Augst 29, 2017

Attention: Atmosphere Chemistry and Physics

Handling Co-Editor
Dr. Jianping Huang

Thank you very much for your handling. We completed to receive all reviewer comments of June 14, 2017, with regard to our manuscript acp-2016-1095-RC1: "**Variations in airborne bacterial communities at high altitudes over the Noto Peninsula (Japan) in response to Asian dust events**" together with the comments from the two reviewers.

I am sending herewith an electric file of our revised manuscript. Our alterations as result of the reviewer's suggestions and our comments for their suggestions are shown in another pages.

I believe the manuscript has been improved satisfactorily and hope it will be accepted for publication of Atmos. Chem. Phys.

Thank you.
Sincerely yours,

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Dear Anonymous Referee #1:

We thank for admitting the value of our manuscript very much. I take your comments into account in our revised manuscript. Additionally, I'm sorry for some technical mistakes. I revised our manuscript with paying attention to the points that you commented. The revised manuscript is attached as supplement file. I described my response for each your comment. The sections [Q] indicate your comments and the sections (A) indicate my responses. The changes introduced in the revised manuscript were indicated by the line numbers at the sections (A).

[Q] The authors should make it clearer to the readers what is dust and non-dust events. This should be emphasized in the figures (2, 3, 4, 5, 6, 7, 9); figure captions; table (I would recommend adding another column for that information); as well as in the result text. Otherwise the data presented is somehow confusing and not clear.

(A) The sampling days of dust or non-dust events have been indicated in Figures and Figure captions in the revised manuscript (Figures 2, 3, 4, 5, 6, and 7). Additional columns defining the dust event days have been inserted into Table 1.

[Q] It would be helpful to add some information on the DAPI-staining colors in the introduction part. Introducing these definitions only in the discussion (line 465) makes it hard to follow along the text beforehand.

(A) Some information on the DAPI-staining colors have been inserted in the Introduction section and the Experiment section in the revised manuscript (lines 89-1091.

[Q] line 103: It is specified that aerosol origin is from continental areas, however, trajectories and analysis shows marine contribution as well. please rephrase.

(A) As this decision, the explanations of aerosol origins over Noto Peninsula were rephrased in the revised manuscript (lines 121-122).

[Q] - line 120: How were the filter sterilized? please add either company cat. number, or

sterilization technique.

(A) In the revised manuscript, we have added the information of filter and the filter -sterilization processes (lines 138-142).

[Q] - line 160: Please add the immersion oil type.

(A) The immersion oil type has been inserted in the revised manuscript (lines 181-182).

[Q] - line 174: Reference for the DNA extraction method: Authors should double check the ref., as the Maki 2008 paper refers to the Maki 2004... And - as in the 2004 paper the extraction is not from air filters, the authors should specify the extraction efficiency from filters using this method in the current paper.

(A) Since gDNA amounts were not enough for the direct determination using light absorbance, the gDNA were determined the PCR products at the first PCR amplification. The extraction efficiency from filters were estimated by the comparison between the PCR products and the particle concentrations by DAPI count, indicating that more 90% of gDNA can be collected by this DNA extraction system. The detail explanations about the DNA extractions have been added to the section of Experiments in the revised manuscript (lines 229-235).

[Q] - section 3.3: The protease treatment is not detailed in the methodology. Although a very important examination, indicative for protein dominance is yellow particle, no documentation of such treatment and detection before and after treatment is presented. The authors should either supply such results and extend methodology, or remove this part.

(A) Although we already have possessed some results about the protease treatments of yellow particles, the data was not sufficient for demonstrating that all yellow particles are composed of protein. Moreover, I think the yellow particle fractions includes unknown organic components. Accordingly, in the revised paper, this part has been removed. The identification of yellow particles are further works.

[Q] - I find it very interesting that marine cyanobacteria contribute to the April 2013, March 2015 events etc. as was also observed by Lang-Yona et al., 2014. This could be relevant for the public health at low altitudes. Please add a discussion on the possible health effects of such species and other gram negative bacteria.

(A) Thank you for your suggestion and the information about valuable reference. We have discussed about the health effects by airborne cyanobacteria with referring to the suggested reference (lines 634-638).

[Q] - section 4.2: Organic particles might indeed represent dead bacteria and fungi, however also anthropogenic and natural SOA (especially when air transport over polluted areas, as in the current study). This should be emphasized in the discussion, as the statement (fraction of dead cells compared to total microbes) based on Fig. S4 could be misleading.

(A) Thank you for your suggestion. I agree to this comments. The anthropogenic and natural SOA were also included in the yellow fluorescent fractions. This topic has been discussed in the revised manuscript (lines 500-506).

[Q] - Line 513: I'm not convinced that cyanobacteria are significantly enriched in dust samples. As described in the result section, cyanobacteria were enriched also in non dust samples. The authors should supply arguments and statistical evidence for this statement.

(A) In the section of previous manuscript, I mistake to describe about cyanobacteria as the dust specific bacteria. Correctly, cyanobacteria are thought to be the bacterial populations in regardless of dust events and originated from marine environments. The name "cyanobacteria" has been removed at the section of dust-specific bacteria in the revised manuscript (lines 528-529).

[Q] - section 4.7: Assuming fluxes of specific bacteria as a representative for the origin of the air mass is a rough estimation and should not be made based on such a study with limited number of sampling points. For example, it is well established that the

aerosolization of cyanobacteria would be dominant during bloom events. Therefore, if the authors make such statement of cyanobacteria represent marine-originated aerosols, they should supply evidence for presence of cyanobacteria in high altitudes seasonally and annually, and correlate with bloom events. In addition, one significant source of airborne cyanobacteria are the fresh water bodies. Many other factors affect the abundance of airborne microorganisms, and therefore I find it hard to accept such statement, where the presence of microbes will reflect the origin of the air mass accurately. Authors are requested to restrain their assumption.

(A) I agree to your comments. We need sufficient information obtained from more numbers of air samples and detail discussion for establishing the air-mass tracking by bacterial compositions. Then this section has been removed and the shortage description about the tracking idea was indicated in the section of Conclusion (lines 659-672).

[Q] - line 671: Please supply reference for this statement.

(A) This parts have been eliminated, because this description about bioaerosol tracking have been shortened and removed to the Conclusion section.

Technical corrections:

[Q] - Section 2.7 should be 2.5.

(A) Section 2.7 has been revised to 2.5 (line 251).

[Q] - line 361-363: Please rewrite this sentence.

(A) I have revised this sentence (lines 378-381).

[Q] - line 421: ": : :their abundance fluctuated between from: : :" please check phrasing.

(A) Sorry for mistake. I have revised this phrase (line 435).

[Q] - line 483: .."ranged from 23.3: : :" – consider rephrasing.

(A) I have rephrased this section in the revised manuscript (lines 495-496).

[Q] - line 505: Mazar et al. reported dust microbial composition over east Mediterranean areas (not European). Please correct.

(A) I'm sorry for errors. " European " has been revised to " east Mediterranean areas " (line 519).

[Q] - Line 513: Please check if "Figure 4" in the text should be corrected.

(A) Sorry for mistake. I have changed to "Figure 4" (line 529).

[Q] - Figure 2 – Caption: should be corrected for black particles denoted in grey color.

(A) The caption has been revised to indicate the matching color (line 1002).

[Q] - Figure 8b: Authors should better defined symbols. It is not clear (from both legend and caption) what are the blue circles (Are they dust samples? non-dust?) The authors should also add information on the statistics significance of the unifrac test. Consider adding dispersion ellipses with 95% standard deviation confidence interval.

(A) I agree to your comment. The definition for each sample was not clear. After the characteristics of samples have been improved to be defined, Figure 8b and its figure caption has been revised to eliminate the confusion relating to symbols (Figure 8b).

[Q] - Figure S4: Please specify in caption/legend what the black and white bars indicate.

(A) The caption of Figure S4 has been improved in the revised manuscript (Figure S4).

Dear Anonymous Referee #2:

We thank for admitting the value of our manuscript very much. I take your comments into account in our revised manuscript. I revised our manuscript with paying attention to the points that you commented. I described my response for each your comment. The revised manuscript is attached as supplement file. The sections [Q] indicate your comments and the sections (A) indicate my responses. The changes introduced in the revised manuscript were indicated by the line numbers at the sections (A).

[Q]1. Introduction: bioaerosols could act as active ice nucleus, consequently affect the microphysical properties of cloud in the atmosphere. Please review some papers about climate effects of bioaerosol, so that the readers are easy to understand the importance of your study.

(A1) The climate effects of bioaerosol has been enhanced using some references in the Introduction section (lines 45-59).

[Q]2. Line 28 in page 3: the authors claimed that aerosols in the two cities directly originate from continental areas. I think it is not rigorous and suitable. There are several sources of aerosols in the Noto Peninsula, such as continental and Ocean area, even from local area, depending on condition of airflows. The word should be changed.

(A2) I agree with this comment. Several sources areas of air-mass transported to Noto Peninsula were explained in the revised manuscript (lines 121-122).

[Q]3. Line 23 in page 4: depolarization ratio is more popular for lidar community that depolarization rates. Please replace it throughout the manuscript.

(A3) The term “depolarization rates” has been changed to “depolarization ratio” in the revised manuscript (entire revised manuscript).

[Q]4. Line 8 in page 5: add ‘number concentration’ to the behind of ‘aerosol’.

(A4) Thank you for your indication. I have revised this part (lines 195-196).

[Q]5. Line 17 in page 6: change ‘dust mineral’ to ‘mineral dust’.

(A5) As your decision, I have changed the term ‘dust mineral’ to ‘mineral dust’ (entire revised manuscript).

[Q]6. Line 7-10 in page 7: the word ‘troposphere’ is not appropriate in the manuscript, please consider ‘tropopause’.

(A6) Thank you for your suggestion. In this section, I have revised to more clear explanation defining the boundary layers over sampling areas (lines 286-288).

[Q]7. Line 25-29 in page 7: please rewrite and cut the paragraph short, it is not necessary to list so many names of the samples. Perhaps the authors can mark dust samples and non-dust samples in Table 1.

(A7) I also think Table 1 can cover the explanation about sample names. Accordingly, this parts explaining about the sample name have been shortened in the revised manuscript (lines 321-325).

[Q]8. Section 3.3: four types of fluorescence particles, such as white, blue, yellow, or black particles, could be seen from fluorescent microscopy. To make the reader easier understand, the author should explain the methods and basis of classification. For example, why the white particles are indicative of mineral dust and yellow particles are organic matter.

(A8) Although some parts of the DAPI staining theory of each fluorescent particles are unclear, they were tried to be explained in the revised manuscript (lines 188-195).

[Q]9. Section 4.1: I suggest move this sentences to Introduction and Section 3.1. Also, I suggest that rewrite the Section 4, and move some sentences to Introduction.

(A9) I agree to your comments. The previous discussion section included some parts which had to be moved to Introduction. In the revised manuscript, the parts were shortened and move to Introduction and the introduction has been modified (in particular lines 455-459, 517-522).

[Q]10. Line 21 in page 12: combine “Maki et al., 2010” and “Maki et al., 2013” to “Maki et al., 2010 and 2013”.

(A10) Thank you for your suggestion. “Maki et al., 2010” and “Maki et al., 2013” have been combined to “Maki et al., 2010 and 2013” in the revised manuscript (line 551).

[Q]11. Line 32 in page 12: add ‘long-range’ in the front of ‘transported’.

(A11) The term ‘long-range’ has been moved to the front of ‘transported’ (line 567).

[Q]12. Figure 1: it is not easy for the readers to understand meaning. Please enlarge four panels of helicopter flight routes and reduce size of the East Asia map. Furthermore, panel (a) can be removed and the location of three cities could be marked in panel (b). N and E should be put at the front of latitude ad longitude, such as 50°N and 120°E.

(A12) The maps in Figure 1 have been improved by depending on your suggestion. Thank you for your comments (Figure 1).

[Q]13. Figure 2: according to the meaning described in the paper, the authors would like to use depolarization ratio of aerosols from lidar measurements, for classifying dust events and non-dust events. But the lidar data as shown in fig. 2 is attenuated backscattering, not depolarization ratio. Same as for the panel (a) in fig. 4 and fig. 5. Please replace the data.

(A13) In the previous manuscript, the data in Figs. 2, 4 and 5 were originated from depolarization ratio, but I showed wrong scale bar and unit. Sorry for causing confusion. The scale bar and unit have been changed to correct ones in the revised manuscript (Figures. 2, 4 and 5).

Furthermore, the explanation about depolarization ratio have been also revised for describing that the ratio means the rates of non-spherical aerosols among all particles (lines 162-164).

[Q]14. In my opinion, more bacteria should be observed during dust events comparing the condition during non-dust events. Because mineral dust usually can be long-range transported with bioaerosols. However, concentration of fluorescent particles (especially blue particles) at near surface (ground level) was lower during dust events (as shown in fig. (a) and (b)) than those during non-dust events. Please explain the reason.

(A14) On our opinion, the fluorescent particles (blue particles and others) are mostly similar each other between fig. (a) and (b), because the particle concentration units of x axis for fig. (a) are one order higher that for fig. (b); fig. (a): 10^6 particles/m^3, and fig. (b): 10^5 particles/m^3. However, I think that the reason for the similar concentrations is needed for this paper and should be inserted in the revised manuscript.

At this sampling periods, the high amounts of bioaerosols would be transported to high altitudes and have not fall down to ground surfaces. On the other hands, the air mass during non-dust events is thought to including high amounts of local aerosols. Accordingly, the microbial concentrations in non-dust events were higher than those of dust events. This explanation has been inserted in the revised manuscript (lines 479-484).

[Q]15. Figure 3: there are several backward trajectories in each panel, but the authors claimed that these three-day backward trajectories only be obtained at two altitudes (2500m and 1200m). Same as for the panel (c) in fig. 4 and fig. 5. Please explain it.

