

Responses to Referee #2's comments

General comment: This work describes the effect of the composition and presence of seed aerosol and gas-phase SO₂ (NO_x) on the SOA formation from guaiacol photooxidation. The topic is of great interest as methoxyphenols are largely emitted by biomass burning (BB) emissions and the evolution of these compounds in the atmosphere in a more complex environment is not fully understood yet. Some paragraphs of the discussion could be improved (in a quantitative way) in order to better appreciate the results provided by the authors.

Response to comment: Many thanks for your constructive comments and valuable suggestions, which would be much helpful to improve the scientific merits of this manuscript. Your concerns have been carefully addressed in the revised manuscript.

Comment 1: The influence of SO₂ in the gas-phase as a source of seed aerosol should be addressed. The sulfate aerosol arising from SO₂ conversion has the potential to impact the surface area available for condensation and therefore enhance SOA formation. This question needs to be addressed in a more quantitative way. SOA yields could be corrected from this estimation and would help the reader to know whether the increase of SOA concentrations is due to the aerosol microphysics or chemical reactions. The AMS used in this study can provide a lot of information about the evolution of the aerosol all along the experiment. The evolution of the seed aerosol concentration for each experiment would help understanding and estimating the influence of both microphysics and chemistry on SOA formation.

Response to comment 1: Thank you very much. Sulfate formed via SO₂ oxidation could serve as seed particles (Jaoui et al., 2008) and increase the surface areas of particles (Xu et al., 2016). These roles are favorable to partition more SOA-forming vapors onto particle phase (Zhang et al., 2014), consequently enhancing SOA yields. Therefore, the time-series variations in the concentrations of sulfate at different SO₂ concentrations were added in the Supplement (Figure R1). As shown in Figure R1, the sulfate concentration increased with the increase of SO₂ concentration, suggesting that more sulfate aerosols were produced via SO₂ oxidation.

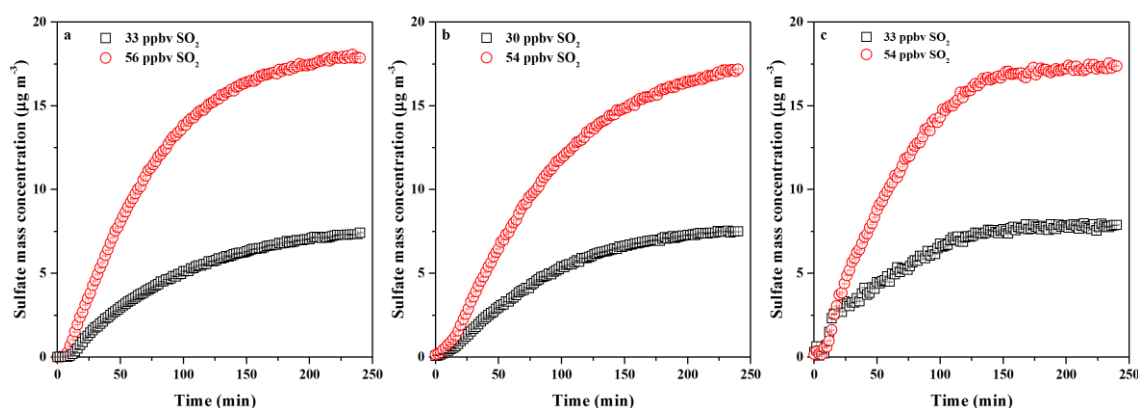


Figure R1. Variations in the concentrations of sulfate as function of irradiation time in the presence of various seed particles and SO₂ concentrations (a: no seed, b: NaCl, c: (NH₄)₂SO₄). The sulfate concentrations shown in Figure R1c is the net concentrations formed via SO₂ oxidation, e.g., do not include the (NH₄)₂SO₄ concentration added in the smog chamber.

In order to further investigate the effect of sulfate formed via SO₂ oxidation on SOA yields, the average gas-particle partitioning timescale ($\bar{\tau}_{g-p}$) over the course of experiment and the vapor wall-loss timescale (τ_{g-w}) under different experimental conditions were estimated and discussed in the revised manuscript. Based on

$\bar{\tau}_{g-p} / \tau_{g-w}$ ratio, the underestimation of SOA yield caused by vapor wall loss could be determined. The calculation methods of $\bar{\tau}_{g-p}$ and τ_{g-w} were added in the Supplement. As shown in Sections 3.2-3.4 in the revised manuscript, the decreasing trend of $\bar{\tau}_{g-p} / \tau_{g-w}$ ratios in the presence of seed particles and SO₂ have been observed (shown in Figure R2), suggesting that more SOA-forming vapors are partitioned onto particle phase, consequently increasing SOA yields. The details about the revisions in the manuscript were pointed out as follows.

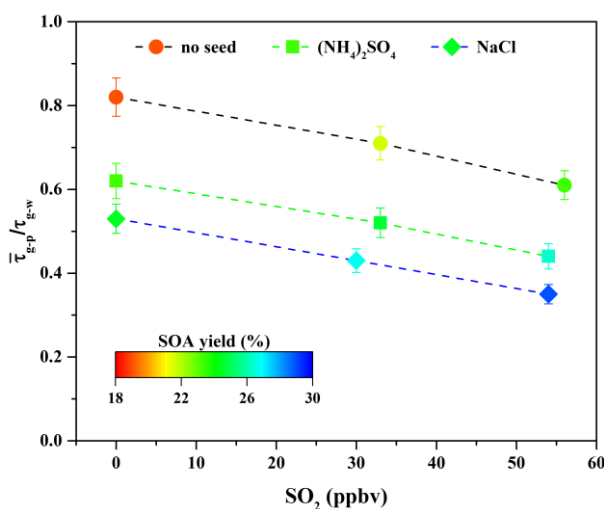


Figure R2. Variations in $\bar{\tau}_{g-p} / \tau_{g-w}$ ratio in the presence of various seed particles as a function of SO₂ concentration.

