Overview and Preliminary Results of the Surface Ocean Aerosol Production (SOAP) campaign

5 Cliff S. Law^{1,2}, Murray J. Smith¹, Mike J. Harvey¹, Thomas G. Bell^{3,4}, Luke T. Cravigan^{5,6}, Fiona C. Elliott¹, Sarah J. Lawson⁷, Martine Lizotte⁸, Andrew Marriner¹, John McGregor¹, Zoran Ristovski⁶, Karl A. Safi⁹, Eric S. Saltzman⁴, Petri Vaattovaara¹⁰, Carolyn F. Walker¹

¹ National Institute of Water and Atmospheric Research, Wellington, New Zealand

⁶ International Laboratory for Air Quality and Health, Queensland University of Technology, Brisbane, Australia

⁸ Université Laval, Department of Biology (Québec-Océan), Québec City, Québec, Canada.

⁹ National Institute of Water and Atmospheric Research, Hamilton, New Zealand

¹⁰ University of Eastern Finland, Kuopio, Finland

Correspondence to: Cliff S. Law (cliff.law@niwa.co.nz)

20

15

Abstract. Establishing the relationship between marine boundary layer (MBL) aerosols and surface water biogeochemistry is required to understand aerosol and cloud production processes over the remote ocean, and represent them more accurately in Earth System Models and global climate projections. This was addressed by the SOAP (Surface Ocean Aerosol Production) campaign, which examined air-sea interaction over biologically-productive frontal waters east of New Zealand. This overview

- 25 details the objectives, regional context, sampling strategy, and provisional findings of a pilot study, PreSOAP, in austral summer 2011, and the following SOAP voyage in late austral summer 2012. Both voyages characterised surface water and MBL composition in three phytoplankton blooms of differing species composition and biogeochemistry, with significant regional correlation observed between chlorophyll-*a* and DMSsw. Surface seawater dimethylsulfide (DMSsw) and associated air-sea DMS flux showed spatial variation during the SOAP voyage, with maxima of 25 nmol L⁻¹ and 100 µmol m⁻² d⁻¹,
- 30 respectively, recorded in a dinoflagellate bloom. Inclusion of SOAP data in a regional DMSsw compilation indicates that the current climatological mean is an underestimate for this region of the South-west Pacific. Estimation of the DMS gas transfer velocity (k_{DMS}) by independent techniques of eddy covariance and gradient flux showed good agreement, although both exhibited periodic deviations from model estimates. Flux anomalies were related to surface warming and sea surface microlayer enrichment, and also reflected the heterogeneous distribution of DMSsw and the associated flux footprint. Other
- 35 aerosol precursors measured included the halides and various volatile organic carbon compounds, with first measurements of the short-lived gases glyoxal and methylglyoxal in pristine Southern Ocean marine air indicating an unidentified local source. The application of a real-time clean-sector, contaminant markers, and a common aerosol inlet facilitated multi-sensor measurement of uncontaminated air. Aerosol characterisation identified variable Aitken mode, and consistent sub-micron sized accumulation and coarse modes. Sub-micron aerosol mass was dominated by secondary particles containing ammonium
- 40 sulfate/bisulfate under light winds, with an increase in sea-salt under higher wind-speeds. MBL measurements and chamber experiments identified a significant organic component in primary and secondary aerosols. Comparison of SOAP aerosol number and size distributions reveals an underprediction in GLOMAP-mode aerosol number in clean marine air masses, suggesting a missing marine aerosol source in the model. The SOAP data will be further examined for evidence of nucleation

² Department of Chemistry, University of Otago, Dunedin, New Zealand

³ Plymouth Marine Laboratory, Prospect Place, The Hoe, Plymouth, UK

⁴ Department of Earth System Science, University of California, Irvine, CA, USA

⁵ Climate Science Centre, Commonwealth Scientific and Industrial Research Organisation, Aspendale, Australia

⁷Commonwealth Scientific and Industrial Research Organisation, Oceans and Atmosphere Flagship, Aspendale, Australia

events, and also to identify relationships between MBL composition and surface ocean biogeochemistry that may provide potential proxies for aerosol precursors and production.

1. Introduction

45

It is recognised that the surface ocean alters the properties of the lower atmosphere, and so atmospheric albedo and climate (McCoy et al., 2015; Seinfeld et al., 2016), via the direct and indirect effects of aerosols (O'Dowd and de Leeuw, 2007). Aerosols are precursors of clouds, which play a major role in the scattering and absorption of incident solar radiation (Carslaw 50 et al., 2013), but the concentration, number and chemical properties of aerosols that act as cloud-condensation nuclei (CCN) can also influence cloud droplet size and number, and consequently precipitation and cloud albedo (Twomey, 1977). Indeed, cloud formation and properties are sensitive to relatively minor changes in aerosol concentration, particularly in remote regions (Carslaw et al., 2013). This is particularly the case in the Southern Ocean, where natural aerosol sources dominate and where CCN concentrations can range from tens per cm³ in winter to hundreds per cm³ in summer (Andreae and Rosenfeld, 2008), 55 leading to seasonally variant trends in cloud albedo. However, the relationship between clouds and aerosols derived from natural sources is poorly understood, and represents a major uncertainty in the representation of low-level marine clouds and feedbacks in climate models (Wang et al., 2013; Stephens, 2005). Current models underestimate cloud over the Southern Ocean, particularly south of 55°S, resulting in excess surface shortwave radiation and a warm bias (Trenberth and Fasullo, 2010; Kay et al., 2016). This discrepancy is potentially attributable to a variety of factors, chief among which is the limited 60 understanding of aerosol-cloud interaction and cloud water phase, compounded by a lack of regional observations and data to advance satellite retrievals and climate model simulations.

Breaking waves and associated bubble formation are a major source of Primary Marine Aerosol (PMA), supplying most the aerosol mass in the marine boundary layer (MBL) over the remote ocean (Andreae and Rosenfeld, 2008), and particularly in regions that experience high winds and breaking waves (de Leeuw et al., 2014). This is reflected in PMA contributing only

- 65 ~10-20% of CCN number concentrations over the remote Pacific Ocean (Blot et al. 2013; Clarke et al. 2013), but up to 55% over the Southern Ocean (McCoy et al. 2015). Although PMA is generally regarded as primarily composed of sea-salt, recent reassessments suggest it is highly enriched in organic matter relative to bulk seawater. Organic material may in fact dominate submicron aerosol mass (Facchini et al., 2008; O'Dowd et al., 2004), with the Primary Organic Aerosol (POA) being of biogenic origin and including bacteria, carbohydrates, polymers and gels (Facchini et al., 2008; Russell et al., 2010). Although
- 70 the contribution of POA to the MBL is uncertain, it may be significant over biologically active oceanic regions, as suggested by correlations between organic aerosol content and surface chlorophyll-a (Chl-a) (O'Dowd et al., 2004). There is also similarity in the composition of aerosol and surface ocean organics, and organically enriched sub-micron particles have been produced experimentally using surface seawater conditions (Quinn and Bates, 2011). Indeed, the degree of organic enrichment may influence both the type and size of aerosols, as well as properties such as aerosol light scattering and water uptake (Vaishya 75 et al., 2012).

80

It is well-established that biologically productive regions are characterised by elevated concentrations and emissions of a range of compounds that may influence aerosol production, composition and properties (Meskhidze and Nenes, 2010; Gantt and Meskhidze 2013; de Leeuw et al., 2014). However, the oceanic influence on atmospheric composition is not only attributable to PMAs but also to secondary marine aerosols (SMAs), that are produced during gas-phase reactions of volatile organic compounds (VOCs). Although SMAs have less impact upon aerosol mass they potentially have a large influence on aerosol number (Meskhidze et al., 2011). The biogeochemical origin of SMAs is reflected in their seasonality, with Aitken and accumulation mode aerosol number concentrations dominated by secondary particles in summertime (Clarke et al. 2013; Cravigan et al. 2015). Research into SMAs has primarily focussed on dimethylsulfide (DMS), the primary natural marine source of volatile sulfur, in response to early hypotheses related to its potential role in climate feedback processes (Charlson

- et al., 1987). The CLAW hypothesis linked the production of the DMS precursor, dimethylsulfoniopropionate (DMSP), by phytoplankton and subsequent DMS emission and oxidation to sulfate aerosol, to CCN formation and changes in cloud cover. Although well-studied, this hypothesis remains unproven and there is a lack of consensus, with a recent review identifying uncertainties regarding the role of DMS in aerosol production in the MBL (Quinn and Bates, 2011). However, there is evidence that DMS may play a role in cloud formation over larger spatial and temporal scales, via entrainment from the free troposphere
- **90** (Carslaw et al., 2010).

The fundamental tenet of the CLAW hypothesis, of feedback between surface ocean biogeochemistry and climate, may be applicable via a broader spectrum of precursor species. Recent research has shown increasing complexity of potential aerosol source pathways, involving a variety of chemical species, processes and interactions (Vaattovaara et al., 2006). In addition to DMS, a variety of other gaseous aerosol precursors that originate from phytoplankton, bacterial and photochemical sources at

- 95 the sea surface may undergo physical and chemical transformation to produce new particles in the MBL (Ciuraru et al., 2015). These SMA precursors include volatile organic species, such as carboxylic acids, isoprene, monoterpenes, halocarbons, iodine oxides and iodine (Vaattovaara et al., 2006; Sellegri et al., 2005). A biological source of these SMAs has been inferred from the spatial and temporal correlation between phytoplankton blooms and cloud microphysics (Meskhidze et al., 2009; Meskhidze and Nenes, 2010; Lana et al. 2012). The presence and concentration of SMA precursors in the MBL may be
- 100 dependent upon plankton abundance and community composition, and consequently their influence on aerosol formation will show spatial and seasonal variability (O'Dowd et al., 2004).

New particle formation may be suppressed by the interaction of aerosol precursors and SMAs with pre-existing aerosol, for example, by absorption of ammonia and gaseous sulfuric acid by coarse mode sea-salt aerosol (SSA; Cainey and Harvey, 2002). Conversely, existing particles may grow via condensation which enhances their CCN capacity (Clarke et al., 2013). It

- 105 has also been proposed that organic acids combine with sulfuric acid to create the critical nucleus required for aerosol formation (Zhang, 2010; Almeida *et al.* 2013). However, nucleation events over the open ocean remain elusive (O'Dowd et al., 2010; Chang et al., 2011; Willis et al., 2016), making it difficult to elucidate the primary pathways and reactants, and consequently they are currently regarded as of low significance to marine aerosol formation. Following nucleation, the aerosol distribution is modified by aerosol-aerosol interaction, heterogenous reactions and removal processes, including coagulation and
- 110 condensation, resulting in the longest-lived aerosol component being in the accumulation mode (0.06-0.4µm). With such a wide variety of potential precursors and inorganic/organic interactions affecting nucleation and CCN activation, the modelling of aerosols and their indirect influence on cloud radiative properties over the remote ocean presents a major challenge (Seinfeld et al., 2016).
- The production and transfer of aerosol precursors from the ocean surface is also dependent upon physical factors. Exchange across the air-sea interface is primarily controlled by near-surface turbulence, which is dependent on wind and waves. For practical purposes, this is represented by a kinetic factor, the transfer velocity k which is generated with wind-speed parameterisations (Nightingale et al., 2000; Ho et al., 2006). Although wind-speed provides a reasonable broad-scale proxy for kinetic transfer, other factors such as fetch, wave development, wind-wave direction and surfactants also influence k, and so generate variation in gas exchange and deviation from k-wind-speed relationships. For example, most k-wind-speed
- 120 parameterisations do not explicitly capture the solubility effects associated with bubbles (Blomquist et al. 2006), although the COAREG gas transfer model incorporates this factor into a physically-based flux algorithm (Fairall et al., 2003; Fairall et al. 2011). Biogeochemical gradients near or at the ocean surface are also not considered, despite their potential to alter the air-sea exchange of gases, PMAs and SMAs (Facchini et al., 2008; Calleja et al., 2013).

Previous related research campaigns have examined the biogeochemical and physical factors influencing oceanic DMS and

- 125 CO₂ fluxes, as summarised in Suppl. Table 1, but few have linked this to the physical controls of air-sea exchange, and variation in aerosol and trace gas composition of the MBL. Similarly, other campaigns with an atmospheric focus, such as MAP (Decesari et al., 2011), have carried out detailed studies of aerosol chemistry, but not interpreted this with regard to surface ocean biogeochemistry. To address this, the Surface Ocean Aerosol Production (SOAP) campaign was initiated, with the primary aim of characterising the variation in aerosol composition and concomitant marine sources, processes and pathways
- in the South-west Pacific. SOAP utilised a multi-disciplinary framework, encompassing surface ocean biology and biogeochemistry, transport and air-sea exchange with characterisation of aerosol number and composition, to establish controls on aerosols and gas exchange. The campaign consisted of two voyages a pilot study, PreSOAP, which carried out a regional survey and established sampling strategies and the following SOAP voyage, in biologically productive frontal waters along the Chatham Rise, east of New Zealand (see Figure 1). Building upon the approaches used in previous studies, the SOAP campaign targeted three phytoplankton blooms of differing plankton community composition, to determine their respective
- influences on biogeochemistry, gas exchange and MBL composition. The following paper details the regional context, sampling strategy, environmental conditions and some preliminary results for the SOAP campaign.

2. Regional context

- The South-west Pacific has many features in common with the Southern Ocean, as it is characterised by low anthropogenic and terrestrial aerosol loading, long ocean fetch and high wind-speed, making it an optimal location for examining the marine contribution to aerosol production. One of the more biologically productive regions lies east of New Zealand, where the Sub-Tropical Front (STF) extends as a tongue of elevated phytoplankton production (Murphy et al., 2001), along 43.0-43.5°S over the Chatham Rise (see Figure 1a). This arises from the confluence of warmer saline subtropical waters that are relatively deplete in macronutrients, with fresher cooler subantarctic waters containing elevated macronutrients but depleted in iron (see
- 145 Figure 1b; Boyd et al., 1999). Mixing across the front alleviates nutrient stress which, combined with a relatively stable water column, promotes primary production (Chiswell et al., 2013). Ocean colour climatologies show a monthly mean Chl-*a* of 0.6 mg m⁻³, reaching ~ 1 mg m⁻³ over the Chatham Rise in spring (Murphy et al., 2001), and the region is characterised by elevated marine particle export, secondary production and fish stocks (Nodder et al., 2007; Bradford-Grieve et al., 1999). In spring the phytoplankton community composition varies with water mass, with diatoms dominating the STF, cryptophytes, prasinophytes
- 150 and dinoflagellates more prevalent in subtropical waters, and photosynthetic nanoflagellates dominating subantarctic waters (Chang and Gall, 1998; Delizo et al., 2007). The STF also supports spatially-extensive coccolithophore blooms (Sadeghi et al., 2012), and is situated on the northern edge of the "Great Calcite Belt" (Balch et al., 2011), a latitudinal band of elevated backscatter attributed to coccolithophore liths. Surface mixed layer nutrients vary spatially in response to mixing of the water masses and seasonally due to phytoplankton uptake, with the evolution of nutrient stoichiometry and grazing determining the succession and duration of different phytoplankton blooms (Chang and Gall, 1998; Delizo et al., 2007). The STF is
- succession and duration of different phytoplankton blooms (Chang and Gall, 1998; Delizo et al., 2007). The STF is characterised by significant gradients in pCO₂ associated with phytoplankton blooms, with current global climatologies indicating the region east of New Zealand as a significant carbon sink (>1mol C m⁻² yr⁻¹, Landschuetzer et al., 2014).

The waters south of New Zealand are characterised by high wind-speeds which drive the disproportionate contribution of this region to global ocean CO₂ uptake. Here, wind, waves and currents develop unhindered by land, and strong persistent westerlies

160 act over long fetch to generate large swells that propagate north-east influencing the wave-climate off New Zealand. While this wave energy is attenuated closer to land in the eastern Chatham Rise, the average wave energy is still 75% of values south of New Zealand where annual mean wave heights exceed 4m. Subantarctic waters south of the Chatham Rise region provided a prime location for a dual tracer release experiment (SAGE; Harvey et al., 2011), aimed at constraining *k* at high wind-speeds. Comparison of the SAGE *k*-wind-speed parameterisation with those generated in other regions, and using different techniques,

- 165 showed generally good agreement (Ho et al., 2006); this may be interpreted as indicating that regional influences on exchange may be less important, supporting the application of a universal wind-speed parameterisation. Nevertheless, other factors, such as wave age, duration and height do influence gas exchange in this region (Smith et al., 2011; Young et al., 2012). The elevated winds also influence the transfer of aerosols and precursors, as reflected by a zonal band of elevated sea spray aerosol mass and water-insoluble organic matter over the Chatham Rise region (Vignati et al., 2010).
- 170 Both models and measurements indicate that DMS is a significant contributor to total non-sea salt sulfate (nssSO4) in the Southern Hemisphere (Gondwe et al., 2003; Korhonen et al., 2008). However, a paucity of observational data in the Southern Ocean has hindered development of global climatologies for surface seawater DMS (DMSsw), with the region south-east of New Zealand represented by only a few data points in a recent DMS climatology (Lana et al., 2011). Despite this shortcoming, this climatology provides a realistic representation of atmospheric DMS and total sulfate when applied in aerosol-climate
- 175 Global Climate Models, particularly over the Southern Ocean (Mahajan et al., 2015). Seasonal variability in atmospheric DMS is apparent at stations in New Zealand and south of 44°S (Blake et al., 1999), with concentrations of 100-200 pptv, and maximal values associated with the transport of DMS from waters to the south in summer (Harvey et al., 1993; de Bruyn et al., 2002; Wylie and de Mora, 1996). Corresponding seasonality in nssSO4 was observed, with a maximum (0.8-1.5 µg m⁻³) in early austral summer at the start of the year, decreasing in late summer to 0.1-0.4 µg m⁻³ through autumn and winter (see Figure. 2;
- Sievering et al., 2004; Allen et al., 1997). For comparison, coarse SSA dominates the aerosol mass at Baring Head, with concentrations of 6-10 µg m⁻³ (Jaeglé et al., 2011; Spada et al., 2015). Similar seasonal cycles of DMS and nssSO4 were recorded at Cape Grim (Ayers, 1991), and the observed diurnal inverse correlation between sulfur dioxide and DMS at Baring Head was applied to estimate yield and potential contribution to aerosols (de Bruyn et al., 2002). Consistent seasonal trends between activated particles and cloud droplet number concentration were also apparent, with a summer maximum over the
- 185 Southern Hemisphere (Boers et al., 1996; 1998), related to phytoplankton production (Thomas et al., 2010). Overall, the temporal trends in aerosol precursors and pathways do not follow that of wind-speed and other physical drivers, but instead reflect biological processes inferring control by surface ocean biogeochemistry (Korhonen et al., 2008).