(A15) Trajectories at two altitudes (2500m and 1200m) were calculated at every hour for 4hr (0hr, 1hr, 2hr, 3hr and 4hr) before the sampling finish time of each sampling periods. Accordingly, there are total 10 trajectories for each panel. This explanation has been inserted in the captions of Figs. 3, 4 and 5 (lines 1005-1006, 1019-1020, 1033-1034).

[Q]16. Figure 5: the title of x-axis in panel (a) should be “March 2015”, please change it.

(A16) Sorry. I have changed “March 2014” to “March 2015” (Figure 5).

[Q]17. The results in the paper give us more information about bioaerosols in the atmosphere, especially during dust events. The authors are encouraged to compare their results with others from previous studies. Please summarize similar results from other papers in response to dust events, and then add a table in Section discussion.

(A17) As your comment, more references have been cited and the bacterial communities differed from the data of previous researches was discussed in the revised manuscript (Sections of Introduction and Discussion, Table 2).

Title:

Variations in airborne bacterial communities at high altitudes over the Noto Peninsula (Japan) in response to Asian dust events

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Abstract

Aerosol particles, including airborne microorganisms, are transported through the free troposphere from the Asian continental area to the downwind area in East Asia and can influence climate changes, ecosystem dynamics, and human health. However, the variations present in airborne bacterial communities in the free troposphere over downwind areas are poorly understood, and there are few studies that provide an in-depth examination of the effects of long-range transport of aerosols (natural and anthropogenic particles) on bacterial variations. In this study, the vertical distributions of airborne bacterial communities at high altitudes were investigated and the bacterial variations were compared between dust events and non-dust events.

Aerosols were collected at three altitudes from ground level to the free troposphere (upper level: 3,000 m or 2,500 m; middle level: 1,200 m or 500 m; and low level: 10 m) during Asian dust events and non-dust events over the Noto Peninsula, Japan, where westerly winds carry aerosols from the Asian continental areas. During Asian dust events, air masses at high altitudes were transported from the Asian continental area by westerly winds, and Laser Imaging Detection and Ranging (LIDAR) data indicated high concentrations of non-spherical particles, suggesting that dust-sand particles were transported from the central desert regions of Asia. The air samples collected during the dust events contained 10–100 times higher concentrations of microscopic fluorescent particles and Optical Particle Counter (OPC) measured particles than in non-dust events. The air masses of non-dust events contained lower amounts of dust-sand particles. Additionally, some air samples showed relatively high levels of black carbon, which were likely transported from the Asian continental coasts. Moreover, during the dust

events, microbial particles at altitudes of >1,200 m increased to the concentrations ranging from 1.2 x 10^6 particles m^{-3} to 6.6 x 10^6 particles m^{-3}. In contrast, when dust events disappeared, the microbial particles at >1,200 m decreased slightly to microbial-particle concentrations ranging from 6.4 x 10^4 particles m^{-3} to 8.9 x 10^5 particles m^{-3}.

High-throughput sequencing technology targeting 16S rRNA genes (16S rDNA) revealed that the bacterial communities collected at high altitudes (from 500 m to 3,000 m) during dust events exhibited higher diversities and were predominantly composed of natural-sand/terrestrial bacteria, such as *Bacillus* members. During non-dust periods, airborne bacteria at high altitudes were mainly composed of anthropogenic/terrestrial bacteria (Actinobacteria), marine bacteria (Cyanobacteria and Alphaproteobacteria), and plant-associated bacteria (Gammaproteobacteria), which shifted in composition in correspondence with the origins of the air masses and the meteorological conditions. The airborne bacterial structures at high altitudes suggested remarkable changes in response to air mass sources, which contributed to the increases in community richness and to the domination of a few bacterial taxa.

1. Introduction

Airborne microorganisms (bioaerosols) associated with desert-sand and anthropogenic particles were transported through free troposphere from the Asian continents to downwind regions of East Asia and can influence climate changes, ecosystem dynamics, and human health (Iwasaka et al., 2009). Natural dust events from the Asian desert regions carry airborne microorganisms, supporting atmospheric microbial dispersals (Griffin et al., 2007; Maki et al., 2010; Pointing and Belnap, 2014). Haze days caused by anthropogenic particles from Asian continents also affect airborne microbial abundance and endotoxin levels (Wei et al., 2016). Some studies demonstrated that Asian dust events, including natural and anthropogenic particles, cause vertical mixture of bioaerosols in downwind areas, such as in Japan (Huang et al., 2015b; Sugimoto et al., 2012; Maki et al., 2015).

Bioaerosols, which include bacteria, fungi, and viruses, are transported from ground environments to the free troposphere and account for a substantial proportion of organic aerosols (Jaenicke, 2005). Bioaerosols are thought to influence atmospheric processes by participating in atmospheric chemical reactions and in the formation of cloud-nucleating particles (Pratt et al., 2009; Morris et al., 2011; Hara et al., 2016b). Indeed, airborne microorganisms act as ice nuclei that are related to ice-cloud formation processes (Möhler et al., 2007; Delort et al., 2010; Creamean et al., 2013; Joly et al., 2013). In particular, ice-nucleation activating proteins of some microorganims, such as *Pseudomonas syringae*, *Xanthomonas campestris* and *Erwinia herbicola*, exhibit high nucleation activities, initiating ice formation at relatively warm temperatures (greater than -5 °C) (Morris et al. 2004) in comparison to the inorganic ice-nucleating particles,

such as potassium feldspar (approximately -8 °C) (Atkinson et al. 2013). Ice-nucleating particles that originate from bioaerosols are believed to activate ice formation more efficiently than inorganic substances (Hoose and Möhler, 2012; Murray et al. 2012), and are primary contributors of rapid ice-cloud formation even at low concentrations in the clouds at temperatures between -8 °C and -3 °C (Hallett and Mossop, 1974). Bioaerosols are key factors for elucidating the detailed mechanisms of ice-cloud formation and precipitation over East Asia (Hara et al., 2016ab), but the microbial characteristics of bioaerosols transported over long distances by Asian-dust events are still unclear. Furthermore, the microorganisms transported by Asian dust events increase the allergenic burden, consequently inducing asthma incidences (Ichinose et al., 2005) and contributing to the dispersal of diseases such as Kawasaki disease (Rodó et al., 2011) and rust diseases (Brown and Hovmøller, 2002).

In downwind areas of East Asia, the atmospheric bacterial dynamics at high altitudes should be investigated in order to understand the ecological and meteorological influences of airborne bacteria as well as their long-range dispersion. Meteorological shifts and dust events can dramatically alter airborne bacterial communities at high altitudes in Japan (Maki et al., 2013 and 2015) because of air masses that originate from heterogeneous environments, including marine, mountainous, urban, and desert areas. The airborne microorganisms around North American mountains (2,700 m above sea level) were also found to increase their species diversities in response to Asian dust events (Smith et al., 2013). High-throughput sequencing technology can generate large numbers of nucleotide sequences and the sequencing database has played an important role for investigation of airborne bacterial compositions (Brodie et al., 2007; Woo et al.,

115 2013). Indeed, the analyses using high-throughput sequencing has demonstrated that
116 airborne bacterial populations at ground levels change in response to pollutants from
117 Beijing (Cao et al., 2014) and African dust events (Mazar et al., 2016). To investigate
118 their long-range transported bacteria while avoiding the ground-surface contaminations,
119 the bioaerosol samples collected at high altitudes by aircrafts were analyzed using
120 high-throughput sequencing, showing the airborne microbial diversities at high altitudes,
121 ranging from 1,000 m to 3,000 m (DeLeon-Rodriguez et al., 2013; Maki et al., 2015).
122 There are also a few studies on the vertical bacterial distribution from the ground level
123 to the troposphere (DeLeon-Rodriguez et al., 2013; Maki et al., 2015). Nonetheless,
124 while some variations were observed, the specific changes in tropospheric bioaerosols
125 over East Asia, and, in particular, differences between Asian dust and non-dust events
126 remain poorly understood.

127 Organic aerosol particles, such as bioaerosols, account for high rates of
128 tropospheric aerosols, ranging from 30 % to 80 % (Jaenicke, 2005), and fluctuate at
129 high concentrations, ranging from 10^3 to 10^5 particles m^{-3}, under the boundary layer at
130 4,000 m above the ground (Twohy et al., 2016). Epifluorescence microscopy using
131 fluorescent-dye staining is a useful tool for observation and determination of microbial
132 particles in the atmosphere, demonstrating that the biomass of airborne microorganisms
133 increased 10– to 100–fold during Asian-dust events (Hara et al., 2012, Maki et al.,
134 2014). Under a fluorescence microscope, DNA in microbial particles fluoresce blue
135 when stained with 4, 6-diamidino-2-phenylindole (DAPI) (Russell et al., 1974), and
136 organic materials aggregated with proteins and microbial cell components were
137 confirmed as yellow fluorescence particles (Mostajir et al., 1995). Mineral particles

(white particles) and black carbon (black particles) can also be observed as background fluorescence in microscopic observation fields (Maki et al., 2014). Accordingly, several DAPI-stained particles could be detected in air samples collected from all over Japan during dust events (Maki et al., 2013) and can be used as indicators for evaluating the amounts of some aerosol species during dust events.

In this study, the bacterial communities from different altitudes around the Japanese islands were compared to identify the potential influences of long-range transported air masses on tropospheric bacteria. We used a helicopter for collecting air samples at altitudes ranging from 1,200 m to 3,000 m over the Noto Peninsula, Japan. Helicopter sampling was used to collect chemical components at high altitudes, which has previously been used to avoid contamination from the downwash created by spinning rotors (Watanabe et al., 2016). This air sampling method can directly collect aerosols moving from Asian continents or marine areas to Japan. We estimated the air mass conditions using the meteorological data obtained during the sampling periods, and determined aerosol amounts by using meteorological monitoring and epifluorescence microscopic observation. Bacterial community structures were analyzed by using high-throughput sequencing targeting bacterial 16S rRNA genes (16S rDNA).

2. Experiments

2.1. Sampling

Aerosol sampling using a helicopter (R44; Robinson, CA, USA) was performed over coastal areas from Uchinada (36º67N, 136º64E) to Hakui (36º92N, 136º76E) in the

Noto Peninsula, Japan. Both cities are located on the western coast of the Noto Peninsula where aerosols arrive from continental areas across the Sea of Japan and are mixed with local aerosols (Fig. 1). The helicopter traveled 20 km northwest from Kanazawa to Uchinada; air sampling was continuously conducted from Uchinada to the northern coastal areas. To compare the vertical distributions of airborne bacteria during dust and non-dust events, air samples were collected using a helicopter at the 1 to 3 altitudes ranging from 500 m to 3,000 m above ground level (Table 1). Air samples from low altitude regions (10 m above ground level) were collected from the roof of a building located at Taki bay in Hakui (36º92 N, 136º76 E). To compare the vertical bacterial distribution, aerosol samples were collected during the daytime (from 9:00 Japanese standard time [JST; UTC + 9 h] to 16:30 JST) on March 19, 2013; April 28, 2013; March 28, 2014; and March 20, 2015. These samples were collected at the following altitude sets; (1) 2,500 m, 1,200 m, and 10 m; (2) 3,000 m, 1,200 m, and 10 m; (3) 3,000 m, 1,200 m, and 10 m; and (4) 2,500 m and 500 m, respectively, and samples were labeled as shown in Table 1. To investigate the bacterial changes at altitudes in response to time, temporal transect at the altitude of 1,200 m was prepared for seven days – the 23rd, 24th, 25th, and 29th of March 2014 and the 16th, 17th, and 21st of March 2015 – and the sample names are showed in Table 1.

Air samples were collected through sterilized polycarbonate filters (0.22-μm pore size; Whatman, Tokyo, Japan) with sterilized filter holders (Swinnex Filter holder; Merck, Darmstadt, German) connected to an air pump. At the sterilization processes, the filters and the filter-holder parts were irradiated separately under UV light for 1.0 h and the filter holders attached with the filters were autoclaved at 121 ºC for 20 min. Air

sampling was performed with a flow rates of 5 L min^{-1} over sampling periods from 0.2 h to 1.0 h. Triplicate sampling filters were obtained for each altitude. During helicopter sampling, outside air was transferred from a window to the bioaerosols-sampling inlet, which was sterilized by autoclaving and UV irradiation. The sterilized filter holders were inserted into the sampling inlet to avoid contamination. To collect air particles at an altitude of 10 m, we used filter holders fixed on a 3 m stick, which was placed on the roof of a building (Maki et al., 2014).

In total, 18 air samples were obtained during the sampling periods (Table 1). Of the two filters used to collect each sample, one filter was used to determine the particulate abundances under fluorescence microscopy, and the other was stored at -80°C before the extraction of genomic DNA for analysis of bacterial compositions.

2.2. Characteristics and trajectories of air masses

Information regarding weather conditions (temperature, relative humidity, and pressure) was gathered. During the helicopter flight, outside air was transferred from a window into the meteorological-measurement inlet, into which the adaptor of the measurement device (TR-73U; T&D Corporation, Matsumoto, Japan) was inserted, and the temperature, relative humidity, and pressures were sequentially measured. The temperature and relative humidity at an altitude of 10 m were also measured on the roof of a building in Hakui. The depolarization ratio, which was measured by Laser Imaging Detection and Ranging (LIDAR) measurements at Toyama, has been used for the detection of non-spherical aerosols, such as mineral dust particles and/or sea salts.

