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Analysis for biomass
burning aerosol**

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Investigation of molar volume and surfactant characteristics of water-soluble organic compounds in biomass burning aerosol

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Abstract

In this study, we characterize the CCN activity of the water-soluble organics in biomass burning aerosol. The aerosol after collection upon filters is dissolved in water using sonication. Hydrophobic and hydrophilic components are fractionated from a portion of the original sample using solid phase extraction, and subsequently desalted. The surface tension and CCN activity of these different samples are measured with a KSV CAM 200 goniometer and a DMT Streamwise Thermal Gradient CCN Counter, respectively. The measurements show that the strongest surfactants are isolated in the hydrophobic fraction, while the hydrophilics exhibit negligible surface tension depression. The presence of salts (primarily $(NH_4)_2SO_4$) in the hydrophobic fraction substantially enhances surface tension depression; their synergistic effects considerably enhance CCN activity, exceeding that of pure $(NH_4)_2SO_4$. For our analysis, average thermodynamic properties (i.e., molar volume) are determined for samples using our newly developed Köhler Theory Analysis (KTA) method. We have found that, the molar mass of the hydrophilic and hydrophobic aerosol components is estimated to be $87\pm26\,\mathrm{g\,mol^{-1}}$ and $780\pm231\,\mathrm{g\,mol^{-1}}$, respectively. KTA also suggests that the relative proportion (in moles) of hydrophobic to hydrophilic compounds in the original sample to be 1:3. For the first time, KTA is applied to an aerosol with this level of complexity and displays its potential for providing physically-based constraints for GCM parameterizations of the aerosol indirect effect.

1 Introduction

Aerosols significantly impact global and regional climate. They can directly reflect the amount of incoming solar radiation into space; by acting as cloud condensation nuclei (CCN), they can indirectly impact climate by affecting cloud albedo (IPCC, 2001). The ability of aerosol to act as CCN is a strong function of their size, composition and physical state. Carbonaceous compounds can constitute up to 70–90% of total aerosol

mass (Andreae and Crutzen, 1997; Cachier et al., 1995; Yamasoe et al., 2000), 10–70% of which can be water-soluble (Facchini et al., 2000; Sullivan et al., 2004; Zappoli et al., 1999). Biogenic and anthropogenic volatile organic carbon oxidation is the primary source of secondary organic aerosol in the troposphere (Kanakidou et al., 2005; Limbeck et al., 2003). Biomass burning is another major source of water-soluble organic carbon (WSOC) (Kanakidou et al., 2005; Sullivan et al., 2006); in pyrogenic air masses, WSOC can account for 45–75% of the total carbon mass (Falkovich et al., 2005; Graham et al., 2002; Mayol-Bracero et al., 2002).

WSOC is a complex mixture of neutral and acidic polar organic compounds (Decesari et al., 2000). They can affect CCN activity by *i*) providing solute, thus reducing the equilibrium water vapor pressure of the droplet, and, *ii*) acting as surfactants capable of depressing surface tension, and potentially, growth kinetics (Decesari et al., 2003; Facchini et al., 1999b; Feingold and Chuang, 2002; IPCC, 2001; Kanakidou et al., 2005; Lance et al., 2004; Nenes et al., 2002; Shulman et al., 1996). Owing to its complexity, WSOC cannot be completely speciated using standard analytical methods, but instead is often characterized using "functional group analysis", using either chromatographic techniques or nuclear magnetic resonance (Sannigrahi et al., 2006; Sullivan and Weber, 2006a, b). WSOC can be classified broadly into hydrophilic and hydrophobic fractions; these are usually operationally defined but correlate with carbon chain length and number of functional groups per molecule. Hydrophilic compounds are typically highly oxygenated low molecular compounds (with potentially numerous carbonyls and carboxyl groups per molecule) and tend to be highly soluble in water and exhibit limited surfactant behavior; hydrophobics can be longer in length, less hygroscopic, and strongly depress surface tension (Kanakidou et al., 2005).