3. Research Programme and Strategy

3.1 PreSOAP

- A pilot study, PreSOAP, was carried out to test technical approaches and confirm the regional source of biogenic aerosols in the Chatham Rise region on the New Zealand research vessel, *Tangaroa*, on 1-12/2/2011 (DoY32-42). The strategy of bloom location using satellite imagery and subsequent mapping of surface properties proved successful, with three blooms of differing DMSsw and pCO₂ signatures located and monitored each for 3-4 days. The first bloom was initially dominated by dinoflagellates with an increase in diatom biomass after 3 days, while the second and third blooms were primarily dominated by coccolithophores and dinoflagellates, respectively. This variability in species composition resulted in significant spatial and temporal variability in DMS concentrations in the MBL (DMSa) and DMSsw. DMSa concentration varied over two orders of magnitude, reaching 1000 ppt on DoY 36 (see Figure 3b), similar in range to that recorded at the Baring Head station near Wellington (Harvey et al., 1993; de Bruyn et al., 2002). There was no significant correlation between DMS in the two phases,
- 200 but there was no significant relationship between DMSsw and Chl-*a*, with the DMSsw maximum of ~10 nmol L^{-1} during the first bloom coinciding with Chl-*a* of ~1 mg m⁻³ (Figure 3d). The observed temporal and spatial variability in DMSa and DMSsw during PreSOAP highlighted the technical challenge of establishing relationships between surface ocean biogeochemistry and

with DMSa showing a stronger relationship with wind-speed (see Figure 3). Surface Chl-a concentrations reached 2 mg m⁻³,

atmospheric composition. Provisional method development was also carried out for measurement of DMS and other parameters in near-surface waters and the sea surface microlayer (SSM).

205 Surface DMSsw and pCO₂ were mapped, and DMSa and CO₂ MBL concentrations and fluxes measured continuously by sensors and collectors mounted on the bow of the vessel. Testing of the eddy covariance (EC) flux technique identified an issue with water vapour interference that dominated the CO₂ signal recorded by an open-path InfraRed Gas Analyser (IRGA). Preliminary studies also identified that residual ship motion dominated over turbulence for the real-time switching of Relaxed Eddy Accumulation measurement of flux under high swell conditions. The logistical challenges of flux measurement at distance from the vessel were also assessed by deployment of a free-floating catamaran supporting a mounted gradient flux sampling system (Smith et al., to be submitted). A temperature microstructure profiler was also deployed to record near-surface temperature and turbulence structure (Stevens et al., 2005), although this was limited to short sampling periods, highlighting the need for a mounted thermistor array on a spar buoy for longer measurement coverage.

The utility of a baseline sector for sampling MBL composition, using relative wind direction and speed, was also tested during
PreSOAP. Measurements showed a tendency for higher condensation nuclei concentration in the "non-baseline" sector, confirming the utility of this approach (Harvey et al., to be submitted). A common aerosol inlet provided clean air from a height of 17.5m above sea level to instruments and sensors in a container laboratory on deck. Particle size distribution and concentration, including ultrafine nuclei concentrations, were continuously monitored using a scanning mobility particle sizer (SMPS) and optical particle counters (OPC's), with bulk ion chemistry samples collected using a high-volume sampler. The composition of primary marine aerosols was also examined using a 0.45m³ bubble chamber, in which sea spray was formed

via the bursting of bubbles produced by passing clean compressed air through sintered glass (Mallet et al., 2016).

3.2 The SOAP voyage

The SOAP voyage employed the strategy successfully piloted on PreSOAP, of identifying phytoplankton blooms in NASA MODIS Aqua and Terra satellite ocean colour images, with subsequent bloom location and mapping using a suite of underway 225 sensors (Chl-a, β_{660} backscatter, pCO₂, DMSsw). The blooms were discrete and coherent areas of elevated ocean colour, that were provisionally characterised by a concentration of 1 mg/m3 Chl-a or higher. For each bloom, a nominal centre was identified, based upon maximum DMSsw and Chl-a concentrations, and marked by deployment of a Spar Buoy. Repeat activities at the bloom centre included characterisation of the surface mixed layer by vertical profiling, collection of SSM samples at distance from the main vessel, and gradient flux on a catamaran. Overnight mapping was carried out to determine 230 changes in bloom magnitude and position. Sampling also took place at stations on the periphery and outside the blooms, as defined by distance from the bloom centre and a clear demarcation in surface biogeochemical variables. The SOAP voyage was nominally divided into three different bloom periods (see Figure 4), with an initial dinoflagellate bloom (B1) located 12 hours into the SOAP voyage that exhibited elevated Chl-a and DMSsw, and pCO₂ drawdown, a coccolithophore bloom (B2) with initially moderate signals that weakened, and a final bloom (B3) of mixed community composition. Following a storm, 235 the surface water column structure and biogeochemistry were significantly different, and so this bloom was subdivided into B3a and B3b.

3.2.1 Environmental conditions during the SOAP voyage

240

Back-trajectory analysis of particle density was calculated for each bloom using the Lagrangian Numerical Atmosphericdispersion Modelling Environment (NAME) for the lower atmosphere (see Figure 5). The meteorological situation evolved over the SOAP voyage from a high-pressure system with light winds during B1, to stronger winds during B2 and B3. The main weather features included a depression crossing the central South Island on DoY 54-55 during B2, and a second depression from the east from DoY 58 onwards. During B3 a vigorous front advanced up the east coast of the South Island on DoY 61 with strong SW winds of 20 m s⁻¹, followed by a depression crossing the lower North Island on DoY 63 that maintained a fresh southerly airflow for the remainder of the voyage. Air and water temperatures during B1were generally similar

- 245 indicating near-neutral stability, whereas B2 experienced a period of warm, moist air and reversal in direction of turbulent heat fluxes, followed by a short period when air temperatures were 2-3°C higher on DoY 56-58 (See Figure 6). Waves were dominated by swell from the south-southwest, with significant wave height mirroring trends in wind-speed, reaching a 5m maximum during the localised storm on DoY 61 (see Figure 6). Wave parameters obtained from NOAA WaveWatch III analyses indicated that wave height was 23% lower during B1 and B3, and 13% lower at B2, relative to wave height south of 250 New Zealand at 50°S.

255

Table 1 summarises the hydrographic and biogeochemical characteristics in the surface mixed layer of the three phytoplankton bloom regions. B1 was a large dinoflagellate bloom with high surface DMSsw (maximum ~30 nmol L⁻¹; mean 16.8 nmol L⁻¹; Bell et al., 2015), and Chl-a (maximum 3.4 mg m⁻³), and significant CO₂ undersaturation with a mean surface pCO₂ of 320 ppmv (see Table 1). B1 was located, south of the Mernoo bank, a deep channel between the western end of Chatham Rise and the east coast of the South Island. This region has been previously identified as a prime location for phytoplankton blooms,

- due to eddy-driven mixing and flow reversals arising from current and topographic interaction, which enhance iron and nutrient supply (Boyd et al., 2004). During B1, winds remained light (see Table 1) with a calm sea state, and the spar buoy drifted north-east primarily under the action of surface currents. Solar irradiance was high and a shallow surface mixed layer developed (see Figure 6), with a significant near-surface temperature gradient (Walker et al., 2016). Mean nitrate and phosphate
- 260 concentrations (5.3 and 0.4 μ mol L⁻¹, respectively) were sufficient for phytoplankton growth, whereas silicate was low (see Table 1), and close to growth-limiting concentrations (Boyd et al., 1999). Although dinoflagellates dominated, coccolithophores biomass was higher at some stations, and nanoeukaryote abundance was generally low. B1 was occupied for 5-6 days, during which broader regional excursions with overnight mapping identified a bloom of high Chl-a but relatively low DMSsw to the south-west.
- 265 The vessel re-located to a coccolithophore bloom, B2, evident at the eastern end of the Chatham Rise in MODIS true colour satellite images (see Figure 4b). Upon arrival on DoY 52 B2 showed an initial mean DMSsw of 9 nmol L⁻¹ and elevated Chla, and was characterised by a relatively warmer, shallower, saltier surface mixed layer of lower nitrate concentration (compared to B1, see Table 1), typical of subtropical water. This appeared to provide optimal conditions for coccolithophores as surface water backscatter (β_{660}) was initially elevated by high lith abundances, with coccolithophores accounting for up to 40% of
- 270 phytoplankton carbon. However, intrusion of warm, moist air associated with north-westerly winds, coincided with a reversal in the direction of turbulent heat fluxes, and was followed by a southwest wind shift strengthening to 17 m s⁻¹ by DoY 56 (see Figure 6). This resulted in deepening and cooling of the surface mixed layer with a corresponding increase in nutrient concentrations which, combined with a decrease in solar irradiance, resulted in a decline in Chl-a and DMSsw (Bell et al., 2015).
- 275 Following the 5-day occupation of B2, the vessel returned to the south of Mernoo bank to assess a bloom that had developed near the original site of B1. Surface biogeochemical signals were initially weak in B3a, with a mixed community of coccolithophores and dinoflagellates and low DMSsw (2.2 nmol L⁻¹) and Chl-a (mean 0.39 mg m⁻³). However, an intense front advanced up the South Island and resulted in strong SW winds that exceeded 20 ms⁻¹ (see Figure 6), after which mixed layer depth and associated nutrients increased. Consequently, stations before and after the storm were physically and
- 280 biogeochemically disparate. B3a stations exhibited similar sea surface temperature to B1, but with a deeper surface mixed layer and a Chl-a half that of B1, whereas B3b stations were significantly cooler (at 13°C) and deeper (41m) than B1 (see Figure 7), with higher silicate concentration due the enhanced vertical mixing. Subsequent stabilisation of the surface mixed layer by light winds combined with elevated nutrients stimulated Chl-a, diatom and coccolithophore abundance in the final B3b stations (see Figures 6 and 7).

285 4. SOAP work programmes and observations

A number of parameters were measured (see Table 2) in three interlinked work programmes during the SOAP voyage, as indicated in Figure 8 and detailed below.

4.1. The distribution and composition of aerosols, precursors and trace gases in the MBL

Aerosol number concentration, size distribution, composition, water uptake and CCN concentration were measured semi-290 continuously during SOAP to address the overall paucity of aerosol observations, and the apparent rarity of nucleation events, over the remote ocean. These were characterised by a suite of instruments covering a particle size range of 0.01 to $10 \,\mu m$ (see Figure 9 and Table 2), which enabled determination of the size-dependent contribution of PMA and nssSO4 to aerosol and CCN concentrations. Aerosol characterisation identified variable Aiken and consistent sub-micron sized accumulation and coarse modes, with the sub-micron aerosol mass dominated by secondary aerosol with ammonium sulfate/bisulfate under light winds, and with an increase in sea-salt proportion as local winds increased. Ongoing data analysis is examining whether significant nucleation events occurred.

295

The operational mode for underway aerosol measurement was to slowly steam at 1-2 kts into the prevailing wind, across an area of high biological productivity or significant air-sea gas gradient, generally between noon and 2:00PM when solar irradiance was maximal. The common aerosol inlet developed during PreSOAP allowed uncontaminated air from above the 300 bridge to be sampled when the wind was on the bow, so minimizing interference from ship stack emissions. Contamination events were screened out using a real-time clean-sector sampling "baseline" flag and switch (Harvey et al., to be submitted), enabling clean collection of integrated samples. Although the vessel exhaust was the primary contaminant, other potential sources included the workboat and recirculation of polluted air around the ship, and longer range terrestrial influences were also assessed. Measurements of black carbon using an aethalometer, and CO₂ by high precision Cavity Ring-Down Laser

- 305 Spectroscopy (CRDS) provided two independent variables for detecting contamination events, and some VOCs, measured by PTR-MS (see Table 2), were also used as indicators of diesel combustion. The vessel was orientated into the wind as often as possible, which resulted in a high frequency (~75%) of baseline sector conditions during the SOAP voyage. Clean marine air periods were defined post-voyage using the baseline wind sector $(225 - 135^{\circ})$ relative to bow and wind speed greater than 3 m s^{-1}), black carbon concentrations (less than 50 ngm⁻³), and back trajectories indicating minimal terrestrial impact (periods when
- 310 the minimum number of hours over land in 72-hour back trajectory is zero), with periods of workboat operations also removed. An ensemble of Hybrid Single-Particle Lagrangian Integrated Trajectory (HYSPLIT) model back trajectories (Draxler and Rolph, 2013) was run for each hour of the voyage, and NAME back trajectories calculated for every three hours (Figure 5, Jones et al., 2007). Figure 10 shows particle number and CCN concentrations, compared to the number of hours the 72-hour back trajectory spent over land calculated from HYSPLIT trajectories. Particle concentrations were generally higher during
- 315 periods of terrestrial influence (see DoY 52 and 60, Figure 10), with average particle number concentrations of 1122 ± 1482 cm⁻³, double that observed for clean marine air. Ion beam analysis also revealed the presence of silicate and aluminium on ambient submicron filter samples suggesting a terrestrial source, supporting the back-trajectory modelling of continental outflow.

During the initial occupation of B1 under light winds, the particulate matter (PM10) total ion mass was 9.5 µg m⁻³ compared 320 to subsequent samples under higher winds in the range 20-50 µg m⁻³. The dominant components of the inorganic mass fraction were sea-salt ions and nssSO4, although a measurable organic fraction was also present (see below). The NaCl mass in light winds during B1 was 6.6 μ g m⁻³ with >95% of > 3 μ m diameter, relative to 32.5 μ g m⁻³ under stronger winds during B3b. Although 72% was > 3μ m, the largest difference in mass occurred in the 1.5 to 3μ m size range. In contrast, the mass of nssSO4 was predominantly sub-micron sized; B1 exhibited the largest nssSO4 mass at 2.0 μ g m⁻³ with 85% in sizes <1 μ m, 325 whereas in B3b, the nssSO4 mass was much lower at $0.6 \ \mu g \ m^{-3}$ with 76% in <1 μm sizes. These results confirm the influence of both physical and biogeochemical processes on aerosol composition.

Voyage particle number concentrations during clean marine periods averaged 534 ± 338 cm⁻³, with CCN concentrations of 178 ± 87 cm⁻³ (± 1 sd) at 0.5% supersaturation, and an average particle fraction activated into CCN of 0.4 ± 0.2 . Bloom average particle number concentrations ranged from a minimum of 385 ± 96 cm⁻³ in B3b to a maximum 830 ± 255 cm⁻³ at the start of

- B2 (Figure 10). B1 displayed the highest CCN activation ratio, of 0.5 ± 0.2 , potentially the combination of low wind speeds, large biogeochemical signals and SMA fluxes. Comparison of the inorganic ion mass determined from high-volume sampler filters between the different blooms does not support the conclusion that the B1 activation ratio was higher simply because particles were larger. As the median particle diameters during clean marine periods were consistent between the three blooms, this suggests that particle composition, secondary organics or coagulation may have impacted CCN activation at B1. These
- findings are supported by preliminary results from an application of the ACCESS-UKCA model (Woodhouse, pers comm.), which simulated the additional impact of emissions of marine secondary organic carbon under the conditions determined during SOAP. In contrast, the average CCN activation ratio for B3a was 0.13 ± 0.06 . Nucleation mode particles (10 nm and 15 nm), were measured by ultra-fine organic tandem differential mobility analyser (UFO-TDMA, Vaattovaara et al., 2005), and Aitken mode particles (50 nm), by UFO-TDMA and a volatility and hygroscopicity tandem differential mobility analyser
- 340 (VH-TDMA, Johnson et al. 2004a; Villani et al., 2008). This analysis typically identified a significant (up to 50% volume fraction) secondary organic component during sunny conditions in bloom regions, particularly during B1. The TDMA results provided further evidence for secondary organic aerosol processing of the dominant secondary nssSO4 mode during B1. Deliquescence measurements (VH-TDMA) indicate that the Aitken mode population is largely comprised of neutralised nssSO4 i.e. ammonium sulfate. Small and sporadic contributions to the Aitken mode from a non-hygroscopic (number fraction)
- 345 up to 0.4) and a highly hygroscopic component (number fraction up to 0.3) were observed in addition to the secondary nssSO4 mode (number fraction of 0.6 1). The water uptake and volatility of the sporadic highly hygroscopic mode indicates this may be composed of PMA.
- The *in-situ* aerosol size, number and composition measurements in the MBL were complemented by *in vitro* chamber measurements of nascent SSA, to determine the PMA organic volume fraction and water uptake properties. Nascent SSA filter
 samples were analysed using Fourier Transform InfraRed spectroscopy (FTIR) for organic functional groups (Russell et al. 2011), and ion beam analysis for inorganic concentrations (Cohen et al. 2004). Measurements of the hygroscopic growth factor and the volatile fraction up to 450°C for 50-150 nm particles using the VH-TDMA were compared with those of reference inorganic samples (e.g. sea salt, ammonium sulfate) to determine their organic volume fractions (Modini et al. 2010). Complementing the VH-TDMA, the UFO-TDMA provided further information on the organic content of particles of 50nm and down to 10 nm. The bubble chamber observations indicate that the PMA contained a substantial primary organic fraction. VH-TDMA results indicate that the Aitken mode PMA was primarily non-volatile (78-93%), with an average organic volume fraction of 51% (ranging from 39 to 68%), and the UFO-TDMA results show an OVF ranging from 35-45%. These results are consistent with observations in the North Pacific and Atlantic, for which an Aitken mode volatile fraction of the order of 15% and OVF of 0.4-0.8 have been observed (Quinn et al. 2014). FTIR analysis indicated that the POA aerosol in the chamber
- 360 experiments was largely composed of hydroxyl functional groups, with minor contributions from alkanes, amines and carboxylic acid groups, consistent with previous PMO observations (Russell et al. 2011).

Although DMS was a primary focus of measurements during SOAP, a wide variety of other VOCs that potentially contribute to secondary organic aerosol formation were also measured. Halogens and halogen oxides were measured using Multi Axis Differential Optical Absorption Spectroscopy (Max-DOAS) and Electron Capture Detector-Gas Chromatography (ECD-GC).

365 Iodine has been identified as a potentially important precursor of nucleation in coastal regions (Sellegri et al., 2005), and SOAP

provided an opportunity to relate the presence of halogen oxides to phytoplankton biomass and composition in the surface ocean, and nucleation events in the MBL. A High Sensitivity Photon Transfer Reaction Mass Spectrometer (PTR-MS) measured continuously in H3O⁺ mode in the range of m/z 21- m/z 155 throughout the voyage (Lawson et al., to be submitted). Aldehydes, ketones and dicarbonyls were measured using 2,4-dinitrophenylhydrazine (2,4-DNPH) cartridges and high

- 370 performance liquid chromatography (HPLC; Lawson et al., 2015), and a range of VOCs were sampled using adsorbent tubes and later analysed via Thermal Desorption-Gas Chromatography - Flame Ionisation Detection - Mass Spectrometry Detection (TD-GC-FID/MS). These measurements identified a positive relationship between DMS (m/z 63), acetone (m/z 59) and methanethiol (m/z 49), indicating common biological drivers (Lawson et al., to be submitted).
- The first in situ measurements of aqueous phase SMA precursors dicarbonyls, glyoxal and methylglyoxal were obtained over the remote Southern Ocean during SOAP (Lawson et al., 2015). Parallel measurements of known dicarbonyl precursors, measured by PTR-MS, were used to calculate the expected yields of glyoxal and methyl glyoxal, which accounted for < 30% of observed mixing ratios indicating an unidentified source of dicarbonyls (Lawson et al., 2015). This was corroborated by inclusion of SOAP glyoxal measurements obtained by Max-DOAS measurement in a global database, which concluded that the missing glyoxal source was an order of magnitude greater than identified sources (Mahajan et al, 2014). Surface mixing ratios of glyoxal converted to vertical columns, were significantly lower than average vertical column densities (VCDs) from satellite retrievals, possibly reflecting the difficulty of retrieving low glyoxal VCDs over the ocean, or incorrect assumptions about the vertical distribution of glyoxal in the atmosphere (Lawson et al., 2015).

