Atmos. Chem. Phys. Discuss., 8, 2129–2141, 2008 www.atmos-chem-phys-discuss.net/8/2129/2008/ © Author(s) 2008. This work is licensed under a Creative Commons License.



**ACPD** 

8, 2129–2141, 2008

Utilising PPO for high exposure solar UVA dosimetry

D. J. Turnbull et al.



# Utilising polyphenylene oxide for high exposure solar UVA dosimetry

# D. J. Turnbull and P. W. Schouten

Faculty of Sciences, University of Southern Queensland, Toowoomba, 4350, Australia Received: 26 November 2007 – Accepted: 20 December 2007 – Published: 6 February 2008 Correspondence to: D. J. Turnbull (turnbull@usq.edu.au)

#### Abstract

Researchers at the University of Southern Queensland have developed a personal UV dosimeter that can quantitatively assess high exposure solar UVA exposures. The chemical polyphenylene oxide has been previously reported on its ability to measure high UVB exposures. This current research has found that polyphenylene oxide, cast in thin film form, is responsive to both the UVA and UVB parts of the solar spectrum. Further to this, the UVB wavelengths were filtered out with the use of mylar. This combined system responded to the UVA wavelengths only and underwent a change in optical absorbance as a result of UVA exposure. Preliminary results indicate that this UVA dosimeter saturates steadily when exposed to sunlight and can measure exposures of more than  $20 \text{ MJ/m}^2$  of solar UVA radiation with an uncertainty level of no more than  $\pm 5\%$ .

#### 1 Introduction

Exposure to UV radiation is known to be a causative factor in the induction of skin cancers and other sun-related disorders. Most acute responses of humans to UV 15 exposure occur as a result of UVB (280 to 315 nm) exposures, as these wavelengths are highly effective in creating a human biological response. However, this does not mean that UVA radiation has no impact on human UV exposures and health. UVA can cause erythema in human skin, yet, the exposures required to create such a response is much larger than UVB radiation. UVA radiation penetrates much deeper into human 20 skin tissue than UVB, resulting in impacts that are not as acute, taking many years to manifest. Past research has shown that UVA (315 to 400 nm) plays a significant role in human skin carcinogenesis, immune suppression, DNA damage, photoageing and wrinkling (Agar et al., 2004; Moan et al., 1999; Garland et al., 2003). Skin cancer is considered the most common malignant neoplasm in Australia and the USA (Kricker 25 and Armstrong, 1996; Glanz and Mayer, 2005; NCI, 2006). Over 1,600 Australians



EGU

die from skin cancer each year and a further 380,000 Australians are treated for skin cancer each year (NCCI, 2003; AIHW, 2004; AIHW, 2005). Estimates put direct health care costs of all types of skin cancer in Australia in the region of \$734.9 million per year, with the indirect costs in the form of sick leave and foregone earnings in the region of

\$1.395 billion per year (Armstrong, 1995). As a result, it is essential to decrease the amount of exposure to damaging solar UV radiation that the population experiences. This requires methods to understand the solar UV radiation environment that humans live in. Quantification of the individual level of solar UV radiation exposure requires personal dosimetry due to changes in the position of people compared to the radiation
 source.

Commonly used UV chemical dosimeters are polysulphone and phenothiazine. Phenothiazine has been used for measuring UVA wavelengths in various environments (Parisi et al., 2005). However, phenothiazine only has the capability to record a cumulative exposure over a small time period, usually three to four hours. This small

- <sup>15</sup> dynamic range greatly reduces the amount of time over which a UVA exposure can be measured in the field. Therefore, a chemical dosimeter that is capable of measuring large amounts of UVA radiation over a long time period would be very useful. Research conducted by Davis et al. (1976) showed that Poly (2,6-dimethyl-1, 4-phenylene oxide) (PPO) film could be employed to measure high levels of UV exposure. Further research
- found PPO to be ideal for this purpose as it would allow for unattended measurements to be made at various locations over extensive amounts of time with high levels of accuracy when compared against radiometric measurements (Davis et al., 1981; Berre and Lala, 1989; Lester et al., 2003). Schouten et al. (2007) found that PPO has a progressively increasing response to UVA wavelengths over time. PPO dosimeters
- have been utilised in the measurement of global UV exposures, however, they have not been used to measure high exposure solar UVA exposures. The aim of this paper was to extend the previous research that has employed PPO dosimeters for the measurement of solar UVB and erythemal exposures to investigate the suitability of PPO for the measurement of high exposure solar UVA irradiances.