An important class of hydrophobic WSOC are Humic-like Substances (HULIS); these are a mixture of high molecular weight compounds that are strong surfactants (Decesari et al., 2000). According to Facchini et al. (2000), HULIS depressed surface tension in fog water samples by 15–20% at a 100 mg C L^{-1} WSOC concentration; Kiss et al. (2005) showed that HULIS dissolved in water at a concentration of 1000 mg L^{-1}

could decrease surface tension by 25–42% or even more if salts like $(NH_4)_2SO_4$ are present in solution. HULIS are present in continental aerosol, urban, rural and smog samples (Decesari et al., 2000; Facchini et al., 2000; Graham et al., 2002; Havers et al., 1998; Kiss et al., 2005; Krivacsy et al., 2000). Despite their importance, the thermody-

5 namic properties of HULIS (e.g., average molecular weight, molar volume, solubility) is poorly constrained. This is largely due to the complexity of HULIS, which do not have a definite chemical structure, but are often described as a "network" of compounds binding together in solution, potentially forming oligomers and macromolecules.

Comprehensively linking carbonaceous aerosol with clouds in aerosol-cloud-climate

10 interaction studies requires knowledge of properties relevant for predicting CCN activity and droplet growth kinetics; these are molar volume (molar mass over density), solubility, effective van't Hoff factor, and water vapor uptake coefficient. Such information, for the complex mixture of organics typical of ambient aerosol does not exist, and few methods until recently were available or obtaining it. A new method, "Köhler

15 Theory Analysis" (KTA; Padró et al., 2007[1]), promises to fill this gap. KTA can constrain the average (effective) molar volume and solubility of a water soluble organic mixture, based on measurements of CCN activity and surface tension combined with an analysis based on Köhler Theory. KTA has been successfully applied for laboratory-generated organic-inorganic mixtures of known composition; the method inferred mo-

20 lar volume for the organic constituents within 20% of the expected value (Padró et al., 2007[1]). KTA has also been successfully applied to infer molar volumes of secondary organic aerosol (SOA) from ozonolysis of alkenes (Asa-Awuku et al., 2007a[2]) and of WSOC of cloudwater samples collected in-situ aboard the CIRPAS Twin Otter during

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[1]Padró, L. T., Asa-Awuku, A., Morrison, R., and Nenes, A.: Inferring Thermodynamic properties from CCN Activation Experiments a), Single and Binary Aerosols, in review, 2007.

[2]Asa-Awuku, A., Nenes, A., Gao, S., Flagan, R. C., and Seinfeld, J. H.: The CCN Activity of Water-Soluble Secondary Organic Aerosols Generated from the Ozonolysis of Alkenes, in preparation, 2007a.

the MASE campaign (Asa-Awuku et al., 2007b[3]).

In this study, a biomass burning sample acquired during a prescribed burning event in Georgia (Sullivan and Weber, 2006b), is fractionated into hydrophilic and hydrophobic components to assess their individual contribution to surface tension depression and CCN Activity. Average molar volumes are inferred from KTA of CCN activation of aerosol generated from the atomization of the original and fractionated biomass burning sample.

2 Experimental methods

Fine Biomass Burning (BB) particulate matter ($PM_{2.5}$) was collected on pre-baked quartz fiber filters with a Thermo Anderson Hi-Volume Air Sampler during a prescribed burning in Augusta, Georgia at Fort Gordon and in Columbus, Georgia at Fort Benning, which are both located in heavily wooded areas, in April 2004 (Sullivan and Weber, 2006b). The freshly burned biomass aerosol is subsequently extracted in water, and fractionated into hydrophobic and hydrophilic fractions. Figure 1 outlines the procedure adopted to characterize the original and fractionated samples. Each step is described in subsequent sections.

2.1 Extraction and fractionation of biomass burning sample

WSOC is extracted from the filter by sonication in a heated water bath (Baumann et al., 2003; Sullivan and Weber, 2006b) for 1.25 h. A portion of this sample (BB) was then fractionated into hydrophilic (HPHIL) and hydrophobic (HPHOB) components using a macro-porous nonionic resin (XAD-8) Solid Phase Extraction (SPE) column (Duarte and Duarte, 2005; Sullivan and Weber, 2006a). With the WSOC solution adjusted to pH=2 using HCl, the resin does not retain hydrophilic compounds. Calibra-

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[3]Asa-Awuku, A., Nenes, A., Sorooshian, A., and Seinfeld, J. H.: CCN Activity of Cloud Processed Organic Aerosol Collected during MASE 2005, in preparation, 2007b.