To track the transport pathways of air masses, 72 h back trajectories were

207 calculated using the National Oceanic and Atmospheric Administration (NOAA)
208 HYbrid Single Particle Lagrangian Integrated Trajectory (HYSPLIT) model
209 (http://www.arl.noaa.gov/HYSPLIT.php). The coordinator of Hakui was used as the
210 back trajectory starting point at several altitudes from 10 m to 3,000 m above ground
211 level to estimate the trajectories of the air masses.
212
213 *2.3. Determination of particle abundance*
214 The air particles at each altitude were measured using an optical particle counter
215 (OPC: Rion, Tokyo, Japan). The OPC device was connected to the
216 meteorological-measurement inlet. The air particles at an altitude of 10 m were also
217 counted using the OPC device placed on the roof of a building.
218 Fluorescent particles stained with DAPI were also counted via epifluorescence
219 microscopy. Within 2 h of sampling, 1 mL of 1 % paraformaldehyde was added to one
220 of the filters to fix the aerosols. After a 1 h incubation, the filter was stained with DAPI
221 at a final concentration of 0.5 μg mL^{-1} for 15 min (Russell et al., 1974). Next, the filter
222 was placed on a slide in a drop of low-fluorescence immersion oil (Type-F
223 IMMOIL-F30CC, Olympus, Tokyo, Japan). A second drop of oil was added, and a
224 coverslip was placed on top. Particles on the filter were observed using a fluorescence
225 microscope (BX-51, Olympus, Tokyo, Japan) with a UV excitation system. A filter
226 transect was scanned, and the four categorized particles, including white fluorescent
227 particles, blue fluorescent particles (microbial particles), yellow fluorescent particles,
228 and black particles, on the filter transect were counted using a previously reported
229 observational technique (Maki et al., 2014). The TA connections in DNA sequences of

230 microbial particles are bound with DAPI, emitting clear blue fluorescence. However, the
231 aggregation of organic matter might also accumulate DAPI at high amounts emitting
232 yellow fluorescence, which is due to formation of a compound with DAPI. Mineral
233 particles often have white autofluorescence or emit weak-blue (mostly white)
234 fluorescence originating from residues of DAPI on the particle surfaces and can be
235 identified on the weak blight background of microscopic observation fields. The black
236 color of black carbon can be identified in the background. The detection limit of aerosol
237 particle concentration was 1.1×10^4 particles m^{-3} of air.
238
239 *2.4. Analysis of bacterial community structures using MiSeq sequencing analysis*
240 *targeting 16S rDNA sequences*
241 After the aerosol particles on the other two filters were suspended in 3 mL of
242 sterile 0.6 % NaCl solution, the particles were pelleted by centrifugation at 20,000 × *g*
243 for 10 min. The genomic DNA (gDNA) was then extracted from the particle pellets
244 using sodium dodecyl sulfate, proteinase K, and lysozyme and purified by
245 phenol-chloroform extraction as previously described (Maki et al., 2008). The bacterial
246 community structure was determined using MiSeq DNA sequencing, which facilitates
247 multiplexed partial sequencing of 16S rDNA. Fragments of 16S rDNA (approximately
248 500 bp) were amplified from the extracted gDNA by PCR using the universal 16S
249 rDNA bacterial primers 515F (5′- Seq A -TGTGCCAGCMGCCGCGGTAA-3′) and
250 806R (5′- Seq B -GGACTACHVGGGTWTCTAAT-3′) (Caporaso et al., 2011), where
251 Seq A and Seq B represent the nucleotide sequences bounded by the second set of PCR
252 primers described below. The PCR amplicon sequences covered the variable region V4

253 of the 16S rRNA gene. Thermal cycling was performed using a thermocycler (Program
254 Temp Control System PC-700; ASTEC, Fukuoka, Japan) under the following
255 conditions: denaturation at 94°C for 1 min, annealing at 52°C for 2 min, and extension
256 at 72°C for 2 min for 20 cycles. Fragments of 16S rDNA in PCR products were
257 amplified again using the second PCR forward primer (5′- Adaptor C - xxxxxxxx - Seq
258 A -3′) and reverse primer (5′- Adaptor D - Seq B -3′), where Adaptors C and D were
259 used for the Miseq sequencing reaction. The sequences "xxxxxxxx" comprise an 8
260 nucleotide sequence tag designed for sample identification barcoding. Thermal cycling
261 was performed under the following conditions: denaturation at 94°C for 1 min,
262 annealing at 59°C for 2 min, and extension at 72°C for 2 min for 15 cycles. PCR
263 amplicons were purified using the MonoFas DNA purification kit (GL Sciences, Tokyo,
264 Japan). PCR amplicons from each sample were pooled at approximately equal amounts
265 into a single sequencing tube on a MiSeq Genome Sequencer (Illumina, CA, USA)
266 machine. The sequences obtained for each sample were demultiplexed based on the tag,
267 including the 8 nucleotide sequence. After removal of the tags, an average read length of
268 450 bp was obtained. Negative controls (no template and extraction products from
269 unused filters) were prepared in the DNA extraction process to check for contamination.
270 The amount of gDNA extracted from air samples ranged from the detection limit (<0.5
271 ng/samples) to approximately 50 ng/samples and cannot be determined directly by light
272 absorbance measurements. Accordingly, quantities of gDNA were estimated using the
273 PCR products after the first amplification step, and compared with the
274 microbial-particle concentrations that were determined by fluorescence microscopic
275 observation. The efficiency of the gDNA extraction from air samples was more than

276 80 %.

277 Before the analysis of bacterial community structures, USEARCH v.8.01623
278 (Edgar, 2013) was used to process the raw Illumina sequencing reads. Anomalous
279 sequences were removed with the following workflow. First, the forward and reverse
280 paired-end reads were merged, and the merged reads with lengths outside of the
281 200-500 bp range or those exceeding 6 homopolymers were discarded using Mothur
282 v1.36.1 (Schloss et al., 2009). Next, the sequences were subjected to Q-score filtering to
283 remove reads with more than one expected error. Reads occurring only once in the
284 entire dataset (singleton) were then removed. Theses sequences were clustered *de novo*
285 (with a minimum identity of 97 %) into 204 operational taxonomic units (OTUs) among
286 the 18 samples. The taxonomy of the representative OTU sequences was assigned using
287 the RDP classifier (Wang et al., 2007) implemented in QIME v1.9.1 (Caporaso et al.,
288 2010). Non-metric multidimensional scaling (NMDS) plot of the pairwise Bray-Curtis
289 distance matrix were used for the classification of all air samples. Greengenes release
290 13_8 (McDonald et al., 2012) was used as the reference taxonomic database.

291

292 *2.5. Accession numbers*

293 All data obtained from MiSeq sequencing data have been deposited in the
294 DDBJ/EMBL/GenBank database (accession number of the submission is
295 PRJEB17915).

296

297 **3. Results**

298

3.1. Air mass analyses using LIDAR measurements, back trajectories, and metrological data

The vertical distributions of the depolarization ratio determined by LIDAR measurements were assessed for the four sampling events (March 19, 2013; March 20, 2015; April 28, 2013; and March 28, 2014). The depolarization ratio increased at the altitude of 3,000 m on March 19, 2013 (Fig. 2a), while it decreased at the middle altitude of 1,000 m. The air mass on March 20, 2015 showed high values of depolarization ratio at altitudes of 2,500 m and 500 m, consistent with the vertical distribution of non-spherical (mineral dust) particles over the Noto Peninsula (Fig. 2d). A 3-day back trajectory analysis indicated that the air mass at 3,000 m on both sampling dates came from the Asian desert region to the Noto Peninsula (Hakui) immediately across the Sea of Japan (Fig. 3). These results indicated the dust event occurrence on March 19, 2013 was specific to the upper altitude of 3,000 m, while the dust event on March 20, 2015 occurred between the altitudes of 2,500 m and 500 m. Moreover, samples collected on April 28, 2013 and March 28, 2014 exhibited low depolarization ratio (Fig. 2b-c), and the air masses on these two sampling dates came from areas of North Asia, including eastern Siberia (Fig. 3).

The air-sampling periods from the March 2014 time series (from the 23rd to the 29th of March 2014) and the March 2015 time series (from the 16th to the 21st of March 2015) showed different patterns of depolarization ratio and air mass trajectory roots between the two series (Figs. 4 and 5). Depolarization ratio from March 2014 maintained lower values (Fig. 4a) and the trajectory lines changed the roots from eastern Siberia to the Korean Peninsula before surrounding the Japanese islands (Fig. 4c). In

322 contrast, the sampling period during March 2015 had substantially higher depolarization
323 ratio, indicating a strong presence of mineral dust particles (Fig. 5a), and air masses at
324 3,000 m consistently originated from the Asian desert regions (Fig. 5c).
325 Temperatures from March 19, 2013; April 28, 2013; March 28, 2014; and March 20,
326 2015 increased from approximately 290 K to approximately 300 K at middle altitudes
327 (500 m and 1,200 m) (Fig. 2). The temperature profile clearly indicated the presence of
328 a thin boundary under the upper altitudes (2,500 m and 3,000 m), which suggested that
329 there is a difference in air qualities between the middle and upper altitudes (Table 1).
330 During the March 2014 time series, temperatures dynamically changed at altitudes of
331 approximately 1,200 m, while those from the March 2015 time series (the 16th, 17th,
332 and 21st of March 2015) were stable at 1,200 m (Figs. S1 and S2). These results
333 indicate that the boundary layers were located at 1,200 m during the March 2014 time
334 series, whereas the tropospheric air transported by westerly winds was suspended at the
335 sampling altitudes (500 m and 1,200 m) used during the March 2015 time series.
336
337 *3.2. Vertical distributions and sequential variations of aerosol particles*
338 Aerosol particle concentrations from the ground level to the troposphere were
339 measured using OPC to compare the vertical distributions of aerosols from the four
340 sampling events. The OPC-measured particles on March 19, 2013 and March 20, 2015
341 maintained similar concentrations below the troposphere (Fig. 2ad), while the
342 concentrations on April 28, 2013 and March 28, 2014 decreased one or two orders of
343 magnitude between the troposphere and ground level (Fig. 2bc). At high altitudes (2,000
344 m to 2,500 m), the course particles (greater 1.0 μm) observed on March 19, 2013 and

March 20, 2015 were one or two orders of magnitude higher (10^5 to 10^6 particles m^{-3}) than those on April 28, 2013 and March 28, 2014 (no more than 1.2×10^4 particles m^{-3}). The fine particles (0.3 μm to 1.0 μm) showed similar concentrations between the four sampling events, fluctuating between 1.2×10^6 to 3.5×10^7 particles m^{-3}. At lower altitudes (130 m to 510), the aerosol particles had similar concentrations and size distributions between the four sampling periods; the course particle concentration ranged from 8.4×10^5 particles m^{-3} to 1.2×10^6 particles m^{-3}, and the fine particles ranged from 1.3×10^7 particles m^{-3} to 1.2×10^8 particles m^{-3}.

OPC measurements indicated that air samples collected at 1,200 m during the March 2015 time series consistently contained course particles at one or two orders of magnitude higher in concentration (1.4×10^6 to 3.4×10^6 particles m^{-3}) than detected in the March 2014 time series, which had concentrations of no more than 1.8×10^5 particles m^{-3} (Fig. 4b). The concentration of relatively large particles (>5.0 μm) in March 2015 maintained relatively higher concentrations (from 1.4×10^4 to 8.2×10^5 particles m^{-3}) than those observed in March 2014 (no more than 3.74×10^3 particles m^{-3}). In contrast, the fine particles measured in March 2014 and March 2015 fluctuated around similar concentrations ranging from 10^7 to 10^8 particles m^{-3}.

Based on the above observations, the sampled air masses that were influenced by Asian dust events and included dust particles were categorized as “dust samples”. The sampled air masses that were not influenced by dust events or contained less dust particles were categorized as “non-dust samples”, in relation to the presence or absence of dust events as the source of the aerosol samples (Table 1).

368 *3.3. Fluorescent microscopic observation of aerosol particles*

369 Using epifluorescence microscopy with DAPI staining, the aerosol particles in the
370 18 air samples emitted several types of fluorescence, categorized as white, blue, yellow,
371 or black (Fig. S3). White fluorescence particles, (white particles) were indicative of
372 mineral particles originating from the sand or soil. Microbial (prokaryotic) particles
373 stained with DAPI emitted blue fluorescence, forming coccoid- or bacilli-like particles
374 with a diameter <3 μm. Yellow fluorescence particles (yellow particles) stained with
375 DAPI were organic matter and ranged from 1.0 μm to 10 μm in diameter. Most of the
376 yellow particles disappeared in the aerosol-particle suspending solutions after protease
377 treatment, suggesting that the yellow particles consisted mainly of proteins. Black
378 particles were indicative of an anthropogenic black carbon originating from East Asian
379 regions, produced by biomass burning, industrial activities, and vehicle exhaust.

380 The dust samples from upper altitudes (2,500 m and 3,000 m) contained 5 to 100
381 times higher concentrations of microbial, organic, and white particles than the
382 concentrations detected in the non-dust samples (Fig. 2). In the upper altitude dust
383 samples, the concentration of mineral particles ranged from 7.77×10^5 particles m^{-3} to
384 1.08×10^6 particles m^{-3} (Fig. 2ad), whereas the concentrations of the non-dust samples
385 ranged from 3.14×10^4 particles m^{-3} to 1.48×10^5 particles m^{-3} (Fig. 2bc). The
386 microbial particles in the high altitude dust samples exhibited concentrations of
387 approximately 1.5×10^6 particles m^{-3} that were two orders of magnitude higher than in
388 the non-dust samples (approximately 6.0×10^4 particles m^{-3}). The organic particles in
389 the high altitude dust samples were also found at higher concentrations of
390 approximately 4.2×10^6 particles m^{-3} than those from the non-dust samples 13H428-u

391 and 14H328-u, which were 2.12 × 10^4 particles m^{-3} and 5.30 × 10^4 particles m^{-3},
392 respectively. In contrast, the air samples collected at the low altitude of 10 m exhibited a
393 random or stochastic pattern between 10^5 and 10^6 particles m^{-3}, regardless of the
394 sampling dates (Fig. 2). Black particles were observed in the four air samples from 10 m
395 and fluctuated around concentrations of less than 8.48 × 10^4 particles m^{-3}. Finally, the
396 percentage of organic particles out of the total number of particles (organic and
397 microbial particles) in the dust samples 13H319-u, 15H320-u, and 15H320-m ranged
398 between approximately 71.5 % and 73.6 %, which was higher than in the non-dust
399 samples, which ranged from 4.6 % to 46.3 % (Fig. S4).

