Interactive comment on "Heterogeneous uptake of amines onto kaolinite in the temperature range of 232-300 K" by Y. Liu et al.

Y. Liu et al.

Referee #1

The authors report uptake coefficients of three amines (methylamine MA, dimethylamine DMA, trimethylamine TMA) on a mineral dust model substance (clay mineral kaolinite) using a Knudsen reactor in the temperature range 232-300 K. Using a simple thermodynamic model for non-reactive systems (P. Davidovits) they go on to interpret the negative T-dependence in terms of adsorption enthalpies and entropies. The paper reads well, however, the authors do not direct their attention to conveying a take-home lesson from their experiments because there are too many loose ends that are not explained or interpreted correctly. Almost all experiments leave wanting to such an extent that the level and quality of effort is not commensurate with the expectations of the readers of acp. The problem is that both kinetics and spectroscopy parts are "light", thus incomplete and misleading. Some of the conclusions are not supported by experimental fact and are left hanging. An example is the assertion that the amines interact with Lewis acid sites on the kaolinite despite the fact that the authors insist on identifying protonated amines in the DRIFTS spectra which would correspond to Bronsted acidic sites on kaolinite, unless I misunderstand. However, only DMA shows protonated base (Figure 2), no evidence is presented for TMA and MA. This is just an example for the severe flaws and shortcomings of this report that prevents the reader from understanding the initial question posed and its eventual resolution. I conclude that his paper is nowhere near to being publishable and I therefore list the most important questions and concerns for the benefit of the authors that need to be answered to the satisfaction of the editor before publication may be envisaged.

Response: Thank you for your comments and suggestions. We reply to your particular concerns about the IR results here. Other questions will be answered point-by-point below.

We stated that "In this work, two bands at 1477-1467 cm⁻¹ and 971 cm⁻¹ were observed, as shown in Fig. 2, and were probably related to the protonated amines (MAH⁺, DMAH⁺ and TMAH⁺). In particular, a strong peak at 1473 cm⁻¹ was observed (Fig. 2B), while the band was very weak for the spectra in Fig. 2A and C. However, this band was very close to the δ (NH) band at ~1470 cm⁻¹ in DMA (NIST database) and/or $\delta(CH_3)$ (Lin et al., 2014) in amines. On the other hand, it should be noted that formation of protonated amines requires surface OH (Brønsted acid). However, as shown in Fig. 2, the surface hydroxyl (-OH) in the range of 3600-3750 cm⁻¹ (Miranda-Trevino and Coles, 2003) was not consumed when the kaolinite was exposed to MA, DMA or TMA. This indicates that the content of reactive Brønsted acid sites on the kaolinite must be very low even if the bands at 1477-1467 cm⁻¹ originated from the protonated amines. Therefore, Lewis acid sites on the kaolinite predominantly contributed to the uptake of amines as observed in Figs. 1 and 2 in this study. This was similar to the adsorption of DMA and N,N-dimethyl formamide (DMF) on TiO₂, for which Lewis acid sites were identified as the reactive sites (Lin et al., 2014)." in lines 210-227.

In Fig. 2, although two IR bands (1473 and 971 cm⁻¹) were close to the characterized bands of protonated TMA ((CH₃)₃NH⁺) at 1484 and 984 cm⁻¹, the band at 1484 cm⁻¹ was also close to the δ (NH) band of Lewis-bound DMA and/or the δ (CH₃) band at ~1470 cm⁻¹ in DMA. In addition, the consumption of OH, which was required to form protonated DMA, was not observed during adsorption of DMA. Therefore, the contribution of protonated amines to the band at 1473 cm⁻¹ was ruled out.

To make our statements more clear, we will revise our manuscript about the IR results as "For MA in Fig. 2A, the band at 1606 cm⁻¹ (-NH₂ deformation band) is the characteristic band for Lewis-bound MA (Auerbach et al., 2003;Nunes et al., 2005). In Fig. 2B, the band at ~1470 cm⁻¹ might have originated from the δ (NH) in Lewis-bound DMA (NIST database) and/or δ (CH₃) (Lin et al., 2014). In a previous work, the IR

bands at 1484 and 984 cm⁻¹ were assigned to the characteristic bands of protonated TMA ((CH₃)₃NH⁺) in an acidic solution containing TMA and on the surface of polyethylene (PE) treated by TMA (Ongwandee et al., 2007). In this work, although two bands at 1477-1467 cm⁻¹ and 971 cm⁻¹ were observed in the spectrum shown in Fig. 2B, and were close to the bands of protonated TMA (TMAH⁺), it was more reasonable to assign them to Lewis-bound DMA for the following reasons. Formation of protonated amines requires the partition of surface OH (Brønsted acid). However, as shown in Fig. 2B, the surface hydroxyl (-OH) in the range of 3600-3750 cm⁻¹ (Miranda-Trevino and Coles, 2003) was not consumed when the kaolinite was exposed to DMA. In particular, the bands at 1477-1467 and 971 cm⁻¹ were not observed for MA and TMA in Fig. 2A and C. At the same time, OH consumption was also unobservable for MA and TMA. Therefore, it can be concluded that Lewis acid sites on the kaolinite predominantly contributed to the uptake of MA, DMA and TMA as observed in Figs. 1 and 2 in this study. This was similar to the adsorption of DMA and N,N-dimethyl formamide (DMF) on TiO₂, for which Lewis acid sites were identified as the reactive sites (Lin et al., 2014)".

I would like to begin with several technical questions that need to be answered such that the interested reader may see for himself rather than taking the author's word for it: trust is good, control is better! I have not found a diagram of the Knudsen reactor in any of the author's papers. I assume, that their machine does not include molecular beam modulation and recovery of the chopped signal using a lock-in amplifier. This is usually a fatal flaw in case one is dealing with "sticky" compounds like the present amines. I strongly suggest that the authors include a block diagram (as an Appendix) on the design of the instrument.

Response: Thank you so much for your good suggestions. Yes, our system does not include molecular beam modulation and a lock-in amplifier. The block diagram (Fig. R1) will be included in Fig. S1. In the experimental details, this will be pointed out as "The schematic diagram is shown in Fig. S1. The parameters of the reactor are summarized in Tables S1.".