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tions suggest this includes saccharides, amines, and carbonyls and aliphatic monocar-boxylic/dicarboxylic/oxocarboxylic acids with less than 4 or 5 carbons, or hydrophobic compounds with pKa less than 2. The column retains the long and short-chained hy-drophobic compounds, which calibrations suggest may be aromatic acids, phenols, organic nitrates, cyclic acids, carbonyls and monocarboxylic and dicarboxylic chains greater than 3 to 4 carbons (Sullivan and Weber, 2006a). The column is then eluted to pH 13 and a large fraction of the adsorbed hydrophobic fraction is removed. The recovered hydrophobic fraction is adjusted to pH 2 with HCl to avoid the oxidation of organic compounds. Of the compounds tested, aromatic acids and phenols had the largest recoveries. These hydrophobic compounds retained on the column at pH 2 and recovered at pH 13 exhibit a dark brown color, suggestive of HULIS (Graber and Rudich, 2006). Similar speciation methods have shown that HULIS in WSOC can be found within the hydrophobic fraction (Decesari et al., 2000; Kiss et al., 2001; Krivacsy et al., 2001; Sullivan and Weber, 2006a).

Part of the BB, HPHOB and HPHIL components are subsequently de-salted with single-use Oasis® HLB Extraction Cartridges (Waters, Milford, Massachusetts), to re-duce the concentration of electrolytes in the samples. Most organics are retained on the resin while inorganic ions pass through. The organics are subsequently eluted with 100% high-purity methanol (Romil, Cambridge, UK). This eluate is dried under a gentle stream of nitrogen gas to remove the methanol and re-dissolved with 18 Mohm ultrapure water. The desalted samples will be referred to as D-HPHOB, D-HPHIL and D-BB (Table 1).

It should be noted that there can be a potential for artifacts in the fractionation pro-cess. During the desalting process low molecular weight organic compounds (i.e. acetic and formic acid) that do not strongly adsorb on the resin may pass through the chromatography column. Hence only when comparing the hydrophilics will the D-HPHIL sample, compared to HPHIL, be somewhat enriched in higher molecular weight organics. In addition, not all hydrophobic compounds are recovered from the column. Based on calibrations (Sullivan and Weber, 2006a), carbonyls and carboxylic

acids greater than roughly C5 and organic nitrates are thus most likely to be under-represented in this analysis.

2.2 Measurement of chemical composition

The WSOC content of the original, fractionated and desalted samples (Table 1) were measured with a Total Organic Carbon (TOC) Analyzer (Sievers Model 800 Turbo, Boulder, CO). A detailed description of the method can be found in Sullivan and Weber (2006a). We also measure anions (SO_4^{2-}, Cl^- and NO_3^-) and cations (Na^+, NH_4^+, Mg^+, Ca^{2+}, K^+) with a Dionex DX-500 ion chromatograph with $Na_2CO_3/NaHCO_3$ eluent and Metrosep A Supp 5–100 analytical column (Metrohm, Switzerland). In samples, ammonium was in excess, suggesting that all the organic and inorganic acids were neutralized.

2.3 Surface tension measurements

All surface tension measurements were performed using the pendant drop method with a KSV CAM 200 optical contact angle and surface tension goniometer. The surface tension at the liquid-air interface, σ, depends on the density jump across the interface, the gravitational constant, the radius of the droplet and the contact angle between the droplet and needle used for forming the pendant drop. Fifty pendant drops were used for each surface tension measurement, requiring in total less than 200 μL of sample; the σ for commercial ultrafine de-ionized ultra-filtered water (Fischer-Scientific, W-2) was measured in between organic samples and were found to agree consistently within 2% of reported literature values at room temperature. This ensures that cross-contamination between samples did not occur. The temperature was measured with a 50 K Ohm Thermoresistor (Digikey ERT-D2FHL503S) thermocouple throughout the experiment.

The surfactant characteristics for each sample (Table 2) were characterized by measuring surface tension as a function of dissolved carbon concentration (at the original

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sample concentration, then at 1:1, 1:2, 1:3 and 1:4 dilution with 18-Mohm ultrapure water). The measurements were then fit to a Szyszkowski -Langmuir (Langmuir, 1917) adsorption isotherm which expresses σ as a function of the soluble carbon,

$$\sigma = \sigma_w - \alpha T \ln(1 + \beta c) \qquad (1)$$

5   where c is the concentration of soluble carbon (mg C L^{1}), α and β are empirical constants, obtained from least square fitting to σ measurements, and σ_w is the surface tension of 18-Mohm ultrapure water at the experiment temperature (obtained by infinitely diluting our sample with de-ionized ultra-filtered water). If the surfactant is composed of a single compound, Eq. (1) describes a Gibbs adsorption isotherm of the partitioning

10   of surfactant between the bulk and droplet surface layer, α and β are related to the orientation of dissolved molecules and their interactions at the surface interface (Langmuir, 1917). Even though WSOC is a complex mixture of compounds, Eq. (1) still reproduces the measurements very well (Fig. 4). Table 2 is a summary of α and β parameters obtained from fitted σ data for the original biomass burning and fractionated

15   samples.