Revisions in the manuscript:

1. Figure R1 has been added in the revised Supplement.
2. Figure R2 has been added in the revised manuscript.
3. **Lines 32-34, Add:** “The decreasing trend of $\bar{\tau}_{g-p} / \tau_{g-w}$ ratio in the presence of seed particles and SO₂ suggested that more SOA-forming vapors were partitioned onto particle phase, consequently increasing SOA yields.”
4. **Experimental section:** The descriptions of vapor wall-loss correction have been

added in the revised manuscript as follows (**Lines 158-167**):

2.4 Vapor wall-loss correction

Previous studies have indicated that the losses of SOA-forming vapors to chamber wall can result in the substantial and systematic underestimation of SOA (Zhang et al., 2014, 2015). Therefore, SOA yields obtained in this work were also corrected by vapor wall loss. The effect of vapor wall deposition on SOA yields mainly depends on the competition between the uptake of organic vapors by aerosol particles and the chamber wall (Zhang et al., 2015). Thus, the ratio of the average gas-particle partitioning timescale ($\bar{\tau}_{g-p}$) over the course of experiment to the vapor wall deposition timescale (τ_{g-w}) could be reasonably used to evaluate the underestimation of SOA yields. The detailed calculation of $\bar{\tau}_{g-p}$ and τ_{g-w} was shown in the Supplement.

5. Lines 248-251, Add: “As illustrated in Fig. 2, the induction period became shorter with the increase of SO₂ concentration. The similar results caused by SO₂ have also been reported previously (Chu et al., 2016; Liu et al., 2016b).”

6. Lines 259-264, Add: “As shown in Fig. 3, $\bar{\tau}_{g-p} / \tau_{g-w}$ ratio decreased from 0.82 to 0.71 and 0.61 when SO₂ concentration increased from 0 to 33 and 56 ppbv. It suggests that the formed sulfate via SO₂ oxidation could serve as seed particles (Jaoui et al., 2008) and increase the surface areas of particles (Xu et al., 2016). These roles are favorable to partition more SOA-forming vapors onto particle phase (Zhang et al., 2014), consequently enhancing SOA yields.”

7. Lines 445-448, Add: “As shown in Fig. 3, $\bar{\tau}_{g-p} / \tau_{g-w}$ ratio had a decreasing trend

when increasing SO₂ concentration in the presence of seed particles, suggesting that the underestimation of SOA yields caused by vapor wall loss was weakened significantly because of the additional sulfate formed from SO₂ oxidation.”

8. Lines 495-497, Add: “The decreasing trend of $\bar{\tau}_{\text{g-p}} / \tau_{\text{g-w}}$ ratio in the presence of seed particles and SO₂ suggested that more SOA-forming vapors were partitioned onto particle phase, consequently increasing SOA yields.”

9. The descriptions of timescale calculation in the Supplement are as follows:

3 Timescale calculation

The average gas-particle partitioning timescale ($\bar{\tau}_{\text{g-p}}$) over the course of experiment could be expressed as Eq. (S1) (Zhang et al., 2014; Seinfeld and Pandis, 2006), and the vapor wall deposition timescale ($\tau_{\text{g-w}}$) is calculated using Eq. (S2) (Zhang et al., 2015).

$$\bar{\tau}_{\text{g-p}} = \frac{1}{2\pi\bar{N}_p\bar{D}_pD_{\text{gas}}F_{\text{FS}}} \quad (\text{S1})$$

$$\tau_{\text{g-w}} = \frac{1}{k_w} \quad (\text{S2})$$

$$k_w = \left(\frac{A}{V}\right) \left(\frac{\alpha_w \bar{c} / 4}{\pi\alpha_w \bar{c} / 8 (D_{\text{gas}} k_e)^{0.5} + 1} \right) \quad (\text{S3})$$

where \bar{N}_p is the average particle number concentration for the whole experimental process since UV lamps were turned on, \bar{D}_p is the number mean diameter, D_{gas} is the gas-phase diffusivity, F_{FS} is the Fuchs-Sutugin correction to the mass transfer flux due to noncontinuum effects and imperfect accommodation (Seinfeld and Pandis, 2006), k_w is the overall wall loss rate of organic vapor (Eq. (S3)), A/V is the surface

to volume ratio of the chamber, α_w is the mass accommodation coefficient of eddy diffusion ($\sim 10^{-5}$) (Zhang et al., 2014; Matsunaga and Ziemann, 2010), \bar{c} is the mean thermal speed of the molecules, and k_e is the coefficient of eddy diffusion (0.015 s^{-1}) (Zhang et al., 2014).

It is assumed that D_{gas} of organic vapor changes with the molecular weight (MW) and is equal to $D_{\text{CO}_2}(\text{MW}_{\text{CO}_2}/\text{MW})$. The value of D_{CO_2} is $1.38 \times 10^{-5} \text{ m}^2 \text{ s}^{-1}$ (Zhang et al., 2014). Conventionally, MW of 98 g mol^{-1} (H_2SO_4) is widely used for the Fuchs-Sutugin correction, but a number more like 300 g mol^{-1} might be more representative of the condensable organic vapors. Thus, MW of 300 g mol^{-1} was selected in this work. The Fuchs-Sutugin correction is expressed as the following equation:

$$F_{\text{FS}} = \frac{0.75\alpha(1 + Kn)}{Kn^2 + Kn + 0.283Kn\alpha + 0.75\alpha} \quad (\text{S4})$$

where α is the mass accommodation coefficient onto particles (~ 0.002) (Zhang et al., 2014) and Kn is the Knudsen number, expressed as follows:

$$Kn = \frac{\lambda}{R_p} \quad (\text{S5})$$

R_p is the particle radius and λ is the gas mean free path, which is calculated using Eq. (S6):

$$\lambda = \frac{3D_{\text{gas}}}{\bar{c}} \quad (\text{S6})$$

$$\bar{c} = \sqrt{\frac{8N_A kT}{\pi \text{MW}}} \quad (\text{S7})$$

where N_A , k , and T are Avagadro's number, Boltzman constant, and temperature, respectively.

Comment 2: I would expect more details in the experimental section specifically for the instruments used (PTR and AMS). Also, it would be worth adding a section for the AMS data analysis (quantification, PMF...)

Response to comment 2: More details about HR-ToF-PTRMS and HR-ToF-AMS have been added in the Experimental section in the revised manuscript. In addition, the analysis of data from HR-ToF-PTRMS and HR-ToF-AMS has also added in the Experimental section. PMF analysis has been added in the Supplement. The detailed revisions in the revised manuscript were listed as follows.