Fig. R1. The schematic diagram of the Knudsen cell-mass spectrometer.

As you know, there are two types of Knudsen cell reactors. 1)With molecular beam modulation and a lock-in amplifier (Seisel et al., 2005;Seisel et al., 2004;Ullerstam et al., 2003;Salgado-Muñoz and Rossi, 2002;Fenter et al., 1994); 2)Without molecular beam modulation and lock-in amplifier (Hoffman et al., 2003;Underwood et al., 2001;Beichert and Finlayson-Pitts, 1996;Liu et al., 2008).

Molecular beam modulation can distinguish between signals in the mass spectrometer arising from background gases in the vacuum system and those produced by the desorbed species by modulating either the adsorbed or desorbed molecular beam and measuring the resulting time-dependent signal in the mass spectrometer. Under this circumstance, it requires a detector capable of recovering signals in the presence of an overwhelming noise background or, alternatively, providing high resolution measurements of relatively clean signals over several orders of magnitude and frequency. Thus, a lock-in amplifier is usually combined with molecular beam modulation. By combination with these techniques, a Knudsen cell reactor with molecular beam modulation and a lock-in amplifier can directly measure the surface lifetime of an adsorbed gas, the order of chemical reaction, and the accommodation coefficient and sticking coefficient of incident molecules (Compton et al., 1984). The uptake experiments using such an instrument can be operated in both a steady flow mode and a pulse mode. By contrast, only the net uptake coefficient can be measured, usually in a steady flow, with a Knudsen cell reactor without molecular beam modulation and a lock-in amplifier (Underwood et al., 2000). This means that a Knudsen cell reactor with molecular beam modulation and a lock-in amplifier should be more powerful than the counterpart without these features. However, the net uptake coefficient measured in this work and those reported in the literature with the second type of Knudsen cell are also meaningful in atmospheric chemistry.

The uptake coefficient (γ_{net}) is defined as,

$$\gamma = \frac{\frac{dn_g}{dt}}{\omega} \quad (R1)$$

where n_g is number of molecules in the gas phase (molecules), *t* is time (s), and ω is gas-surface collision frequency (s⁻¹). When the pressure in the reactor cell is kept low enough, the mean free path of the molecules within the cell is greater than the cell dimensions, so that boundary layer effects and homogeneous gas-phase collisions are minimized and can be neglected. Gas-surface collision frequency is given by

$$\omega = \frac{\overline{v}A_s}{4}\frac{n}{V} = \frac{vA_s}{4}\frac{4F}{A_hv} = \frac{FA_s}{A_h} \quad (R2)$$

where \bar{v} is the average molecular speed (m s⁻¹); A_s is the area that the flux of molecules impinges upon or the geometric area of the sample holder (m²); (*n*/*V*) is the number density of the reactant gas (molecules m⁻³); *F* is the flow of molecules out of the cell (molecules s⁻¹); A_h is the area of the exit aperture (m²).

In a steady-state flow, the number of the reactant molecules that is "lost" to the surface $(-dn_g/dt)$ is equal to the change in flow out of the cell, $(F_0 - F)$, where F_0 and F represent the gas-phase flow out of the cell with the sample covered and exposed, respectively (Underwood et al., 2000). Therefore,

$$\gamma = \frac{A_h}{A_s} \frac{F_0 - F}{F} \quad (R3)$$

Once the gas-phase flow out of the cell is proportional to the measured mass spectral intensity of the reactant gas (I),

$$\gamma = \frac{A_h}{A_s} \frac{I_0 - I}{I} \quad (R4)$$

Therefore, Eq. (R4) is correct and without any additional limitations if the uptake experiment is performed in a molecular flow (or the mean free path of the molecules within the cell is greater than the cell dimensions) and in a steady-state flow, and the gas flow out of the cell is proportional to the measured mass spectral intensity of the reactant gas (I).

In this work, the working pressure was $\sim 3.5 \times 10^{-4}$ Torr. The mean free path of gas was greater than 12 cm even at 232 K. This value was greater than the maximal dimensions of the cell (5.6 cm). By adjusting the variable leak valve, the mass signal was calibrated against the pressure in the cell, which was proportional to the flow rate (Table R1). As shown in Fig. R2, the signal intensity of TMA linearly responded to the flow rate or the pressure in the reactor. Similar results were also obtained for MA and DMA.

No.	Pressure in the	Signal intensity	$\sigma(m/z 50)$	Signal intensity	σ(m/z 58)	
	reactor (Torr)	(m/z 59)	O(III/Z 39)	(m/z 58)		
P1	1.00E-03	5.90E+04	1.16E+03	2.91E+04	7.76E+02	
P2	1.30E-04	8.27E+03	4.38E+02	4.02E+03	3.02E+02	
P3	2.50E-04	1.72E+04	5.97E+02	8.43E+03	4.28E+02	
P4	3.70E-04	2.38E+04	7.16E+02	1.17E+04	5.20E+02	
P5	7.40E-04	3.79E+04	1.09E+03	1.86E+04	6.83E+02	
P6	3.90E-04	2.35E+04	7.10E+02	1.15E+04	5.02E+02	
P7	3.10E-04	1.80E + 04	6.19E+02	8.83E+03	4.53E+02	
P8	1.80E-04	1.16E+04	5.45E+02	5.66E+03	3.73E+02	
P9	1.00E-04	6.30E+03	3.89E+02	3.02E+03	2.70E+02	
P10	6.00E-05	3.40E+03	3.03E+02	1.61E+03	1.96E+02	
P11	2.40E-05	9.21E+02	1.59E+02	3.73E+02	1.21E+02	

Table R1. Calibration of the MS signal of TMA to the flow rate.



Fig. R2. The relationship between the MS signal and the pressure in the reactor or the flow rate of TMA. "I" and "F" means the MS signal intensity and the flow rate of TMA.