In addition to measurements of the original and extracted samples, we also measured the effect of adding electrolytes on surface tension; limited by the amount of fractionates available, we performed this exercise on the original BB sample. The concentration of carbon was kept constant at 850 mg L^{-1} and premeasured amounts of

20   electrolytes were dissolved. $(NH_4)_2SO_4$ and NaCl were used in these experiments, typical of ionic species found in the atmosphere.

2.4   CCN activity measurements

Aerosol generated from the original BB sample and fractions were activated into cloud droplets to determine their CCN activity. 80 to 100 μL of organic sample were placed in

25   3–5 ml of commercial ultrafine de-ionized ultra-filtered water (Fischer-Scientific, W-2) and atomized with a collision-type (University of Minnesota) atomizer (Fig. 2) operated at 3.5 psig pressure. The polydisperse droplets are subsequently dried by passing

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them through two silica gel dryers (Fig. 2), neutralized by a Kr-85 bipolar ion source and classified using a Differential Mobility Analyzed (DMA 3081) (Fig. 2). The classified aerosol is then split and passed though a TSI 3010 Condensation Particle Counter (CPC) for measuring aerosol number concentration, CN. CCN concentrations were
5 measured in the other stream by a Droplet Measurement Technologies Continuous-Flow Streamwise Thermal Gradient CCN Counter (STGC) (Lance et al., 2006; Roberts and Nenes, 2005).

   CCN activity of the aerosol was characterized as follows. For a fixed supersaturation the "activation curves" (i.e., ratio of CCN to CN) was measured between 10 and
10 250 nm dry mobility diameter. Activation ratios are then determined from 0.2% to 1.0% supersaturation. Figure 3 shows an example of activation curves obtained for D-HPHIL activation. The activation curves are then fit to a sigmoidal curve, neglecting the impact of doubly charged particles which appear as a characteristic secondary peak to the left of the main sigmoid (Fig. 3). The particle dry diameter size, d, at which 50% of the par-
15 ticles were CCN represent the dry diameter of the particle with critical supersaturation equal to the instrument supersaturation. The dependence of d with respect to supersaturation can be used to infer solute molar volume and the presence of surfactants using KTA (Padró et al., 2007[1]) (Sect. 3).

3 Description of köhler theory analysis

20 ### 3.1 Single component CCN theory

Each particle requires a discrete amount of water vapor supersaturation to activate into cloud droplets. This "critical supersaturation", s_c, for simple (water-soluble) single-component aerosol is given by (Köhler, 1936; Seinfeld and Pandis, 1998),

$$s_c = \left(\frac{4A^3}{27B}\right)^{1/2}, \text{ and } A = \left(\frac{4M_w\sigma}{RT\rho_w}\right), B = \left(\frac{6n_sM_w\upsilon}{\pi\rho_w}\right) \tag{2}$$

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where M_w, ρ_w are the molecular weight and density of water, respectively, R is the universal gas constant, n_s are the moles of solute dissolved in the droplet, υ is the effective van't Hoff factor of the solute, T is the ambient temperature and σ is the droplet surface tension at the point of activation. Single-component Köhler theory can be aptly applied to soluble inorganic salts and has been successfully applied to soluble low molecular weight organics as well (Cruz and Pandis, 2000; Facchini et al., 1999a; Raymond and Pandis, 2002).

3.2 Multi-component CCN theory

For multi-component CCN, the contribution of solute from each organic and inorganic component can be accounted for as a modification of the Raoult term (B term in Eq. 2) as:

$$B = \sum_i B_i = \sum_i \left(\frac{\rho_i}{\rho_w}\right)\left(\frac{M_w}{M_i}\right)\varepsilon_i\upsilon_i d^3 = d^3\left(\frac{M_w}{\rho_w}\right)\sum_i\left(\frac{\rho_i}{M_i}\right)\varepsilon_i\upsilon_i \tag{3}$$

where d is the dry diameter of the CCN, and ρ_i, ε_i, υ_i, M_i are the density, volume fraction, effective van't Hoff factor and molar mass of the solute i, respectively. ε_i is related to the mass fraction of i, m_i, as

$$\varepsilon_i = \frac{m_i/\rho_i}{\sum_i m_i/\rho_i} \tag{4}$$

m_i are obtained from measurements of chemical composition. For a single component $\varepsilon_i=1$ and Eq. (3) reduces to Eq. (2).