Revisions in the manuscript:

1. Lines 128-134, Add: “The HR-ToF-PTRMS with a time resolution of 1 min was used online to measure the gas phase concentration of guaiacol, and its m/z range was 10–500 in the process of data acquisition. Before data collection, the peaks of the protonated water ($[\text{H}_3^{18}\text{O}]^+$) and protonated acetone ($[\text{C}_3\text{H}_7\text{O}]^+$) ions at m/z 21.0246 and 59.0491 were used for mass calibration, with the aim to obtain accurate mass determination during experimental process. All data obtained by the HR-ToF-PTRMS were analyzed with the PTR-MS Viewer software (version 3.1.0, IONICON Analytik).”

2. Lines 137-143, Add: “For all experiments, the acquisition time of the HR-ToF-AMS was 2 min. The inlet flow rate, ionization efficiency, and particle sizing of the HR-ToF-AMS were calibrated at regular intervals, according to the standard protocols using the size-selected pure ammonium nitrate particles (Drewnick et al., 2005; Jimenez et al., 2003). All data obtained by the HR-ToF-AMS were

analyzed by the ToF-AMS analysis toolkit SQUIRREL 1.57I/PIKA 1.16I version, in Igor Pro version 6.37.”

3. Supplement, PMF analysis has been added as follows:

2 Positive matrix factorization (PMF) analysis

Positive Matrix Factorization (PMF) is a receptor model and a multivariate factor analysis tool, which could decompose a matrix of speciated sample data into two matrices, namely factor contributions and factor profiles (Paatero, 1997). In recent years, the PMF model was widely used for the analysis of high resolution (HR) mass spectra data, which could provide better separation of different organic components (DeCarlo et al., 2010; Docherty et al., 2008). This model is expressed as a bilinear factor model, namely, $x_{ij} = \sum_p g_{ip} f_{pj} + e_{ij}$, where i and j refer to values of j species in i samples, respectively, p is the number of factors in the solution and is used a least-squares fitting process to minimize the quality of fit parameter. In this work, the HR mass spectra (m/z 12–115) was analyzed by the PMF software coupled with a modified version of the CU AMS PMF Execute Calcs Tool version 2.06 developed by Ulbrich et al. (2009). The concentration and uncertainty matrices input into the PMF analysis were generated from the PIKA version 1.15D. Ions were classified and down-weighted according to the signal to noise ratios (SNR). $0.2 < \text{SNR} < 2$ was classified as the weak ions and down-weighted by a factor of 2, $\text{SNR} < 0.2$ was bad ions and removed from the analysis, and the uncertainty values of CO_2^+ -related peaks at m/z 16 (O), 17 (HO), 18 (H_2O), 28 (CO), and 44 (CO_2) were down-weighted.

Comment 3: The authors used the AMS which is an instrument known to highly fragment molecules and does not allow molecular level chemical characterization for organics. Organosulfates and organonitrates can be positively identified only using appropriate instrumentation. I would suggest to use: organic sulfur-containing compounds and organic nitrogen-containing compounds (or something similar).

Response to comment 3: Thank you for your valuable suggestion. Organosulfates and organonitrates have been rewritten to be organic S-containing compounds and organic N-containing compounds, respectively.

Revisions in the manuscript:

1. Lines 39, 231, 233, 499, Change “organic nitrates” To “organic N-containing compounds”
2. Lines 41, 329, 354, Change “organosulfates” To “organic S-containing compounds”

Comment 4: For some results, an evaluation of the uncertainties would help knowing if the variability is due to the uncertainty of the measurement or really represents a difference in a physico-chemical process. It would be worth providing uncertainties for M_o , SOA yields and OS_c . In Figure 7, the larger SOA mass in the first 2 hours of the experiment with NaCl only comparing with NaCl + 30 ppbv of SO_2 could be due to the uncertainty of the experiment. If not, this behavior needs to be discussed by the authors.

Response to comment 4: The uncertainties for M_o , SOA yields and OS_c have been

added in the revised manuscript as follows.

Revisions in the manuscript:

1. Line 24, Change “(9.46–26.37%)” **To** “ranged from $(9.46 \pm 1.71)\%$ to $(26.37 \pm 2.83)\%$ ”

2. Lines 202-203, Change “9.46–26.37%” **To** “ $(9.46 \pm 1.71)\%$ to $(26.37 \pm 2.83)\%$ ”

3. Line 252, Change “63.62 to 71.88 and 78.59” **To** “ (63.62 ± 1.71) to (71.88 ± 1.43) and (78.59 ± 2.06) ”

4. Line 254, Change “The corresponding SOA yield increased by 14.05% and 23.66%, respectively.” **To** “The corresponding SOA yields were $(9.46 \pm 1.71)\%$, $(21.60 \pm 1.27)\%$, and $(23.42 \pm 1.80)\%$, respectively.”

5. Lines 365-366, Change “63.62 to 79.44 and 84.91” **To** “ (63.62 ± 1.71) to (79.44 ± 1.86) and (84.91 ± 2.01) ”

6. Lines 367-368, Change “The corresponding SOA yield increased by 23.06% and 29.57%, respectively.” **To** “The corresponding SOA yields were $(23.31 \pm 1.59)\%$ and $(24.54 \pm 1.73)\%$, respectively.”

7. Lines 439-440, Change “M₀ was enhanced by 41.43% and 53.47%” **To** “M₀ increased from (63.62 ± 1.71) to (90.89 ± 2.28) and $(98.86 \pm 2.11) \mu\text{g m}^{-3}$.”

8. Lines 441-442, Change “the corresponding SOA yield increased by 41.43% and 53.47%” **To** “the corresponding SOA yields were $(26.78 \pm 1.97)\%$ and $(29.06 \pm 1.82)\%$.”

9. Lines 442-445, Change “M₀ was enhanced by 32.58% for 33 ppb SO₂ and 41.34% for 54 ppb SO₂, respectively, and the corresponding SOA yield increased by 29.78%”

and 39.24%.” To “ M_o increased from (63.62 ± 1.71) to (84.35 ± 2.09) for 33 ppbv SO_2 and $(89.92 \pm 2.31) \mu g m^{-3}$ for 54 ppbv SO_2 , enhanced by 32.58% and 41.34%, respectively, and the corresponding SOA yields were $(24.58 \pm 1.78)\%$ and $(26.37 \pm 1.98)\%$.”