We agree with you that amines are sticky molecules. We compared the escape rate of amines with that of N₂ (in the same experiment). The escape rate (k_{Esc}) was calculated by

$$\ln \frac{I - I'}{I_0 - I'} = -k_{Esc} t \quad (R5)$$

Where I_0 and I' is the signal intensity at the initial and the final steady state when the flow rate into the reactor was changed using the variable leak valve. As shown in Fig. R3, the measured escape rate of TMA was slightly smaller than that of N₂ at 300 K. However, if the effective area of the escape hole (A_h) was calculated based on amines but not N₂, the effect of the viscosity of amines on the measurement of uptake coefficient should be ruled out according to Eq. (R4) because the kinematic viscosity has been factored into the k_{Esc} or A_h calculation. The A_h was obtained by

$$A_{h} = \frac{4V}{\tau \overline{\nu}} = \frac{4Vk_{Esc}}{\overline{\nu}} \quad (R6)$$

Table R2 summarizes the measured effective A_h of these three amines after each uptake experiment was completed. The effective A_h of amines were significantly smaller than that of N₂. In particular, the mean A_h of MA was the smallest one among these three amines. This is in agreement with the larger coefficient of kinematic viscosity of MA (13.9 m² s⁻¹) compared with that of DMA (8.7 m² s⁻¹) and TMA (6.7 m² s⁻¹) calculated at 300 K and 3.5×10^{-4} Torr based on the Peng-Robinson model (Fan and Wang, 2006).



Fig. R3. Escape rate of TMA and N₂ measured at 300 K.

Exp. No.	MA	N_2	DMA	N_2	TMA	N_2
1	0.28	0.89	0.29	0.80	0.76	0.91
2	0.32	1.12	0.42	0.89	0.61	0.93
3	0.30	1.15	0.41	0.97	0.81	1.08
4	0.28	0.94	0.44	0.87	0.75	0.83
5	0.25	0.77	0.38	0.78	0.62	0.82
Average	0.28±0.03	0.97±0.16	0.39±0.06	0.86±0.07	0.71±0.09	0.92±0.10

Table R2. The area of escape hole (A_h) of amines measured at 300 K. Unit: mm².

In the original manuscript, we calculated the uptake coefficients based on the A_h of N₂. This would overestimate the uptake coefficients of amines. In the revised manuscript, all the uptake coefficients have been corrected based on the measured $A_{\rm h}$ of the amines at the corresponding temperature and pressure. The corrections will also be done in our previous work. In the experimental section, it will be discussed in the section 3.2 as "It should be pointed out that the escape rate of amines from the reactor should be lower than that of N₂ because amines are sticky molecules. This will lead to a smaller effective A_h of amines compared with N₂. Table S2 summarizes the summarized $A_{\rm h}$ for these amines at 300 K and 3.5×10⁻⁴ Torr. The effective $A_{\rm h}$ of amines were significantly smaller than that of N₂. At the same time, MA showed the smallest A_h among these three amines. This is in agreement with the larger coefficient of kinematic viscosity of MA (13.9 $m^2 s^{-1}$) compared with that of DMA (8.7 $m^2 s^{-1}$) and TMA (6.7 m² s⁻¹) calculated at 300 K and 3.5×10^{-4} Torr based on the Peng-Robinson model (Fan and Wang, 2006). Therefore, the effective A_h of amines but not N₂ measured at the corresponding temperature and pressure were used to calculate the uptake coefficients according to Eq. (1). The Ah at different temperatures are summarized in Tables S2-S3".

The authors do not seem to ever perform (or report the results of) reference (blank) experiments with the empty sample compartment exposed to the amines. Owing to the day-long (8 h) saturation of the internal vessel walls with the amines one cannot be sure about the interpretation of the signal lasting just 10 to 20 minutes during which "saturation" of the substrate takes place. This is definitely the wrong instrument to tackle the problem at hand.

Response: We have performed blank experiments for all of these amines. Fig. R4 shows the results (raw data) for the blank experiments. This experiment was carried out at 232 K. When the steady state of MA or TMA signals was reached, the Teflon sample holder (without particles) was exposed to MA or TMA vapor by opening the sample cover. As shown in Fig. R4, the signals of MA or TMA did not change. This means the contribution of the fresh surface (including the sample holder and the inner surface of

the sample cover) to the uptake of amines by kaolinite is negligible. The blank experiment results will be added in the supporting information as Fig. S2. In the manuscript, it will also be pointed out that "Blank experiments showed that the fresh surface (including the sample holder and the inner surface of the sample cover) did not contribute to the signal drops observed for amines when the sample cover was opened (Fig. S2)."



Fig. R4. Blank experiments (raw data) for (A) MA and (B) TMA at 232 K.

In the experimental section, we mentioned that the sample was out-gassed at 298 K in the Knudsen cell reactor for 8 h (lines 127-130, in the original manuscript) but not passivated with amines for 8 h. We also pointed out that the reactor chamber was passivated with amines while the sample was isolated from the reactant gas by the sample cover, until a steady state QMS signal was established (lines 133-135, in the original manuscript). The passivation time was usually 1 h. In the revised manuscript, this will be described as "The reactor chamber was passivated with amines while the sample cover, until a steady state QMS signal was established with amines while the sample was isolated from the reactant gas by the sample was isolated from the reactant gas by the sample was isolated from the reactant gas by the sample cover, until a steady state QMS signal was established (~ 60 min)".

I am missing a Table with all salient parameters of the Knudsen reactor such as gas wall collision frequency, volume and surface of the flow reactor, sample surface, used escape rate constant, flow rate calibrations, MS sensitivities, etc.

Response: Thank you for your suggestion. The volume (*V*), surface of the inner wall (A_w) , the diameter (Φ), the height (*H*) and the sample surface (A_s) of the reactor, the area of the escape hole (A_h), the escape rate constant (k_{esc}), the flow rate of amine (*F*) and the MS sensitivity have been summarized in Table R3. These parameters of the Knudsen cell reactor will be included in the supporting information Table S1. As discussed above, the A_h of amines varied among these three amines and temperature, while it was constant within the uncertainty for N₂ in the range of 232-300 K. Therefore, the A_h of N₂ was listed in this table. The A_h of amines will be summarized in Tables S2 and S3. In the manuscript, this will be pointed out as "The parameters of the reactor are summarized in Tables S1".