Full Screen / Esc

**Printer-friendly Version** 

Interactive Discussion

EGU

#### 2 Materials and methods

#### 2.1 PPO

PPO film of approximately 40  $\mu$ m thickness was attached with adhesive tape to a plastic holder. The holders for the dosimeters were 3×3 cm in size and fabricated from thin polyvinylchloride with an aperture of approximately 1.2×1.6 cm (Fig. 1). A UV/visible 5 spectrophotometer (model 1601, Shimadzu Co., Kyoto, Japan) was used to measure the pre and post exposure absorbance of the dosimeter. The uncertainty range for optical absorbance measurements in the spectrophotometer has been stated as  $\pm 0.002$ by the manufacturer. PPO is responsive to both the UVA and UVB part of the solar spectrum, therefore the UVB wavelengths were filtered by placing  $120 \,\mu m$  thick my-10 lar film (Cadillac Plastics, Australia) on top of the PPO film. For each dosimeter, the change in optical absorbance ( $\Delta A_{320}$ ) due to UV exposure was measured at 320 nm (with mylar removed) at four different sites over the dosimeter in order to minimise any errors due to any possible minor variations in the PPO film over the size of the dosimeter. The change in optical absorbance was measured at 320 nm as previous 15 studies have shown that this is the wavelength where the greatest overall UV energy induced change occurs (Lester et al., 2003; Schouten et al., 2007). The post exposure absorbance was measured at a standardized time following exposure to minimize any error associated with the post exposure "dark reaction" of the PPO (Lester et al., 2003; Schouten et al., 2007). Lester et al. (2003) has shown previously that the PPO 20 dosimeter is not a temperature dependent system, so fluctuations in temperature over the exposure time period will not have any influence upon the subsequent measure-

2.2 Calibration

ment error.

<sup>25</sup> The dosimeters were calibrated for UVA exposures by exposing a series of dosimeters on a horizontal plane, to relatively clear sky solar UV from approximately 0800 to 1600 h

# **ACPD**

8, 2129-2141, 2008

## Utilising PPO for high exposure solar UVA dosimetry



Australian Eastern Standard Time (EST) for a total of 22 days. These calibrations ran over the months of April and May at a subtropical Southern Hemisphere site at the University of Southern Queensland, Toowoomba, Australia (27.6° S, 151.9° E, altitude 693 m). The solar zenith angle (SZA) ranged from 30° to 75°. The PPO dosimeters
<sup>5</sup> were calibrated on a horizontal plane with a UVA meter (501 Biometer, Solar Light Co., Philadelphia, USA). The UVA meter was calibrated against a scanning spectrora-diometer (Bentham Instruments, Ltd, Reading, UK). The spectroradiometer is based on a double grating monochromator, a UV sensitive detector and amplifier with software variable gain provided by a programmable high voltage power supply. The interior
<sup>10</sup> of the spectroradiometer enclosure is temperature stabilised to 23.0± 0.5° C, using a Peltier heater/cooler unit. The input optics of the spectroradiometer are provided by a PTFE (polytetrafluoro ethylene) diffuser and connected by an optical fibre to the input

slit of the monochromator. The spectroradiometer is programmed to start scanning the global UV spectrum in 0.5 nm increments from dawn, and thereafter every 10 min till dusk. The instrument is wavelength calibrated to the UV spectral lines of a mercury lamp and irradiance calibrated to a 150 Watt quartz tungsten halogen lamp with calibration traceable to the National Physical Laboratory, UK standard.