10. Line 487, Change “9.46–26.37%” To “ $(9.46 \pm 1.71)\%$ to $(26.37 \pm 2.83)\%$ ”

11. Tables and Figures: The uncertainties of M_o , SOA yields, and OS_c have been added in Tables 1 and 2, as well as Figures 1, 2, 5, 6, and 8 in the revised manuscript.

Comment 5: 1-51 and elsewhere in the introduction: Specify the type of fuel used for the given emission factors.

Response to comment 5: The type of fuel has been specified in the revised manuscript.

Revisions in the manuscript:

Lines 56 and 60, Change “fuel” To “wood”

Comment 6: 1-62 SOA formation from guaiacol photooxidation in the presence of NO_x has been studied in the past (Lauraguais et al., 2014; Yee et al., 2013). See your discussion paragraph. Experimental section: The experimental section could be improved by adding details about the the AMS/PTR-MS instrumental settings (experimental conditions) and data analysis (N:C, O:C, H:C, sulfur and nitrogen-containing compounds concentrations, PMF).

Response to comment 6: In previous studies, the significant SOA formation from

guaiacol oxidation by OH radicals has been reported, and OH radicals are produced from the photolysis of the OH precursors (i.e., H₂O₂ and CH₃ONO) (Ahmad et al., 2017; Lauraguais et al., 2014; Yee et al., 2013). In order to describe accurately, this sentence has been rewritten in the revised manuscript.

More details about HR-ToF-PTRMS and HR-ToF-AMS have been added in the Experimental section in the revised manuscript. In addition, the analysis of data from HR-ToF-PTRMS and HR-ToF-AMS has also been added in the Experimental section. PMF analysis and the calculation of organosulfate concentration have been added in the Supplement. The supplementary revisions about HR-ToF-PTRMS, HR-ToF-AMS, and PMF analysis have been pointed out in the responses to the Referee #2's Comment 2. Thus, these revisions were not listed in the following revisions. Other detailed revisions in the revised manuscript were listed as follows.

Revisions in the manuscript:

1. Lines 65-68, Change “Meanwhile, several studies have reported the significant SOA formation from guaiacol oxidation by OH radicals.” **To** “Meanwhile, several studies have reported the significant SOA formation from guaiacol oxidation by OH radicals, produced from the photolysis of the OH precursors (i.e., H₂O₂ and CH₃ONO) (Ahmad et al., 2017; Lauraguais et al., 2014b; Yee et al., 2013).”

2. Supplement, organosulfate concentration calculation have been added as follows:

4 Organosulfate concentration calculation

Considering that methyl sulfate is the simplest organosulfate, the fraction of C_xH_yO_zS is more likely to be lower than that of methyl sulfate (Huang et al., 2015). In addition,

more information about the varieties of organosulfates could not be obtained in this work. Therefore, a conservative low-bound of organosulfate concentration has been estimated according to the method described by Huang et al. (2015), as shown in Eq. (S8).

$$C_{OS_{min}} = f_{OS} \times (C_{org} + C_{SO_4}) \quad (S8)$$

$$f_{OS} = \frac{\sum_h I_{OS,h}}{RIE_{OS}} \quad (S9)$$

$$\frac{\sum_n I_{org,n}}{RIE_{org}} + \frac{\sum_t I_{SO_4,t}}{RIE_{SO_4}}$$

where $C_{OS_{min}}$ is the minimum concentration of organosulfates; f_{OS} is the factor that represents for the fractional contribution of organosulfates to the lumped species of organics and sulfate in W-mode data; C_{org} and C_{SO_4} are the concentrations of organics and sulfate calculated from V-mode data, respectively; h , n , and t indicate the fragments of organosulfates, organics, and sulfate, respectively; $\sum_h I_{OS,h}$, $\sum_n I_{org,n}$, and $\sum_t I_{SO_4,t}$ are the total signal intensities of organosulfates, organics, and sulfate, respectively, obtained from W-mode data; RIE_{OS} is the relative ionization efficiency for organosulfates, which is estimated as the average of $RIE_{org} = 1.4$ and $RIE_{SO_4} = 1.2$ (Barnes et al., 2006).

Comment 7: 1. 165-167 Do you see any low volatility products in the gas phase based on the PTRMS data?

Response to comment 7: According to the products of guaiacol oxidation by OH radicals in the presence of NO_x reported by Lauraguais et al. (2014) and Yee et al. (2013), we did not observe these low volatility products by the HR-ToF-PTRMS. The

reason should be from the configuration of the HR-ToF-PTRMS (Ionicon Analytik GmbH) because a capillary interface was used. This leads to a very low transmission efficiency of low volatile compounds. In fact, these compounds can be measured using an API-ToF-CIMS. Unfortunately, the API-ToF-CIMS is unavailable in our laboratory.

Comment 8: 1.171 and all the paper: What is the estimated uncertainties of the reported SOA yields?

Response to comment 8: The uncertainties of SOA yields have been added in the revised manuscript. The corresponding revisions have been pointed out in the responses to the Referee #2's Comment 4. Thus, the detailed revisions were not listed.

Comment 9: 1.170 How would you explain the difference in SOA yield between your study and Lauraguais et al., 2014? The SOA formation in their study is 3 times higher with the same guaiacol and OH concentrations and without seed particles.

Response to comment 9: In the original manuscript, the vapor wall-loss was not considered when calculating SOA yields. In the revised manuscript, the vapor wall-loss was corrected and the SOA yields were well consistent with the results reported by Lauraguais et al. (2014). Thus, the difference in SOA yields should be reasonably resulted from vapor wall loss which was not considered in the original manuscript. The vapor wall loss has been corrected in our revised manuscript.

Revisions in the manuscript:

1. Lines 203-210, Add: “According to the ratios of $\bar{\tau}_{g-p} / \tau_{g-w}$ (0.61–0.93), the determined SOA yields were underestimated by a factor of ~2 times, suggesting that vapor wall loss in the chamber could significantly affect SOA formation. The similar results were reported previously by Zhang et al. (2014), who indicated that SOA yields for toluene photooxidation were substantially underestimated by factors as much as 4 times, caused by vapor wall loss. As shown in Fig. 1, the vapor wall-loss corrected SOA yields were in the range of $(15.24 \pm 0.85)\%$ to $(50.89 \pm 2.87)\%$, and could also be reproduced by a one-product model ($R^2 = 0.96$).”