Parameter	value		
$V(\mathrm{cm}^3)$	93.90		
$A_{ m w}~(m cm^2)$	116.20		
$\Phi(m cm)$	5.60		
<i>H</i> (cm)	3.50		
$A_{\rm s}({\rm cm}^2)$	3.26		
$A_{\rm h}({ m N}_2,{ m mm}^2)$	0.90±0.15		
$k_{ m esc} ({ m N}_2, { m s}^{-1})$	1.14±0.14		
F of amines (molecules s ⁻¹)	~1.2×10 ¹³		
	~1.1×10 ¹² (MA),		
MS sensitivity of amines $(3\sigma, molecules s^{-1})$	~1.4×10 ¹² (DMA),		
	~1.7×10 ¹² (TMA)		
Dp of kaolinite (μm)	0.56		

 Table R3. The parameters of the Knudsen cell reactor.

Although the surface of the wall was larger than that of the sample holder, the wall was inactive for amine uptake after being passivated in the steady flow as shown in Fig. R4.

Regarding the low-temperature runs I am missing a detailed sketch (again in the Appendix) of the cooling module: are the feeding lines for the coolant insulated inside the Knudsen reactor? Are the authors sure that the amines are not exposed to additional cold surfaces in low temperature runs? Again, let the reader see ALL the DETAILS! Let the reader make the decision as to the validity of the chosen experimental set-up. Most importantly, I have not seen any blanks with the empty cold cell at the lowest temperatures used. This is a must in order to instill a minimum of confidence in your experiments. In addition, the authors withhold any experimental uptake curves at low temperatures which would be of significant interest to many colleagues!

Response: Thank you for your suggestion. The detailed sketch of the cooling module was included in Fig. R1. There was a small isolated chamber cooled with low-temperature ethanol. Temperature was measured with a Pt resistance thermometer embedded in the top ceiling of the cooling chamber on which the sample holder sat. When the sample was exposed to amines, there was no additional cold surface except for the inner surface of the sample cover and the sample holder. The results of the blank experiments have been shown in Fig. R4. However, as shown in Fig. R4 in the blank experiment performed at 232 K, the fresh surface of the sample cover and the sample cover and the sample holder did not contribute to the signal changes in the presence of amines. The uptake curve of amines at 232 K is shown in Fig. R5. This figure has been added in the SI as Fig. S6. In the text, a paragraph will be added as "Fig. S6 shows the uptake curves of amines on kaolinite at 232 K. The signals of all three amines in Fig. S6 decreased more than those observed at 300 K (Fig. 1) after the samples were exposed to amines. For example, the I/I_0 of MA, DMA and TMA were 0.32, 0.30 and 0.22, respectively, compared with the values of 0.45, 0.63, and 0.47 at 300 K" in section 3.3.



Fig. R5. The uptake curves of (A) MA on 21.0 mg kaolinite, (B) DMA on 20.7 mg kaolinite and (C) TMA on 20.5 mg kaolinite at 232 K.

Have the authors checked whether or not the uptake corresponds to a first-order rate law? If the rate law is more complex, and I suspect it is, how good an approximation is a first-order rate law? The Knudsen reactor is a suitable instrument to check out the rate law: one has to vary the rate constant of escape by varying the orifice diameter: if the rate constant for uptake is independent of the orifice diameter, thus the gas residence time, then we have a first-order rate law. Have the authors varied the flow rate? By the way, what was the flow rate of the amine into the flow reactor? One may see that these questions cannot remain unanswered for a halfway complete and reasonable experimental kinetic study on the uptake of amines on kaolinite.

Response: Thank you for your suggestion. We have measured the uptake coefficient of TMA on kaolinite with different escape rates. As shown in Fig. R6, the uptake coefficient of TMA was independent of the escape rate under our experimental conditions, which indicates a first-order rate law.



Fig. R6 The dependence of uptake coefficient on escape rate of TMA. T: 300 K, mass of kaolinite: 20 mg.

It should be pointed out that the flow rate into the reactor was too small (~10⁻⁹ L min⁻¹) to directly measure because we do not have a sufficiently sensitive apparatus to do the flow rate calibration. The flow rate could be measured based on the pressure differential across the constriction produced by an orifice plate if the size of the orifice is known. However, the accurate orifice size was unavailable because a variable leak valve with non-circular hole was used in this work. Therefore, we estimated the flow rate based on the law of conservation of mass. According to the pumping rates of the turbomolecular pumps, the corresponding pressure and the vapor pressure of amines over the solution, the flow rate of amines was estimated to be ~1.2×10¹³ molecules s⁻¹. The MS sensitivity was estimated to be ~1.1×10¹², ~1.4×10¹² and ~1.7×10¹² molecules s⁻¹ for MA, DMA and TMA, respectively, based on the standard deviation of the MS signal (3 σ). In the experimental section, a sentence will be added as "Based on the law of conservation of mass, the flow rate of amines into the reactor was around 1.2×10¹³ molecules s⁻¹ according to the pumping rates of the turbomolecular pumps, the

corresponding pressure and the saturation vapor pressure of the amines" in section 2.1.

First and foremost I am missing a calibration of the residual gas MS signals in terms of amine concentrations. To that end I do not understand why the authors use an aqueous solution of the amines. In order to calibrate the amine signals they must use pure amines which are commercially available. One cannot interpreted saturation curves such as Figure 1 if one does not have the slightest idea how many molecules of amine it takes to saturate the kaolinite: Does the saturation level correspond to a fraction of a monolayer, one or several layers? These are important questions in order to interpret and grasp the meaning of these saturation experiments.

Response: e agree with you that pure amines should be used to calibrate the MS signals. Actually, a permeate tube is required to perform calibration for amines and ammonia (Freshour et al., 2014;Neuman et al., 2003). However, the pure amine cylinders are unavailable in China, and a permeate tube for amines is also unavailable in our laboratory at the present time. Thus, we used the amine solutions to generate amine vapors.