400 All types of fluorescence particles were also observed in the sequentially collected
401 air samples at 1,200 m in the March 2015 time series (except for 2,500 m on March
402 20th) and the March 2014 series. The dust samples examined from the March 2015
403 series had higher concentrations of total particles than the non-dust samples of the
404 March 2014 series (Figs. 4 and 5). The mineral particles detected in the March 2014
405 series fluctuated at low concentrations from 3.39 × 10^4 particles m^{-3} to 2.62 × 10^5
406 particles m^{-3} (Fig. 4), while in the March 2015 series the mineral particles showed
407 higher values from 1.80 × 10^5 particles m^{-3} to 1.77 × 10^7 particles m^{-3} (Fig. 5). High
408 levels of organic particles were detected in the March 2015 series samples, ranging from
409 3.13 × 10^5 to 3.75 × 10^7 particles m^{-3}, which decreased to below 2.28 × 10^5 particles m^{-3}
410 in the March 2014 series samples. The microbial particle concentrations in the March
411 2015 series samples (ranging from 4.75 × 10^5 to 2.06 × 10^6 particles m^{-3}) were higher
412 than those of in the March 2014 series samples (ranging from 3.31 × 10^5 to 1.25 × 10^6
413 particles m^{-3}). The ratio of organic particles to the total number of organic and microbial

414 particles detected during March 2015 (71.5 % to 95.6 %) were higher than those during
415 March 2014 series (8.0 % to 36.2 %) (Fig. S4). The black particles were randomly
416 observed in all samples from March 2015 and March 2014.
417
418 *3.4. Analysis of bacterial communities using MiSeq sequencing analysis*
419 For the analysis of the prokaryotic composition in the 18 samples, we obtained
420 645,075 merged paired-end sequences with the lengths ranging from 244 bp to 298 bp
421 after quality filtering, and the sequence library size for each sample was normalized at
422 1,500 reads. The 16S rDNA sequences were divided into 204 phylotypes (sequences
423 with >97 % similarity). Phylogenetic assignment of sequences resulted in an overall
424 diversity of 16 phyla and candidate divisions, 32 classes (and class-level candidate taxa),
425 and 72 families (and family-level candidate taxa). The majority (>90 %) of the
426 sequences were represented by 9 bacterial classes and 33 families (Figs. 6 and 7). The
427 bacterial compositions varied during the sampling periods and included the phylotypes
428 belonging to the classes Cyanobacteria, Actinobacteria, Bacilli, Bacteroidetes, SBRH58,
429 and Proteobacteria (Alpha, Beta, Gamma, and Deltaproteobacteria), which are typically
430 generated from atmospheric, terrestrial and marine environments. On the box plots, the
431 numbers of bacterial species estimated by Chao I were similar at average levels between
432 the dust samples and non-dust samples, while the Chao I and Shannon values of the
433 non-dust samples showed a wider range than that of dust samples (Fig. 8a). A
434 non-metric multidimensional scaling (NMDS) plot demonstrated the distinct clustering
435 of prokaryotic communities separating the dust samples and the non-dust samples (Fig.
436 8b). For the PCR-analysis steps, negative controls (no template and template from

437 unused filters) did not contain 16S rDNA amplicons demonstrating the absence of
438 artificial contamination during experimental processes.
439
440 *3.5. Vertical distributions of bacterial communities in dust and non-dust samples*
441 The vertical distributions of bacterial compositions showed different patterns
442 between dust event days and non-dust days (Fig. 6). In the dust samples collected at
443 upper altitudes, phylotypes belonging to the phylum Bacilli accounted for more than
444 60.5 % of the total and were mainly composed of members of the families Bacillaceae
445 and Paenibacilliaceae (Fig. 6). Bacterial numbers from the phylum Bacilli decreased at
446 lower altitudes during dust events, and the phylotypes of Cyanobacteria, Actinobacteria,
447 and Protobacteria increased in relative abundance in the samples collected at middle and
448 low altitudes (13H319-m, 13H319-l, and 15H320-m).
449 Cyanobacteria, Actinobacteria, and Proteobacteria sequences also dominated in the
450 air samples collected during non-dust events (13H428-m, 14H328-u, 14H328-m, and
451 14H328-l). Specifically, Actinobacteria phylotypes increased in their relative abundance,
452 ranging from 14.1 % to 24.7 % in the non-dust samples collected on March 28, 2014.
453 Proteobacteria phylotypes containing several bacterial families occupied a high relative
454 abundance, ranging from 60.5 % to 85.3 % in the non-dust samples 13H428-u,
455 13H428-m, 14H328-u, 14H328-m, and 14H328-l. In particular, the non-dust samples
456 collected on March 28, 2014 included the Alphaproteobacteria phylotypes, which have
457 composed of members of the families Phyllobacteriaceae and Sphingomonadaceae.
458 Most Betaproteobacteria, phylotypes including the families Oxalobacteraceae and
459 Comamonadaceae, were specific to the non-dust samples collected at 1,200 m and 2,500

460 m on April 28, 2013.

461 Cyanobacteria phylotypes, which were randomly detected from both dust samples
462 and non-dust samples, particularly increased in both the non-dust sample collected at 10
463 m on April 28, 2013 and the dust sample collected at 3,000 m on March 20, 2015, with a
464 relative abundance of 15.3 % and 74.6 %, respectively. Bacteroidia phylotypes also
465 randomly appeared in several air samples, regardless of the dust event influences and
466 were present at maximal levels in the non-dust sample 13H319-m, with a relative
467 abundance of 35.6 %.

468

469 *3.6. Variations in bacterial communities during dust events and non-dust events*

470 Sequential variations in the bacterial composition of air samples at altitudes of
471 1,200 m or 2,500 m were compared between dust event periods (March 2015 series) and
472 non-dust periods (March 2014 series). During the March 2015 dust event, phylotypes of
473 the family Bacillaceae in the class Bacilli occupied more than 53.0 % of the relative
474 abundance in the four dust samples collected (Fig. 7). Cyanobacteria phylotypes related
475 to the marine cyanobacterium Synechococcaceae uniquely appeared in the dust samples
476 of the March 2015 series; their abundance fluctuated the values ranging from 12.5 % to
477 14.8 % between the 16th and the 20th of March 2015 before decreasing to 1.5 % on
478 March 20.

479 During the non-dust periods of the March 2014 series at the middle altitude, the
480 relative abundance of Actinobacteria phylotypes belonging to the family
481 Micrococcaceae was occupied 59.9 % on March 23, decreased to 19.5 % on March 24,
482 and disappeared from samples collected on March 29. Corresponding to the decrease in

Actinobacteria phylotypes, Alpha and Gammaproteobacteria phylotypes showed an increasing trend from 30.6 % to 96.8 % between the 23rd and the 29th of March 2014 (Fig. 7a). Alphaproteobacteria phylotypes belonging to the families Sphingomonadaceae, and Phyllobacteriaceae, consistently appeared throughout the sampling periods of the March 2014 series and occupied a maximum relative abundance of 72.9 % and 22.3 % respectively. For Gammaproteobacteria, the Xanthomonadaceae sequences dominated at a relative abundance of 18.3 % and 5.4 % in the non-dust samples 14H325-m and 14H329-m, respectively, during the air mass was suspended the Japanese islands for a few days.

4. Discussion

4.1 Air mass conditions during Asian dust and non-dust events

Westerly winds blowing over East Asia disperse airborne microorganisms associated with dust mineral particles (Maki et al., 2008) and anthropogenic particles (Cao et al., 2014; Wei et al., 2016), influencing the abundances and taxon compositions of airborne bacteria at high altitudes over downwind areas, such as Noto Peninsula (Maki et al., 2013). In this investigation, the increases in aerosol particles (dust particles) and associated microbial particles were observed over the Noto Peninsula during the dust events of March 19, 2013 and March 20, 2015 (Figs. 2 and 4). At the two sampling dates, the air mass including microbial particles had traveled from the Asian desert region throughout the anthropogenic polluted areas (Fig. 2), and the dust particles entered the Japanese troposphere and were maintained at high altitudes (March

506 19, 2013) or mixed with the ground-surface air (March 20, 2015). During non-dust days,
507 the air masses at high altitudes came from several areas, including the eastern region of
508 Siberia, Asian continental coasts (Korean Peninsula), the Sea of Japan, or surrounding
509 Japanese islands, and mixed with ground-surface air over the Noto Peninsula. The air
510 samples collected during dust and non-dust events were valuable for understanding the
511 westerly wind influences on vertical distributions and sequential dynamics of airborne
512 bacteria at high altitudes over the downwind regions.
513
514 *4.2 Aerosol dynamics during Asian dust and non-dust event*
515 The microscopic fluorescence particles of all samples could be separated into four
516 categories: mineral (white), microbial (blue), organic (yellow), and black-carbon (black)
517 particles (Fig. S3), which were observed in the previous air samples collected during
518 dust events (Maki et al., 2015). The amount of microbial particles increased at high
519 altitudes during dust events, suggesting that the dust events directly carried bacterial
520 particles to the troposphere over downwind areas. At low altitudes, similar
521 concentrations of fluorescent particles were observed in air samples collected between
522 dust events (13H319-l) and non-dust events (13H428-l) (Fig. 2) because the dust
523 particles did not reach the ground surface on the dust-event days. In the absence of the
524 influences of dust-events, the aerosols mainly originated from local environments in
525 Japanese areas.
526 Organic particles also increased during dust events and in the ratios between all
527 particles related to the dust events. The organic particles originate from proteins and
528 other biological components (Mostajir et al., 1995). The tropospheric aerosols would be

529 composed of organic particles at high rates ranging from 30 % to 80 % (Jaenicke, 2005),
530 and organic particle concentrations fluctuated from 10^3 to 10^5 particles m^{-3} at high
531 altitudes of 4,000 m above the ground (Twohy et al., 2016). The dead-phase cells of
532 microbial isolates obtained from aerosol samples mainly irradiated yellow fluorescence
533 instead of blue fluorescence (Liu et al., 2014). When fungi (*Bjerkandera adusta*) and
534 bacteria (*Bacillus* spp.) isolated from aerosol samples were incubated, the dead-phase
535 microbial cells mainly irradiated yellow fluorescence instead of blue fluorescence (Liu
536 et al., 2014; Fig. S3). The relative numbers of organic particles to the total number of
537 microbial and organic particles in the dust samples showed significantly higher values
538 (82.9 ± 32.3 %) than in the non-dust samples (23.3 ± 13.7 %) (Fig. S4). Hara and Zhang
539 reported that dust events in Kyushu, Japan, resulted in an increased ratio of damaged
540 microbial cells in the air at the ground-surface and that the ratio increased to
541 approximately 80 % (Hara and Zhang, 2012). Furthermore, organic molecules
542 associated with dust aerosols are reported to be composed of mannitol, glucose, and
543 fructose, which are part of cell components of airborne microorganisms and contribute
544 to the formation of secondary organic aerosols (SOA) (Fu et al., 2016). Microbial cells
545 or their components coming from Asian continents to Japan would be exposed to air at
546 high-altitudes during their long-range transport, increasing the ratios of damaged and
547 dead cells or SOA.

548 The appearance of black carbon most likely originated from anthropogenic
549 activities, such as biomass burning, industrial activities, and vehicle exhaust (Chung and
550 Kim, 2008). In the anthropogenic regions of eastern China, anthropogenic particles
551 originating from human activities are expected to comprise more than 90 % of dust

552 particles (Huang et al., 2015a). When the westerly winds are strongly blowing over the
553 Noto Peninsula, the black carbon particles at upper altitudes (3,000 m) are thought to
554 mainly derive from continental anthropogenic regions.
555
556 *4.3 Comparing the community structures of atmospheric bacteria between Asian dust*
557 *and non-dust events*
558 Dust events and air-pollutant occurrences changed the airborne bacterial
559 communities over the downwind areas, such as Beijing (Jeon et al., 2011; Cao et al.,
560 2014) and east Mediterranean areas (Mazar et al., 2016). The westerly winds blowing
561 over East Asia would transport airborne bacteria to the high-altitude atmosphere over
562 the Noto Peninsula (Maki et al., 2015) and North American mountains (Smith et al.,
563 2013). Our box plots analysis suggested that changes in the bacterial diversity in the
564 dust samples would be more stable than in the non-dust samples (Fig. 8a). Furthermore,
565 using a NMDS plot, the bacterial compositions in the dust samples could be
566 distinguished from non-dust samples (Fig. 8b). Thus, the aerosol particles transported
567 by Asian dust events changed the atmospheric bacterial composition at higher altitudes
568 over downwind areas.
569 The phylotypes in the dust samples were predominately clustered into the class
570 Bacilli (Fig. 4a), while the non-dust samples mainly included the phylotypes of the
571 classes Alpha, Beta, and Gammaproteobacteria and Actinobacteria. Our previous
572 investigations indicated that the bacterial communities at an altitude of 3,000 m over the
573 Noto Peninsula included more than 300 phylotypes, which were predominantly
574 composed of Bacilli phylotypes (Maki et al., 2015). Bacterial groups belonging to

575 Bacilli, Proteobacteria, and Actinobacteria have been reported as airborne bacteria
576 around European mountains (Vaïtilingom et al., 2012) as well as over Asian rural
577 regions (Woo et al., 2013). Some Bacilli isolates were found to act as ice-nucleating
578 agents and to be involved in ice cloud (Matulova et al., 2014; Mortazavi et al., 2015).
579 Isolates of Gammaproteobacteria isolates were obtained from mineral dust particles
580 (Hara et al., 2016a), glaciated high-altitude clouds (Sheridan et al., 2003), and plant
581 bodies (Morris et al., 2008), and some isolate species, such as *Pseudomonas*, were
582 confirmed to have the ice-nucleation activity. Accordingly, Bacilli and Proteobacteria
583 members associated with dust events could potentially contribute to climate change
584 resulting from dust events.
585
586 *4.4 Dominant bacterial populations in the air masses transported from Asian continents*
587 In some dust-event samples collected at high altitudes (13H319-u, 15H320-u, and
588 15H320-m), Bacilli sequences accounted for more than 52.7 % of the total number of
589 sequences (Fig. 6). Back trajectories on March 19, 2013 and March 20, 2015 came from
590 the Asian desert region to the Noto Peninsula. Some *Bacillus* species were
591 predominantly detected at high altitudes above the Taklimakan Desert (Maki et al.,
592 2008) and above downwind areas during Asian dust events (Maki et al., 2010 and 2013;
593 Smith et al., 2013; Jeon et al., 2011; Tanaka et al., 2011). *Bacillus* species are the most
594 prevalent isolates obtained from mineral dust particles collected over downwind areas
595 (Hua et al., 2007; Gorbushina et al., 2007).
596 Bacilli members can form resistant endospores that support their survival in the
597 atmosphere (Nicholson et al., 2000). The *Bacillus* isolates obtained from atmospheric

598 samples showed higher-level resistance to UV irradiation than normal isolates
599 (Kobayashi et al., 2015). In the Gobi Desert, dust events increase the diversity of
600 airborne microbial communities; after dust events, spore-forming bacteria, such as
601 *Bacillus*, increase in their relative abundances (Maki et al., 2016). Accordingly, in the
602 atmosphere, selected Bacilli members associated with dust particles would be
603 transported over long distances.