4.2. Rates and controls of volatile and precursor emissions at the air-sea interface

- DMS measurements were made using three different instruments during SOAP (see Table 2); an Atmospheric Pressure
 Ionisation-Chemical Ionisation Mass Spectrometer (API-CIMS) continuously monitored DMS in both phases (Bell et al., 2015), a PTR-MS monitored DMSa (Lawson et al., to be submitted), and discrete water measurements were made using a Sulfur Chemiluminescence Detector Gas Chromatograph (SCD-GC; Walker et al., 2016). Intercomparison of sulfur measurements is not easily or routinely performed (Bell et al., 2012), particularly at sea. Seawater DMS measurements (CIMS and SCDGC) compared well during SOAP (Walker et al., 2016) and the SCD-GC technique also compared well with traditional gas chromatography (with flame photometric detector) in an international intercalibration exercise (Swan et al., 2014). Intercomparison of the PTR-MS and SCD during SOAP, involved analysis of two air samples and two diluted DMS gas standards with a concentration range of 158 354 ppt. The instruments showed very good agreement, with a mean difference of 5% and maximum 10%.
- Although the majority of DMS flux estimates to date have been derived by applying an independently determined transfer
 velocity (k) to the measured DMS gradient at the ocean surface (ΔDMS), there has been a recent increase in direct micrometeorological measurements of DMS flux. Measurements at 10-30-minute resolution show considerable variability in flux, which may reflect methodological artefacts or inherent variability in the distribution of DMS. SOAP provided a platform for comparing eddy covariance (EC) flux measurements of DMS using API-CIMS (Bell et al., 2015), with a gradient flux technique using a drogued catamaran within one kilometre of the vessel (Smith et al., to be submitted). The gradient flux technique is less direct than EC but provides an alternative reference on a platform that is relatively free of shipboard air-flow distortion. The EC system sampled from an intake on the ships bow, with flux instruments mounted on the foremast 12.6m above sea level, and the air pumped to a containerised laboratory on the foredeck. Additional meteorological measurements were obtained from a weather station above the bridge. Both sites are subject to airflow distortion which is azimuthally
- 405 on a 5.3m mast, sampled closer to the water surface where gas gradients are largest. Flux measurements were augmented by

dependent (Popinet et al., 2004). The catamaran sampling framework, which consisted of four air intakes distributed vertically

continuous near-surface measurement of physical parameters using a range of sensors attached to a Spar Buoy, with stratification determined by temperature sensors at 0.5 m intervals (Walker et al., 2016), turbulence determined by a Vector acoustic Doppler Velocimeter at 0.6 m depth. This permitted comparison of k_{DMS} estimates with near-surface upper-ocean turbulence at a distance from the vessel (Smith et al., to be submitted). Wave-breaking whitecap coverage was monitored using

410 a Campbell Scientific 5-megapixel camera (cc5mpx) located on the starboard side of the vessel (Scanlon and Ward, 2016). This provided an indicator of bubble entrainment, which contributes to the differential transfer rate of DMS and CO₂ due to their different solubilities (Blomquist et al., 2006; Bell et al., 2017).

Although SOAP primarily focussed on DMS fluxes, EC measurements of CO₂ flux were an important adjunct measurement for providing insight into gas exchange mechanisms and controls, and improving gas transfer algorithms for gases of differing solubilities. Four Licor infrared gas analyzers were used for eddy covariance flux measurements of CO₂ during SOAP,

- following the initial trials on PreSOAP. Comparison of EC measurements with wet and dry incoming gas streams, and an empirically-based post-processing correction, indicated that only gas stream drying produced robust CO₂ flux and k_{CO2} estimates (Landwehr et al., 2014). A detailed examination of ship motion and airflow distortion effects resulted in a significant reduction of the scatter in the CO2 eddy covariance data (Landwehr et al., submitted). The EC-derived k_{CO2} estimates provided
- 420 a better correlation with a linear fit to the EC friction velocity than with the 10-metre neutral wind speed (u10N), and showed good agreement with dual tracer-derived estimates from the SAGE experiment conducted in this region in March-April 2004 (Ho et al, 2006). Measurement of DMS and CO₂ fluxes also provided further constraint of k parameterisations based upon wind-speed, and the opportunity to assess the influence of bubbles on gas exchange at high wind speeds. DMS fluxes derived by EC and gradient flux techniques showed good agreement (Bell et al., 2015; Smith et al., to be submitted), and confirmed
- 425 previous observations that gas transfer is a linear function of wind-speed at low to intermediate winds (Blomquist et al., 2006; Yang et al., 2011). However, despite winds reaching 20 m s⁻¹ during the latter part of SOAP insufficient data was obtained to draw conclusions regarding the reported deviation of k_{DMS} under high winds (Bell et al., 2015). However, SOAP provided a novel estimate of the size of the EC flux footprint, and the temporal-spatial mismatch between DMSsw and shipboard measured fluxes, highlighting the importance of considering skew in flux estimates arising from non-linear distribution of DMSsw (Bell et al., 2015).

430

435

445

415

A further objective of SOAP was comparison of measured DMS fluxes with calculated estimates from the COAREG model (Fairall et al. 2011) based on ΔDMS , to assess potential discrepancies with modelled fluxes (Marandino et al., 2008; Walker et al., 2016). Potential factors examined here included air and water stability, and the influence of the SSM. Despite the agreement between DMS flux estimates by the two micrometeorological techniques, there was significant departure from COAREG predictions (Fairall et al., 2011) on occasions, suggesting the influence of unidentified processes (Smith et al., to be

- submitted). One potential example was the suppressed DMS flux during a period of atmospheric stability and reversed heat flux during B2. Concurrent EC flux measurement for DMS and CO₂ also provided an opportunity to assess other influences on k. The DMS flux data indicate that the k_{DMS} -wind speed relationship was relatively insensitive to surface biogeochemistry or wave action during SOAP (Bell et al., 2015). In addition, SOAP data was used to parameterise whitecap coverage against
- 440 wind-speed, and identify that maturing waves may obscure and lead to underestimate of the variability of breaking waves (Scanlon and Ward, 2016).

4.3. Surface ocean biogeochemical influences on aerosols and volatiles

Surface mapping of DMSsw and pCO2, using API-CIMS and IRGA, respectively (Bell et al., 2015) were critical to the SOAP voyage strategy and the aims of the two workpackages discussed above. These measurements also provided insight into the covariance of DMS sources and CO₂ sinks in surface waters, and established the importance of this region to global budgets.

The New Zealand Coastal province (NEWZ), which includes the frontal region (STF) studied during SOAP, is characterised in the global DMS climatology by year-round low DMS concentrations with a maximum <2 nmol L⁻¹ (Lana et al., 2011). This infers that this region has some of the lowest global DMSsw concentrations, in marked contrast to the adjacent South Subtropical Convergence (SSTC) province, which occupies the remainder of the 35-50°S latitude band and accommodates the

- 450 STF, which is characterised by a mid-summer maximum of 10 nmol L^{-1} DMS. This discrepancy between the two regions likely reflects the low number of DMS observations for the NEWZ province in the climatology (n=6; Lana et al., 2011). Previous DMSsw measurements in subantarctic waters south of the Chatham Rise, and east of Tasmania in the SSTC biome (Archer et al., 2011; Griffiths et al., 1999), are consistent with this climatological estimate, whereas larger unpublished surveys have recorded elevated surface DMSsw during austral spring (October 2000), with a mean DMSsw of 4.5 (+/- 6.8) nmol L^{-1} on the
- 455 Chatham Rise (Harvey et al., pers. comm). Combining these measurements with data from the SOAP campaign (mean DMSsw = $6.6 \text{ nmol } \text{L}^{-1}$) gives a weighted-mean DMSsw of 5.3 nmol L^{-1} (n=5300, see Table 3), confirming that DMSsw in the NEWZ province is currently underestimated, and is in fact more typical of the SSTC province. Although the PreSOAP and SOAP sampling strategy of focussing on phytoplankton blooms may introduce bias towards higher DMSsw, the BOX voyage, which had broad spatial coverage of subtropical and subAntarctic waters between 39.5-47°S, gave a similar mean DMSsw to the
- weighted mean for all voyages. The elevated DMSsw was reflected in the EC flux measurements during SOAP, which recorded maximum and mean fluxes of 100 and 16.3 µmol S m⁻²d⁻¹, respectively, (Bell et al., 2015), which exceed the climatological mean of >10 µmol S m² d⁻¹ for the SSTC region (Lana et al., 2011). In addition, the high MBL DMS concentrations of 1000 ppt recorded during SOAP exceed DMSa at coastal stations on the New Zealand North Island in summer (Harvey et al., 1993; de Bruyn et al., 2002; Wylie and de Mora, 1996). Although seasonally constrained, the SOAP measurements provide evidence that regional DMS emissions are significant in the South West Pacific. The large dataset of regional concentrations and flux will allow further refinement of global climatologies, such as the Global Surface Water DMS Database and the Surface Ocean

CO2 Atlas (SOCAT).

485

The spatial variability of DMSsw was related to surface ocean biogeochemistry and bloom type by measurement of a suite of ancillary parameters in underway mode, including temperature and salinity, Chl-*a*, chromophoric dissolved organic matter

- 470 (CDOM), β_{660} backscatter, dissolved oxygen and pCO₂ (see Tables 1 and 2). The vertical variability of DMSsw, and the dissolved and particulate pools of its precursor DMSP, were quantified in the surface mixed layer at stations within each bloom, and related to plankton biomass and community composition, nutrient and organic composition and physical drivers (see Suppl. Table 2). Process studies of DMSP cycling included deck incubations examining the bacterially-mediated pathways of DMSP cleavage and demethylation in relation to different bloom dynamics (Lizotte et al., submitted). DMSP concentrations
- were relatively high, reaching a maximum of 160 nmol L⁻¹, and showed significant correlation with phytoplankton biomass during SOAP. However, the yield of DMS from bacterial conversion of dissolved DMSP was variable with no spatial trend, although a correlation with leucine incorporation indicates that DMSP was an important carbon source for bacteria. Overall, gross DMS production by bacteria in deck incubations of near-surface water was relatively low, at < 6 nmol L⁻¹ d⁻¹, inferring that phytoplankton-mediated conversion of DMSP was likely a significant near-surface source of DMS (Lizotte et al., submitted).

The SSM is a potentially important interface controlling MBL and aerosol composition, as it is the interface across which material exchanges between atmosphere and ocean. Physical and biogeochemical processes within this thin layer have the potential to alter transfer via factors, such as the concentration of organic material and enhanced biological and photochemical processing. Near-surface CO₂ gradients have been observed (Calleja et al., 2005), and several studies report DMS enrichment in the SSM (see summary in Walker et al., 2016). If DMS consumption or production in the SSM is significant this represents

a potential source of discrepancy in comparison of measured fluxes with that calculated by the COAREG model (see above). The biogeochemistry of the SSM and the upper 1.6m surface water were characterised at 10 stations during SOAP at distance

from the research vessel, to determine the spatial variability in composition within, and between, different phytoplankton blooms (Walker et al., 2016). Near-surface DMS gradients were generally negligible, except during B1 where low wind-speed,

490 near-surface stratification and high dinoflagellate abundance may have combined to enhance DMS in the SSM relative to subsurface waters. The observed DMS enrichment factors in the SSM during B1, ranging from 1.4 to 5.3, are some of the highest reported to date. The anomaly between measured DMS fluxes and COAREG estimated was also greatest during B1, inferring that DMS emissions, and associated k-wind-speed parameterisations, may be sensitive to DMS production in the SSM under certain conditions. However, the observations also raise questions as to how such significant DMS enrichment is maintained in the SSM, as high DMS production would be required to ameliorate loss processes (Walker et al., 2016).

5. Conclusions

The SOAP voyage has identified new questions in important areas of SOLAS-related research, including the influence of the SSM on DMS emissions, implications for secondary aerosol formation, and unidentified sources of organic aerosol precursors, all of which are potentially influenced by photochemistry in the surface ocean and MBL (Lawson et al., 2015). It has also 500 addressed confounding technical challenges including small-scale heterogeneity in surface waters, clean air baseline sampling, and discrepancies between existing techniques and models. An overarching aim of the SOAP campaign was to assess potential relationships between surface water biogeochemistry and corresponding or related species in the MBL, to identify the factors influencing aerosol precursors, and their potential as analogues. Chl-a is an indicator of phytoplankton biomass that is readily retrievable by satellite, and consequently has been investigated as a potential proxy for DMSsw (Lana et al., 2011). The SOAP 505 voyage provided a platform to validate this observation, particularly as it took place in the 40-60°S latitude band which exhibits the most significant regional correlation between Chl-a and DMSsw (Vallina et al., 2006). Overall there was a weak, but significant, correlation (r = 0.12, p < 0.005) between Chl-a and DMSsw in the underway surface data during SOAP, but also significant variability in the slope and the sign of this relationship between the different blooms. Correlations were also apparent between Chl-a and DMSP (Lizotte et al., submitted), and Chl-a and DMSa, but there was no relationship between Chl-a and DMS flux, as expected, due to the short timescales and flux footprint identified by Bell et al., 2015. Correlations

- 510 Chl-*a* and DMS flux, as expected, due to the short timescales and flux footprint identified by Bell et al., 2015. Correlations have been reported previously for Chl-*a* with CCN (Meskhidze and Nenes, 2006), and aerosol organic enrichment (Gantt et al., 2011), although other assessments have shown variable results (Russell et al., 2010; Rinaldi et al., 2013). The measurement of PMA and SMA composition and number coincident with multi-species characterisation of MBL and surface water composition during SOAP has provided a broad database with which to assess and develop these relationships for potential
- 515 application in remote sensing and Earth System Models. The first step towards this is the inclusion of SOAP aerosol and tropospheric data in the global ACCESS-UKCA model (Woodhouse et al., 2015), containing the GLOMAP-mode aerosol scheme (Mann et al., 2010, 2012), which shows very good agreement with observed distributions of condensation nuclei (Woodhouse et al, pers. comm.)

6. Data availability

520 The underway DMSsw can be downloaded at <u>http://saga.pmel.noaa.gov/dms/select.php</u>. The remaining data is available by request email to cliff.law@niwa.co.nz

7. Supplement link

8. Author contribution

525 The SOAP campaign was led and coordinated by CSL, MJS & MJH. All authors developed the analytical methods and instruments used during the SOAP campaign, and CSL, MJS, MJH, TGB, LTC, FCE, SJL, ML, AM, JM, KAS, PV and CFW made measurements during the SOAP voyage. All authors were involved in analysis and interpretation of data. CSL led the manuscript production, with contributions from MJS, MJH, TGB, LTC, SJL, ML, ZR, KA, ES, PV and CFW.

9. Competing interests

530 The authors declare that they have no conflict of interest.

10. Acknowledgements

We acknowledge the invaluable assistance of the Captain, officers and crew of the R/V Tangaroa. We thank Kim Currie, Steve George, Matt Walkington, Mark Gall, Marc Mallet, Greg Olsen, Gus Olivares and Nick Talbot for collection and analysis of samples; Christa Marandino, Warren DeBruyn and Cyril McCormick for assistance with API-CIMS measurements; Sebastian 535 Landwehr, Scott Miller and Brian Ward for CO₂ flux measurements; Paul Johnson, Karin Kreher and Neil Harris for halocarbon and VOC measurements; Jason Ward for assistance with CCN measurement; Gordon Brailsford for Picarro set up; Paul Selleck and Min Cheng for analysis of VOC samples, Sarah Connors for the back-trajectory suite using the UK Met Office NAME model, and Matt Woodhouse for analysis of SOAP data using the global ACCESS-UKCA model. We also acknowledge Maurice Levasseur and Melita Keywood for their comments, and their support of the microbial DMSP cycling 540 and VOC measurements, respectively. This research was supported by funding from NIWA's Climate and Atmosphere Research Programme 3 – Role of the oceans (2015/16 SCI), and a Postdoctoral Fellowship (CO1X0911) for CW from the New Zealand Ministry for Business, Innovation and Employment (MBIE). Flux measurements were supported by a NSF Atmospheric Chemistry Program (grant nos. 08568, 0851472, 0851407 and 1143709), and VOC and CCN measurements supported by CSIRO's Capability Development Fund. PV participation was supported by EU COST action 735, the Academy 545 of Finland through the Centre of Excellence, and via a Finnish Academy visiting grant no. 136841. SL participation was supported by Science Foundation Ireland as part of the US-Ireland R&D Partnership Programme under grant number

11. Figures

Figure 1. a) An ocean colour image (10/2/11) during the PreSOAP voyage, showing phytoplankton blooms on the western Chatham Rise region along 44°S (data courtesy of NASA) b) The SOAP voyage track in the Chatham Rise region, overlain by Sea Surface Temperature (°C), with the study region (box) indicated in the inset bathymetric map of New Zealand.

Engineering Research Council of Canada (NSERC) for supporting the microbial DMSP cycling research.

Figure 2. Non-sea salt sulfate concentrations plotted against Day of Year at different New Zealand coastal atmospheric monitoring sites.

08/US/I1455, and a SFI Short-term Travel Fellowship under grant 09/US/I1758-STTF-11. We thank the Natural Sciences and

Figure 3. Continuous measurements during PreSOAP of a) wind speed (m s⁻¹), b) atmospheric DMS (ppt), c) surface water DMS (nmol L⁻¹), and d) surface chlorophyll-*a* (mg m⁻³; quenched data removed).

Figure 4. 8-day composite images of surface chlorophyll-*a* (MODIS, 4 km resolution) during the SOAP voyage for a) 10-17 Feb 2012 (DoY 41-48), b) 18-25 Feb 2012 (DoY 49-56), and c) 26th Feb-4th March (DoY 57-64), showing bloom locations

(red dots), with the colour scale (mg m⁻³) above figures a) - c), and daily true colour images for d) Bloom 1 (16 Feb 2012., DoY 47), e) Bloom 2 (18 Feb 2012, DoY 49) and f) Bloom 3 (3 March, DoY 65) (MODIS Aqua data courtesy of NASA).

Figure 5. a)-c) Synoptic meteorology summary for each bloom period during the SOAP voyage. Surface pressure and wind plots (colour scale to the left of figures a)-c)) are derived from the NZ local area Unified Model NZLAM, with the bloom location indicated by a red dot. d)-f) Back-trajectory analyses for each bloom period during the SOAP voyage. This was calculated using the Lagrangian Numerical Atmospheric-dispersion Modelling Environment (NAME) for the lower atmosphere (0-100m) as time integrated particle density (g s m⁻³, colour scale below figures). Each plot shows the back-

trajectory of 8 "releases", i.e. one every three hours over 24 hours for the actual ship position.

Figure 6. Meteorological and hydrodynamic variables during the SOAP voyage, including a) Wind speed (WS, ms⁻¹); b) Direction (Dir., °); Wind (blue) and wave (cyan); c) Temperature (Temp., °C); Air (black) and surface water (green); d) Irradiance (Irrad., Wm⁻²) and e) Significant wave height (Hs, m). Bloom occupation periods are indicated by the red horizontal bars and bloom label in the upper panel.

570

560

565

Figure 7. Surface water properties (2-10m) recorded at each station during the SOAP voyage: Temperature (Temp, °C), Mixed Layer Depth (ML Depth, m), Chlorophyll-a (Chl-a, mg m⁻³), and nitrate concentration (µmol L⁻¹), plotted against Day of Year (DoY), with the occupation period for each bloom indicated by the vertical shaded bars and bloom labels at the top of the figure.

575 Fig. 8. Conceptual figure of the parameters, processes and vertical range measured during SOAP, with the integrated work programmes (WP) indicated on the left of the figure.