2. Lines 216-219, Add: “Overall, the vapor wall-loss corrected SOA yields in this work are well in agreement with those reported previously (Lauraguais et al., 2014b; Yee et al., 2013), but the determined SOA yields are much lower. Therefore, the effect of vapor wall loss on SOA formation should be seriously taken into account.”

Comment 10: 1. 179 Sun et al., 2010 presented results of aqueous-phase oxidation of guaiacol with and without an added source of OH. Their results are not really comparable to this study.

Response to comment 10: According to your valuable suggestion, the comparison has been deleted in the revised manuscript.

Revision in the manuscript:

Lines 180-182, Delete: “For example, Sun et al. (2010) have reported that SOA mass formed from the aqueous-phase photochemical reaction of guaiacol in the presence of H₂O₂ is about one-fold higher than that in the absence of H₂O₂.”

Comment 11: l. 189-197 Organonitrates (-ONO₂) are different than nitro-organics (nitro-aromatics, nitro-compounds...) containing a (-NO₂) chemical group. The formation of nitroaromatics from the photooxidation of guaiacol under high NO_x conditions has already been reported (i.e. nitroguaiacol, nitrocatechol) (Ahmad et al., 2017; Lauraguais et al., 2014).

Response to comment 11: According to your valuable suggestion, organonitrates have been rewritten to be organic N-containing compounds, respectively.

Revisions in the manuscript:

Lines 39, 231, 233, 499, Change “organic nitrates” To “organic N-containing compounds”

Comment 12: In this study, the reported NO⁺/NO₂⁺ is clearly different from the ammonium nitrate standard. This difference could be explained by the presence of both nitro and nitrate organics (Farmer et al., 2010; Sato et al., 2010). I would suggest to use organic nitrogen-containing compounds (or something similar) instead of “organonitrates” unless the authors provide molecular chemical characterization clearly showing the formation of organonitrates.

Response to comment 12: According to your valuable suggestion, the difference between NO⁺ / NO₂⁺ ratio observed in this work and that for ammonium nitrate was explained. In addition, organonitrates have been rewritten to be organic N-containing compounds, respectively. The detailed revisions were pointed out as follows.

Revisions in the manuscript:

1. Lines 39, 231, 233, 499, Change “organic nitrates” To “organic N-containing compounds”

2. Lines 227-231, Change “The average $\text{NO}^+ / \text{NO}_2^+$ ratio of SOA from guaiacol photooxidation is 4.08, which is within the range of 3.82–5.84 for organic nitrates of SOA from the photooxidation of aromatics (Sato et al., 2010). In this work, the measured $\text{NO}^+ / \text{NO}_2^+$ ratios for inorganic nitrates are in the range of 2.06 to 2.54, determined by HR-ToF-AMS using ammonium nitrate as calibration sample.” To “The average $\text{NO}^+ / \text{NO}_2^+$ ratio of SOA from guaiacol photooxidation is 4.08, which is higher than that (2.06–2.54) for ammonium nitrate, determined by the HR-ToF-AMS in this work. The possible explanation might be that nitro-organics and organonitrates both exist in SOA (Farmer et al., 2010; Sato et al., 2010).”

Comment 13: 1. 224 The results in Figure 3 arise from one specific condition or represent an average of MS for the three different conditions? Unclear. In the MS in Figure 3, we can see a large contribution of the m/z 44 in the factor 1. At t=0 min, the factor 1 dominates the aerosol fraction which means that before the photooxidation experiment there is already a large fraction of m/z 44 in the aerosol. Is there already oxidized organics and SOA in the aerosol before the photooxidation experiments? Does it correspond to guaiacol condensation on the aerosol phase? Something else? To illustrate these experiments, I would recommend to provide the evolution of the organics, sulfate, nitrate, over the course of an experiment (before and after lights on and corrected with wall loss). This would help visualizing the change in sulfate an

organics concentrations. The impact of sulfate formation on SOA formation due to an increase of condensation or due to chemical processes needs to be addressed in a more quantitative way.

Response to comment 13: Figure 3 in the original manuscript is obtained by applying PMF analysis to the AMS data, collected at different SO₂ concentrations over the course of experiments. Thus, it represents an average of mass spectrum over the course of experiments at three different SO₂ concentrations. We are very sorry to make a mistake, e.g., Figure S4 in the original Supplement is wrong, caused by our carelessness. The right figure has been added in the Supplement (Figure R3). As shown in Figure R3, the relative fraction of Factor 1 at different SO₂ concentrations at the beginning of experiments was close to 0, which increased firstly and then decreased along with prolonging irradiation time.

In addition, the time-series variations in the concentrations of sulfate and nitrate over the course of experiments have been added in the Supplement (Figure R4). The time-dependent growth curves of SOA mass concentration for guaiacol photooxidation at different SO₂ levels are shown in Figure 2 in the manuscript (Figure R5).

In the revised manuscript, the quantitative analysis about the effect of sulfate formed via SO₂ oxidation on SOA yield has been discussed, according to the $\bar{\tau}_{g-p} / \tau_{g-w}$ ratios obtained at different SO₂ concentrations. The detailed revisions have been pointed out in the responses to the Referee #2's Comment 1, which were not listed as follows.

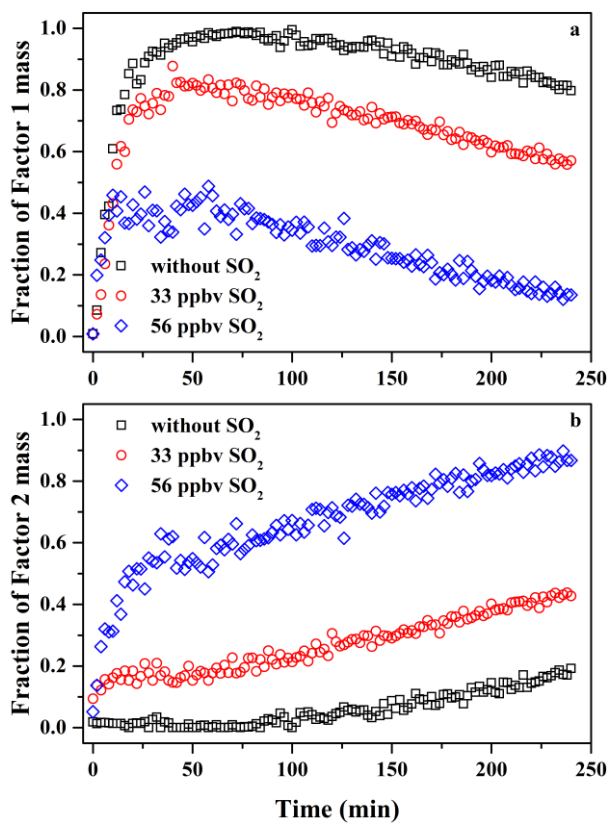


Figure R3. Time-dependent curves of Factor 1 (a) and Factor 2 (b) at three different SO₂ concentrations (without seed particles).