It should be noted that the relative concentration of amine is required to measure the uptake coefficient according to Eq. (R3) or Eq. (R4). Once the MS signal linearly responds to the flow rate or concentration of amine, Eq. (R4) is correct. As shown in Fig. R2, the MS signal was linearly correlated to the flow rate of amine into the reactor or the pressure in the reactor. This means that the measured uptake coefficients of amines in this work are credible.

Of course, the absolute flow rate or concentration of amine is required to calculate the uptake capacity of amine on kaolinite. As stated above, the flow rate of the amines into the reactor was estimated to be $\sim 1.2 \times 10^{13}$ molecules s⁻¹ based on the law of conservation of mass with the pumping rates, the corresponding pressures and the saturation vapor pressure of the amines. Therefore, the uptake capacity can be roughly estimated. We carefully checked the calculation method for the uptake capacity. An improper integration method was utilized in our original manuscript and a conversion factor was missed. The correct uptake capacities of MA, DMA and TMA on kaolinite

were ~ 1.4×10^{14} , ~ 6.3×10^{13} and ~ 3.7×10^{13} molecules mg⁻¹, respectively. This has been corrected in the revised manuscript. In addition, the total amines lost on the kaolinite surface (in Fig. 1) were ~ 2.7×10^{15} , ~ 1.3×10^{15} and ~ 7.4×10^{14} molecules, respectively, under our experimental conditions. With the assumption of the cross-sectional area of MA (0.243 nm²), DMA (0.323 nm²), and TMA (0.394 nm²) (Liu et al., 2012), the adsorbed amines would cover 6.6 (MA), 4.2 (DMA) and 2.9 cm² (TMA) in a monolayer. These values were comparable to the geometric area of the sample holder (3.26 cm^2) and less than the surface area of the kaolinite (1.4 m^2) . Therefore, the saturation level shown in Fig. 1 might correspond to a fraction of a monolayer under our conditions. In the revised manuscript, we will add the related discussion as "The integrated uptake capacities were obtained based on the uptake curves shown in Fig. 1 once the MS signal intensities were converted to the flow rate of amines into the reactor. The uptake capacities were ~2.7×10¹⁵, ~1.3×10¹⁵ and ~7.4×10¹⁴ molecules for MA, DMA and TMA, respectively, under our experimental conditions. The areas of the adsorbed amines in a monolayer would be 6.6 (MA), 4.2 (DMA) and 2.9 cm² (TMA). These values are comparable to the geometric area of the sample holder (3.26 cm^2) and less than the surface area of the kaolinite (1.42 m^2) . This means that the saturation level in Fig. 1 might correspond to a fraction of a monolayer under our conditions".

The following questions are more general:

Pg. 9, top: What is the typical lifetime of MA, DMA or TMA on the stainless steel vessel walls? How does the signal vary with time if you interrupt the flow of amine at once? This should provide the characteristic residence time of the amines on the vessel walls, at least at the beginning because the MS signal decays are complex, unimolecular at the beginning, and more complex at the end.

Response: The residence time of N₂ (τ =1/ k_{esc}) was 0.90±0.15 s⁻¹ in the temperature range of 323-300 K, while the values were slightly dependent on temperature for MA, DMA and TMA (Fig. R7). The mean residence time (τ), the escape rate (k_{esc}) and the area of the escape hole (A_h) for amines and N₂ at different temperatures based on the data fitting are summarized in Table R4. This table has been added in the supporting

information as Table S3.



Fig. R7. Temperature dependence of the residence time in the temperature range of 232-300 K.

Table R4. The mean residence time (τ), the escape rate (k_{esc}) and the area of the escape hole (A_h) in the temperature range of 232-300 K.

T(K)	τ(s)			$k_{\rm esc}(s^{-1})$			$A_{\rm h}({\rm mm^2})$					
	MA	DMA	TMA	N ₂	MA	DMA	TMA	N_2	MA	DMA	TMA	N ₂
232	2.22	1.73	1.27		0.46	0.56	0.79		0.43	0.63	1.02	
237	2.28	1.84	1.29		0.45	0.54	0.78		0.42	0.61	1.00	
248	2.41	2.07	1.32		0.43	0.50	0.76		0.39	0.56	0.97	
258	2.53	2.29	1.36	1.14	0.41	0.47	0.75	0.90	0.37	0.51	0.93	0.93
263	2.58	2.39	1.37	-	0.40	0.45	0.74		0.36	0.49	0.92	-
278	2.76	2.71	1.42		0.38	0.40	0.72		0.33	0.42	0.87	
300	3.02	3.18	1.49		0.34	0.32	0.69		0.28	0.32	0.79	

Pg. 10, lines 224-226: What is the experimental evidence for the interaction of the amines with Lewis acidic sites on the kaolinite. The same message is recurrent: See also pg. 15, line 359, pg. 16, line 379 and pg. 18, line 412. What is the experimental

proof or physical evidence for this? Protonated amines originate from the neutralization of Bronsted, not Lewis acid sites! Furthermore, it is unjustified to postulate protonated MA and TMA from Figure 2. There are no peaks at 1467-1477 cm-1 for these two amines that I can see! To that effect, the authors must amplify and expand the spectrum such that one may distinguish the noise from a potential absorption FTIR signal in the above spectral range.

Response: As a primary amine, Lewis-bound MA gives rise to a characteristic δ band at 1606 cm⁻¹ (-NH₂ deformation band) (Auerbach et al., 2003;Nunes et al., 2005). In Fig. 2A, the band at 1606 cm⁻¹ was observable for the uptake of MA on kaolinite. Lewis-bound DMA shows the δ (NH) band at ~1470 cm⁻¹ (Lin et al., 2014), which was observed in Fig. 2B. Lewis-bound TMA does not have the typical δ band of NH_x except for the IR bands related to CH₃ and NC₃. For all of these amines, however, Lewis-bound amines can be indirectly confirmed by the absence of OH consumption during uptake. As you noted, no peaks at 1467-1477 cm⁻¹ were observable for MA and TMA (Fig. 2A and C). This further confirmed that they were adsorbed on the Lewis acid sites. Physisorption has been confirmed by repeat uptake experiments (Fig. R10) and will be discussed below.