604 The Bacilli sequences showed different vertical variations between the two dust
605 events on March 19, 2013 and March 20, 2015. On March 19, 2013 (13H319-m), the
606 relative abundances of Bacilli sequences decreased dynamically from 3,000 m to 1,200
607 m, while unstable atmospheric layers on March 20, 2015 most likely mixed the
608 long-range transported bacteria with the regional bacteria over the Noto Peninsula. A
609 previous investigation also demonstrated the vertical mixture of airborne bacteria over
610 Suzu in the Noto Peninsula (Maki et al., 2010).

611 Actinobacteria sequences decreased in relative abundance between the 23rd and
612 29th of March 2014 corresponding with changes in the air mass trajectory roots from
613 north Asian regions, such as eastern Siberia and Japan (Fig. 7). Furthermore,
614 Actinobacteria sequences appeared in the samples collected from air masses that were
615 transported throughout the Korean Peninsula on March 19, 2013; April 28, 2013; and
616 March 20, 2015. Actinobacteria members are frequently dominant in terrestrial
617 environments but seldom survive in the atmosphere for a long time, because they cannot
618 form spores (Puspitasari et al., 2015). However, the family Micrococcaceae in
619 Actinobacteria was primarily detected from anthropogenic particles collected in Beijing,
620 China (Cao et al., 2014). Over anthropogenic source regions for Asian continents,

621 anthropogenic particles occupy more than 90 % of dust particles and originate from
622 soils disturbed by human activities in cropland, pastureland, and urbanized regions
623 (Huang et al., 2015a; Guan et al., 2016). Air masses transported from the continental
624 coasts are expected to include a relatively high abundance of Actinobacteria members
625 associated with anthropogenic particles.

626 Natural dust particles from Asian desert areas (Taklimkan and Gobi Deserts) are
627 transported in the free troposphere (Iwasaka et al., 1988) and vertically mixed with
628 anthropogenic particles during the transportation processes (Huang et al., 2015a). In
629 some cases, short-range transport of air masses would carry only anthropogenic
630 particles to Japan, because the anthropogenic particles are often dominant in Asian
631 continental coasts (Huang et al., 2015a). Actinobacteria members may have been
632 transported with anthropogenic particles from continental coasts.

633

634 *4.5 Dominant bacterial populations in the air masses originated from marine*
635 *environments and Japanese islands*

636 Proteobacteria sequences increased in their relative abundances at high altitudes
637 during non-dust sampling dates (13H428-u, 13H428-m, 14H328-u, 14H328-m, and
638 March 2014 series), when air mass origins at 1,200 m changed from the Korean
639 Peninsula to Japan (Fig. 7). Proteobacteria members were the dominate species in the
640 atmosphere over mountains (Bowers et al., 2012; Vaïtilingom et al., 2012; Temkiv et al.,
641 2012), in the air samples collected on a tower (Fahlgren et al., 2010), and from the
642 troposphere (DeLeon-Rodriguez et al., 2013; Kourtev et al., 2011). In the phylum
643 proteobacteria, the families Phyllobacteriaceae, Methylobacteriaceae, and

644 Xanthomonadaceae were predominately detected from the non-dust samples and are
645 associated with plant bodies or surfaces (Mantelin et al., 2006; Fürnkranz et al., 2008;
646 Khan and Doty, 2009; Fierer and Lennon, 2011). The Betaproteobacteria sequences in
647 the non-dust samples mainly contained the Oxalobacteraceae and Comamonadaceae
648 families, which are commonly dominate in freshwater environments (Nold and Zwart,
649 1998) as well as on plant leaves (Redford et al., 2010). In addition, the class
650 Alphaproteobacteria in the non-dust samples also included marine bacterial sequences
651 belonging to the family Sphingomonadaceae (Cavicchioli et al., 2003). Bacterial
652 populations originating from marine areas are prevalent in cloud droplets (Amato et al*.,*
653 2007), in air samples collected from the seashores of Europe (Polymenakou et al., 2008),
654 in storming troposphere (DeLeon-Rodriguez et al., 2013), and at high altitudes in
655 Japanese regions (Maki et al., 2014), suggesting that the marine environments represent
656 a major source of bacteria in clouds. The air masses suspended over the Sea of Japan or
657 Japanese islands during non-dust events (the March 2014 series) could include a high
658 relative abundance of airborne bacteria, which were transported from the surface-level
659 air over the marine environments and the regional phyllosphere.
660
661 *4.6. Bacterial populations commonly detected during dust events and the non-dust*
662 *events*
663 Sequences originating from Synechococcaceae (in the class Cyanobacteria)
664 randomly appeared in the MiSeq sequencing databases results obtained from air samples,
665 regardless of dust event occurrences. *Synechococcus* species in the family
666 Synechococcaceae can eliminate excess peroxide from photosynthesis to resist UV

radiation and oxygenic stress (Latifi et al., 2009), suggesting that these bacteria resist environmental stressors in the atmosphere. In a previous study, the air samples transported from marine environments to Japan predominately contained *Synechococcus* species (Maki et al., 2014), which were dominant marine bacteria in the Sea of Japan and the East China Sea (Choi and Noh, 2009). The cloud water at approximately 3,000 m above ground level was also dominated by Cyanobacteria populations, indicating their atmospheric transport (Kourtev et al., 2011). In addition to Alphaproteobacteria, marine cyanobacterial cells can be transported from seawater to the atmosphere, thereby contributing to the airborne bacterial variations over the Noto Peninsula. Marine bioaerosols originated from cyanobacteria and gram-negative bacteria (including Alphaproteobacteria) are reported to contribute the increase of endotoxin levels in coastal areas influencing human health by inflammation and allergic reaction (Lang-Yona et al., 2014).

Bacteroidetes sequences were detected in some air samples collected during Asian dust and non-dust events. Members of the phylum Bacteroidetes, which were composed of the families Cytophagaceae, associate with organic particles in terrestrial and aquatic environments (Turnbaugh et al., 2011; Newton et al., 2011). Furthermore, these bacterial populations dominate the atmosphere and sand of desert areas, where plant bodies and animal feces are sparsely present (Maki et al., 2016). These bacterial groups possibly originated from organic-rich microenvironments in several areas, such as desert and marine areas.

5. Conclusion

690 Air samples including airborne bacteria were sequentially collected at high
691 altitudes over the Noto Peninsula during dust events and non-dust events. The sampled
692 air masses could be categorized based on sample types with (dust samples) and without
693 (non-dust samples) dust event influences. Bacterial communities in the air samples
694 displayed different compositions between dust events and non-dust events. The dust
695 samples were dominated by terrestrial bacteria, such as Bacilli, which are thought to
696 originate from the central desert regions of Asia, and the bacterial compositions were
697 similar between the dust samples. In contrast, the air masses of non-dust samples came
698 from several areas, including northern Asia, continental coasts, marine areas, and Japan
699 regional areas, showing different variations in bacterial compositions between the
700 sampling dates. Some scientists have attempted to apply airborne bacterial composition
701 as tracers of air mass sources at ground level (Bowers et al., 2011; Mazar et al., 2016).
702 In this study, the terrestrial bacteria, such as Bacilli and Actinobacteria members (Bottos
703 et al., 2014), were dominant populations in the air samples transported from Asian
704 continental areas. The air samples when the air mass was suspended around Japanese
705 islands, mainly included the members of the classes Alpha (Phyllobacteriaceae and
706 Methylobacteriaceae), Gamma, and Betaproteobacteria, which are commonly
707 dominated in phyllosphere (Redford et al., 2010; Fierer and Lennon, 2011) or
708 freshwater environments (Nold and Zwart, 1998). The atmospheric aerosols transported
709 via marine areas include a high relative abundances of marine bacteria belonging to
710 classes Cyanobacteria (Choi and Noh, 2009) and Alphaproteobacteria
711 (Sphingomonadaceae) (Cavicchioli et al., 2003). This study suggested that bacterial
712 compositions in the atmosphere can be used as air mass tracers, which could identify the

levels of mixed air masses transported from different sources.

However, one limitation of our investigation is that the number of samples analyzed was not sufficient to cover entire changes in airborne bacteria at high altitudes over the Noto Peninsula. Although the airborne bacterial composition during non-dust periods was found to change dynamically, only a few types of variation were followed in this investigation. In the future, greater numbers of samples, which are sequentially collected at high altitudes using this sampling method, will need to be originated to more accurately evaluate bioaerosol tracers. Since helicopter sampling procedures require sophisticated techniques and are expensive, the sample numbers at high altitudes are difficult to increase. Air sampling at high altitudes should be combined with sequential ground-air sampling to advance the understanding of the influence of westerly winds on airborne bacterial dynamics in downwind areas. Metagenomic analyses and microbial culture experiments would also provide valuable information about airborne microbial functions relating to ice-nucleation activities, chemical metabolism, and pathogenic abilities.

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743

744 **Competing Interests**

745 The authors declare that they have no conflict of interest.
746

747 **References**

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Figure Legends

Fig. 1. Sampling location (a) and helicopter flight routes during the sampling periods on March 19, 2013, and April 28, 2013 (b); the 23rd, 24th, 25th, and 29th of March 2014 (c); and the 16th, 17th, 20th, and 21st of March 2015 (d).

Fig. 2. LIDAR observation of the depolarization ratio in Toyama city as well as vertical changes in temperature, relative humidity, and potential temperature, and vertical distributions of concentrations of OPC-counted particles and DAPI-stained particles from the four sampling events on March 19, 2013 (a); April 28, 2013 (b); March 28, 2014 (c); and March 20, 2015 (d). The red circles in the LIDAR images indicate that the sampling air included dust mineral particles (solid line) or that dust-event influences are absent at the altitudes on the sampling time (dotted line). OPC-counted particles were categorized according to diameter sizes of 0.3–0.5 μm (closed squares), 0.5–0.7 μm (closed triangles), 0.7–1.0 μm (closed circles), 1.0–2.0 μm (closed diamonds), 2.0–5.0 μm (crosses), and >5.0 μm (open circles). DAPI-stained particles were classified into microbial particles (blue bars), white particles (white bars), yellow fluorescent particles (yellow bars), and black carbon (gray bars).

Fig. 3. Trajectories 3 days ago of aerosols that arrived at 2,500 m (blue-type lines) and 1,200 m (red-type lines) in Hakui, Japan, every hour for 5 h before the completion of sampling time at the four dates; March 19, 2013; April 28, 2013; March 28, 2014; and March 20, 2015.

Fig. 4. (a) LIDAR observation of the depolarization ratio in Toyama city and concentrations of OPC-counted particles and DAPI-stained particles during no-dust days from 0:00 UTC on March 23 to 0:00 UTC on March 30, 2014. The red circles with dotted lines in the LIDAR images indicate dust-event influences are absent at the altitudes on the sampling time. (b) OPC-counted particles were categorized according to diameter sizes of 0.3–0.5 μm (closed squares), 0.5–0.7 μm (closed triangles), 0.7–1.0 μm (closed circles), 1.0–2.0 μm (closed diamonds), 2.0–5.0 μm (crosses), and >5.0 μm (open circles). DAPI-stained particles were classified into microbial particles (blue bars), white particles (white bars), yellow particles (yellow bars), and black particles (gray bars). (c) Trajectories 3 days ago of aerosols that arrived at 2,500 m (blue-type lines) and 1,200 m (red-type lines) in Hakui, Japan, every hour for 5 h before the completion of sampling time during sampling periods on the 23rd, 24th, 25th, 28th, and 29th of March 2014.

