SO_4), sea-salt, and dust. nss-
484 SO_4^{2-} is calculated from the measured SO_4^{2-} minus the sea-salt fraction of SO_4^{2-} ($nssSO_4^{2-} = SO_4^{2-} - ssSO_4^{2-}$
485 where $ssSO_4^{2-} = 0.252 \times Na^+$) according to Seinfeld and Pandis (1997). The sea-salt fraction is calculated
486 according to Putaud et al. (2010): $sea - salt = Cl^- + 1.47 \times Na^+$. Finally the dust fraction is estimated thanks
487 to Putaud et al. (2004b) as: $dust = (nss - Ca^{2+}) \times 5.6$ with nss-Ca^{2+} stands for non-sea salt Ca^{2+} and is
488 computed thanks to $nss - Ca^{2+} = Ca^{2+} - Na^+/26$. We note that the conversion coefficient provided by Putaud
489 et al. (2004b) may be influenced by an extreme value and then gives only a low estimate of dust resuspension.

490 Figure 11 presents the normalized average contributions of these 7 classes to the PBOA mass for the 16 sites with
491 PMF modelling. It clearly reveals that the PBOA factor is dominated by contributions from OM (78 ± 9 %),
492 followed by EC (9 ± 7 %), and only a minor contribution from the dust class (3 ± 4 %).

493 The large value for the contribution of EC is driven by two high values obtained at the sites of Strasbourg (that
494 reaches 25%) and Chamonix (18%) both influenced by direct and indirect traffic emissions. However, 6 other sites
495 present no EC in PBOA. Moreover, the uncertainties of EC in the PBOA profile of Strasbourg and Chamonix is
496 rather high (between 5 to 30% of PM mass at Strasbourg, see SI-7). On a yearly average, EC apportioned by this
497 factor (0 to 400 ng m^{-3} depending on the site) is close to the rural EC background in France of about 300 ng m^{-3}
498 (Golly et al., 2018).

499 This result on the general chemical profile of the PBOA factor, with a low crustal fraction, tends to infirm the
500 hypothesis of an emission process associating PBOA material with mineral dust resuspension. Indeed, our findings
501 rather suggest that a main part of PBOA (and polyols) is most likely associated with biological particle direct
502 emissions. It leaves only a minor fraction that could be linked to the mechanical resuspension of PBOA with crustal
503 elements. Some minor fraction of EC in this factor could come from resuspended EC-containing dust particles
504 being accumulated in topsoil as demonstrated in previous works (Forbes et al., 2006; Hammes et al., 2007; Zhan
505 et al., 2016). Hence, the origin of the larger fraction of the contribution of EC remains unknown. Our conclusions
506 are in good agreement with those made by Jia and Fraser (2011), based on the concentrations of these chemicals
507 in different types of samples: i.e., size-fractionated (equivalent to PM$_{2.5}$ and PM$_{10}$) soil, plant, fungi, atmospheric
508 PM$_{2.5}$ and PM$_{10}$. They found that the ambient concentrations of primary saccharide compounds at the suburban
509 site of Higley (USA) are typically dominated by contributions of biological materials rather than resuspension of
510 soil dust particles and associated microbiota.

511

	OM	EC	NO₃⁻	NH₄⁺	nss-SO₄²⁻	sea-salt	dust
mean	78	9	2	1	6	1	3
std	9	7	5	2	7	1	4
min	55	0	0	0	0	0	0
max	91	25	20	5	21	3	11

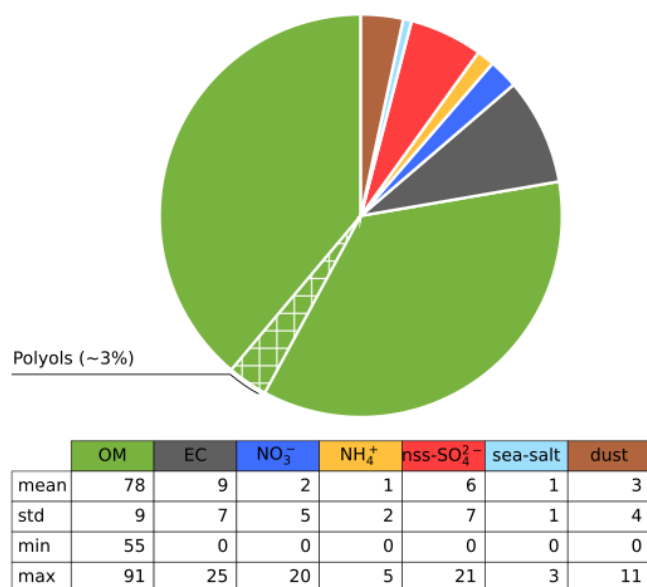



Figure 11: Average contribution (%) of species in the PBOA factor for the sites in SOURCES and DECOMBIO. The hatched area represents the proportion of the OM apportioned by the polyols (see text for reconstruction method).