We will revise the description of the IR spectra as "For MA in Fig. 2A, the band at 1606 cm⁻¹ (-NH₂ deformation band) is the characteristic band for Lewis-bound MA (Auerbach et al., 2003;Nunes et al., 2005). In Fig. 2B, the band at ~1470 cm⁻¹ might have originated from the δ (NH) in Lewis-bound DMA (NIST database) and/or δ (CH₃) (Lin et al., 2014). In a previous work, the IR bands at 1484 and 984 cm⁻¹ were assigned to the characteristic bands of protonated TMA ((CH₃)₃NH⁺) in an acidic solution containing TMA and on the surface of polyethylene (PE) treated by TMA (Ongwandee et al., 2007). In this work, although two bands at 1477-1467 cm⁻¹ and 971 cm⁻¹ were observed in the spectrum shown in Fig. 2B, and were close to the bands of protonated DMA (DMAH⁺), it was more reasonable to assign them to Lewis-bound DMA for the following reasons. Formation of protonated amines requires the partition of surface OH (Brønsted acid). However, as shown in Fig. 2B, the surface hydroxyl (-OH) in the range of 3600-3750 cm⁻¹ (Miranda-Trevino and Coles, 2003) was not consumed when the

kaolinite was exposed to DMA. For MA and TMA, the bands at 1477-1467 and 971 cm⁻¹ are discernable in Fig. 2A and C. At the same time, OH consumption was also unobservable when kaolinite was exposed to MA or TMA. Therefore, it can be concluded that Lewis acid sites on the kaolinite predominantly contributed to the uptake of MA, DMA and TMA as observed in Figs. 1 and 2 in this study. This is similar to the adsorption of DMA and N,N-dimethyl formamide (DMF) on TiO₂, for which Lewis acid sites were identified as the reactive sites (Lin et al., 2014)".

How did the authors evaluate the numbers on pg. 11, line 250 (molecules mg-1) in the complete absence of any quantitative calibration of the amine MS signals?

Response: T These numbers were estimated using the flow rate of amines based on the law of conservation of mass in the light of the pumping rates of the turbomolecular pumps, the corresponding pressure and the vapor pressure of the amines over the solution. As pointed out above, standard gas sources of amines were unavailable. The flow rate was roughly estimated here. Using a similar method, we have estimated the flow rate into the reactor of SO₂, COS and amines in our previous work (Ma et al., 2012;Liu et al., 2012;Liu et al., 2008). Although the uncertainties in the flow rate from the pumping rate, pressure and vapor pressure still remain, the trend of the estimated adsorption capacity among these three amines into the reactor will be added in the experimental section as "Based on the law of conservation of mass, the flow rate of amines into the reactor was around 1.2×10^{13} molecules s⁻¹ according to the pumping rates of the turbomolecular pumps, the corresponding pressure and the saturation vapor pressure of the amines".

Regarding Figure 3: The plateau of gamma(obs) seems to be reached for a mass of kaolinite ranging from 20 to 100 mg. Why does the situation change that much for COS/kaolinite (ref. 2010b) in Figure 4 (or reference 2010b) where a sample mass of 20 mg is definitely at the beginning of the linear mass regime (LMR)? In contrast to V. Grassian and coworkers the LMR may be interpreted also in the sense that the mass is

not sufficient to cover up the sample surface area with a coherent material layer because there are "holes" in the substrate layer. This depends of course on the particle size. What was the particle size in this study? This information should go into the technical Table requested above.

Response: Thank you. This difference can be explained by the different diffusion depths of the reactant gas in the multilayer particles. As discussed in our manuscript (**section 3.2**), the probe depth of a reactant in a packed powder sample is governed by the bulk density, the true density, the tortuosity factor and the particle diameter of the sample. It is also determined by the effective uptake coefficient (γ_{eff}) of the reactant. According to Eq. (8) and (9), η (a factor to account for the effect of gas-phase diffusion into the underlying layers) can be simplified as

$$\eta = C \cdot \gamma_{eff}^{-1/2} \cdot \tanh(\gamma_{eff}^{1/2})$$
 (R7)

where *C* is a constant and a function of the bulk density, the true density, the tortuosity factor and the particle diameter of the sample, the sample mass and the geometric area of the sample holder. A large γ_{eff} , which is determined by the properties of both the reactant and the substrate (particles), corresponds to a small η . An η with zero value means that the underlying layers of the packed powder sample do not contribute to the uptake of the reactant gas.

The minimal *m* was 20 mg; ρ_b was measured as 0.65 g cm⁻³; ρ_t was 2.62 g cm⁻³ (Wardhana et al., 2014); d_p was 0.56 µm; A_g was 3.26 cm²; τ was 3 (Matthews et al., 1996). Therefore, the relationship between η and γ_{eff} can be calculated with these parameters. Fig. R8 shows the relationship between η and γ_{eff} . If the γ_{eff} of amines on kaolinite was ~10⁻³ as discussed in this paper, the η value should be ~0.1. Strictly speaking, a nonzero η indicates that underlying layers can contribute to the uptake of reactant gas. However, this cannot be measured if the slope of the observed uptake coefficient against the sample mass is too small, as observed in this study. Therefore, we conclude that the contribution of underlying layers of kaolinite to the uptake of amines was negligible. If the parameters of mineral oxides in our previous work were similar to those of kaolinite in this study, the γ_{eff} of COS of ~10⁻⁶-10⁻⁷ (Liu et al., 2010a)

should correspond to an η of ~1. Therefore, different responses of γ_{obs} against sample mass can be explained by the different values of η between the two reaction systems. In the manuscript, a paragraph will be added as "According to the minimal sample mass (m=20 mg), the measured ρ_b (0.65 g cm⁻³), the ρ_l (2.62 g cm⁻³) (Wardhana et al., 2014) and the measured d_p (0.56 µm) of kaolinite, the A_g (3.26 cm²) and the τ (3) of kaolinite reported in the literature (Matthews et al., 1996), the relationship between η and γ_{eff} was obtained based on Eq. (8)-(10) and is shown in Fig. S5. If the γ_{eff} of amines are close to their γ_{obs} reported here, the corresponding η values should be ~0.1. On the other hand, based on the saturated adsorption capacity of amines estimated above, only 1~2 monolayers of kaolinite particles contribute to the uptake of amines. This is consistent with the assumption that uptake of amines dominantly takes place on the first layer of the kaolinite sample and the contribution of the underlying layers to amine uptake is negligible. Therefore, it is reasonable to conclude that the γ_{eff} of amines on kaolinite are close to or equal to the γ_{obs} in this study".