Fig. 5. (a) LIDAR observation of the depolarization ratio in Toyama city and concentrations of OPC-counted particles and DAPI-stained particles during dust event days from 0:00 UTC on March 16 to 0:00 UTC on March 23, 2015. The red circles with solid lines in the LIDAR images indicate that the sampling air included dust mineral particles. (b) OPC-counted particles were categorized based on diameter sizes of 0.3–0.5 μm (closed squares), 0.5–0.7 μm (closed triangles), 0.7–1.0 μm (closed circles), 1.0–2.0 μm (closed diamonds), 2.0–5.0 μm (crosses), and >5.0 μm (open circles). DAPI-stained particles were classified into microbial particles (blue bars), white

1072 particles (white bars), yellow particles (yellow bars), and black particles (gray bars). (c)
1073 Trajectories 3 days ago of aerosols that arrived at 2,500 m (blue-type lines) and 1,200 m
1074 (red-type lines) in Hakui, Japan, every hour for 5 h before the completion of sampling
1075 time during sampling periods on the 16th, 17th, 20th, and 21st of March 2015.
1076
1077 Fig. 6. Vertical variations in bacterial compositions at (a) the class level and (b) the
1078 family level of the partial sequences obtained in the MiSeq sequencing database (ca.
1079 400 bp) obtained from air samples collected at different altitudes over the Noto
1080 Peninsula at dust-event days (March 19, 2013; March 20, 2015) and non-dust-event
1081 days (March 19, 2013; March 20, 2015).
1082
1083 Fig. 7. Changes in bacterial compositions at (a) the class level and (b) the family level
1084 of the partial sequences obtained in the MiSeq sequencing database (ca. 400 bp) from
1085 air samples collected at altitudes of 1,200 m (except for the sample collected at 500 m
1086 on March 20, 2015) over the Noto Peninsula during dust-event days from the 16th to the
1087 23rd of March 2015 and during non-dust-event days from the 23rd to the 29th of March
1088 2014.
1089
1090 Fig. 8. Comparison of the bacterial compositions among all air samples collected over
1091 the Noto Peninsula. (a) Box plots of Chao 1 and Shannon analyses indicating the
1092 bacterial diversity observed in dust samples and non-dust samples. Species were binned
1093 at the 97 % sequence similarity level. (b) NMDS of the pairwise Bray-Curtis distance
1094 matrix displaying clustering by all the air samples. Red indicates the samples that were

collected during dust-events and blue indicates those collected during non-dust-events as determined by meteorological data. Circle indicates that the sample contained dust particles as identified via microscopic observation, and triangle indicates that dust particles were absent from the sample. The confidence ellipses are based on a multivariate t-distribution, and represents the 95 % confidence interval of the occurrence of dust vs. non-dust events when the samples were collected.

Table 1 Sampling information during the sampling periods.

Sample name	Sampling date	Collection time (JST)	Total time (min)	Air volume	Sampling method	Sampling location[*1]	Free troposphere[*2]
13H319-u	19 March 2013	14:04 − 15:04	60	700 L	helicopter	2500m	FT
13H319-m		15:19 − 16:19	60	700 L	helicopter	1200m	ABL
13H319-l		14:25 − 15:25	60	700 L	building	10m	GL
13H428-u	28 April 2013	12:10 − 13:04	56	653 L	helicopter	2500m	FT
13H428-m		13:13 − 14:03	50	583 L	helicopter	1200m	ABL
13H428-l		12:03 − 13:03	60	700 L	building	10m	GL
14H328-u	28 March 2014	12:50 − 13:50	60	700 L	helicopter	3000m	FT
14H328-m		14:04 − 15:04	60	700 L	helicopter	1200m	ABL
14H328-l		13:00 − 14:00	60	700 L	building	10m	GL
15H320-u	20 March 2015	12:26 − 13:23	47	548 L	helicopter	2500m	FT
15H320-m		13:39 − 14:40	60	711 L	helicopter	500m	ABL
14H323-m	23 March 2014	10:45 − 11:02	17	11.1 L	helicopter	1200m	ABL
14H324-m	24 March 2014	9:09 − 9:30	21	13.7 L	helicopter	1200m	ABL
14H325-m	25 March 2014	9:31 − 9:50	29	18.9 L	helicopter	1200m	ABL
14H328-m	28 March 2014	14:04 − 15:04	60	700 L	helicopter	1200m	ABL
14H329-m	29 March 2014	9:06 − 9:24	15	9.75 L	helicopter	1200m	PT
15H316-m	16 March 2015	11:21 − 11:43	22	14.3 L	helicopter	1200m	FT
15H317-m	17 March 2015	11:04 − 11:31	27	17.6 L	helicopter	1200m	FT
15H320-u	20 March 2015	12:26 − 13:23	47	548 L	helicopter	2500m	FT
15H321-m	21 March 2015	15:35 − 15:55	20	13.0 L	helicopter	1200m	FT

***1 Height above the ground.**

***2 Free troposhere: FT, Atmospheric boundary layer: ABL, Phase transiens: PT, GL: Ground level**

Table 2. Researches targeting bacterial communities associated with Asian-dust events

Sampling area[*1]	Sample	Location	Altitudes (m)	Sampling place	Sampling method	Analytical method for microorganisms	Dominated bacteria[*2] 1st	2nd	3rd	references
Dust source area	Soil	Taklamakan Desert, China	0	Ground surface	soil sampling	clone libarary	Bacteroidetes (Sphingobacteriia)	Actinobacteria (Actinobacteria)	Proteobacteria (Alpha, Beta, Gamma)	Yamaguchi et al. 2012
Dust source area	Soil	Gobi Desert, China	0	Ground surface	soil sampling	clone libarary	Actinobacteria (Actinobacteria)	Proteobacteria (Beta)	Bacteroidetes (Sphingobacteriia)	Yamaguchi et al. 2012
Dust source area	Soil	Taklamakan Desert, China	0	Ground surface	soil sampling	pyrosequencing	Firmicutes (Bacilli)†	Actinobacteria	Proteobacteria (Gamma)	An et al. 2013
Dust source area	Soil	Gobi Desert, China	0	Ground surface	soil sampling	pyrosequencing	Firmicutes (Bacilli)†	Proteobacteria (Gamma)	Bacteroidetes	An et al. 2013
Dust source area	Soil	Taklamakan, China	0	Ground surface	soil samples	clone libarary	Actinobacteria (Actinobacteria)	Firmicutes (Bacilli)	Proteobacteria	Puspitasari et al. 2016
Dust source and deposition area	Soil	Loess plateau, China	0	Ground surface	soil sampling	clone libarary	Proteobacteria (Beta, Gamma)	Actinobacteria (Actinobacteria)	Bacteroidetes (Sphingobacteriia)	Yamaguchi et al. 2012
Dust source and deposition area	Soil	Loess plateau, China	0	Ground surface	soil sampling	PCR-DGEE	Proteobacteria	Bacteroidetes	Gemmatimonadetes	Kenzaki et al. 2010
Dust source area	Air	Tsogt-Ovoo, Mongolia	3	Ground surface	filtration	MiSeq sequencing	Proteobacteria (Alpha)	Firmicutes (Bacilli)	Actinobacteria (Actinobacteria)	Maki et al. 2017
Dust source area	Air	Dunhuang, China	10	Top of building	filtration	clone libarary	Firmicutes (Bacilli)†	Proteobacteria	Bacteroidetes	Puspitasari et al. 2016
Dust source area	Air	Dunhuang, China	800	Balloon	filtration	PCR-DGEE	Firmicutes (Bacilli)†	-	-	Maki et al. 2008
Dust source area	Air	Dunhuang, China	800	Balloon	filtration	clone libarary	Proteobacteria (Gamma)	Firmicutes (Bacilli)	-	Kakikawa et al. 2009
Dust deposition area	Air	Noto peninsula, Japan	3000	Aircraft	filtration	clone libarary	Firmicutes (Bacilli)†	Bacteroidetes (Bacteroidia)	Proteobacteria (Gamma)	Maki et al. 2013
Dust deposition area	Air	Noto peninsula, Japan	3000	Aircraft	filtration	MiSeq sequencing	Firmicutes (Bacilli)†	Actinobacteria (Actinobacteria)	Proteobacteria (Alpha&Beta)	Maki et al. 2015
Dust deposition area	Air	Mt. Bachelor Observatory, USA	2700	Mt. Bachelor	filtration	culture	Firmicutes (Bacilli)†	Actinobacteria (Actinobacteria)	Proteobacteria (Gamma)	Smith et al. 2012
Dust deposition area	Air	Mt. Bachelor Observatory, USA	2700	Mt. Bachelor	filtration	Microarray	Proteobacteria (Beta&Gamma)	Actinobacteria (Actinobacteria)	Firmicutes (Bacilli)†	Smith et al. 2013
Dust deposition area	Snow	Mt. Tateyama, Japan	2450	Mt. Tateyama	Snow sampling	PCR-DGEE	Firmicutes (Bacilli)†	Proteobacteria (Beta, Gamma)	Actinobacteria (Actinobacteria)	Tanaka et al. 2011
Dust deposition area	Snow	Mt. Tateyama, Japan	2450	Mt. Tateyama	Snow sampling	PCR-DGEE	Firmicutes (Bacilli)†	Proteobacteria (Beta)	Actinobacteria (Actinobacteria)	Maki et al. 2011
Dust deposition area	Air	Noto peninsula, Japan	1200	Helicopter	filtration	MiSeq sequencing	Firmicutes (Bacilli)†	Proteobacteria (Alpha, Gamma)	Cyanobacteria	This study
Dust deposition area	Air	Suzu, Japan	1000	Balloon	filtration	MiSeq sequencing	Firmicutes (Bacilli)†	Proteobacteria (Alpha)	Deinococcus-Thermus (Deinococci)	Maki et al. 2015
Dust deposition area	Air	Osaka, Japan	900	Air craft	filtration	clone libarary	Firmicutes (Bacilli)	Bacteroidetes (Sphingobacteriia)	Actinobacteria (Actinobacteria)	Yamaguchi et al. 2012
Dust deposition area	Air	Suzu, Japan	800	Balloon	filtration	clone libarary	Firmicutes (Bacilli)†	Bacteroidetes (Bacteroidia)	Proteobacteria (Gamma)	Maki et al. 2013
Dust deposition area	Air	Suzu, Japan	600	Balloon	filtration	PCR-DGEE	Firmicutes (Bacilli)†	-	-	Maki et al. 2010
Dust deposition area	Air	Seoul, South Korea	25	Top of building	liquid impiger	pyrosequencing	Actinobacteria (Actinobacteria)	Proteobacteria (Alpha, Gamma)	Firmicutes (Bacilli)†	Cha et al. 2017
Dust deposition area	Air	Osaka, Japan	20	Top of building	filtration	pyrosequencing	Actinobacteria (Actinobacteria)	Cyanobacteria	Acidobacteria (Acidobacteria)	Park et al. 2016
Dust deposition area	Air	Seoul, South Korea	17	Top of building	filtration	PCR-DGEE	Actinobacteria (Actinobacteria)	Firmicutes (Bacilli)†	Proteobacteria (Gamma)	Lee et al. 2011
Dust deposition area	Air	Beijing, China	15	Top of building	filtration	pyrosequencing	Firmicutes (Bacilli)	Proteobacteria (Gamma)	Bacteroidetes (Flavobacteriia)	Wei et al. 2016
Dust deposition area	Air	Beijing, China	10	Top of building	filtration	HiSeq sequencing	Actinobacteria (Actinobacteria)	Proteobacteria (Alpha, Beta, Gamma)	Chloroflexi (Thermomicrobia)	Cao et al. 2014
Dust deposition area	Air	Seoul, South Korea	10	Top of building	filtration	clone libarary	Firmicutes (Bacilli)†	Actinobacteria	Bacteroidetes	Jeon et al. 2011
Dust deposition area	Air	Suzu, Japan	10	Top of building	filtration	MiSeq sequencing	Firmicutes (Bacilli)†	Deinococcus-Thermus (Deinococci)	Proteobacteria (Alpha)	Maki et al. 2015
Dust deposition area	Air	Goyang, South Korea	-	Top of building	filtration	pyrosequencing	Actinobacteria (Actinobacteria)	Proteobacteria (Gamma)	Firmicutes (Bacilli)†	Cha et al. 2016
Dust deposition area	Air	Kanazawa, Japan	10	Roof of building	filtration	MiSeq sequencing	Firmicutes (Bacilli)†	Cyanobacteria	Proteobacteria (Alpha)	Maki et al. 2014
Dust deposition area	Air	Western Pacific Ocean	-	Ship board	filtration	pyrosequencing	Firmicutes (Bacilli)†	Proteobacteria (Beta, Gamma)	Cyanobacteria	Xia et al. 2015

***1 Dust source area: the areas providing dust mineral particles, Dust deposition area: the area where the dust mineral paticles deposit**