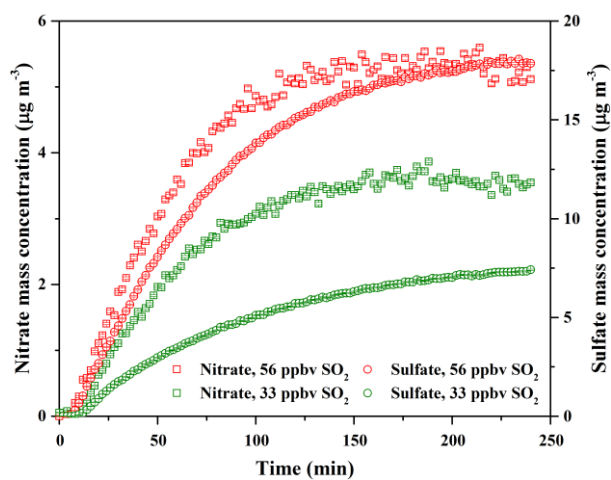


Figure R4. Variations in the concentrations of sulfate and nitrate in the presence of various SO₂ concentrations as a function of reaction time (without seed particles).

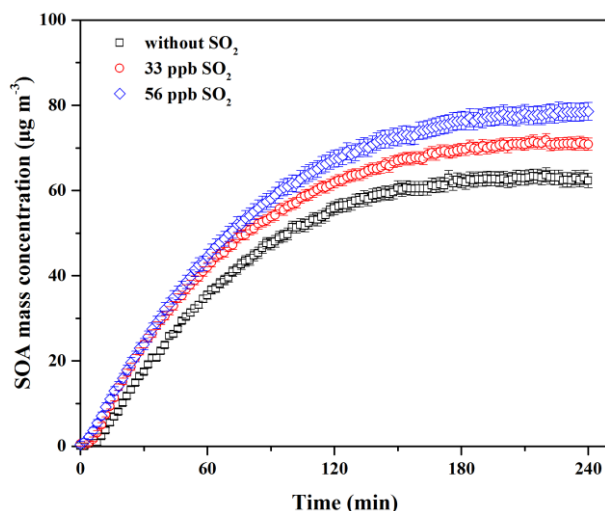


Figure R5. Time-dependent growth curves of SOA mass concentration for guaiacol photooxidation at different SO₂ levels.

Revisions in the manuscript:

1. Figures R3 and R4 have been added in the Supplement.
2. The caption of Figure 4 was changed to be “Mass spectra of Factor 1 (a) and Factor 2 (b) for the formed SOA identified by applying PMF analysis to the AMS data, obtained at different SO₂ concentrations over the course of experiments.”

Comment 14: 1 243-248 From the MS presented in Figure S5 we clearly see that in the presence of SO₂, low-MW compounds are more abundant. It also appears that high-MW compounds are in higher abundance in the presence of SO₂ (33 ppb vs no SO₂ and 56 ppb vs 33 ppb). Do you have an idea of the role of SO₂ in the formation of these compounds?

Response to comment 14: As shown in Figure S5 in the original manuscript, the signal fractions from the low-MW species were enhanced significantly in the presence of SO₂, and were much higher than those from the high-MW species ($m/z > 300$). In

other words, SO_2 played a more important role in the formation of organic S-containing compounds and the formation or uptake of low-WM species. For example, we observed that acetic acid concentration decreased in the presence of SO_2 and seed particles compared with the corresponding control experiments, which suggesting that the uptake of acetic acid might be enhanced. These results are in good agreement with those reported by Liggio et al. (2005) and Liu et al. (2010), who observed that that the uptake of organic compounds under acidic conditions would be enhanced significantly. The time-series changes in the gas-phase concentrations of acetic acid under different conditions were discussed in the revised manuscript and added in the Supplement (shown in Figure R6).

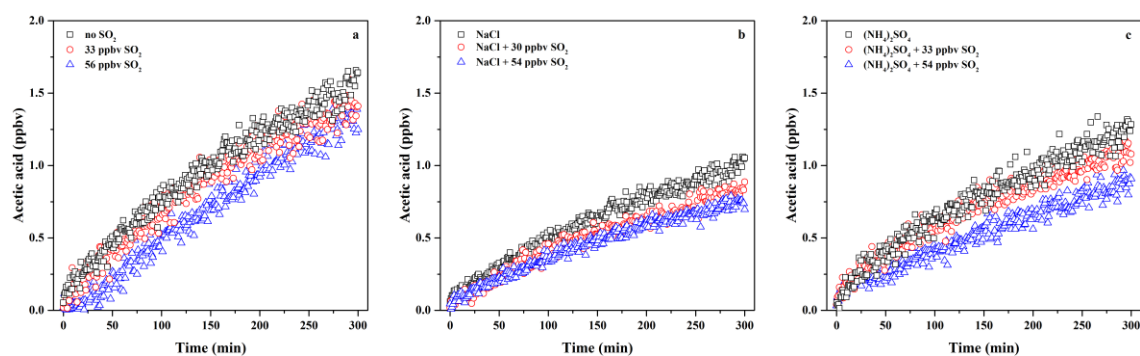


Figure R6. Variations in the gas-phase concentrations of acetic acid as function of irradiation time in the presence of various seed particles and SO_2 concentrations (a: no seed, b: NaCl, c: $(\text{NH}_4)_2\text{SO}_4$).