To produce the calibration curve, the UVA exposure (MJ/m<sup>2</sup>) was measured over specific intervals with the UVA meter. This measurement was then calibrated to the scan-<sup>20</sup> ning spectroradiometer by employing a transfer equation with a calculated R<sup>2</sup> value of no less than 0.99. After each interval, the  $\Delta A_{320}$  for a single PPO dosimeter was measured by the spectrophotometer to provide a single data point, where  $\Delta A_{320}$  was calculated by the following

$$\Delta A_{320}{=}A_{320}^{\mathsf{FINAL}}{-}A_{320}^{\mathsf{INITIAL}}$$

where  $A_{320}^{FINAL}$  is the final optical absorbance measurement after exposure taken at 320 nm and  $A_{320}^{INITIAL}$  is the initial absorbance measurement before exposure taken at 320 nm. After the absorbency was measured, the dosimeter was then removed from the batch.

# **ACPD**

8, 2129–2141, 2008

#### Utilising PPO for high exposure solar UVA dosimetry



#### 2.3 Reproducibility

To test the reproducibility of the dosimeters for the measurement of solar UVA, ten dosimeters (with mylar filter) were exposed simultaneously to solar UV over a three hour period on a horizontal plane. These exposures were conducted on an unshaded sports oval in spring for clear sky conditions.

#### 3 Results

5

10

#### 3.1 Filtered exposures

The spectral transmission of the mylar film was measured pre-exposure and postexposure to solar UV to test for any significant changes. The change in spectral transmission of the mylar film is provided in Fig. 2. The maximum change was approximately 13% from 331 to 337 nm. The change in absorbency of the PPO film preand post-exposure is provided in Fig. 3.

#### 3.2 Calibration

The calibration of the PPO dosimeters for solar UVA exposure is shown in Fig. 4. The data points are the averages of the four ΔA's measured for each dosimeter and the error bars on the x-axis values are the standard deviation of the four measurements. A power law was fitted to the calibration data with the form of:

UVA=11.7( $x^{0.51}$ )MJ/m<sup>2</sup> where x is the change in absorbency. The resulting  $R^2$  for the calibration was greater than 0.99.

20 3.3 Reproducibility

For reproducibility tests, ten dosimeters were placed on a horizontal plane and exposed to solar UV. All dosimeters received the same exposure of solar UV producing a mean

8, 2129–2141, 2008

Utilising PPO for high exposure solar UVA dosimetry



 $\Delta A_{320}$  of 0.598 with a standard deviation of no more than 3% and a coefficient of variation equal to approximately 5%. This variation may be due minor variations over the surface of the sheet of PPO film from which the dosimeters were fabricated and the influence of dust particles that accumulated on the surface of the dosimeters during the exposure period.

#### 4 Discussion

5

Preliminary results indicate that this UVA dosimeter saturates reasonably slowly when exposed to sunlight and can measure exposures of more than 20 MJ/m<sup>2</sup> of solar UVA radiation with an uncertainty level of no more than ±5%. The size and lightweight
properties of the dosimeter means that it can be attached to any anatomical site on the human body in different environments such as underneath shade or in a vehicle in order to measure the solar UVA exposures with a level of ease, simplicity and cost-effectiveness not associated with counterpart radiometric measurements. The usage of the dosimeter requires the calibration against a calibrated UVA meter. The profile
of the calibration curve will vary with the season and this can be overcome by calibrating the dosimeter in the season that it will be employed to measure the solar UVA exposures.

*Acknowledgements.* This research was supported by a Project Grant awarded by the Cancer Council Queensland.

#### 20 References

Agar, N. S., Halliday, G. M., Barnetson, R. S., Ananthaswamy, H. N., Wheeler, M., and Jones, A. M.: The basal layer in human squamous tumors harbors more UVA than UVB fingerprint mutations: a role for UVA in human skin carcinogenesis, Proc. Nat. Acad. Sci., 101, 4954– 4959, 2004.