Figure 9. Aerosol characterisation during SOAP indicating size spectral (red) and total counts (black) range for each instrument, relative to aerosol size and mode. Ambient RH measurement was used for RH correction of the PCASP, Hi Vol and SMPS, and diffusion driers (Silica Gel) were used on the inlet of the UFO-TDMA and VH-TDMA.

580 Figure 10. a) Marine boundary layer CN concentrations (cm⁻³, top, CPC3772 in blue, CPC3010 in red), b) CCN concentrations (middle, cm⁻³) and c) number of hours over land indicated by 72 hour back-trajectory (bottom, 27-member ensemble average). Bloom occupation periods are indicated by the vertical shaded bars and bloom labels at the top of the figure.

12. References

- 585 Allen, A. G., Dick, A. L., and Davison, B. M.: Sources of atmospheric methanesulphonate, non-sea-salt sulphate, nitrate and related species over the temperate South Pacific, Atmospheric Environment, 31, 191-205, doi:10.1016/1352-2310(96)00194-x, 1997.
 - Almeida, J., Schobesberger, S., Kürten, A., Ortega, I. K., Kupiainen-Maatta, O., Praplan, A. P., Adamov, A., Amorim, A., Bianchi, F., Breitenlechner, M., David, A., Dommen, J., Donahue, N. M., Downard, A., Dunne, E., Duplissy, J., Ehrhart,
- 590 S., Flagan, R. C., Franchin, A., Guida, R., Hakala, J., Hansel, A., Heinritzi, M., Henschel, H., Jokinen, T., Junninen, H., Kajos, M., Kangasluoma, J., Keskinen, H., Kupc, A., Kurtén, T., Kvashin, A. N., Laaksonen, A., Lehtipalo, K., Leiminger, M., Leppa, J., Loukonen, V., Makhmutov, V., Mathot, S., McGrath, M. J., Nieminen, T., Olenius, T., Onnela, A., Petäjä, T., Riccobono, F., Riipinen, I., Rissanen, M., Rondo, L., Ruuskanen, T., Santos, F. D., Sarnela, N., Schallhart, S., Schnitzhofer, R., Seinfeld, J. H., Simon, M., Sipilä, M., Stozhkov, Y., Stratmann, F., Tomé, A., Trostl,
- 595 J., Tsagkogeorgas, G., Vaattovaara, P., Viisanen, Y., Virtanen, A., Vrtala, A., Wagner, P. E., Weingartner, E., Wex, H., Williamson, C., Wimmer, D., Ye, P., Yli-Juuti, T., Carslaw, K. S., Kulmala, M., Curtius, J., Baltensperger, U., Worsnop, D. R., Vehkamaki, H. and Kirkby, J.: Molecular understanding of sulphuric acid-amine particle nucleation in the atmosphere, Nature, 502(7471), 359–363, doi:10.1038/nature12663, 2013.
- Andreae, M. O., and Rosenfeld, D.: Aerosol-cloud-precipitation interactions. Part 1. The nature and sources of cloud-active 600 aerosols, Earth-Science Reviews, 89, 13-41, doi:10.1016/j.earscirev.2008.03.001, 2008.
 - Archer, S. D., Safi, K., Hall, A., Cummings, D. G., and Harvey, M.: Grazing suppression of dimethylsulphoniopropionate (DMSP) accumulation in iron-fertilised, sub-Antarctic waters, Deep-Sea Research Part Ii-Topical Studies in Oceanography, 58, 839-850, doi:10.1016/j.dsr2.2010.10.022, 2011.

- Ayers, G. P., Ivey, J. P., and Gillett, R. W.: Coherence between seasonal cycles of dimethylsulfide, methanesulfonate, and sulfate in Marine Air, Nature, 349, 404-406, 1991.
 - Balch, W. M., Drapeau, D. T., Bowler, B. C., Lyczskowski, E., Booth, E. S., and Alley, D.: The contribution of coccolithophores to the optical and inorganic carbon budgets during the Southern Ocean Gas Exchange Experiment: New evidence in support of the "Great Calcite Belt" hypothesis, Journal of Geophysical Research-Oceans, 116, doi:10.1029/2011jc006941, 2011.
- 610 Bell, T. G., De Bruyn, W., Marandino, C. A., Miller, S. D., Law, C. S., Smith, M. J., and Saltzman, E. S.: Dimethylsulfide gas transfer coefficients from algal blooms in the Southern Ocean, Atmos. Chem. Phys., 15, 1783-1794, doi:10.5194/acp-15-1783-2015, 2015.
- Bell, T. G., Landwehr, S., Miller, S. D., de Bruyn, W. J., Callaghan, A. H., Scanlon, B., Ward, B., Yang, M., and Saltzman, E. S.: Estimation of bubble-mediated air-sea gas exchange from concurrent DMS and CO2 transfer velocities at intermediate–high wind speeds, Atmos. Chem. Phys., 17, 9019-9033, 10.5194/acp-17-9019-2017, 2017.
 - Bell, T. G., Malin, G., Lee, G. A., Stefels, J., Archer, S., Steinke, M., and Matrai, P.: Global oceanic DMS data intercomparability, Biogeochem, 110, 147-161, 10.1007/s10533-011-9662-3, 2012.
- Blake, N. J., Blake, D. R., Wingenter, O. W., Sive, B. C., Kang, C. H., Thornton, D. C., Bandy, A. R., Atlas, E., Flocke, F., Harris, J. M., and Rowland, F. S.: Aircraft measurements of the latitudinal, vertical, and seasonal variations of NMHCs, methyl nitrate, methyl halides, and DMS during the First Aerosol Characterization Experiment (ACE 1), Journal of
 - Geophysical Research-Atmospheres, 104, 21803-21817, doi:10.1029/1999jd900238, 1999.
 Blomquist, B.W., Fairall, C.W., Huebert, B.J., Kieber, D.J., Westby, G.R.: DMS sea-air transfer velocity: Direct measurements by eddy covariance and parameterization based on the NOAA/COARE gas transfer model. Geophysical Research Letters, *33*(7), 2006.
- 625 Blot, R., Clarke, A. D., Freitag, S., Kapustin, V., Howell, S. G., Jensen, J. B., Shank, L. M., McNaughton, C. S. and Brekhovskikh, V.: Ultrafine sea spray aerosol over the southeastern Pacific: open-ocean contributions to marine boundary layer CCN, Atmos. Chem. Phys., 13(14), 7263–7278, doi:10.5194/acp-13-7263-2013, 2013.
 - Boers, R., Jensen, J. B., Krummel, P. B., and Gerber, H.: Microphysical and radiative structure of wintertime marine stratocumulus clouds over the Southern Ocean, Q. Jl R. met. Soc., 122, 1307-1339, 1996.
- 630 Boers, R., Jensen, J. B., and Krummel, P. B.: Microphysical and shortwave radiative structure of stratocumulus clouds over the Southern Ocean: summer results and seasonal differences, Q. Jl R. met. Soc., 124, 151-168, 1998.
 - Boyd, P., LaRoche, J., Gall, M., Frew, R., and McKay, R. M. L.: Role of iron, light, and silicate in controlling algal biomass in subantarctic waters SE of New Zealand, Journal of Geophysical Research-Oceans, 104, 13395-13408, doi:10.1029/1999jc900009, 1999.
- 635 Boyd, P. W., McTainsh, G., Sherlock, V., Richardson, K., Nichol, S., Ellwood, M., and Frew, R.: Episodic enhancement of phytoplankton stocks in New Zealand subantarctic waters: Contribution of atmospheric and oceanic iron supply, Global Biogeochemical Cycles, 18, C8 GB1029, doi:10.1029/2002GB002020 2004.
- Bradford-Grieve, J. M., Boyd, P. W., Chang, F. H., Chiswell, S., Hadfield, M., Hall, J. A., James, M. R., Nodder, S. D., and Shushkina, E. A.: Pelagic ecosystem structure and functioning in the Subtropical Front region east of New Zealand in austral winter and spring 1993, Journal of Plankton Research, 21, 405-428, doi:10.1093/plankt/21.3.405, 1999.
 - Cainey, J., and Harvey, M.: Dimethylsulfide, a limited contributor to new particle formation in the clean marine boundary layer (DOI 10.1029/2001GL014439), Geophysical Research Letters, 29, 32, doi:DOI: 10.1029/2001GL014439, 2002.
- Calleja, M. L., Duarte, C. M., Navarro, N., and Agustí, S.: Control of air-sea CO2 disequilibria in the subtropical NE Atlantic by planktonic metabolism under the ocean skin, Geophysical Research Letters, 32, L08606, doi:doi:10.1029/2004GL022120, 2005.
- Calleja, M. L., Duarte, C. M., Alvarez, M., Vaquer-Sunyer, R., Agusti, S., and Herndl, G. J.: Prevalence of strong vertical CO2 and O-2 variability in the top meters of the ocean, Global Biogeochemical Cycles, 27, 941-949, doi:10.1002/gbc.20081, 2013.
- Carslaw, K. S., Boucher, O., Spracklen, D. V., Mann, G. W., Rae, J. G. L., Woodward, S., and Kulmala, M.: A review of natural aerosol interactions and feedbacks within the Earth system, Atmospheric Chemistry and Physics, 10, 1701-1737, doi:10.5194/acp-10-1701-2010, 2010.
 - Carslaw, K. S., Lee, L. A., Reddington, C. L., Pringle, K. J., Rap, A., Forster, P. M., Mann, G. W., Spracklen, D. V., Woodhouse, M. T., Regayre, L. A., and Pierce, J. R.: Large contribution of natural aerosols to uncertainty in indirect forcing, Nature, 503, 66-71, 2013.
- 655 Chang, F. H., and Gall, M. Phytoplankton assemblages and photosynthetic pigments during winter and spring in the Subtropical Convergence region near New Zealand, New Zealand Journal of Marine and Freshwater Research, 32, 515-530, 1998.

660

- Chang, F. H., Williams, M. J. M., Schwarz, J. N., Hall, J. A., Maas, E. W., and Stewart, R.: Spatial variation of phytoplankton assemblages and biomass in the New Zealand sector of the Southern Ocean during the late austral summer 2008, Polar Biology, 36, 391-408, doi:10.1007/s00300-012-1270-8, 2013.
- Chang, R.Y.W., Sjostedt, S.J., Pierce, J.R., Papakyriakou, T.N., Scarratt, M.G., Michaud, S., Levasseur, M., Leaitch, W.R. and Abbatt, J.P. Relating atmospheric and oceanic DMS levels to particle nucleation events in the Canadian Arctic. *Journal of Geophysical Research: Atmospheres*, 116(D17), 2011.
- Charlson, R. J., Lovelock, J. E., Andreae, M. O., and Warren, S. G.: Oceanic phytoplankton, atmospheric sulfur, cloud albedo
 and climate, Nature, 326, 655-661, doi:10.1038/326655A0 1987.

- Chiswell, S. M., Bradford-Grieve, J., Hadfield, M. G., and Kennan, S. C.: Climatology of surface chlorophyll a, autumn-winter and spring blooms in the southwest Pacific Ocean, Journal of Geophysical Research-Oceans, 118, 1003-1018, doi:10.1002/jgrc.20088, 2013.
- Ciuraru, R., Fine, L., Pinxteren, M. V., D'Anna, B., Herrmann, H., and George, C.: Unravelling new processes at interfaces:
 Photochemical isoprene production at the sea surface. Environmental Science and Technology, 49(22), 13199-13205, 2015.
- Clarke, A. D., Freitag, S., Simpson, R. M. C., Hudson, J. G., Howell, S. G., Brekhovskikh, V. L., Campos, T., Kapustin, V. N. and Zhou, J.: Free troposphere as the dominant source of CCN in the Equatorial Pacific boundary layer: long-range transport and teleconnections, Atmospheric Chemistry and Physics Discussions, 13(1), 1279–1326, doi:10.5194/acpd-13-1279-2013, 2013.
 - Cohen, D. D., Stelcer, E., Hawas, O. and Garton, D.: IBA methods for characterisation of fine particulate atmospheric pollution: a local, regional and global research problem, The 13th International Conference on Particle Induced X-ray Emission (PIXE 2013), 219–220 IS -, 145–152, 2004.
- Cravigan, L. T., Ristovski, Z., Modini, R. L., Keywood, M. D. and Gras, J. L.: Observation of sea-salt fraction in sub-100 nm diameter particles at Cape Grim, J. Geophys. Res. Atmos., 2014JD022601, doi:10.1002/2014JD022601, 2015.
 - de Bruyn, W. J., Harvey, M., Cainey, J. M., and Saltzman, E. S.: DMS and SO2 at Baring Head, New Zealand: Implications for the yield of SO2 from DMS, Journal of Atmospheric Chemistry, 41, 189-209, doi:10.1023/a:1014252106572, 2002.
- de Leeuw, G., Guieu, C., Arneth, A., Bellouin, N., Bopp, L., Boyd, P. W., van der Gon, H. A. C. D., Desboeufs, K. V., Dulac, F., Facchini, M. C., Gantt, B., Langmann, B., Mahowald, N. M., Maranön, E., O'Dowd, C., Olgun, N., Pulido-Villena, E., Rinaldi, M., Stephanou, E. G., and Wagener, T.: Ocean–Atmosphere Interactions of Particles, P.S. Liss and M.T. Johnson (Eds.), Ocean-Atmosphere Interactions of Gases and Particles, Springer Earth System Sciences, 171-246, doi:10.1007/978-3-642-25643-1_4, 2014.

690

- Decesari, S., Finessi, E., Rinaldi, M., Paglione, M., Fuzzi, S., Stephanou, E.G., Tziaras, T., Spyros, A., Ceburnis, D., O'Dowd, C. and Dall'Osto, M.: Primary and secondary marine organic aerosols over the North Atlantic Ocean during the MAP experiment. Journal of Geophysical Research: Atmospheres, 116(D22), 2011.
- Delizo, L., Smith, W. O., and Hall, J.: Taxonomic composition and growth rates of phytoplankton assemblages at the Subtropical Convergence east of New Zealand, Journal of Plankton Research, 29, 655-670, doi:10.1093/plankt/fbm047, 2007.
- Draxler, R. R. and Rolph, G. D., Eds.: HYSPLIT(HYbrid Single-PArticle Lagrangian Intergrated Trajectory) Model, NOAA
 Air Resources Laboratory, College Park, MD. [online] Available from: http://www.arl.noaa.gov/HYSPLIT.php (Accessed 1 September 2013), 2013.
- Facchini, M. C., Rinaldi, M., Decesari, S., Carbone, C., Finessi, E., Mircea, M., Fuzzi, S., Ceburnis, D., Flanagan, R., Nilsson, E. D., de Leeuw, G., Martino, M., Woeltjen, J., and O'Dowd, C. D.: Primary submicron marine aerosol dominated by insoluble organic colloids and aggregates, Geophysical Research Letters, 35, L17814, doi:10.1029/2008GL034210, 2008.
 - Fairall, C. W., Bradley, E. F., Hare, J. E., Grachev, A. A., and Edson, J. B.: Bulk parameterization of air-sea fluxes: Updates and verification for the COARE algorithm, J Climate, 16, 571-591, 2003.
 - Fairall, C. W., Yang, M. X., Bariteau, L., Edson, J. B., Helmig, D., McGillis, W., Pezoa, S., Hare, J. E., Huebert, B., and Blomquist, B.: Implementation of the Coupled Ocean-Atmosphere Response Experiment flux algorithm with CO2, dimethyl sulfide, and O-3, Journal of Geophysical Research-Oceans, 116, doi:10.1029/2010jc006884, 2011.
 - Gantt, B., and Meskhidze, N.: The physical and chemical characteristics of marine primary organic aerosol: a review, Atmos. Chem. Phys., 13, 3979-3996, doi:10.5194/acp-13-3979-2013, 2013.
- Gantt, B., Meskhidze, N., Facchini, M. C., Rinaldi, M., Ceburnis, D., and O'Dowd, C. D.: Wind speed dependent size-resolved parameterization for the organic mass fraction of sea spray aerosol, Atmospheric Chemistry and Physics, 11, 8777-8790, 2011.
 - Gondwe, M., Krol, M., Gieskes, W., Klaassen, W., De Baar, H.: The contribution of ocean-leaving DMS to the global atmospheric burdens of DMS, MSA, SO2, and NSS SO4=. Global Biogeochemical Cycles, *17*(2), 2003.
- Griffiths, F. B., Bates, T. S., Quinn, P. K., Clementson, L. A., Parslow, J. S.: Oceanographic context of the First Aerosol Characterization Experiment (ACE 1): A physical, chemical, and biological overview, Journal of Geophysical Research-Atmospheres, 104, 21649-21671, doi:10.1029/1999jd900386, 1999.
- Harvey, M. J., Wylie, D. J., Martin, R. J., Clarkson, T. S., and de Mora, S. J.: Dimethylsulphide measurements at Baring Head, New Zealand, in: Dimethylsulphide, Oceans, Atmosphere and Climate, edited by: Restelli, G., and Angeletti, G., Kluwer Academic, Dordrecht, 143-151, 1993.
- Harvey, M. J., Law, C. S., Smith, M. J., Hall, J. A., Abraham, E. R., Stevens, C. L., Hadfield, M. G., Ho, D. T., Ward, B., Archer, S. D., Cainey, J. M., Currie, K. I., Devries, D., Ellwood, M. J., Hill, P., Jones, G. B., Katz, D., Kuparinen, J., Macaskill, B., Main, W., Marriner, A., McGregor, J., McNeil, C., Minnett, P. J., Nodder, S. D., Peloquin, J., Pickmere, S., Pinkerton, M. H., Safi, K. A., Thompson, R., Walkington, M., Wright, S. W., and Ziolkowski, L. A.: The SOLAS air-sea gas exchange experiment (SAGE) 2004, Deep Sea Research Part II: Topical Studies in Oceanography, 58, 753-763, doi:10.1016/j.dsr2.2010.10.015, 2011.
- 725 Harvey M.J., McGregor J.A., Lawson S. J., Cravigan L., Talbot N., Olivares G., Conners S., Woodhouse M.: Aerosol and tracer measurement at sea defining the baseline. Atmospheric Chemistry and Physics, to be *submitted*

- Ho, D. T., Law, C. S., Smith, M. J., Schlosser, P., Harvey, M., and Hill, P.: Measurements of air-sea gas exchange at high wind speeds in the Southern Ocean: Implications for global parameterizations. doi:10.1029/2006GL026817, Geophys. Res. Lett., 33, L16611, 2006.
- 730 Jaeglé, L., Quinn, P. K., Bates, T. S., Alexander, B., and Lin, J. T.: Global distribution of sea salt aerosols: new constraints from in situ and remote sensing observations, Atmos. Chem. Phys., 11, 3137-3157, doi:10.5194/acp-11-3137-2011, 2011.

Johnson, G., Ristovski, Z., and Morawska, L.: Application of the VH-TDMA technique to coastal ambient aerosols, Geophys. Res. Lett., 31, doi:10.1029/2004GL020126, 2004a.

735 Jones, A. R., Thomson, D. J., Hort, M., and Devenish, B.: The UK Met Office's next-generation atmospheric dispersion model, NAME III, in: Proceedings of the 27th NATO/CCMS International Technical Meeting on Air Pollution Modelling and its Application, edited by: Borrego, C., and Norman, A.-L., Springer, pp 580 - 589, 2007.