As shown in Figure S5 in the original Supplement, the formation of high-MW species (i.e., oligomers) was observed, which should be reasonably produced via the acid-catalyzed heterogeneous reactions (Cao and Jang, 2007; Jaoui et al., 2008; Liu et al., 2016; Xu et al., 2016). But, compared to low-MW species, the formation of

oligomers should not be predominant due to the much lower signal fractions.

Revisions in the manuscript:

1. Figure R6 has been added in the Supplement.
2. **Lines 330-332, Add:** “compared to the formation of high-MW species (i.e., oligomers) that should be reasonably produced via the acid-catalyzed heterogeneous reactions (Cao and Jang, 2007; Jaoui et al., 2008; Liu et al., 2016b; Xu et al., 2016).”

Comment 15: 1. 254 Do you see water soluble low-MW compounds in the gas-phase using the PTRMS and an increase of the uptake of these compounds with increasing SO₂ concentrations?

Response to comment 15: The time-series changes in the gas-phase concentrations of acetic acid under different conditions were discussed in the revised manuscript and added in the Supplement (shown in Figure R6). The results showed that acetic acid concentration decreased in the presence of SO₂, suggesting that the uptake of acetic acid might be enhanced.

Revisions in the manuscript:

1. Figure R6 has been added in the Supplement.
2. **Lines 342-349, Add:** “This is well supported by the time-series variations in the concentrations of acetic acid at different SO₂ concentrations measured by the HR-ToF-PTRMS (Fig. S15a), which shows that acetic acid concentration decreased with the increase of SO₂ concentration (0–56 ppbv). These results were in good agreement with those reported by Liggió et al. (2005) and Liu et al. (2010), who

observed that that the uptake of organic compounds under acidic conditions would be enhanced significantly. Recently, Huang et al. (2016) have also reported that acetic acid is present in SOA formed via α -pinene ozonolysis and its uptake would increase in the presence of seed particles.”

Comment 16: L 253 How did you quantify the amount of organosulfates? In Figure S6: Is this MS from a standard or from smog chamber experiment? Unclear. In Huang et al., 2015, they quantified using a commercial standard. What is the uncertainty in your measurement considering that the C_5H_8O and $C_5H_8O_2$ are highly abundant (and the resolution seems lower comparing to Huang et al. 2015) and can induce error during the peak fitting?

Response to comment 16: The calculation method of organosulfate concentration has been added in the Supplement, which has already been pointed out in the responses to Referee #2’s Comment 6. Thus, it was not listed in the following revisions.

Figure S6 in the original Supplement is the mass spectra obtained from smog chamber experiment. In addition, the uncertainties of the estimated concentration of organosulfate have been added in the revised manuscript.

Revisions in the manuscript:

Line 334, Change “2.1–4.3 ng m⁻³” **To** “(2.1 ± 0.8) to (4.3 ± 1.7) ng m⁻³”

Line 479, Change “2.2–4.6 ng m⁻³” **To** “(2.2 ± 0.7) to (4.6 ± 1.8) ng m⁻³”

Comment 17: L 261 Would it be possible to provide a potential chemical mechanism

that could lead to the formation of organosulfate (or organic sulfur containing compounds) through heterogeneous uptake of SO₂?

Response to comment 17: According to the results reported previously, organic S-containing compounds might be produced by the nucleophilic addition of HSO₄⁻ or the electrophilic addition of H₂SO₄ on the corresponding groups of organic molecules (Liggio and Li, 2006; Surratt et al., 2008), or through the reactions initiated by sulfate radicals (SO₄^{-•}) which are formed by sulfates under UV irradiation (Nozière et al., 2010). Considering that organic S-containing compounds are not identified in this work due to the lack of analytical instruments, it is very difficult for us to propose a chemical mechanism about the formation of organic S-containing compounds.

Comment 18: L 323 Do you see chlorine containing compounds (organics) or fragments using the AMS or the PTR-MS? Do you think they could participate to SOA formation?

Response to comment 18: Chlorine-containing compounds or fragments are not detected by HR-ToF-PTRMS and HR-ToF-AMS. Its concentration might be below the detection limits of analytical instruments.

Comment 19: 1-362 Do you see a higher SO₂ uptake in the presence of NaCl comparing with (NH₄)₂SO₄? This could illustrate the enhanced SO₂ uptake and explain the higher SOA formation.

Response to comment 19: The time-series decays of SO₂ with different seed particles

are shown in Figure R7, which shows that there is no significant difference of SO₂ decay with different seed particles. However, Figure R6 shows that acetic acid concentration decreases in the presence of SO₂ and seed particles, suggesting that the uptake of acetic acid might be enhanced.

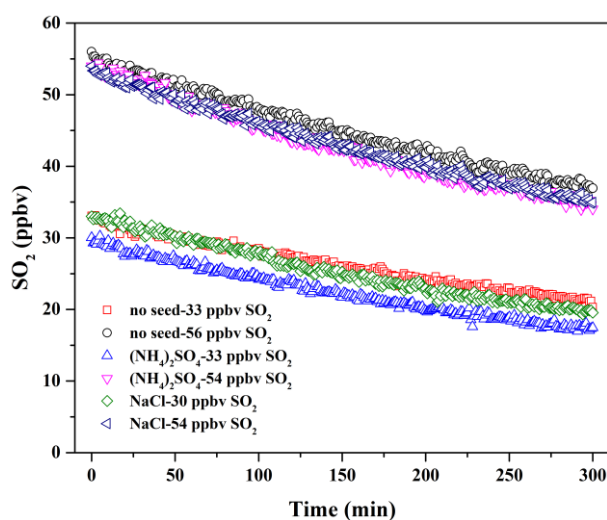


Figure R7. Decays of SO₂ as a function of irradiation time with different seed particles.

Revisions in the manuscript:

1. Figures R6 and R7 have been added in the Supplement.
2. Lines 467-471, Add: “This hypothesis could be supported by the variations in acetic acid concentration in the presence of different seed particles and SO₂ concentrations (Fig. S15), which shows that acetic acid concentration decreased with the increase of SO₂ concentration (0–54 ppbv).”

Comment 20: 1. 368 Do you see a difference in the PTR-MS signal for low MW compounds in the presence of SO₂ for NaCl and (NH₄)₂SO₄? (a larger uptake of these molecules in the presence of NaCl?).