4. Conclusion

The contribution of primary biogenic organic aerosols to PM is barely documented in the scientific literature. The present study aimed at providing a large overview of the spatial and temporal evolution of concentrations and contributions to aerosol organic matter of dominant primary sugar alcohols and saccharide compounds, for a large selection of environmental conditions in France. With 28 sites and more than 5,340 samples, it is to our knowledge the most comprehensive dataset for these compounds. The main results obtained indicate that:

- among the identified polyols, arabitol together with mannitol are the major species by mass, with lesser amounts of others polyols (e.g., erythritol, inositol, glycerol, sorbitol, and xylitol). Glucose is the dominant primary monosaccharide and its relative abundance is comparable to the sum of arabitol and mannitol;

- the two main polyols (arabitol and mannitol) together with glucose are mainly present within the coarse aerosol mode;

- at nearly all sites, ambient levels of the main polyols and glucose displayed clear seasonal variation cycles, with a gradual increase from spring and maximum in summer and autumn aerosols, followed by a sudden decrease in late autumn, and a winter minimum;

- atmospheric concentrations of the main polyols and glucose fluctuate according to site and season, and account each for between 0.1 to 2.1 % of OM on a seasonal average basis at these French sites;

- ambient mass concentrations of arabitol and mannitol are comparable. Meanwhile, they display very good temporal covariation, with ratios varying between sites. Conversely, linear correlations between the main polyols and glucose concentrations are much lower, suggesting different atmospheric sources, or atmospheric processes;

- arabitol and mannitol are efficient organic markers for PBOA. PMF studies of the yearly series from 16 sites give contributions of the primary biogenic emission (traced with the main polyols) to the total OM around 13±6 % on a yearly average and 26±12 % during summer, thereby showing that PBOA is an important source of total OM in PM_{10} for all sites across France. Furthermore, the average PBOA chemical source profile is made out of a very large fraction of OM (78±9 % of the total PBOA mass on average), suggesting it is mainly related to direct biogenic emissions from biological particles. Noteworthy, the presence of BSOA within the PBOA factor, particularly during summer could not be fully ruled out and further works using additional organic tracers (such as 3-methylbutanecarboxylic acid, pinic acid, and/or cellulose) are still needed to solve this issue. Additionally, the low crustal fraction indicates that this factor is weakly linked to soil dust resuspension associated with biological material;

- however, the PBOA source remains chemically poorly characterized as the main polyols represent only a small fraction of its total OM mass (3.0±1.5 % on average);

- despite comparable high concentrations in the atmosphere, the sources and processes leading to glucose concentrations and seasonal evolutions are still elusive. Indeed, the different PMF performed with glucose in input variable do not lead to a statistically stable solution;

Further investigations of the emission pathways and chemical characterization of the PBOA source associated with polyols are on-going, which may improve our understanding of its dynamic at various geographical scales, for a potential implementation in emission models in the future.

Author contributions: JLJ was the supervisor for the PhD for AS, FC, SW, and for the post-doc of DS. He directed all the personnel who performed the analysis at IGE. He was coordinator or principal investigator (PI) of the programs that generated the data for 18 of the 28 sites in this study (OPE-ANDRA, Part'Aera, CAMERA, SRN 2013, 3 Villes PACA, DECOMBIO, QAMECS) and co-PI for programs for 5 other sites. He is the coordinator for the CNRS LEFE-EC2CO CAREMBIOS program that is funding the work of AS. GU was the co-supervisor for the PhD of AS and SW. OF is the coordinator of the CARA program, (co-)funding and supervising the filter sampling and chemical analyses at 12 of the 28 sites. EP, OF, and VR supervised the PhD of DMO who investigated the 5 sites in northern France. Finally, JLB was the coordinator (program Lanslebourg) or partner of several programs whose data were used in this study (OPE-ANDRA, Part'Aera, 3 Villes PACA, DECOMBIO), and OF was the coordinator of the SOURCES program, which includes the work of DS as a post-doctoral fellow under the supervision of JLJ to gather and prepare most of the datasets used in the present studies.

All authors from the ANDRA (#5) and AASQA (#6 to 13) are representatives for each network that conducted the sample collection and the general supervision of the sampling sites.

FC and DS ran the PMF analysis. AS, SW and JLJ processed the data and wrote up the manuscript. All authors reviewed and commented on the manuscript.

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Competing interests: The authors declare that they have no conflict of interest.

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