- Kay, J. E., Wall, C., Yettella, V., Medeiros, B., Hannay, C., Caldwell, P., and Bitz, C.: Global Climate Impacts of Fixing the Southern Ocean Shortwave Radiation Bias in the Community Earth System Model (CESM), Journal of Climate, 29, 4617-4636, doi:doi:10.1175/JCLI-D-15-0358.1, 2016.
- Kiene, R. P., Kieber, D. J., Slezak, D., Toole, D. A., del Valle, D. A., Bisgrove, J., Brinkley, J., and Rellinger, A.: Distribution and cycling of dimethylsulfide, dimethylsulfoniopropionate, and dimethylsulfoxide during spring and early summer in the Southern Ocean south of New Zealand, Aquatic Sciences, 69, 305-319, doi:10.1007/s00027-007-0892-3, 2007.
- Korhonen, H., Carslaw, K. S., Spracklen, D. V., Mann, G. W., and Woodhouse, M. T.: Influence of oceanic dimethyl sulfide emissions on cloud condensation nuclei concentrations and seasonality over the remote Southern Hemisphere oceans: A global model study, Journal of Geophysical Research-Atmospheres, 113, doi:10.1029/2007jd009718, 2008.
 - Lana, A., Bell, T. G., Simo, R., Vallina, S. M., Ballabrera-Poy, J., Kettle, A. J., Dachs, J., Bopp, L., Saltzman, E. S., Stefels, J., Johnson, J. E., and Liss, P. S. 2011. An updated climatology of surface dimethlysulfide concentrations and emission fluxes in the global ocean, Global Biogeochemical Cycles, 25, doi:10.1029/2010gb003850, 2011.
- 750 Lana, A., Simo, R., Vallina, S. M., and Dachs, J.: Potential for a biogenic influence on cloud microphysics over the ocean: a correlation study with satellite-derived data, Atmospheric Chemistry and Physics, 12, 7977-7993, doi:10.5194/acp-12-7977-2012, 2012.
 - Landschuetzer, P., Gruber, N., Bakker, D. C. E., and Schuster, U.: Recent variability of the global ocean carbon sink, Global Biogeochemical Cycles, 28, 927-949, doi:10.1002/2014gb004853, 2014.
- 755 Landwehr, S., Miller, S. D., Smith, M. J., Saltzman, E. S., and Ward, B.: Analysis of the PKT correction for direct CO2 flux measurements over the ocean, Atmospheric Chemistry and Physics, 14, 3361-3372, 2014.
 - Landwehr, S., Miller, S. D., Smith, M. J., Saltzman, E. S., Bell, T. G., and Ward, B. Using Eddy Covariance to Measure the Dependence of Air-Sea CO2 Exchange Rate on Friction Velocity, Atmospheric Chemistry and Physics *submitted*
- Lawson, S. J., Selleck, P. W., Galbally, I. E., Keywood, M. D., Harvey, M. J., Lerot, C., Helmig, D., and Ristovski, Z.: Seasonal in situ observations of glyoxal and methylglyoxal over the temperate oceans of the Southern Hemisphere, 15, 223-240, doi:10.5194/acp-15-223-2015, 2015.
 - Lawson S.J., Selleck P.W., Cheng M. and M. Harvey: VOCs in marine air over the biologically productive Chatham Rise. Atmospheric Chemistry and Physics, to be submitted
- Lizotte, M., Levasseur, M., Law, C. S., Walker, C. F., Safi, K. A., Marriner, A., and Kiene, R. P.: Dimethylsulfoniopropionate (DMSP) and dimethylsulfide (DMS) cycling across contrasting biological hotspots of the New Zealand Subtropical Front, Ocean Sci. Discuss., 2017, 1-49, doi:10.5194/os-2017-32, 2017
 - Mahajan, A. S., Fadnavis, S., Thomas, M. A., Pozzoli, L., Gupta, S., Royer, S.-J., Saiz-Lopez, A., and R., S.: Quantifying the impacts of an updated global dimethyl sulfide climatology on cloud microphysics and aerosol radiative forcing, J. Geophys. Res. Atmos., 120, 2524-2536, doi:10.1002/2014JD022687, 2015.
- 770 Mahajan, A. S., et al.: Glyoxal observations in the global marine boundary layer, J. Geophys. Res. Atmos., 119, 6160–6169, doi:10.1002/2013JD021388, 2014
 - Mallet, M., L. Cravigan, B. Miljevic, P. Vaattovaara, E. Deschaseaux, H. Swan, G. Jones, *and* Z. Ristovski (2016), Sea spray aerosol in the Great Barrier Reef and the presence of nonvolatile organics, J. Geophys. Res. Atmos., 121, 7088–7099, *doi:*10.1002/2016JD024966.
- 775 Mann, G. W., Carslaw, K. S., Spracklen, D. V., Ridley, D. A., Manktelow, P. T., Chipperfield, M. P., Pickering, S. J. and Johnson, C. E. Description and evaluation of GLOMAP-mode: a modal global aerosol microphysics model for the UKCA composition-climate model Geoscientific Model Development, 3, 519-551, 2012.
- Mann, G. W., Carslaw, K. S., Ridley, D. A., Spracklen, D. V., Pringle, K. J., Merikanto, J., Korhonen, S., Schwarz, J. P., Lee, L. A., Manktelow, P. T., Woodhouse, M. T., Schmidt, A., Breider, T. J., Emmerson, K. M., Chipperfield, M. P. and Pickering, S. J. Intercomparison of modal and sectional aerosol microphysics representations within the same 3-D global chemical transport model Atmospheric Chemistry and Physics, 12, 4449-4476, 2012.
 - Marandino, C. A., De Bruyn, W. J., Miller, S. D., and Saltzman, E. S.: DMS air/sea flux and gas transfer coefficients from the North Atlantic summertime coccolithophore bloom, Geophysical Research Letters, 35, doi:10.1029/2008gl036370, 2008.
- 785 McCoy, D. T., Burrows, S. M., Wood, R., Grosvenor, D. P., Elliott, S. M., Ma, P.-L., Rasch, P. J., and Hartmann, D. L.: Natural aerosols explain seasonal and spatial patterns of Southern Ocean cloud albedo, Science Advances, 1, doi:10.1126/sciadv.1500157, 2015.
 - Meskhidze, N., Xu, J., Zhang, Y., Gantt, B., Ghan, S., Nenes, A., Liu, X., Easter, R., and Zaveri, R.: Effect of marine biogenic organic aerosols on cloud properties: Modeling study, Geochimica Et Cosmochimica Acta, 73, A874-A874, 2009.

790 Meskhidze, N., and Nenes, A.: Phytoplankton and cloudiness in the Southern Ocean. Science, 314(5804), 1419-1423, 2006. Meskhidze, N., and Nenes, A.: Effects of Ocean Ecosystem on Marine Aerosol-Cloud Interaction, Advances in Meteorology, 2010, doi:10.1155/2010/239808, 2010.

795

800

- Meskhidze, N., Xu, J., Gantt, B., Zhang, Y., Nenes, A., Ghan, S. J., Liu, X., Easter, R., and Zaveri, R.: Global distribution and climate forcing of marine organic aerosol: 1. Model improvements and evaluation, Atmospheric Chemistry and Physics, 11, 11689-11705, doi:10.5194/acp-11-11689-2011, 2011.
- Modini, R. L., Harris, B. and Ristovski, Z.: The organic fraction of bubble-generated, accumulation mode Sea Spray Aerosol (SSA), Atmos. Chem. Phys., 10(6), 2867–2877, 2010.
 - Murphy, R. J., Pinkerton, M. H., Richardson, K. M., Bradford-Grieve, J. M., and Boyd, P. W.: Phytoplankton distributions around New Zealand derived from SeaWiFS remotely-sensed ocean colour data, New Zealand Journal of Marine and Freshwater Research, 35, 343-362, 2001.
- Nightingale, P. D., Malin, G., Law, C. S., Watson, A. J., Liss, P. S., Liddicoat M.I., Boutin, J., and Upstill-Goddard, R. C.: In situ evaluation of air-sea gas exchange parameterizations using novel conservative and volatile tracers, Global Biogeochemical Cycles, 14, 373-387, 2000.
- Nodder, S. D., Duineveld, G. C. A., Pilditch, C. A., Sutton, P. J., Probert, P. K., Lavaleye, M. S. S., Witbaard, R., Chang, F.
 H., Hall, J. A., and Richardson, K. M.: Focusing of phytodetritus deposition beneath a deep-ocean front, Chatham Rise, New Zealand, Limnology and Oceanography, 52, 299-314, 2007.
 - O'Dowd, C., Monahan, C., and Dall'Osto, M.: On the occurrence of open ocean particle production and growth events, Geophysical Research Letters, 37, doi:10.1029/2010gl044679, 2010.
- O'Dowd, C. D., Facchini, M. C., Cavalli, F., Ceburnis, D., Mircea, M., Decesari, S., Fuzzi, S., Yoon, Y. J., and Putaud, J.-P.:
 Biogenically driven organic contribution to marine aerosol, Nature, 431, 676-680, 2004.
 - O'Dowd, C. D., and de Leeuw, G.: Marine aerosol production: a review of the current knowledge, Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences, 365, 1753-1774, 2007.
- Popinet, S., Smith, M., and Stevens, C.: Experimental and Numerical Study of the Turbulence Characteristics of Airflow around a Research Vessel, Journal of Atmospheric and Oceanic Technology, 21, 1575-1589, doi:10.1175/1520-0426(2004)021<1575:EANSOT>2.0.CO;2 2004.
 - Quinn, P. K., and Bates, T. S.: The case against climate regulation via oceanic phytoplankton sulphur emissions, Nature, 480, 51-56, 2011.
 - Quinn, P.K., Bates, T.S., Schulz, K.S., Coffman, D.J., Frossard, A.A., Russell, L.M., Keene, W.C., Kieber, D.J.: Contribution of sea surface carbon pool to organic matter enrichment in sea spray aerosol. Nature Geoscience, 7(3), 228-232, 2014.
- 820 Rinaldi, M., Fuzzi, S., Decesari, S., Marullo, S., Santoleri, R., Provenzale, A., von Hardenberg, J., Ceburnis, D., Vaishya, A., O'Dowd, C. D., and Facchini, M. C.: Is chlorophyll-a the best surrogate for organic matter enrichment in submicron primary marine aerosol?, Journal of Geophysical Research-Atmospheres, 118, 4964-4973, doi:10.1002/jgrd.50417, 2013.
- Russell, L. M., Bahadur, R. and Ziemann, P. J.: Identifying organic aerosol sources by comparing functional group composition
 in chamber and atmospheric particles, Proceedings of the National Academy of Sciences, 2011.
 - Russell, L. M., Hawkins, L. N., Frossard, A. A., Quinn, P. K., and Bates, T. S.: Carbohydrate-like composition of submicron atmospheric particles and their production from ocean bubble bursting, Proceedings of the National Academy of Sciences of the United States of America, 107, 6652-6657, doi:10.1073/pnas.0908905107, 2010.
- Sadeghi, A., Dinter, T., Vountas, M., Taylor, B. B., Altenburg-Soppa, M., Peeken, I., and Bracher, A.: Improvement to the PhytoDOAS method for identification of coccolithophores using hyper-spectral satellite data, Ocean Science, 8, 1055-1070, doi:10.5194/os-8-1055-2012, 2012.
 - Scanlon, B., and Ward, B.: The influence of environmental parameters on active and maturing oceanic whitecaps, Journal of Geophysical Research: Oceans, 121, doi:10.1002/2015JC011230, 2016.
- Seinfeld, J. H., Bretherton, C., Carslaw, K. S., Coe, H., DeMott, P. J., Dunlea, E. J., Feingold, G., Ghan, S., Guenther, A. B.,
 Kahn, R., Kraucunas, I., Kreidenweis, S. M., Molina, M. J., Nenes, A., Penner, J. E., Prather, K. A., Ramanathan, V.,
 Ramaswamy, V., Rasch, P. J., Ravishankara, A. R., Rosenfeld, D., Stephens, G., and Wood, R.: Improving our fundamental understanding of the role of aerosol–cloud interactions in the climate system, Proceedings of the National Academy of Sciences, 113, 5781-5790, doi:10.1073/pnas.1514043113, 2016.
- Sellegri, K., Loon, Y. J., Jennings, S. G., O'Dowd, C. D., Pirjola, L., Cautenet, S., Chen, H. W., and Hoffmann, T.:
 Quantification of coastal new ultra-fine particles formation from in situ and chamber measurements during the BIOFLUX campaign, Environmental Chemistry, 2, 260-270, doi:10.1071/en05074, 2005.
- Sievering, H., Cainey, J., Harvey, M., McGregor, J., Nichol, S., and Quinn, P.: Aerosol non-sea-salt sulfate in the remote marine boundary layer under clear-sky and normal cloudiness conditions: Ocean-derived biogenic alkalinity enhances sea-salt sulfate production by ozone oxidation, Journal of Geophysical Research-Atmospheres, 109, doi:10.1029/2003jd004315, 2004.
- Smith, M. J., Ho, D. T., Law, C. S., McGregor, J., Popinet, S., and Schlosser, P.: Uncertainties in gas exchange parameterization during the SAGE dual-tracer experiment, Deep-Sea Research Part II-Topical Studies in Oceanography, 58, 869-881, doi:10.1016/j.dsr2.2010.10.025, 2011.
- Smith M.J., Walker C.F., Bell T.G., Harvey M.J., Law C.S., Saltzman E.S. Air-sea gradient fluxes of DMS during the Surface
 Ocean Aerosol Production (SOAP) experiment. Ocean Sciences, *submitted*

- Spada, M., Jorba, O., Pérez García-Pando, C., Janjic, Z., and Baldasano, J. M.: On the evaluation of global sea-salt aerosol models at coastal/orographic sites, Atmospheric Environment, 101, 41-48, doi:http://dx.doi.org/10.1016/j.atmosenv.2014.11.019, 2015.
- Stephens, G. L.: Cloud feedbacks in the climate system: A critical review, Journal of climate 18, 237-273, 2005.
- 855 Stevens, C. L., Hill, P., Smith, M. J., and Popinet, S.: SUCA: An engine for repetitive autonomous profiling near the ocean surface, Limnol Oceanogr-Meth, 3, 300-307, 2005.
 - Swan, H. B., Armishaw, P., Lavetz, R., Alamgir, M., Davies, S. R., Bell, T. G., and Jones, G. B.: An interlaboratory comparison for the quantification of aqueous dimethylsulfide, Limnology and Oceanography-Methods, 12, 784-794, doi:10.4319/lom.2014.12.784, 2014.
- 860 Thomas, M. A., Suntharalingam, P., Pozzoli, L., Rast, S., Devasthale, A., Kloster, S., Feichter, J., and Lenton, T. M.: Quantification of DMS aerosol-cloud-climate interactions using the ECHAM5-HAMMOZ model in a current climate scenario, Atmospheric Chemistry and Physics, 10, 7425-7438, 2010.
 - Trenberth, K. E., and Fasullo, J. T.: Simulation of present-day and twenty-first-century energy budgets of the southern oceans, J. Climate, 23, 440-454, 2010.
- 865 Twomey, S.: The influence of pollution on the shortwave albedo of clouds. Journal of the atmospheric sciences, *34*(7), 1149-1152, 1977.
 - Vaattovaara, P., Räsänen, M., Kühn, T., Joutsensaari, J., and Laaksonen, A.: A method for detecting presence of organic fraction in nucleation mode sized particles, Atmospheric Chemistry and Physics, 5, 3277-3287, 2005.
- Vaattovaara, P., Huttunen, P. E., Yoon, Y. J., Joutsensaari, J., Lehtinen, K. E. J., O'Dowd, C. D., and Laaksonen, A.: The composition of nucleation and Aitken modes particles during coastal nucleation events: evidence for marine secondary organic contribution, Atmos. Chem. Phys., 6, 4601-4616, doi:10.5194/acp-6-4601-2006, 2006.
 - Vaishya, A., Jennings, S. G., and O'Dowd, C.: Wind-driven influences on aerosol light scattering in north-east Atlantic air, Geophysical Research Letters, 39, 2012.
- Vallina, S.M., Simó, R., Gassó, S.: What controls CCN seasonality in the Southern Ocean? A statistical analysis based on satellite-derived chlorophyll and CCN and model-estimated OH radical and rainfall. Global Biogeochemical Cycles, 20(1), 2006
 - Vignati, E., Facchini, M.C., Rinaldi, M., Scannell, C., Ceburnis, D., Sciare, J., Kanakidou, M., Myriokefalitakis, S., Dentener, F., O'Dowd, C.D.: Global scale emission and distribution of sea-spray aerosol: Sea-salt and organic enrichment. Atmospheric Environment, 44(5), 670-677, 2010.
- 880 Villani, P., Picard, D., Michaud, V., Laj, P., and Wiedensohler, A.: Design and Validation of a Volatility Hygroscopic Tandem Differential Mobility Analyzer (VH-TDMA) to Characterize the Relationships Between the Thermal and Hygroscopic Properties of Atmospheric Aerosol Particles, Aerosol Science and Technology, 42, 729-741, doi:10.1080/02786820802255668, 2008.
- Walker, C. F., Harvey, M. J., Smith, M. J., Bell, T., Saltzman, E., Marriner, A. S., McGregor, J. A., and Law, C. S.: Assessing
 the potential for DMS enrichment at the sea-surface and its influence on sea-to-air flux Ocean Sciences, 12, 1033-1048, doi:doi:10.5194/os-12-1033-2016., 2016.
 - Wang, H., Easter, R. C., Rasch, P. J., Wang, M., Liu, X., Ghan, S. J., Y., Q., JH., Y., P.L., M., and Vinoj, V.: Sensitivity of remote aerosol distributions to representation of cloud–aerosol interactions in a global climate model, Geoscientific Model Development, 6.3, 765-782, 2013.
- 890 Willis, M.D., Burkart, J., Thomas, J.L., Köllner, F., Schneider, J., Bozem, H., Hoor, P.M., Aliabadi, A.A., Schulz, H., Herber, A.B. and Leaitch, W.R.: Growth of nucleation mode particles in the summertime Arctic: a case study. *Atmospheric Chemistry and Physics*, 16(12), 7663-7679, 2016.
 - Woodhouse, M. T., Luhar, A. K., Stevens, L., Galbally, I., Thatcher, M., Uhe, P., Wolff, H., Noonan, J., and Molloy, S. Australian reactive-gas emissions in a global chemistry-climate model and initial results Air Quality and Climate Change, 49, 31-38, 2015.
 - Wylie, D. J., and de Mora, S. J.: Atmospheric dimethylsulfide and sulfur species in aerosol and rainwater at coastal site in New Zealand, Journal of Geophysical Research, 101, 21041-21049, 1996.
- Yang, M., Blomquist, B. W., Fairall, C. W., Archer, S. D., and Huebert, B. J.: Air-sea exchange of dimethylsulfide in the Southern Ocean: Measurements from SO GasEx compared to temperate and tropical regions, J. Geophys. Research, 116,, doi:10.1029/2010JC006526, 2011.
 - Young, I. R., Vinoth, J., Zieger, S., and Babanin, A. V.: Investigation of trends in extreme value wave height and wind speed, J Geophys Res-Oceans, 117, 2012.
 - Zhang, R.: Getting to the Critical Nucleus of Aerosol Formation, Science, 328, 1366-1367, doi:10.1126/science.1189732, 2010.