Response to comment 20: In this work, we observed that acetic acid concentration decreased with increasing SO₂ concentration when using NaCl and (NH₄)₂SO₄ as seed particles, suggesting that the uptake of acetic acid might be enhanced. The time-series changes in the gas-phase concentrations of acetic acid under different conditions are shown in Figure R6. Compared to (NH₄)₂SO₄ as seed particles, the uptake of acetic acid on NaCl in the presence of SO₂ was larger.

Revisions in the manuscript:

1. Figure R6 has been added in the Supplement.
2. Lines 467-471, Add: “This hypothesis could be supported by the variations in acetic acid concentration in the presence of different seed particles and SO₂ concentrations (Fig. S15), which shows that acetic acid concentration decreased with the increase of SO₂ concentration (0–54 ppbv).”

Comment 21: Typing errors: l. 164 values l. 166 partitioned l. 335 inorganic SI l. 31 Formation Figure S5: signal.

Response to comment 21: All type errors have been corrected in the revised manuscript.

Revisions in the manuscript:

1. Line 196, Change “vaules” To “values”
2. Line 198, Change “partitioned” To “partitioned”
3. Line 356, Change “inorganic” To “inorganic”
4. Supplement, Line 80, Change “Fomration” To “Formation”

5. Supplement, Figures S13, S21 and S22, Change “Singal” To “Signal”

References

- Ahmad, W., Coeur, C., Tomas, A., Fagniez, T., Brubach, J.-B., and Cuisset, A.: Infrared spectroscopy of secondary organic aerosol precursors and investigation of the hygroscopicity of SOA formed from the OH reaction with guaiacol and syringol, *Appl. Opt.*, 56, E116-E122, doi: 10.1364/ao.56.00e116, 2017.
- Cao, G., and Jang, M.: Effects of particle acidity and UV light on secondary organic aerosol formation from oxidation of aromatics in the absence of NO_x, *Atmos. Environ.*, 41, 7603-7613, doi: 10.1016/j.atmosenv.2007.05.034, 2007.
- Jaoui, M., Edney, E. O., Kleindienst, T. E., Lewandowski, M., Offenberg, J. H., Surratt, J. D., and Seinfeld, J. H.: Formation of secondary organic aerosol from irradiated alpha-pinene/toluene/NO_x mixtures and the effect of isoprene and sulfur dioxide, *J. Geophys. Res.-Atmos.*, 113, doi: 10.1029/2007jd009426, 2008.
- Lauraguais, A., Coeur-Tourneur, C., Cassez, A., Deboudt, K., Fourmentin, M., and Choel, M.: Atmospheric reactivity of hydroxyl radicals with guaiacol (2-methoxyphenol), a biomass burning emitted compound: Secondary organic aerosol formation and gas-phase oxidation products, *Atmos. Environ.*, 86, 155-163, doi: 10.1016/j.atmosenv.2013.11.074, 2014.
- Liggio, J., and Li, S.-M.: Organosulfate formation during the uptake of pinonaldehyde on acidic sulfate aerosols, *Geophys. Res. Lett.*, 33, doi:10.1029/2006GL026079, 2006.
- Liggio, J., Li, S.-M., and McLaren, R.: Reactive uptake of glyoxal by particulate matter, *J. Geophys. Res.-Atmos.*, 110, doi:10.1029/2004JD005113, 2005.

- Liu, T., Wang, X., Hu, Q., Deng, W., Zhang, Y., Ding, X., Fu, X., Bernard, F., Zhang, Z., Lu, S., He, Q., Bi, X., Chen, J., Sun, Y., Yu, J., Peng, P., Sheng, G., and Fu, J.: Formation of secondary aerosols from gasoline vehicle exhaust when mixing with SO₂, *Atmos. Chem. Phys.*, 16, 675-689, doi: 10.5194/acp-16-675-2016, 2016.
- Liu, Z., Ge, M., Yin, S., and Wang, W.: Uptake and reaction kinetics of α -pinene and β -pinene with sulfuric acid solutions, *Chem. Phys. Lett.*, 491, 146-150, doi: 10.1016/j.cplett.2010.04.004, 2010.
- Nozière, B., Ekström, S., Alsberg, T., and Holmström, S.: Radical-initiated formation of organosulfates and surfactants in atmospheric aerosols, *Geophys. Res. Lett.*, 37, doi:10.1029/2009GL041683, 2010.
- Surratt, J. D., Gómez-González, Y., Chan, A. W. H., Vermeylen, R., Shahgholi, M., Kleindienst, T. E., Edney, E. O., Offenberg, J. H., Lewandowski, M., Jaoui, M., Maenhaut, W., Claeys, M., Flagan, R. C., and Seinfeld, J. H.: Organosulfate formation in biogenic secondary organic aerosol, *J. Phys. Chem. A*, 112, doi: 8345-8378, 10.1021/jp802310p, 2008.
- Xu, L., Middlebrook, A. M., Liao, J., de Gouw, J. A., Guo, H., Weber, R. J., Nenes, A., Lopez-Hilfiker, F. D., Lee, B. H., Thornton, J. A., Brock, C. A., Neuman, J. A., Nowak, J. B., Pollack, I. B., Welti, A., Graus, M., Warneke, C., and Ng, N. L.: Enhanced formation of isoprene-derived organic aerosol in sulfur-rich power plant plumes during Southeast Nexus, *J. Geophys. Res.-Atmos.*, 121, 11137-11153, doi: 10.1002/2016jd025156, 2016.

Yee, L. D., Kautzman, K. E., Loza, C. L., Schilling, K. A., Coggon, M. M., Chhabra, P. S., Chan, M. N., Chan, A. W. H., Hersey, S. P., Crouse, J. D., Wennberg, P. O., Flagan, R. C., and Seinfeld, J. H.: Secondary organic aerosol formation from biomass burning intermediates: phenol and methoxyphenols, *Atmos. Chem. Phys.*, 13, 8019-8043, doi: 10.5194/acp-13-8019-2013, 2013.

Zhang, X., Cappa, C. D., Jathar, S. H., McVay, R. C., Ensberg, J. J., Kleeman, M. J., and Seinfeld, J. H.: Influence of vapor wall loss in laboratory chambers on yields of secondary organic aerosol, *Proc. Natl. Acad. Sci. U. S. A.*, 111, 5802-5807, doi: 10.1073/pnas.1404727111, 2014.