Fig. R8. The relationship between η and γ_{eff} on kaolinite.

Fig. R9 shows the particle size of kaolinite used in this work. The mean diameter was 0.56 μ m with a half-width of 0.47 μ m. This value will be added in Table S1. The TEM results will also be added in Fig. S3. A sentence will be added in the revised manuscript as "The mean diameter of kaolinite particles was 0.56 μ m with a half-width of 0.47 μ m (Fig. S3) measured with a transmission electron microscope (TEM, Hitachi H7500 with an accelerating voltage of 80 kV)."





Fig. R9. (A) TEM image of kaolinite; (B) size distribution of kaolinite particles.

With the measured bulk density (0.65 g cm⁻³), the depth of the sample was 0.09 mm in the sample holder for 20 mg of kaolinite. Kaolinite particle samples with multilayers should be prepared. We tried to prepare a sample with lower mass. However, it was difficult to obtain an even film covering all of the sample holder surface. We agree with you that LMR may be interpreted in the sense that the mass is not sufficient to cover up the sample surface area with a coherent material layer because there are "holes" in the substrate layer. This was the reason why we did not perform uptake experiments with sample masses lower than 20 mg. In our work, we are sure that all of the sample holder was covered by the samples.

On pg. 13 the authors evaluate gamma(eff) using the KML theory. What are the values of the parameters dp, "tau", _, ', etc. so as to be able to follow the authors in their calculation.

Response: In Eq. (9), the minimum *m* was 20 mg; ρ_b was measured as 0.65 g cm⁻³; ρ_t was 2.62 g cm⁻³ (Wardhana et al., 2014); d_p was 0.56 µm; A_g was 3.26 cm²; τ was 3

(Matthews et al., 1996). We have added these parameters and the corresponding discussion to the revised manuscript as mentioned above.

Pg. 14, line 324 and following: What is the reason the particle size plays such a large role for the magnitude of the uptake coefficient? This question comes up several times without the authors giving an answer.

Response: According to the definition of uptake coefficient (Eq. R1), γ is proportional to the net loss rate of amine. Surface reactions take place between amines and ammonium salts. Therefore, ammonium salts with smaller particle size should lead to a higher loss rate of amines due to the larger specific area when compared with the counterpart. The above discussion will be added in the revised manuscript as "According to the definition of uptake coefficient, γ is proportional to the net loss rate of amine, and consequently, the specific area of the particles. Therefore, ammonium salts with smaller particle size should lead to the larger specific area of amines due to the larger specific area of amines due to the larger specific area of amines due to the larger specific area when compared with the counterpart".

It does not make sense to correlate γ_{eff} with pKb of the amines in Figure 4. The latter parameter is dominated by solvent effects because DMA is a stronger base than TMA in solution whereas one expects the inverse. What the authors should take is either the proton affinity (PA or enthalpy of protonation in the gas phase) or gas phase basicity (gB or equilibrium constant). The values are: NH3 (853.6/819.0 kJ/mol corresponding to PA/gB), MA (899.0/864.5), DMA (929.5/896.5), TMA (948.9/918.1). In this series TMA is clearly the strongest base which is an intrinsic property of the molecule compared to MA and DMA.

Response: Thank you for your good suggestion. The proton affinities (PA) are 879, 924 and 942 kJ mol⁻¹ for MA, DMA and TMA, respectively (Tunon et al., 1992). These values are close to those you mentioned. Therefore, the relationship between the reactivity of amines and the basicity of gaseous amines (Fig. R10B) has been discussed as you suggested. The corresponding discussion will be revised in the manuscript as "On the other hand, the basicity of amines might be another factor affecting the

reactivity. The proton affinity (PA), which is a measurement of the basicity in the gas phase, is 879, 924 and 942 kJ mol⁻¹ for MA, DMA and TMA (Tunon et al., 1992), respectively. The sequence of the reactivity is consistent with the PA of amines in the temperature range of 232-300 K. Therefore, the higher reactivity of TMA on kaolinite can be explained by its strong basicity, and vice versa for MA."



Fig. R10 (A) Box chart for the γ_{eff} of amines on kaolinite measured at 300 K. (B) Relationship between the γ_{eff} and the proton affinity of amines.

Pg. 17, enthalpy and entropy of vaporization: The resulting thermodynamic parameters do not make any sense at all as they are at least a factor of three too small compared to the experimental heat of vaporization of the amines: MA (25.6 kJ/mol), DMA (26.4), TMA (22.94). If the values of the present study were true, then why should the amines interact with kaolinite at all? They certainly will prefer to condense unto itself onto the stainless steel walls into small droplets rather than to adsorb on kaolinite! The reason is that equation (2) is too simple a model for this reactive system. Rather, one must distinguish physisorption from chemisorption. Davidovits did not develop his simple model to a reactive system, therefore, it seems that the simple model is totally

inadequate and yields unphysical results.

Response: The ΔH_{obs} of MA, DMA and TMA on kaolinite were -11.2 ±0.6, -17.2 ±3.7 and -12.1 ±1.3 kJ mol⁻¹, respectively. These values are slightly lower than the heat of vaporization (ΔH_{vap}) of amines. The ΔH_{vap} represents the evaporation heat of amines from liquid phase to gas phase, while the ΔH_{obs} is the adsorption heat from the gas phase to surface-adsorbed amines. These are two different processes. As discussed above and in the manuscript, a fraction of a monolayer of amines formed on the kaolinite surface. Therefore, ΔH_{obs} reflects the strength for the interaction between the amines and kaolinite, but not the intermolecular interaction among amine molecules. On the other hand, the fresh sample holder did not lead to a decrease of MS signals, as shown in Fig. R4 in the blank experiment. All experiments were performed in a steady flow after the reactor wall had been passivated by amines. The reactor walls should also not contribute to the signal changes when the sample was exposed to amines. Therefore, the enthalpy and entropy of adsorption are meaningful. It is also useful to obtain an empirical equation of γ_{eff} with temperature.