905

895

13. Tables

	Time/Location				Meteorological				Hydrodynamic			Biogeochemical				
Bloom	Start NZST (DoY UTC)	End NZST (DoY UTC)	Bloom Centre Lat.	Bloom Centre Long.	Atmos Press. mb	Irrad W m ⁻²	U ₁₀ Range m s ⁻¹	Hs m	MLD m *	SST °C #	Sal. #	Nitrate/ Phosphate/ Silicate µmol L ⁻¹ *	Chl-a mg/m ³	pCO ₂ ppmv #	DMSsw nmolL ⁻¹ #	Dominant Phytoplankton *
B1	14/02/12 2:00 (44.6)	19/02/12 12:00 (50.0)	-44.34 -44.61	174.2 174.78	1019.1 ± 2.9	232 <1061	6.6 (5- 7.6)	2.0	14.5 ±1	14.5 ±0.4	34.48	5.3 ± 0.9 0.43 ± 0.2 0.35 ± 0.1	0.84 ±0.2 <3.4	320 ±24	16.8 ±1.5	Dinoflagellate
B2	21/02/12 16:15 (52.2)	26/02/12 12:00 (57.0)	-43.55 -43.71	180.16 180.32	1011.5 ± 5.3	196 <1079	10.4 (6.9- 12.4)	2.9	24.0 ±9	15.8 ±0.2	34.6	$\begin{array}{c} 1.7 \pm 1.0 \\ 0.27 \pm \\ 0.07 \\ 0.41 \pm \\ 0.33 \end{array}$	0.67 ±0.3 <1.0	339 ±9	9.1 ±2.9	Coccolithophore Dinoflagellate
B3a	27/02/12 10:00 (57.9)	1/03/12 04:00 (60.67)	-44.11 -44.61	174.47 174.88	1010.0 ± 8.2	242 <1212	10.3 (8.1- 12.1)	2.6	28.6 ±1.7	14.4 ±0.2	34.32	$\begin{array}{r} 3.7 \pm 1 \\ 0.34 \pm \\ 0.06 \\ 0.3 \pm 0.16 \end{array}$	$\begin{array}{r} 0.44 \ \pm \\ 0.17 \\ {<}0.92 \end{array}$	333 ±14	5.37 ±1.5	Mixed
B3b	02/03/12 06:00 (61.7)	5/03/12 17:00 (65.2)	-44.19 -44.78	174.3 174.93	1008.6 ± 9.4	182 <1016	12.6 (8.5- 14.9)	3.6	41.1 ±6	13.2 ±0.4	34.32	$\begin{array}{rrr} 4.2 \pm 1.1 \\ 0.39 \pm 0.1 \\ 0.48 & \pm \\ 0.05 \end{array}$	0.59 ±0.2 <1.1	340 ±8	3.1 ±1.2	Mixed

Table 1. Summary of surface water characteristics during each bloom period. All values are Mean ± 1 standard deviation, except where maximum value also shown by <. * indicates value derived from 2-10m depth on all stations during bloom occupation; # indicates continuous measurement in surface waters (nominal 6m depth). Abbreviations: Lat: Latitude; Long.: Longitude; Atmos. Press.; Atmospheric Pressure; Irrad.: Irradiance; U₁₀: Wind speed adjusted to 10m height (uncorrected for vessel flow distortion); Hs: Significant wave Height; MLD: Mixed Layer Depth; SST: Sea Surface Temperature; Sal>: Surface salinity; Chl-*a*: Chlorophyll-*a*.