# ACPD 8, 2129-2141, 2008 Utilising PPO for high exposure solar UVA dosimetry D. J. Turnbull et al. **Title Page** Introduction Abstract Conclusions References **Tables Figures** Back Full Screen / Esc **Printer-friendly Version** Interactive Discussion EGU

- AIHW (Australian Institute of Health and Welfare) and Australasian Association of Cancer Registries (AACR): Cancer in Australia 2001, Cancer Series Number 28, Canberra: AIHW, 2004, availible at: http://www.aihw.gov.au/publications/index.cfm/title/10083, last access: 30 January 2006.
- 5 AIHW (Australian Institute of Health and Welfare): Health system expenditures on cancer and other neoplasms in Australia, 2000–2001, Health and Welfare Expenditure Series Number 22, Canberra: AIHW, 2005, available at: http://www.aihw.gov.au/publications/index.cfm/title/ 10129, last access: 30 January 2006.

Armstrong, B.K.: Skin Cancer, Dermatol. Clin., 13, 583-594, 1995.

- <sup>10</sup> Berre, B. and Lala, D.: Investigation on photochemical dosimeters for ultraviolet radiation, Sol. Ener., 42, 405–416, 1989.
  - Davis, A., Deane, G. W. H., Gordon, D., Howell G. V., and Ledbury, K. J.: A world-wide program for the continuous monitoring of solar UV radiation using poly (phenylene oxide) film and consideration of the results, J. Appl. Polym. Sci., 20, 1165–1174, 1976.
- <sup>15</sup> Davis, A., Diffey, B. L., and Tate, B. K.: A personal dosimeter for biologically effective solar UV-B radiation, Photochem. Photobiol., 34, 283–286, 1981.
  - Garland, C. F., Garland F. C. and Gorham, E. D.: Epidemiologic evidence for different roles of ultraviolet A and B radiation in melanoma mortality rates, Ann. Epidemiol., 13, 395–404, 2003.
- <sup>20</sup> Glanz, K. and Mayer, J. A.: Reducing ultraviolet radiation exposure to prevent skin cancer, Am. J. Prev. Med., 29, 131–142, 2005.
  - Kricker, A. and Armstrong, B.: International trends in skin cancer, Can. Forum, 20, 192–194, 1996.

Lester, R. A., Parisi, A. V., Kimlin, M. G., and Sabburg, J.: Optical properties of poly(2,6-

- dimethyl-1,4-phenylene oxide) film and its potential for a long term solar ultraviolet dosimeter, Phys. Med. Biol., 48, 3685–3698, 2003.
  - Moan, J., Dahlback, A., and Setlow, R. B.: Epidemiological support for an hypothesis for melanoma induction indicating a role for UVA radiation, Photochem. Photobiol., 70, 243–247, 1999.
- <sup>30</sup> NCCI (National Cancer Control Initiative): The 2002 national non-melanoma skin cancer survey, in: A report by the NCCI Non-melanoma Skin Cancer Working Group, edited by: M. P. Staples Melbourne: NCCI, 2003.

NCI (National Cancer Institute): http://www.cancer.gov/cancertopics/commoncancers, last ac-

8, 2129–2141, 2008

## Utilising PPO for high exposure solar UVA dosimetry

Title Page	
Abstract	Introduction
Conclusions	References
Tables	Figures
14	ÞI
•	•
Back	Close
Full Screen / Esc	
Printer-triendly Version	
Interactive Discussion	

cess: 15 December 2006.

5

Parisi, A. V., Kimlin, M. G., Turnbull, D. J., and Macaranas, J.: Potential of phenothiazine as a thin film dosimeter for UV-A exposures, Photochem. Photobiol. Sci., 4, 907–910, 2005.
Schouten, P. W., Parisi, A. V., and Turnbull, D. J.: Evaluation of a high exposure solar UV dosimeter for underwater use, Photochem. Photobiol., 83, 1–7, 2007.

# ACPD 8, 2129-2141, 2008 **Utilising PPO for high** exposure solar UVA dosimetry D. J. Turnbull et al. Title Page Abstract Introduction Conclusions References **Tables Figures** 14 ►L. ◄ ► Back Close Full Screen / Esc **Printer-friendly Version** Interactive Discussion



Fig. 1. A sample PPO dosimeter with the mylar film attachment.

# ACPD

8, 2129–2141, 2008

## Utilising PPO for high exposure solar UVA dosimetry





**ACPD** 







EGU

Full Screen / Esc

**Printer-friendly Version** 

Interactive Discussion

**ACPD**