It should be pointed out that the enthalpy of physisorption is characterized by change in enthalpy less than 40 kJ mol⁻¹, while the enthalpy of chemisorption is greater than 40 kJ mol⁻¹ (De Moor et al., 2008). Therefore, the uptake of amines on kaolinite can be ascribed to physisorption but not chemisorption. On the other hand, we performed repeat uptake experiments after the kaolinite with adsorbed amines was evacuated overnight. The uptake curve in the second run coincided well with that in the first run (Fig. R11 and Fig. S4). This means that uptake of amines on kaolinite. In the revised manuscript, the following paragraph "Fig. S4 shows the repeat uptake curve in the second run. The uptake curve in the second run coincided well with that second run coincided well with that in the first run coincided well with that in the first run (Fig. R11 and Fig. S4) is valid to describe the uptake of amines on kaolinite. In the revised manuscript, the following paragraph "Fig. S4 shows the repeat uptake curves of MA after the sample that had adsorbed amines in the first run was evacuated overnight and exposed to amines again in the second run. The uptake curve in the second run coincided well with that in the first run. This means amines are reversibly adsorbed on kaolinite" will be added before Eq. (2). Before Eq. (13), we will also revise the sentence as "The relatively small ΔH_{obs} values of amines on kaolinite demonstrate that amines physically adsorb on kaolinite, because the characteristic ΔH of

physisorption is less than 40 kJ mol⁻¹ (De Moor et al., 2008). This is in agreement with the reversible nature of adsorption, as shown in Fig. S4."



Fig. R11. Repeat uptake curves of MA after the used sample was evacuated overnight at 300 K. The sample mass was 19.8 mg.

Pg. 17, line 404: What did you fit in order to obtain equations (13) to (15)? **Response**: These equations were obtained from fitting the uptake coefficients against temperature using Eq. (12). This will be pointed out as "Based on Eq. (12), the empirical

formulas relating γ_t of amines on kaolinite and temperature are given as".

Pg. 19, middle: What is the saturation behavior of the amines at low temperature? **Response**: The saturation time was extended, as shown in Fig. R5, when compared with that in Fig. 1. This figure has been also added in the SI as Fig. S6.

Pg. 18, line 419-420: "...the uptake of amines was predominantly ascribed to mass

accommodation" is hard to understand because mass accommodation is seldom rate limiting, but transition over a barrier is.

Response: In general, gas-particle interaction includes the following elemental steps in sequence. 1) Mass transfer (diffusion) of the reactant(s) from the bulk fluid to the external surface of the particle; 2) Diffusion of the reactant from the pore mouth through the particle pores to the immediate vicinity of the internal surface; 3) Adsorption of reactant A onto the particle surface; 4) Reaction on the surface; 5) Desorption of the products from the surface; 6) Diffusion of the products from the interior of the particle to the pore mouth at the external surface; 7) Mass transfer of the products from the external particle surface to the bulk fluid. The mass accommodation includes the first two steps. As discussed in the manuscript, only physisorption of amines occurs on the surface of kaolinite. It is a barrierless step. According to Eq. (11), mass accommodation is possible if the surface interaction is fast enough. Actually, this has been observed in many reaction systems including uptake of carbonyl sulfide on kaolinite (Liu et al., 2010b), N₂O₅ on (NH₄)₂SO₄ (Griffiths and Anthony Cox, 2009), and N₂O₅ on sulfuric acid aerosol (Hallquist et al., 2000).

Table 2, pg. 26: Show raw data at low temperatures!

Response: This will be added in the Fig. S6 as mentioned above.

Pg. 32, Figure 5: The TMA data lie on a curve, NOT on a straight line! There are important deviations.

Response: Thank you. The data in Fig. 5 were linearly fitted. The correlation coefficients were 0.99 (MA), 0.94 (DMA), and 0.98 (TMA). We think it is reasonable to depict as a line but not a curve. The correlation coefficients will be added in the figure caption in the revised manuscript as "The correlation coefficients are 0.99, 0.94 and 0.98 for MA, DMA and TMA, respectively".

Some of less important items:

Pg. 3, line 41: What are "anthropological" emissions?

Response: Thank you. The word will be replaced with "anthropogenic".

Pg. 6, line 138: Tabor et al., 1994, Ullerstam et al., 2003: references are missing.**Response**: Thank you so much. These references will be added.

Pg. 13, 295: Salgado-Muñoz and,,,,also in bibliographic list at the end (line 623)! **Response**: Thank you. This will be corrected as "(Salgado-Muñoz and Rossi, 2002)"

Pg. 14, line 327: cluster has the wrong polarity!

Response: We double checked this cluster in the reference (Bzdek et al., 2010). It was $[(NH_4)_3(HSO_4)_2]^+$ instead of $[(NH_4)_3(SO_4)_2]^+$. This will be corrected in the manuscript.

Pg. 14, line 332: Larger than what? The author's comparison only has one leg!! **Response**: Thank you. This sentence will be revised as "H₂SO₄ solution with high H₂SO₄ content also showed a slightly larger γ of amines than that with low H₂SO₄ concentration".

Pg. 15, line 344: What is TEAH sulfate?

Response: It should be TEA-sulfate. It will be corrected in the manuscript.

Pg. 20, line 473: mineral dust is a bad reservoir or no reservoir at all! The authors should cut out qualitative or meaningless talk.

Response: Thank you. The last sentence "In this process, mineral dust takes on the role of a carrier or reservoir of amines" will be deleted in the manuscript.

Pg. 23, line 653: "Physiochemical"?**Response**: Yes, it is. The title of this paper is:





Physiochemical Properties of Alkylaminium Sulfates: Hygroscopicity, Thermostability, and Density

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Pg. 25, Table 1: Under DMA: fifth entry from the top of DMA field has wrong polarity!

Response: Thank you. It should be "[(NH₄)₃(HSO₄)₂]⁺" and will be corrected in the manuscript.

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