WP1 Amospheric Organic nuclei production C* UtraFine Organic-Tandem Differential Mobility Analyser (UFO-TDMA) Aerosol water uptake and volatility C* Volatility Humidity Differential Mobility Analyser (VH-TDMA) Nucleation/Aitken mode size spectra Condensation nuclei counts C* Scanning Mobility Particle Sizer (SMPS) Condensation nuclei counts C* Consensation Particle Counter (CPC) Accumulation mode aerosol number C PCASP Condensation nuclei counts C* CN spectrometer Actosol filter chemistry – Major ions C Hi-Vol, cascade, Ion Chromatograph Black carbon D Sun photometer (Microtops II) Nascosol filters C Organic functional groups by FTIR and inorganic composition by IBA Column aerosol D Sun photometer (Microtops II) Nascost sea spray composition via D Chamber experiments bubble bust set seaver as spray composition via D Chamber experiments OVCS (Acctone, DMS, Acctonitrile, Monoterpnes, Acetaldehyde) Proconcentration and TD-GC-FID/MS VOCS (Acctone, DMS, Acctonitrile, Monoterpnes, Acetaldehyde) D Pre-concentration and TD-GC-FID/MS	Measurement	Mode	Instrument				
Organic nuclei production C* UltraFine Organic-Tandem Differential Mobility Analyser (UFO-TDMA) Aerosol water uptake and volatility C* Volatility Humidity Differential Mobility Analyser (UFI-TDMA) Nucleation/Aitken mode size spectra C* Scanning Mobility Particle Sizer (SMPS) Condensation nuclei counts C* Condensation Particle Counter (CPC) Accumulation mode acrosol number C* CASP Cloud condensation nuclei C* CASP Cloud condensation nuclei C* CASP Cloud condensation nuclei C* CASP Cloum acrosol D Sun photometer (Microtops II) Nascent sea spray composition via bubble burst of sea-water samples D Chamber experiments DMS C mesocIMS (Chemical Ionization Mass Spectrometry) Colarn acrosol D Sun photometer (Microtops II) Nascent sea spray composition via bubble burst of sea-water samples D Chamber experiments DMS C mesocIMS (Chemical Ionization Mass Spectrometry) Oclaamber experiments C Proton Transfer Reaction Mass Spectrometry) OXG & Cy to C15 D Pre-concentration and TD-GC-FID/MS Aldehydes, ketones (incl. D Derivatisation and HPLC Methanol, Michameter motion sensors DMS <	WP1 Atmospheric						
Aerosol water uptake and volatility C* Analyser (UF-ČTDMA) Aerosol water uptake and volatility C* Volatility Humidity Differential Mobility Analyser (UH-TDMA) Nucleation / Aitken mode size spectra Condensation nuclei counter C* Scanning Mobility Particle Sizer (SMPS) Condensation nuclei counter C* Condensation Particle Counter (CPC) Accoundiation mode aerosol number C PCASP Cloud condensation nuclei C* CY spectrometer Accoundiation acrosol D Sun photometer (Microtops II) Nascent sea spray composition via bubble burst of sca-water samples D Sun photometer (Microtops II) DMS C mesoCIMS (Chemical Ionization Mass Spectrometry) CO2 ad methane C Picarro CRDS Halocarbons, Iodine & halogen mesoCIMS (Chemical Ionization Mass Spectrometry) CO3 ad methane C Pre-concentration and TD-GC-FID/MS Addelydes, ketones (incl. D D Pre-concentration and TD-GC-FID/MS Addelydes, ketones D C Mass Spectrometry) VOS C Ac to C is D P Creansers Solic anemometer motion sensors DMS flux C Licers, Sonic anemometer motion sensors DMS gradient flux D Catamaran, SCD-GC Near surface trubulence <	Organic nuclei production	C*	UltraFine Organic-Tandem Differential Mobility				
Aerosol water uptake and volatility C* Volatility Humidity Differential Mobility Analyser (VH-TDMA) Nucleation/Aitken mode size spectra C* Scanning Mobility Particle Sizer (SMPS) Condensation nuclei counts C* Condensation Particle Counter (CPC) Accumulation mode acrosol number C PCASP Cloud condensation nuclei C* CCASP Cloud condensation nuclei C* CCASP Cloud condensation nuclei C* CASP Cloum aerosol D Sun photometer Mascent sea spray composition via bubble burst of sea-water samples D Chamber experiments DMS C mesoCIMS (Chemical Ionization Mass Spectrometry) CO2 and methane C Pricaro CRDS Halocarbons, Iodine & halogen oxides C Proton Transfer Reaction Mass Spectrometer (PTR- MS) Methanol, Methanetholi, Isoprene, Monotrepenes, Acetaldehyde) D Priconcentration and TD-GC-FID/MS Aldehydes, ketones (incl. dicatoryls), C ₂ to C D Pre-concentration and TD-GC-FID/MS Aldehydes, ketones (incl. dicatoryls), C ₂ to C D Catamaran, SCD-GC WP2 Physits D Catamaran, SCD-GC Sectrol	Contraction of the second seco		Analyser (UFO-TDMA)				
Nucleation/Aitken mode size spectra C* CVH-TDDA) Nucleation/Aitken mode size spectra C* Scanning Mobility Particle Sizer (SMPS) Condensation nuclei counts C* Consensation Particle Counter (CPC) Accumulation mode acrosol number C PCASP Condensation nuclei C* CS spectrometer Accoundation nuclei C* CN spectrometer Accoundation nuclei C Organic functional groups by FTIR and inorganic composition by IBA Column aerosol D Sun photometer (Microtops II) Nascent sea spray composition via D Chamber experiments DMS C mesoCIMS (Chemical Ionization Mass Spectrometry) CO2 and methane C Picarro CRDS Halocarbons, Iodine & halogen oxidas D VOCs (Acctone, DMS, Acetaldehyde) VOCs (Acctone, Spectroscopy (Max-DOAS) VOCs (C to C1s D D Proton Transfer Reaction Mass Spectrometry) OLi acarbonyls, Octo C1s D <td>Aerosol water untake and volatility</td> <td>C*</td> <td>Volatility Humidity Differential Mobility Analyser</td>	Aerosol water untake and volatility	C*	Volatility Humidity Differential Mobility Analyser				
Nucleation/Aitken mode size spectra Condensation nuclei countsC* Scamming Mobility Particle Sizer (SMPS) Condensation Particle Counter (CPC) PCASPAccumulation mode aerosol number Cloud condensation nucleiC*CASP PCASPCloud condensation nucleiC*CCASP CCN spectrometerAerosol filter tenmistry – Major ions Black carbonC*CASP AetholometerPM aerosol filtersCOrganic functional groups by FTIR and inorganic composition by IBAColumn aerosolDSun photometer (Microtops II)Nascent sea spray composition via bubble burst of sca-water samplesDDMSCPicarro CRDSCO2 and methaneCPicarro CRDSHalocarbons, Iodine & halogen oxidesCProton Transfer Reaction Mass SpectrometryVOCs (Acctone, DMS, Acctonitrile, Moonterpnes, Acetaldelyde)DPre-concentration and TD-GC-FID/MSVOCs (Acctone, CIS VOCs (s to C15DDerivatisation and HPLCWP2 PhysicsDPre-concentration and TD-GC-FID/MSDMS gradient fluxCmesoCIMS (Chemical Ionization Mass Spectrometry)CO2 EC fluxCTDNear-surface stratificationCSpar buoy – temperature array, microcatsNear-surface stratificationCSpar buoy – temperature array, microcatsNear-surface traditionsCAWSPMS predient fluxDCatamera, SCD-GCNear-surface traditionsCAWSDMS gradient fluxDRadiosondePWP PhysicsCNOAA Wavewatch III <td>relosor water uptake and volutility</td> <td>e</td> <td>(VH-TDMA)</td>	relosor water uptake and volutility	e	(VH-TDMA)				
Nucleation Article Induces Size SpectraCScanning Hooling Failed Counter (CPC)Condensation nuclei courtsC*Condensation Particle Counter (CPC)Accumulation nuclei courtsCPCASPCloud condensation nucleiC*CCN spectrometerAerosol filter chemistry – Major ionsCHi-Vol, cascade, Ion ChromatographBlack carbonC*AetholometerPM1 aerosol filtersCOrganic functional groups by FTIR and inorganic composition by IBAColumn aerosolDSun photometer (Microtops II)Nascent sea spray composition via bubble burst of sea-water samplesDPMSCmesoCIMS (Chemical Ionization Mass Spectrometry)CO2 and methaneC μ -Dirac ECD-GC and Multi-Axis Differential Optical Absorption; Spectroscopy (Max-DOAS)VOCs (Acetone, DMS, Acetonitrile, Methanol, Methanethiol, Isoprene, Monterpenes, Acctaldehyde)DPre-concentration and TD-GC-FID/MSAldehydes, ketones (incl. DDerivatisation and HPLC dicarboryls), C ₂ to C ₃ Tere-concentration and HPLC dicarboryls), C ₂ to C ₃ DMS fluxCmesoCIMS (Chemical Ionization Mass Spectrometry)CO2 EC fluxCLicors, Sonic aneometer motion sensorsDMS gradient fluxDCatamaran, SCD-GCNear-surface turbulenceCVector, FastCat actamaran, SCD-GCNear-surface turbulenceCNOAA Wavewatch IIIWhitecap coverageDCameraBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP); <td< td=""><td>Nucleation/Aitkan mode size spectra</td><td>C*</td><td>(VII-IDMA) Scanning Mobility Particle Sizer (SMPS)</td></td<>	Nucleation/Aitkan mode size spectra	C *	(VII-IDMA) Scanning Mobility Particle Sizer (SMPS)				
Concensation nuclei counts C C Concensation Particle Counter (CPC) Accumulation mode aerosol number C PCASP Cloud condensation nuclei C C CASP CCN spectrometer CCN spectrometer C C Status C C C Status C C C Status C C C C Status C C C C C C C C C C C C C C C C C C C	Condensation pueloi counta	C*	Condensation Derticle Counter (CDC)				
Accumulation mode aeroson number C PCASP Cloud condensation nuclei C* CCN spectrometer Aerosol filter chemistry – Major ions C Hi-Vol, cascade, Ion Chromatograph Black carbon C* Aetholometer PM ₁ aerosol filters C Organic functional groups by FTIR and inorganic composition by IBA Column aerosol D Sun photometer (Microtops II) Nascent sea spray composition via D Chamber experiments bubble burst of sea-water samples DMS C mesoCIMS (Chemical lonization Mass Spectrometry) CO2 and methane C Picarro CRDS Halocarbons, Iodine & halogen oxides C Proton Transfer Reaction Mass Spectrometer (PTR- Methanol, Methanethiol, Isoprene, Monoterpenes, Acetaldehyde) VOCS C ₁ to C ₁₅ D Pre-concentration and TD-GC-FID/MS Aldehydes, ketones (incl. D Derivatisation and HPLC dicarbonyls), C ₂ to C ₈ WP2 Physics DMS flux C C Near-surface turbulence C Vector, FastCat Near-surface strafification C Spar buoy – temperature array, microcats Near-surface turbulence C Vector, FastCat Sea state C NOAA Wavewatch III Whitecap coverage D Camera Meteorological conditions C AWS Bulk fluxes C Eppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP); MBL Height and stability D Radiosonde WT3 Occan biogeochemistry Chorophyll-a D, W Spectrophotometer DIC D Nutrients D, W Colorimetric Autoanalyser POC/PON/PC/PN/13C/15N D Mass Spectrometry Dissolved DMS C, miniCIMS (Chemical lonization Mass Spectrometry) Dissolved DMS D, W SCD Figments D, W CD Figments D, W CD Figments D, W CD Figments D, W CD Fi	A summer lation mucher counts	C*	Condensation Particle Counter (CPC)				
Cloud condensation nucleiC*CCN spectrometerAcrosof filter chemistry – Major ionsCHi-Vol, cascade, Ion ChromatographBlack carbonC*AetholometerPM1 aerosol filtersCOrganic functional groups by FTIR and inorganic composition by IBAColumn aerosolDSun photometer (Microtops II)Nascent sea spray composition via bubble burst of sea-water samplesDDMSCPicarro CRDSDMSCPicarro CRDSMato and methaneCPicarro CRDSHalocarbons, Iodine & halogen oxides μ -Dirac ECD-GC and Multi-Axis Differential Optical Absorption; Spectroscopy (Max-DOAS)VOCs (Acetone, DMS, Acetonitrile, Monoterpenes, Acetaldehyde)DVOCs (Acetone, Acetaldehyde)Pre-concentration and TD-GC-FID/MSAldehydes, carboryls, C ₂ to C ₅ DPre-concentration and HPLCWP2 PhysicsDPrevatination and HPLCOZ 2E C fluxCLicors, Sonic anemometer motion sensorsDMS gradient fluxDCatamaran, SCD-GCNear-surface trafdSDCTDNear-surface tradeloneCW2 PhysicsDDMS gradient fluxDRear surface tradeloneCNear-surface tradeloneCNear-surface tradeloneCNear-surface tradeloneCNear-surface tradeloneCNear-surface tradeloneCNear-surface tradeloneCNear-surface tradeloneCDCambraBulk flux	Accumulation mode aerosol number	C C*	PCASP				
Acrosol fulter chemistry – Major ions C Hi-Vol, cascade, Ion Chromatograph Black carbon C* Actholometer PM ₁ aerosol filters C Organic functional groups by FTIR and inorganic composition by IBA Column aerosol D Sum photometer (Microtops II) Nascent sea spray composition via bubble burst of sea-water samples DMS C Picarro CRDS Halocarbons, Iodine & halogen oxides C Picarro CRDS Halocarbons, Iodine & halogen oxides C Picarro CRDS C Picarro CRDS Monoterpenes, Acetaldehyde) VOCs (Acetone, DMS, Acetonitrile, Monoterpenes, Acetaldehyde) VOCs (C to C ₁₅ D Pre-concentration and TD-GC-FID/MS Aldehydes, ketones (incl. D Derivatisation and HPLC dicarbonyls), C ₂ to C ₈ WP2 Physics DMS Tux C mesoCIMS (Chemical Ionization Mass Spectrometry) CO2 EC flux C Licors, Sonic anemometer motion sensors DMS gradient flux D Catamaran, SCD-GC Near-surface tradification C Spar buoy – temperature array, microcats Near-surface stratification C Spar buoy – temperature array, microcats Near-surface tradifies C AWS Meteorological conditions C C AWS Bulk fluxes C Expley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP); MBI. Height and stability D Radiosonde WP3 Ocean biogeochemistry Chorophyll- <i>a</i> Back-scatter β_{860} backscatter C C, D, W Ecotriplet PCO2 C D, W Colorimetric Autoanalyser POC/PON/PC/PN/13C/15N D Mass Spectrometer) Disolved DMS C, miniCIMS (Chemical Ionization Mass Spectrometry) Disolved DMS C, miniCIMS (Chemical Mass Spectrometer) Disolved DMS D, W Spectrophotometer POC/PON/PC/PN/13C/15N D Mass Spectrometer) Disolved DMS C, miniCIMS (Chemical Ionization Mass Spectrometry) Dissolved DMS D, W SCD/FPD DMSP & processes D, W SCD Figments D, W Colorimetric Autoanalyser PO/CPON/PC/PN/13C/15N D Mass Spectrometer) Disolved DMS D, W SCD/FPD DMSP & processes D, W SCD Figments D W Optical Microscopy	Cloud condensation nuclei	C*	CCN spectrometer				
Black carbon C* Actholometer PM1 aerosol filters C Organic functional groups by FTIR and inorganic composition by IBA Column aerosol D Sun photometer (Microtops II) Mascent sea spray composition via bubble burst of sea-water samples D Chamber experiments DMS C mesoCIMS (Chemical Ionization Mass Spectrometry) CO2 and methane C picarro CRDS Halocarbons, Iodine & halogen oxides C Proton Transfer Reaction Mass Spectrometer (PTR-Methanol, Methanethiol, Isoprene, MS) Monoterpenes, Acetaldehyde) VOCs (Acetone, DMS, Acetonitrile, C D Pre-concentration and TD-GC-FID/MS VOCs (C to C15 D Pre-concentration and TD-GC-FID/MS Aldehydes, ketones (incl. D Derivatisation and HPLC VO2 EC flux C Licors, Sonic anemometer motion sensors D Catamaran, SCD-GC DMS gradient flux D Catamaran, SCD-GC Near-surface stratification C Spar buoy – temperature array, microcats Near-surface turbulence C NOAA Wavewatch III Whitecap coverage D Camera Meteorological conditions C Agaiosonde Spectraplyranometer (PSP); MEI deigh and s	Aerosol filter chemistry – Major ions	C	Hi-Vol, cascade, Ion Chromatograph				
PM ₁ acrosol filters C Organic functional groups by FTIR and inorganic composition by IBA Column aerosol D Sun photometer (Microtops II) Nascent sea spray composition via bubble burst of sea-water samples D Chamber experiments DMS C mesoCIMS (Chemical Ionization Mass Spectrometry) CO2 and methane C Picarro CRDS Halocarbons, Iodine & halogen C μ-Dirac ECD-GC and Multi-Axis Differential Optical Absorption; Spectroscopy (Max-DOAS) VOCs (Acetone, DMS, Acetonitrile, Motorepnes, Acetaldehyde) Proton Transfer Reaction Mass Spectrometer (PTR-Methanol, Methanethiol, Isoprene, MS) Monoterpnes, Acetaldehyde) VOCs (5 to C ₁₅ D Pre-concentration and TD-GC-FID/MS Aldehydes, ketones (incl. D Derivatisation and HPLC dicarbonyls), C ₂ to C _k WP2 Physics DMS gradient flux D Catamaran, SCD-GC Near-surface tranulication Mass Spectrometry) CO2 EC flux C NoAA Wavewatch III Mear-surface tranulication C Spar buoy – temperature array, microcats Near-surface tranulence C NoAA Wavewatch III Mear-surface turbulence C NoAA Wavewatch III Near-surface turbulence C NoAA Wavewatch III	Black carbon	C*	Aetholometer				
composition by IBA Column acrosol D Sun photometer (Microtops II) Nascent sea spray composition via bubble burst of sea-water samples D Chamber experiments DMS C mesoCIMS (Chemical Ionization Mass Spectrometry) CO2 and methane C Picarro CRDS Halocarbons, Iodine & halogen C µ-Dirac ECD-GC and Multi-Axis Differential Optical Absorption; Epectroscopy (Max-DOAS) VOCs (Acctone, DMS, Acetonitrile, Monoterpenes, Acetaldehyde) C Proton Transfer Reaction Mass Spectrometer (PTR- Methanol, Methanethiol, Isoprene, MS) VOCs C ₅ to C ₁₅ D Pre-concentration and TD-GC-FID/MS Aldehydes, ketones (incl. D Derivatisation and HPLC V022 FC fux C Licors, Sonic anemometer motion sensors DMS gradient flux D Catamaran, SCD-GC Near-surface stratification C Spar buoy – temperature array, microcats Near-surface turbulence C NoAA Wavewatch III Whitecap coverage D Camera Meteorological conditions C AWS Bulk fluxes C Ecotriplet Back-scatter & β ₆₆₀ ba	PM_1 aerosol filters	С	Organic functional groups by FTIR and inorganic				
Column aerosolDSun photometer (Microtops II)Nascent sea spray composition via bubble burst of sea-water samplesDChamber experimentsDMSCmesoCIMS (Chemical Ionization Mass Spectrometry)CO2 and methaneCµ-Dirac ECD-GC and Multi-Axis Differential Optical Absorption; Spectroscopy (Max-DOAS)VOCs (Acetone, DMS, Acetonitrile, Monoterpenes, Acetaldehyde)CProton Transfer Reaction Mass Spectrometer (PTR- MS)Monoterpenes, Acetaldehyde)VOCs (C to C15DPre-concentration and TD-GC-FID/MSAldehydes, ketones (incl. dicarbonyls), C to C5DPre-concentration and HPLCdicarbonyls, C to C5DPre-concentration and MSDMS fluxCmesoCIMS (Chemical Ionization Mass Spectrometry)CO2 EC fluxCLicors, Sonic anemometer motion sensorsDMS gradient fluxDCatamaran, SCD-GCNear-surface turbulenceCVector, FastCatSea stateCNOAA Wavewatch IIIWhitecap coverageDCameraMeteorological conditionsCAWSBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MEL Height and stabilityDRadiosondeWP3 Ocean biogeochemistryCNewDOCDWDOCDWDOCDPDOCDPDOCDFireDOCDFireDOCDHTCODOCDHTCOD			composition by IBA				
Nascent sea spray composition via bubble burst of sea-water samplesDChamber experimentsDMSCmesoCIMS (Chemical Ionization Mass Spectrometry)CO2 and methaneC μ -Dirac ECD-GC and Multi-Axis Differential Optical Absorption; Spectroscopy (Max-DOAS)VOCs (Acetone, DMS, Acetonitrile, Methanoth, Isoprene, Monoterpenes, Acetaldehyde)CProton Transfer Reaction Mass Spectrometer (PTR- MS)Workethanethic, Isoprene, Monoterpenes, Acetaldehyde)DPre-concentration and TD-GC-FID/MSVOCs C; to C15DDPer-concentration and HPLCdicarbonyls), C2 to C3CLicors, Sonic anemometer motion sensorsVP2 PhysicsDCatamaran, SCD-GCDMS gradient fluxDCatamaran, SCD-GCNear-surface tradificationCSpar buoy – temperature array, microcatsNear-surface tradificationCAWSNear-surface turbulenceCAWSSea stateCNOAA Wavewatch IIIWhitecap coverageDCameraMetorological conditionsCAWSBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MBL Height and stabilityDRadiosondePCO2DWCocon biogeochemistryCChlorophyll-aC, D, WEquation of the sector of the	Column aerosol	D	Sun photometer (Microtops II)				
bubble burst of sea-water samplesmesoCIMS (Chemical Ionization Mass Spectrometry)DMSCPicarro CRDSHalocarbons, Iodine & halogenCµ-Dirac ECD-GC and Multi-Axis Differential Optical Absorption; Spectroscopy (Max-DOAS)VOCs (Acetone, DMS, Acetonitrile, Methanol, Methanethiol, Isoprene, Monoterpenes, Acetaldehyde)CProton Transfer Reaction Mass Spectrometer (PTR- MS)VOCs (C Acetone, DMS, Acetonitrile, Monoterpenes, Acetaldehyde)DPre-concentration and TD-GC-FID/MSAldehydes, ketones (incl.DPer-concentration and HPLCdicarbonyls), C ₂ to C_sWDWP2 PhysiosDDMS fluxCmesoCIMS (Chemical Ionization Mass Spectrometry)CO2 EC fluxCLicors, Sonic anemometer motion sensorsDMS gradient fluxDCatamaran, SCD-GCNear-surface tarbulenceCVector, FastCatSea stateCNOAA Wavewatch IIIWhitecap coverageDCameraMeteorological conditionsCAWSBulk fluxesCEcotripletPOCCRGAPHC, D, WSpectrophotometerDOCD, WTCONutrientsD, WColorimetric AutoanalyserDOCD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerDOCD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerDOCD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass Spectrometer	Nascent sea spray composition via	D	Chamber experiments				
DMSCmesoCIMS (Chemical Ionization Mass Spectrometry)CO2 and methaneCPicarro CRDSHalocarbons, Iodine & halogenCPicarro CRDSvidesCu-Dirac ECD-GC and Multi-Axis Differential Optical Absorption; Spectroscopy (Max-DOAS)VOCs (Acetone, DMS, Acetonitrile, Methanol, Methanethiol, Isoprene, Monoterpenes, Acetaldehyde)CProton Transfer Reaction Mass Spectrometer (PTR- Most Spectroscopy (Max-DOAS)VOCs C ₅ to C ₁₅ DDPre-concentration and TD-GC-FID/MSAldehydes, ketonosyls, C ₂ to C ₈ DDerivatisation and HPLCWP2 PhysicsDDerivatisation and HPLCDMS fluxCmesoCIMS (Chemical Ionization Mass Spectrometry)CO2 EC fluxCLicors, Sonic anemometer motion sensorsDMS gradient fluxDCatamaran, SCD-GCNear-surface taradSDCTDNear-surface turbulenceCVector, FastCatSea stateCNOAA Wavewatch IIIWhitecap coverageDCameraMeteorological conditionsCAWSBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);WP3 Ocean biogeochemistryCIRGAPCDNDOCDWPOCD,WSpectrophotometerDOCD,WSpectrophotometerDOCD,WSpectrophotometerDOCD,WSpectrophotometerDOCD,WSpectrophotometerDOCD,WSpectropho	bubble burst of sea-water samples						
CO2 and methane C Picarro CRDS Halocarbons, Iodine & halogen oxides μ-Dirac ECD-GC and Multi-Axis Differential Optical Absorption; Spectroscopy (Max-DOAS) VOCs (Acetone, DMS, Acetonitrile, Methanol, Methanethiol, Isoprene, Monoterprenes, Acetaldehyde) C Proton Transfer Reaction Mass Spectrometer (PTR-Methanol, Methanethiol, Isoprene, Most) Works C 1s D Derivatisation and TD-GC-FID/MS Aldehydes, ketones (incl. D Derivatisation and HPLC dicarbonyls, C ₂ to C ₃ C mesoCIMS (Chemical Ionization Mass Spectrometry) CO2 EC flux C Licors, Sonic anemometer motion sensors DMS gradient flux D Catamaran, SCD-GC Near surface tandS D CTD Near-surface turbulence C Vector, FastCat Sea state C NOAA Wavewatch III Whitecap coverage D Camera Metoorological conditions C AWS Bulk fluxes C Ecotriplet PCO2 C IRGA PH C, D, W Spectrophotometer PCO2 D Radiosonde Water-surface turbulence C IRGA	DMS	С	mesoCIMS (Chemical Ionization Mass Spectrometry)				
Halocarbons, Iodine & halogen oxidesC µ-Dirac ECD-GC and Multi-Axis Differential Optical Absorption; Spectroscopy (Max-DOAS)VOCs (Acetone, DMS, Acetonitrile, Methanol, Methanethiol, Isoprene, Monoterpenes, Acetaldehyde)C Proton Transfer Reaction Mass Spectrometer (PTR- MS)WoCs C ₅ to C ₁₅ DPre-concentration and TD-GC-FID/MS Aldehydes, ketones (incl. DD Per-concentration and HPLCdicarbonyls), C ₂ to C ₃ CmesoCIMS (Chemical Ionization Mass Spectrometry) CO2 EC fluxCMS fluxCLicors, Sonic anemometer motion sensorsDMS gradient fluxDCatamaran, SCD-GCNear-surface stratificationCSpar buoy - temperature array, microcatsNear-surface turbulenceCVector, FastCatSea stateCNOAA Wavewatch IIIWhitecap coverageDCameraMeteorological conditionsCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MBL Height and stabilityDRadiosondeWP3 Ocan biogeochemistryC, D, WEcotripletCO2CIRGApHC, D, WSpectrophotometerDICDNutrientsDQCD, WSpectrophotometerDOCD, WSpectrophotometerDOCD, WSpectrophotometerDOCD, WSpectrophotometerDICDMass Spectrometry)Dissolved DMSD, WDissolved DMSD, WDissolved DMSD, WDissolved DMSD, W	CO2 and methane	С	Picarro CRDS				
oxides Absorption; Spectroscopy (Max-DOAS) VOCs (Acetone, DMS, Acetonitrile, Methanol, Methanethiol, Isoprene, Monoterpenes, Acetaldehyde) Absorption; Spectroscopy (Max-DOAS) VOCs (Acetone, DMS, Acetonitrile, Methanol, Methanethiol, Isoprene, Monoterpenes, Acetaldehyde) D VOCs Cs to C15 D Aldehydes, ketones (incl. dicarbonyls), C2 to C3 D WP2 Physics D DMS gradient flux D Col C2 E flux C C2 E flux C Near-surface stratification C Near-surface stratification C Casa state C Witeroological conditions C Bulk fluxes C Chlorophyll-a C, D, W Bulk fluxes C Chlorophyll-a C, D, W Bulk fluxes C, D, W Spectrophotometer DIC D Nattrients D, W COCO D, W Chorophyll-a C, D, W Spectrophotometer POC D, W CDOM D, W Spectrophotometer POC/2001/PC/PN/13C/15N D Mass Spectrometer DCC D, W Spectrophotometer POC/PON/PC/PN/13C/15N <	Halocarbons, Iodine & halogen	С	u-Dirac ECD-GC and Multi-Axis Differential On				
VOCs (Acetone, DMS, Acetonitrile, Monoterpenes, Acetaldehyde)CProton Transfer Reaction Mass Spectrometer (PTR- MS)Monoterpenes, Acetaldehyde)VOCs C, to C15DPre-concentration and TD-GC-FID/MSAldehydes, ketones (incl. dicarbonyls), C2 to C8DPre-concentration and HPLCMV2 PhysicsDDerivatisation and HPLCDMS fluxCmesoCIMS (Chemical Ionization Mass Spectrometry)CO2 EC fluxDCLow arface TandSDCTDNear-surface stratificationCSpar buoy – temperature array, microcatsNear-surface turbulenceCVector, FastCatSea stateCNOAA Wavewatch IIIWhitecap coverageDCameraBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MEI Height and stabilityDRadiosondeWP3 Ocean biogeochemistryC, D, WEcotripletCO2CIRGApC02D, WColorimetric AutoanalyserpC02D, WSpectrophotometerDICDNDOCD, WSpectrophotometerDICD, WSpectrophotometerDICD, WSpectrophotometerDICD, WSpectrophotometerDICD, WSpectrophotometerDICD, WSpectrophotometerDICD, WSpectrophotometerDICD, WSpectrophotometerDICD, WSpectrophotometerDICD, WSpectr	oxides	-	Absorption: Spectroscopy (Max-DOAS)				
Notion (1)ConstraintsConstraintsMSMethanol, Methanethiol, Isopreno, Actaldehyde)Notar Transfer Teatrich This Septement (1)VOCs C ₅ to C ₁₅ DPre-concentration and TD-GC-FID/MSAldehydes, ketones (incl.DDirivatisation and HPLCdicarbonyls), C ₂ to C ₈ WP2 PhysicsDMS fluxCCO2 EC fluxCLicors, Sonic anemometer motion sensorsDMS gradient fluxDCatamaran, SCD-GCNear-surface stratificationCNear-surface stratificationCNear-surface stratificationCNear-surface stratificationCNear-surface turbulenceCVector, FastCatSea stateCMeteorological conditionsCAWSBulk fluxesCWP3 Ocean biogeochemistryChlorophyll-aC, D, WpC02CRGApHC, D, WSpectrophotometerDICDNutrientsD, WOCCD, WDOCD, WDOCD, WDOCD, WDOCD, WDOCD, WDOCD, WDissolved DMSC,D, WSpectrophotometerDICDDMSD, WDissolved DMSD, WDMS PercessesD, WDMSP & processesD, WDMSP & processesD, WDMSP & processesD	VOCs (Acetone DMS Acetonitrile	С	Proton Transfer Reaction Mass Spectrometer (PTR-				
Inclumot, IntermediatelydeIntermediatelydeWonoterprenes, AcetaldehydeDVOCs C ₅ to C ₁₅ DAldehydes, ketones (incl.DDerivatisation and TD-GC-FID/MSAldehydes, ketones (incl.DDerivatisation and HPLCW2 PhysicsDMS gradient fluxCmesoCIMS (Chemical Ionization Mass Spectrometry)CO2 EC fluxCLicors, Sonic anemometer motion sensorsDMS gradient fluxDNear-surface taradifCNear-surface turbulenceCVector, FastCatSea stateCMeteorological conditionsCBulk fluxesCWP3 Ocean biogeochemistryChlorophyll-aC, D, WPC02CRea-surface fluxDC2CRea-surface fluxDRadiosondeWP3 Ocean biogeochemistryChlorophyll-aDCDDICDNutrientsD, WCDCD, WCDOCD, WCDOCD, WDOCD, WDOCD, WDissolved DMSC, miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WDissolved DMSD, WDMS processesD, WPigmentsDHD, WOC/PONPC/PN/13C/15NDDissolved DMSD, WDissolved DMSD, WDMSP & processesD, WPigmentsDHPLC <td>Methanol Methanethiol Isonrene</td> <td>C</td> <td>MS)</td>	Methanol Methanethiol Isonrene	C	MS)				
Monotriptics, Notabelight D Proconcentration and TD-GC-FID/MS Aldehydes, ketones (incl. D dicarbonyls), C ₂ to C ₈ WP2 Physics DMS flux C mesoCIMS (Chemical Ionization Mass Spectrometry) CO2 EC flux D Near surface TandS D Near surface stratification C Near-surface turbulence C VeCtor, FastCat Sea state C Whitecap coverage D Bulk fluxes C Bulk fluxes C WP3 Ocean biogeochemistry D Chorophyll-a C, D, W Back-scatter & β ₆₆₀ backscatter C pCQ C IRGA pH C, D, W Spectrophotometer DC D P OCC D, W TCO PH C, D, W Spectrophotometer DC D Radiosonde PH C, D, W Spectrophotometer DIC D N Nutrients D, W Colorimetric Autoan	Monoterpanes Acetaldebyde)		WIS)				
WOS CS US CJS D Preconcentration and TD-OC-FID/MIS dicarbonyls), C2 to C8 D Derivatisation and HPLC W2 Physics D Derivatisation and HPLC DMS flux C mesoCIMS (Chemical Ionization Mass Spectrometry) CO2 EC flux C Licors, Sonic anemometer motion sensors DMS gradient flux D Catamaran, SCD-GC Near-surface TandS D CTD Near-surface turbulence C Vector, FastCat Sea state C NOAA Wavewatch III Whitecap coverage D Camera Meteroological conditions C Eppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP); MBL Height and stability D Radiosonde WP3 Ocean biogeochemistry C Ecotriplet PGO2 C IRGA pH C, D, W Spectrophotometer DC D Natisets D/C D Mass Spectrometer PGO2 C IRGA pH C, D, W Spectrophotometer DOC D Mass Spectrometer	VOCa C, to C	D	Pro-concentration and TD CC EID/MS				
Andersydes, Ketones (nct. D Derivatisation and HPLC W22 Physics mesoCIMS (Chemical Ionization Mass Spectrometry) CO2 EC flux C Licors, Sonic anemometer motion sensors DMS gradient flux D Catamaran, SCD-GC Near-surface TandS D CTD Near-surface stratification C Spar buoy - temperature array, microcats Near-surface turbulence C NOAA Wavewatch III Whitecap coverage D Camera Meteorological conditions C AWS Bulk fluxes C Expley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP); MBL Height and stability D Radiosonde WP3 Ocean biogeochemistry C IRGA Chlorophyll-a C, D, W Ecotriplet Back-scatter & β ₆₆₀ backscatter C IRGA PH C, D, W Spectrophotometer DOC D, W HTCO DOM D, W Spectrophotometer POC/PON/PC/PN/13C/15N D Mass Spectrometer Fatty Acids and Alkanes D, W SCD DMSS de processes	$\begin{array}{c} VOCS C_5 \text{ to} C_{15} \\ Aldebase \\ Aldebase \\ betamas \\ \mathsf$	D	Devices tiestics and UDLC				
Intertorphylic, C 2 to C gWP2 PhysicsDMS fluxCmesoCIMS (Chemical Ionization Mass Spectrometry)CO2 EC fluxDCatamaran, SCD-GCNear surface TandSDCTDNear-surface stratificationCSpar buoy – temperature array, microcatsNear-surface turbulenceCVector, FastCatSea stateCNOAA Wavewatch IIIWhitecap coverageDCameraMeteorological conditionsCAWSBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MBL Height and stabilityDRadiosondeWP3 Ocean biogeochemistryCIRGAChlorophyll-aC, D, WEcotripletBack-scatter & β_{660} backscatterCIRGApHC, D, WSpectrophotometerDICDNutrientsDOCD, WMass SpectrometerDOCD, WMass SpectrometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WSCD/FPDDMSP & processesD, WSCD/FPDDMSP & processesD, WSCDPigmentsDHPLCMicrotopial ktorn identification/countsD, WOtical MicroscopyMicrotopial hteroscopy	Aldenydes, ketones (incl.	D	Derivatisation and HPLC				
WP2 PhysicsDMS fluxCmesoCIMS (Chemical Ionization Mass Spectrometry)CO2 EC fluxCLicors, Sonic anemometer motion sensorsDMS gradient fluxDCatamaran, SCD-GCNear-surface TandSDCTDNear-surface turbulenceCVector, FastCatSea stateCNOAA Wavewatch IIIWhitecap coverageDCameraMeteorological conditionsCAWSBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MBL Height and stabilityDRadiosondeWP3 Ocean biogeochemistryCEcotripletChlorophyll-aC, D, WEcotripletBack-scatter & β_{660} backscatterCEcotripletpCO2CIRGApHC, D, WSpectrophotometerDICDNutrientsDOCD, WSpectrophotometerDOCD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WSCD/FPDDMSP & processesD, WSCD/FPDDMSP & processesD, WSCD/FPDDMSP & processesD, WFlow CytometryPigmentsDHPLCMicrotopilankton identification/countsD, WOptical MicroscopyMicrotopilankton identification/countsD, WOptical Microscopy	dicarbonyis), C_2 to C_8						
DMS fluxCmesoCIMS (Chemical lonization Mass Spectrometry)CO2 EC fluxCLicors, Sonic anemometer motion sensorsDMS gradient fluxDCatamaran, SCD-GCNear surface TandSDCTDNear-surface stratificationCSpar buoy – temperature array, microcatsNear-surface turbulenceCVector, FastCatSea stateCNOAA Wavewatch IIIWhitecap coverageDCameraMeteorological conditionsCAWSBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MBL Height and stabilityDRadiosondeWP3 Ocean biogeochemistryCIRGAChlorophyll-aC, D, WEcotripletBack-scatter & β ₆₆₀ backscatterCIRGADICDNutrientsDICDMass SpectrometerDICDMass SpectrometerDICDMass SpectrometerDOCD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerPatty Acids and AlkanesD, WSCD/FPDDissolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSDHPLCMicrobal community abundanceD, WPigmentsDHPLCMicrobal community abundanceD, WPhytoplankton identification/countsD, WOrderial MicroscopyMicroscopy	WP2 Physics	a					
CO2 EC fluxCLicors, Sonic anemometer motion sensorsDMS gradient fluxDCatamaran, SCD-GCNear surface TandSDCTDNear-surface stratificationCSpar buoy – temperature array, microcatsNear-surface turbulenceCVector, FastCatSea stateCNOAA Wavewatch IIIWhitecap coverageDCameraMeteorological conditionsCAWSBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MBL Height and stabilityDRadiosonde WP3 Ocean biogeochemistry CEcotripletChlorophyll-aC, D, WEcotripletBack-scatter β_{660} backscatterCEcotripletpCO2CIRGApHC, D, WSpectrophotometerDICDNutrientsDOCD, WHTCOCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WSCD/FPDDMSP & processesD, WSCDPigmentsDHPLCMicrozooplankton identification/countsD, WPhytoplankton identification/countsD, WOverlankton identification/countsD, WOverlankton identification/countsD, WOverlankton identification/countsD, WOverlankton identification/countsD, WOverlankton identification/countsD, WOverlankton identification/countsD, W <td>DMS flux</td> <td>С</td> <td>mesoCIMS (Chemical Ionization Mass Spectrometry)</td>	DMS flux	С	mesoCIMS (Chemical Ionization Mass Spectrometry)				
DMS gradient fluxDCatamaran, SCD-GCNear surface TandSDCTDNear-surface stratificationCSpar buoy – temperature array, microcatsNear-surface turbulenceCVector, FastCatSea stateCNOAA Wavewatch IIIWhitecap coverageDCameraMeteorological conditionsCAWSBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MBL Height and stabilityDRadiosondeWP3 Ocean biogeochemistryCIRGAChlorophyll-aC, D, WEcotripletBack-scatter & β_{660} backscatterCIRGApCO2CIRGApHC, D, WSpectrophotometerDICDNutrientsDOCD, WTCOCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerPossolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WSCD/FPDDMSP & processesD, WSCDPigmentsDHPLCMicrozoonlanktonD, WFlow CytometryPhytoplankton identification/countsD, WOptical Microscopy	CO2 EC flux	С	Licors, Sonic anemometer motion sensors				
Near surface TandSDCTDNear-surface stratificationCSpar buoy – temperature array, microcatsNear-surface stratificationCSpar buoy – temperature array, microcatsNear-surface stratificationCVector, FastCatSea stateCNOAA Wavewatch IIIWhitecap coverageDCameraMeteorological conditionsCAWSBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MBL Height and stabilityDRadiosondeWP3 Ocean biogeochemistryCEcotripletChlorophyll-aC, D, WEcotripletBack-scatter & β_{660} backscatterCIRGApHC, D, WSpectrophotometerDICDDNutrientsD, WColorimetric AutoanalyserDOCD, WHTCOCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerPosloved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WSCD/FPDDMSP & processesD, WSCDPigmentsDHPLCMicrozonlanktonD, WFlow CytometryPhytoplankton identification/countsD, WOptical Microscopy	DMS gradient flux	D	Catamaran, SCD-GC				
Near-surface stratificationCSpar buoy – temperature array, microcatsNear-surface turbulenceCVector, FastCatSea stateCNOAA Wavewatch IIIWhitecap coverageDCameraMeteorological conditionsCAWSBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MBL Height and stabilityDRadiosonde WP3 Ocean biogeochemistry CEcotripletChlorophyll-aC, D, WEcotripletBack-scatter & β_{660} backscatterCIRGApHC, D, WSpectrophotometerDICDDNutrientsD, WColorimetric AutoanalyserDOCD, WHTCOCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WScD/FPDDMSP & processesD, WSCD/FPDDMSP & processesD, WSCDPigmentsDHPLCMicrobial community abundanceD, WFlow CytometryPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonD, WOptical Microscopy	Near surface TandS	D	CTD				
Near-surface turbulenceCVector, FastCatSea stateCNOAA Wavewatch IIIWhitecap coverageDCameraMeteorological conditionsCAWSBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MBL Height and stabilityDRadiosondeWP3 Ocean biogeochemistryCEcotripletChlorophyll-aC, D, WEcotripletBack-scatter & β ₆₆₀ backscatterCEcotripletpCO2CIRGApHC, D, WSpectrophotometerDICDDNutrientsD, WColorimetric AutoanalyserDOCD, WHTCOCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerPotsolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WSCD/FPDDMSP & processesD, WSCDPigmentsDHPLCMicrozonlankton identification/countsD, WOptical MicroscopyMicroscopyMicrozonlanktonD, WOptical MicroscopyMicroscopy	Near-surface stratification	С	Spar buoy – temperature array, microcats				
Sea stateCNOAA Wavewatch IIIWhitecap coverageDCameraMeteorological conditionsCAWSBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MBL Height and stabilityDRadiosondeWP3 Ocean biogeochemistryCEcotripletChlorophyll-aC, D, WEcotripletBack-scatter & β660 backscatterCEcotripletpCO2CIRGApHC, D, WSpectrophotometerDICDNutrientsDoCD, WColorimetric AutoanalyserDOCD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerPosolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WSCD/FPDDMSP & processesD, WSCDPigmentsDHPLCMicrobial community abundanceD, WOptical MicroscopyPhytoplankton identification/countsD, WOptical Microscopy	Near-surface turbulence	С	Vector, FastCat				
Whitecap coverageDCameraMeteorological conditionsCAWSBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MBL Height and stabilityDRadiosondeWP3 Ocean biogeochemistryCEcotripletChlorophyll-aC, D, WEcotripletBack-scatter & β ₆₆₀ backscatterCEcotripletpCO2CIRGApHC, D, WSpectrophotometerDICDNutrientsDOCD, WColorimetric AutoanalyserDOCD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WScD/FPDDMSP & processesD, WSCD/FPDDMSP & processesDHPLCMicrobial community abundanceD, WFlow CytometryPhytoplankton identification/countsD, WOntical MicroscopyMicrozoonlanktonD, WOntical Microscopy	Sea state	С	NOAA Wavewatch III				
Meteorological conditionsCAWSBulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MBL Height and stabilityDRadiosonde WP3 Ocean biogeochemistry C, D, WEcotripletChlorophyll-aC, D, WEcotripletBack-scatter & β_{660} backscatterCEcotripletpCO2CIRGApHC, D, WSpectrophotometerDICDNutrientsDOCD, WColorimetric AutoanalyserDOCD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WSCD/FPDDMSP & processesD, WSCDPigmentsDHPLCMSP & processesD, WSCDPigmentsDHPLCMicrozoonlanktonD, WOptical MicroscopyMicrozoonlanktonD, WOptical Microscopy	Whitecap coverage	D	Camera				
Bulk fluxesCEppley radiometers, raingauge; Eppley Precision Spectral Pyranometer (PSP);MBL Height and stabilityDRadiosondeWP3 Ocean biogeochemistryDRadiosondeChlorophyll-aC, D, WEcotripletBack-scatter & β_{660} backscatterCEcotripletpCO2CIRGApHC, D, WSpectrophotometerDICDNutrientsD, WColorimetric AutoanalyserDOCD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WScD/FPDDissolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WSCD/FPDMSP & processesD, WSCDPigmentsDHPLCMicrobial community abundanceD, WFlow CytometryPhytoplanktonD, WOntical MicroscopyMicrozoonlanktonD, WOntical Microscopy	Meteorological conditions	С	AWS				
MBL Height and stability D Radiosonde WP3 Ocean biogeochemistry Chlorophyll-a C, D, W Ecotriplet Back-scatter & β ₆₆₀ backscatter C Ecotriplet pCO2 C IRGA pH C, D, W Spectrophotometer DIC D Nutrients D, W Colorimetric Autoanalyser DOC D, W Spectrophotometer D DOC D, W Spectrophotometer D OOC D, W Spectrophotometer D DOC D, W Spectrophotometer D OOC D, W Spectrophotometer D POC/PON/PC/PN/13C/15N D Mass Spectrometer Fatty Acids and Alkanes D, W SCD/FPD Dissolved DMS D, W SCD/FPD DMSP & processes D, W SCD Pigments D HPLC Microbial community abundance D, W Flow Cytometry Phytoplankton identification/counts D, W Optical Microscopy Microzoonlankton D W Optical Microscopy <td>Bulk fluxes</td> <td>С</td> <td>Eppley radiometers, raingauge; Eppley Precision</td>	Bulk fluxes	С	Eppley radiometers, raingauge; Eppley Precision				
MBL Height and stabilityDRadiosondeWP3 Ocean biogeochemistryC, D, WEcotripletChlorophyll-aC, D, WEcotripletBack-scatter & β ₆₆₀ backscatterCEcotripletpCO2CIRGApHC, D, WSpectrophotometerDICDNutrientsD, WColorimetric AutoanalyserDOCD, WHTCOCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WDissolved DMSC, miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WPigmentsDHPLCHPLCMicrobial community abundanceD, WPhytoplankton identification/countsD, WOutcal MicroscopyD			Spectral Pyranometer (PSP);				
MDL Height and stabilityDRadioSolideWP3 Ocean biogeochemistryC, D, WEcotripletChlorophyll-aC, D, WEcotripletBack-scatter & β_{660} backscatterCEcotripletpCO2CIRGApHC, D, WSpectrophotometerDICDNutrientsD, WColorimetric AutoanalyserDOCD, WHTCOCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WDissolved DMSC, miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WScold PigmentsDHPLCMicrobial community abundanceD, WPhytoplankton identification/countsD, WOptical MicroscopyDMicroscoplanktonD, WOntical Microscopy	MBI Height and stability	D	Radiosonde				
W13 Occan biogeothemistryChlorophyll-aC, D, WEcotripletBack-scatter & β_{660} backscatterCEcotripletpCO2CIRGApHC, D, WSpectrophotometerDICDNutrientsD, WColorimetric AutoanalyserDOCD, WHTCOCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WDissolved DMSC, miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WScolved DMSD, WPigmentsDHPLCMicrobial community abundanceD, WPhytoplankton identification/countsD, WOwOptical MicroscopyMicrozooplanktonD, WOwOptical Microscopy	WD2 Ocean biogeochemistry	D	Radiosolide				
ChorophysicalC, D, WEconipletBack-scatter & β_{660} backscatterCEcotripletpCO2CIRGApHC, D, WSpectrophotometerDICDNutrientsD, WColorimetric AutoanalyserDOCD, WHTCOCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WDissolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WSCD/FPDDMSP & processesD, WSCDPigmentsDHPLCMicrobial community abundanceD, WFlow CytometryPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonD, WOptical Microscopy	Chlorophyll a	CDW	Factriplat				
Back-scatterCEcompletpCO2CIRGApHC, D, WSpectrophotometerDICDNutrientsD, WColorimetric AutoanalyserDOCD, WHTCOCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WDissolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WSCD/FPDDMSP & processesD, WSCDPigmentsDHPLCMicrobial community abundanceD, WFlow CytometryPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonD, WOptical Microscopy	Children a	C, D, W	Ecomplet				
pCO2CIKGApHC, D, WSpectrophotometerDICDNutrientsD, WColorimetric AutoanalyserDOCD, WHTCOCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WDissolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WScolved DMSD, WScolved DMSD, WScolved DMSD, WScolved DMSD, WScolved DMSD, WPigmentsDHPLCMicrobial community abundanceD, WPhytoplankton identification/countsD, WOntical MicroscopyMicrozoonlanktonDWOntical Microscopy	nCO2	C C					
pHC, D, WSpectrophotometerDICDNutrientsD, WDOCD, WCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerPOC/PON/PC/PN/13C/15NDMass SpectrometerPotophysicFatty Acids and AlkanesD, WDissolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WSCD/FPDDMSP & processesD, WPigmentsDHPLCMicrobial community abundanceD, WPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonDWOptical Microscopy	pCO2		IRGA				
DICDNutrientsD, WColorimetric AutoanalyserDOCD, WHTCOCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WDissolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WScD/FPDDMSP & processesD, WPigmentsDHPLCMicrobial community abundanceD, WPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonDWOntical Microscopy	рн	C, D, W	Spectrophotometer				
NutrientsD, WColorimetric AutoanalyserDOCD, WHTCOCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WDissolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WScD/FPDDMSP & processesD, WPigmentsDHPLCMicrobial community abundanceD, WPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonDWOptical Microscopy	DIC	D					
DOCD, WHTCOCDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WDissolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WScD/FPDDMSP & processesD, WPigmentsDHPLCMicrobial community abundanceD, WPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonDWOptical Microscopy	Nutrients	D, W	Colorimetric Autoanalyser				
CDOMD, WSpectrophotometerPOC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WDissolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WSCD/FPDDMSP & processesD, WSCDPigmentsDHPLCMicrobial community abundanceD, WFlow CytometryPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonDW	DOC	D, W	НТСО				
POC/PON/PC/PN/13C/15NDMass SpectrometerFatty Acids and AlkanesD, WDissolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WSCD/FPDDMSP & processesD, WPigmentsDHPLCMicrobial community abundanceD, WPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonDWOptical Microscopy	CDOM	D, W	Spectrophotometer				
Fatty Acids and AlkanesD, WDissolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WSCD/FPDDMSP & processesD, WSCDPigmentsDHPLCMicrobial community abundanceD, WFlow CytometryPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonD, WOptical Microscopy	POC/PON/PC/PN/13C/15N	D	Mass Spectrometer				
Dissolved DMSC,miniCIMS (Chemical Ionization Mass Spectrometry)Dissolved DMSD, WSCD/FPDDMSP & processesD, WSCDPigmentsDHPLCMicrobial community abundanceD, WFlow CytometryPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonD, WOptical Microscopy	Fatty Acids and Alkanes	D, W					
Dissolved DMSD, WSCD/FPDDMSP & processesD, WSCDPigmentsDHPLCMicrobial community abundanceD, WFlow CytometryPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonD, WOptical Microscopy	Dissolved DMS	С,	miniCIMS (Chemical Ionization Mass Spectrometry)				
DMSP & processesD, WSCDPigmentsDHPLCMicrobial community abundanceD, WFlow CytometryPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonD, WOptical Microscopy	Dissolved DMS	D, W	SCD/FPD				
PigmentsDHPLCMicrobial community abundanceD, WFlow CytometryPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonD, WOptical Microscopy	DMSP & processes	D, W	SCD				
Microbial community abundanceD, WFlow CytometryPhytoplankton identification/countsD, WOptical MicroscopyMicrozoonlanktonD, WOptical Microscopy	Pigments	D	HPLC				
Phytoplankton identification/counts D, W Optical Microscopy Microzoonlankton D W Optical Microscopy	Microbial community abundance	D, W	Flow Cytometry				
Microzoonlankton D W Ontical Microscopy	Phytoplankton identification/counts	D. W	Optical Microscopy				
	Microzooplankton	D, W	Optical Microscopy				