***2 The bacterial phyla in the orders of large abundance rates in each samples.**

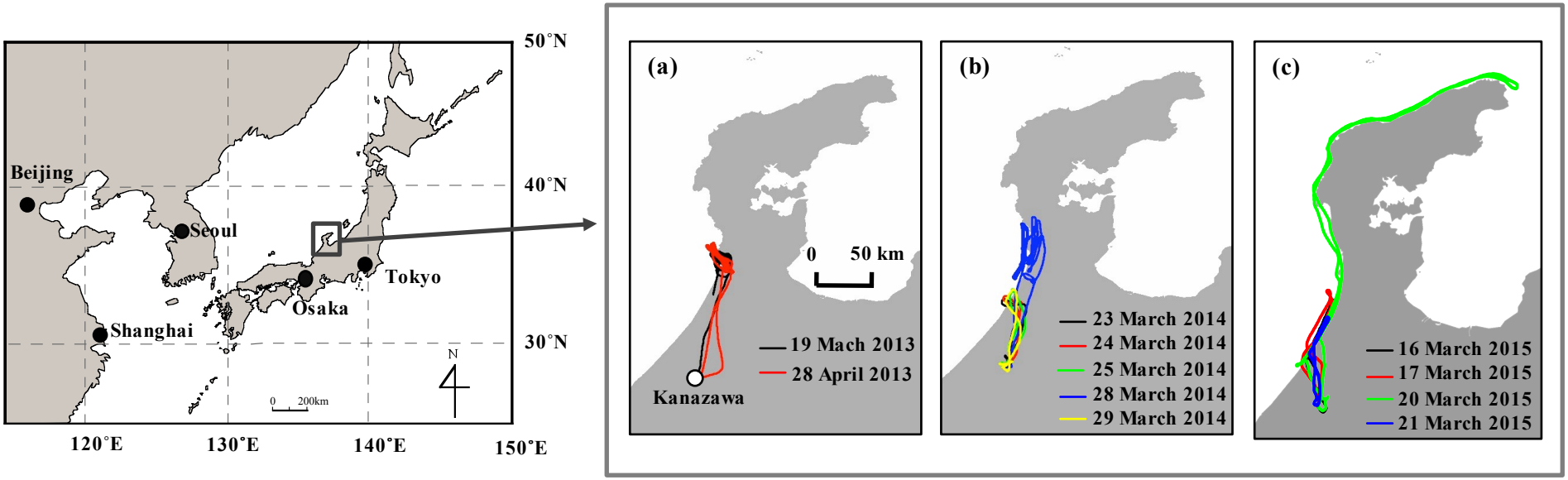
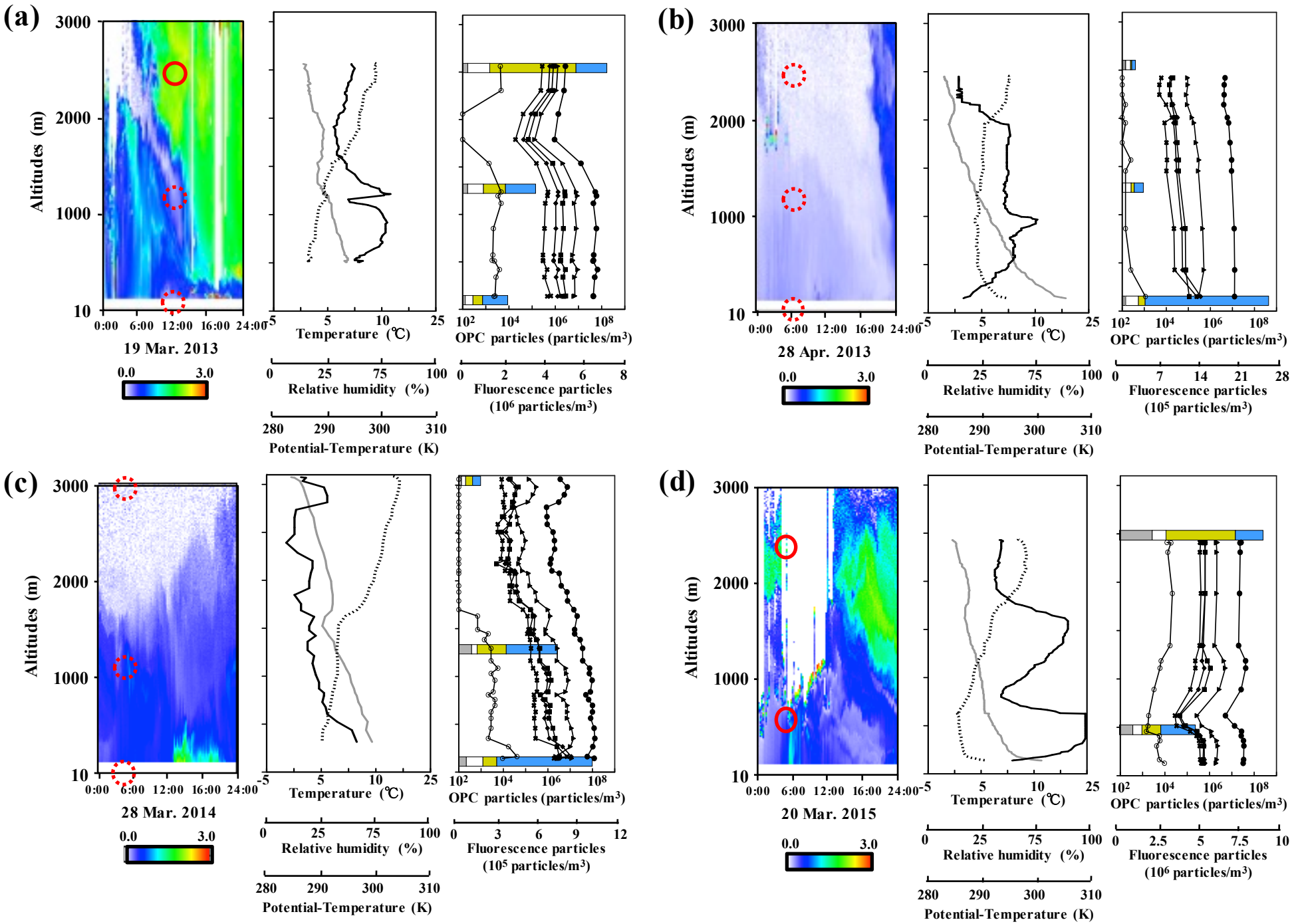


Fig. 1 T. Maki et al.



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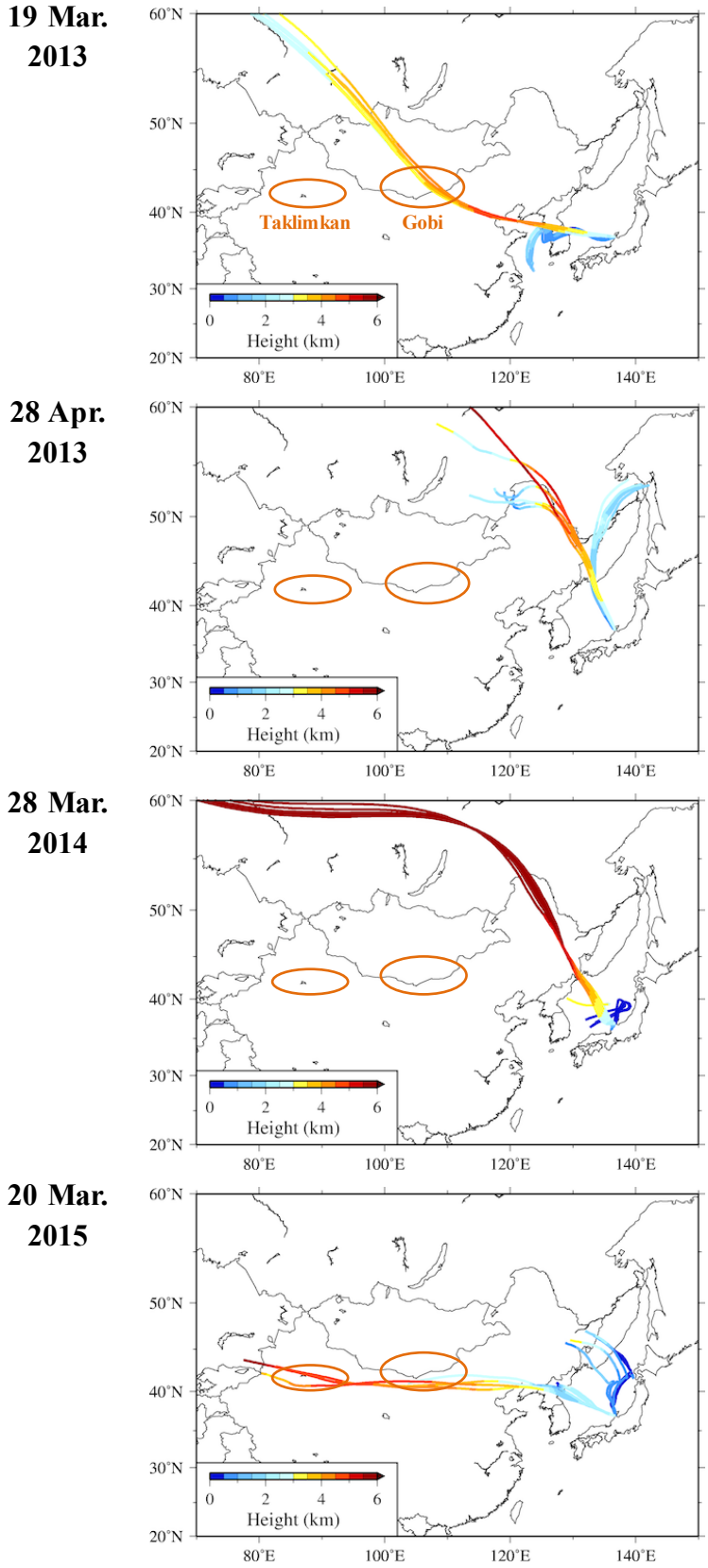

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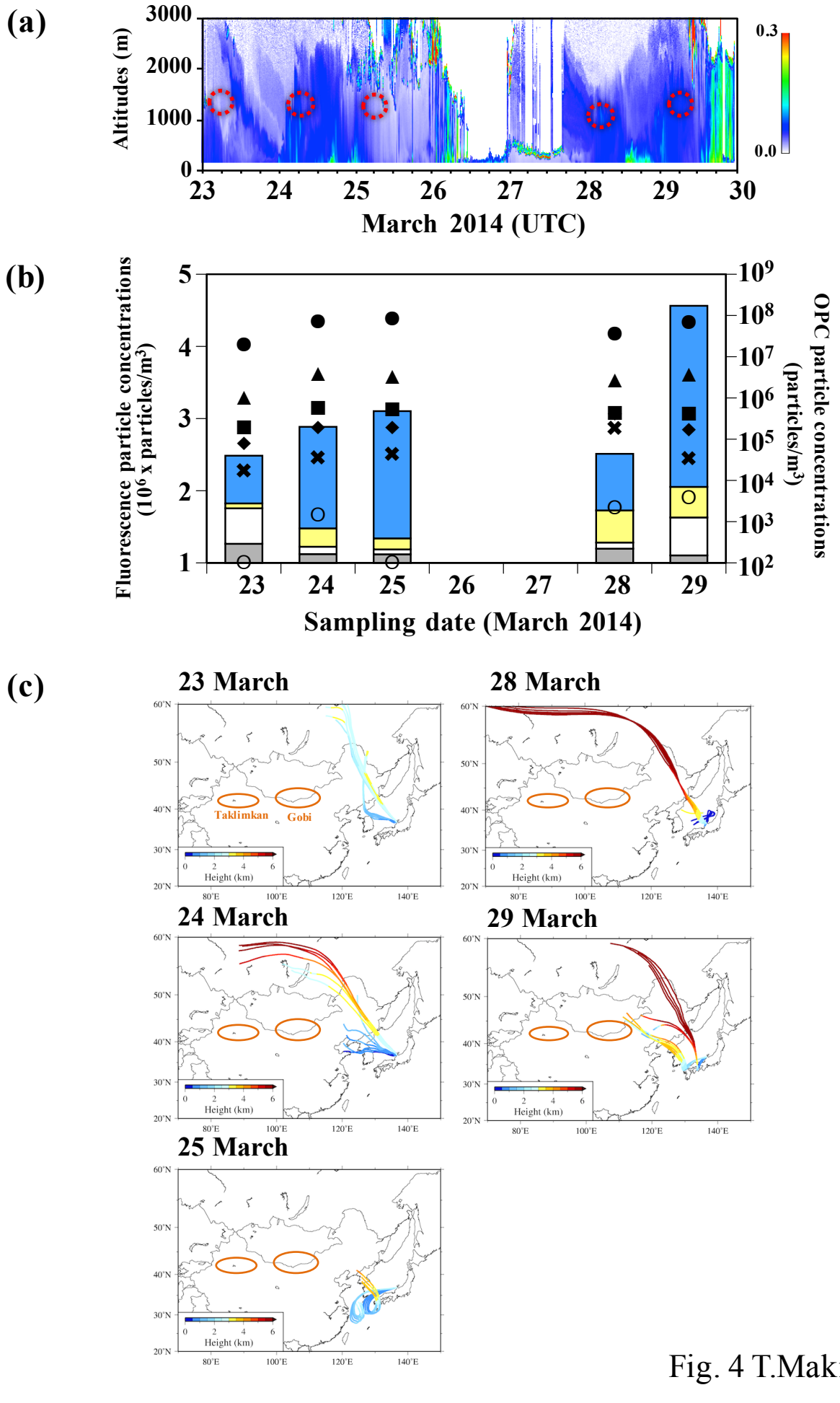