Table 2. Parameters sampled during the SOAP voyage. Key: C - Continuous, D - Discrete, W – Workboat, *indicates instrument sampling on common aerosol inlet.

				Mean DMS	Std				
Voyage	Date	Latitude	Longitude	(nmol L ⁻¹)	Dev	n	Method	Reference	
BOX	October 2000	39.5-47°S	170-179°E	4.55	6.8	482	FPD-GC	this paper	
	November 2005	49-50°S	175°E	1.75	-	2	FPD-GC	Kiene et al., 2007	
SAGE	April 2006	41-46.6°S	172.5-78.5E	1.06	0.9	6	PFPD	Archer et al., 2011	
PreSOAP	February 2011	42.5-44°S	174E-178°W	2.2	2.0	736	MIMS	this paper	
SOAP	March-April 2012	41.7-46.5°S	172E-179°W	6.36	4.4	4132	miniCIMS	Bell et al., 2015	
SOAP	March-April 2012	41.7-46.5°S	172E-179°W	11.5	9.2	22	SCD	Walker et al. 2016	
CW									
S.W. Decific	Weighted Meen	20 5 5005	170 170°W	56		5280		this paper	
Facilic	weighted Mean	39.3-30 S	1/0-1/9 W	5.0		5560		uns paper	
NEWZ		35-55S	170E-170W	0.05-2.0		6	Climatol.	Lana et al., 2011	
SSTC		35-50S	170W-170E	0.05-10		n/a	Climatol.	Lana et al., 2011	

Table 3. DMS data for the S.W. Pacific region east of New Zealand. Std Dev = standard deviation, n = number of measurements, FPD-GC Flame Photometric Detector-Gas Chromatograph, PFPD Pulsed Flame Photometric Detector, MIMS Membrane Inlet Mass Spectrometer, miniCIMS (Atmospheric Pressure Chemical Ionization Mass Spectrometer), SCD Sulfur Chemiluminescent Detector, Climatol. Climatology.











DoY 47

DoY 49

DoY 65