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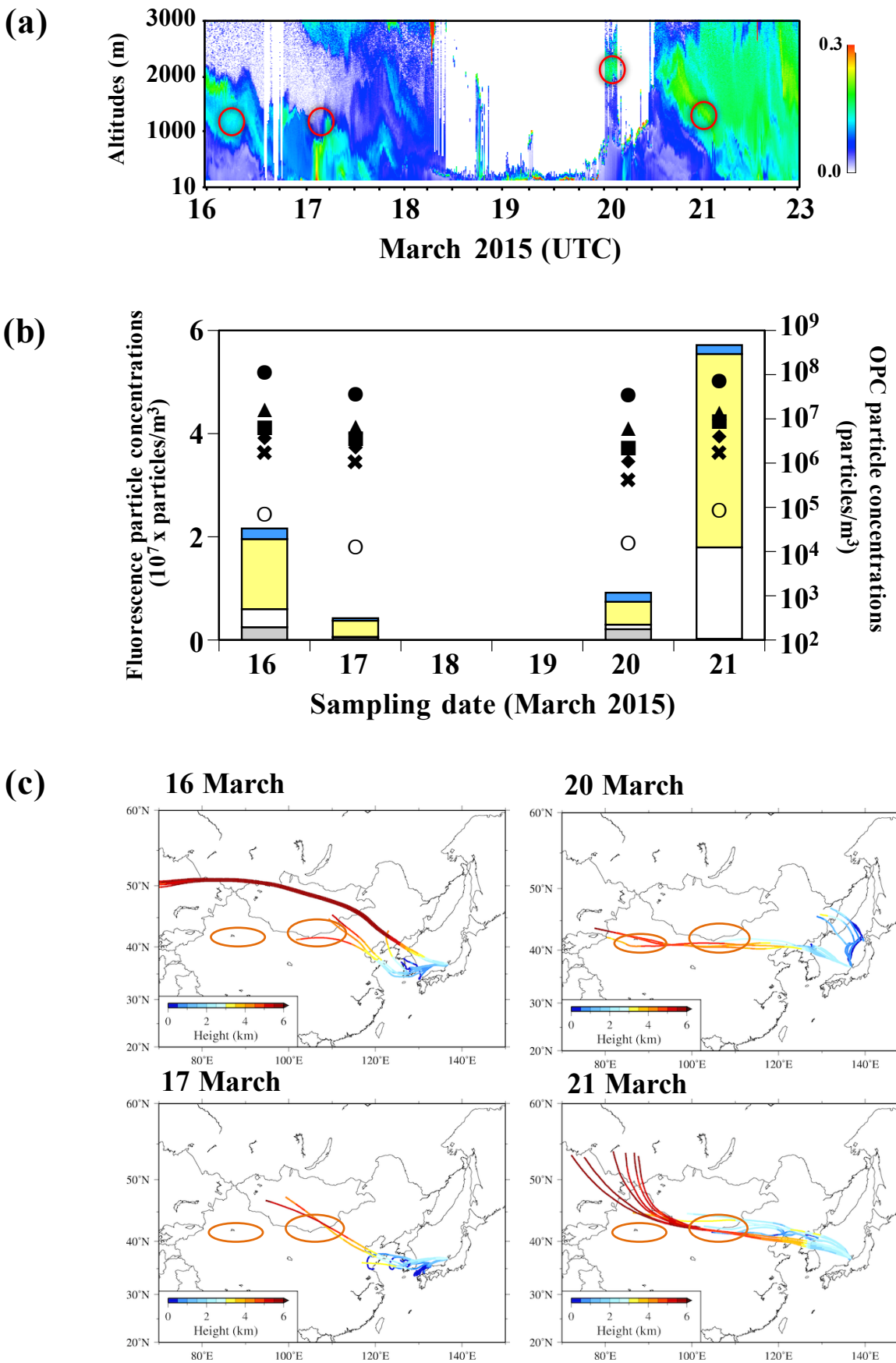

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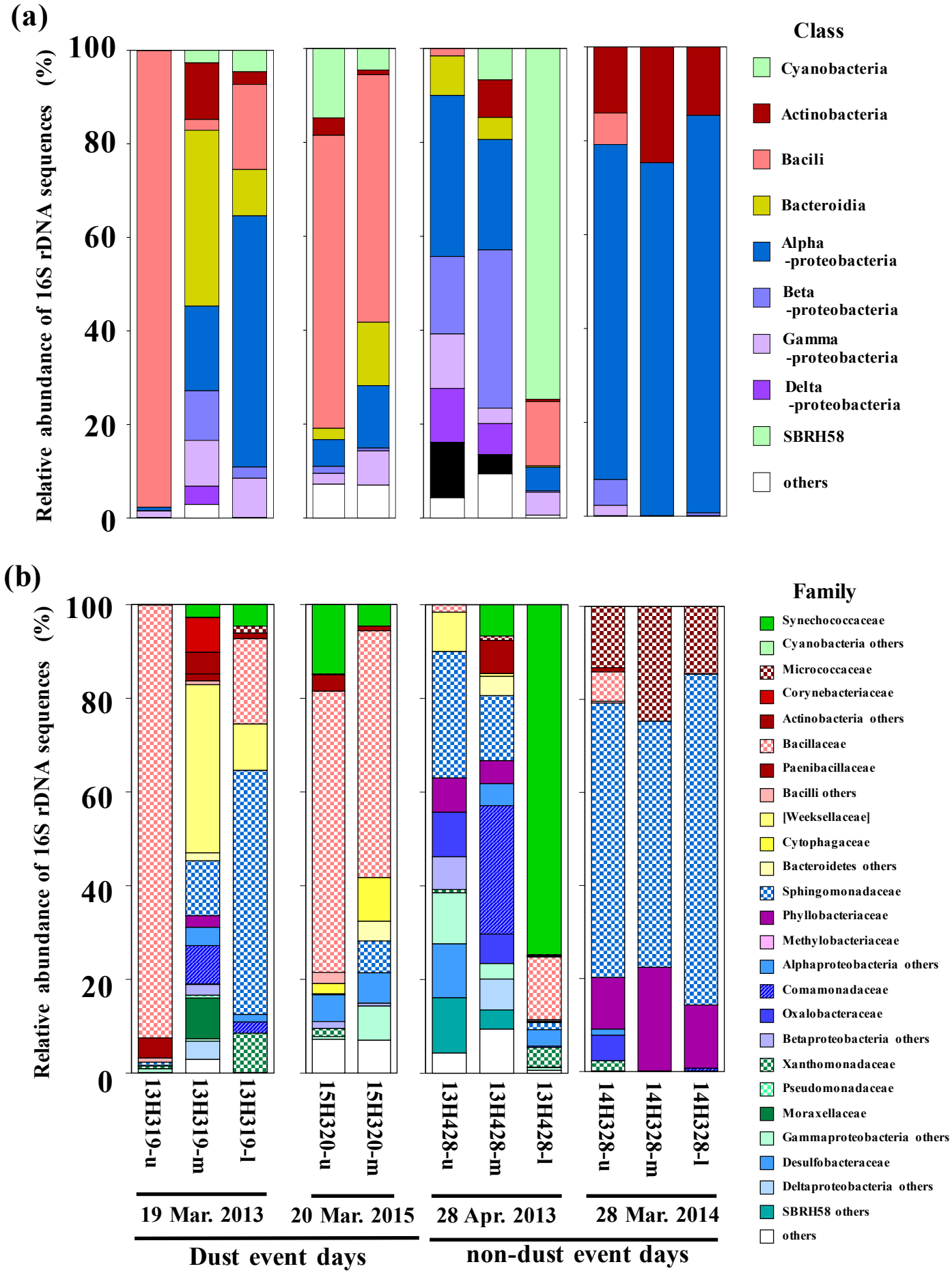

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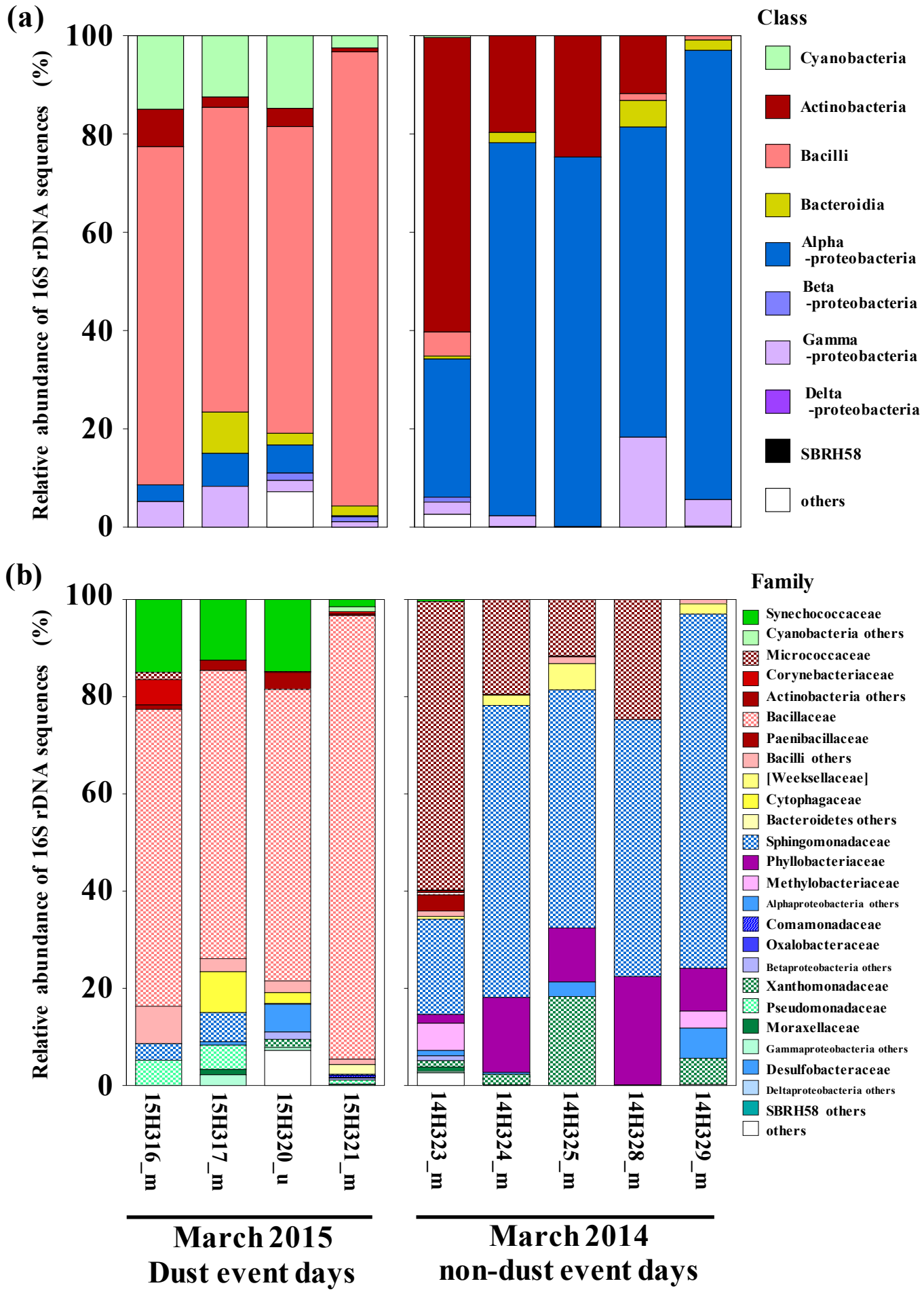

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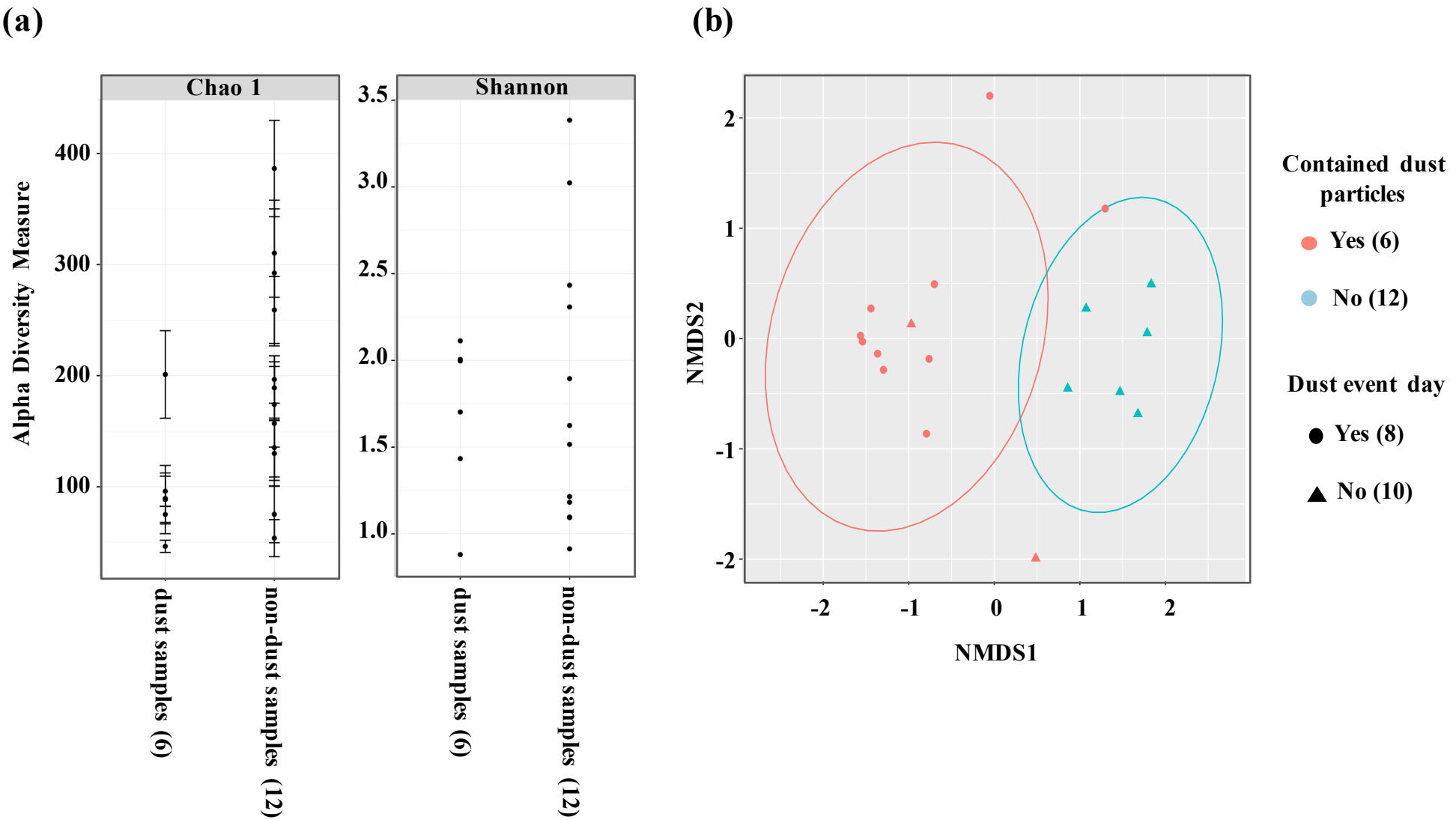

Fig. 8 T. Maki et al.