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3.3 Köhler theory analysis of ambient CCN

Assuming that all solute dissolves, Eq. (2) can be written as

$$s_c = \left(\frac{256 M_w^3 \sigma^3}{27 R^3 T^3 \rho_w^3} \right)^{1/2} \left[\sum_i \left(\frac{M_w}{\rho_w} \right) \left(\frac{\rho_i}{M_i} \right) \varepsilon_i \upsilon_i \right]^{-1/2} d^{-3/2} = \omega d^{-3/2} \tag{5}$$

where

$$\omega = \left(\frac{256 M_w^3 \sigma^3}{27 R^3 T^3 \rho_w^3} \right)^{1/2} \left[\sum_i \left(\frac{M_w}{\rho_w} \right) \left(\frac{\rho_i}{M_i} \right) \varepsilon_i \upsilon_i \right]^{-1/2} \tag{6}$$

The molar volume of the organic component, $\left(\frac{M_j}{\rho_j} \right)$, is explicitly solved for by rearranging Eq. (6) as follows:

$$\frac{M_j}{\rho_j} = \frac{\varepsilon_j \upsilon_j}{\frac{256}{27} \left(\frac{M_w}{\rho_w} \right)^2 \left(\frac{1}{RT} \right)^3 \sigma^3 \omega^{-2} - \sum_{i \neq j} \frac{\rho_i}{M_i} \varepsilon_i \upsilon_i} \tag{7}$$

where j is used to denote the organic constituent and all other components i, refer to the inorganic components present in the aerosol. Equation (7) is the basis of KTA (Padró et al., 2007[1]). In this study, KTA is applied as follows:

a. *If there are no strong surfactants present* (i.e., the surface tension of the CCN does not depend on the concentration of solute at the point of activation), ω does not depend on d and its value can be determined from a power law fit between measured s_c and d (Table 3). Application of Eq. (7) then infers the average molar volume of the water-soluble organics, provided that the volume fractions of all constituents are measured (from WSOC for organics and IC for inorganics) and the composition of the inorganics are known.

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b. *If there are strong surfactants present* (i.e., the surface tension of the CCN does not depend on the concentration of solute at the point of activation), ω depends on d, so method "a." cannot be used to infer the molar volume. Instead, two alternate methods can be used:

Method b_1. Since ω depends weakly on d at low supersaturation (because the CCN are dilute enough at the point of activation so that surface tension is approximately constant) method "a" can be used for a subset of the activation experiments. The appropriate supersaturation range can be determined from the experimental data, by examining the slope of s_c vs. d.

Method b_2. For each supersaturation, directly apply Eq. (7) and an estimate of surface tension at the point of activation (obtained by using Köhler theory to estimate water volume and from there, WSOC concentration) to infer molar volume. Then compute the average molar volume over the range of supersaturations examined.

3.4   Molar volume sensitivity analysis

The uncertainty in inferred organic molar volume, $\Delta\left(\frac{M_j}{\rho_j}\right)$, is quantified as

$$\Delta\left(\frac{M_j}{\rho_j}\right) = \sqrt{\sum_{\text{for all } x} (\Phi_x \Delta x)^2} \tag{8}$$

where Φ_x is the sensitivity of molar volume to each of the measured parameters x (i.e., any of σ, ω, ε_i, ε_j, υ_i, and υ_j)

$$\Phi_x = \frac{\partial}{\partial x}\left(\frac{M_j}{\rho_j}\right) \tag{9}$$

and Δx is the uncertainty in x. The Φ_x for Eq. (9) is obtained by differentiating Eq. (7) and are shown in Table 5.

4   Experimental results

4.1   WSOC and inorganic composition

Table 1 is a summary of the composition for all samples analyzed in this study. The original 850 mg C L^{-1} BB sample had to be fractionated twice to yield enough carbon mass in samples for surface tension and KTA. The HPHOB, D-HPHIL, and D-HPHOB fractions shown in Table 1 are from this later fractionation. The desalted original sample contained significant amounts of ions after the desalting process (17% of the original anion fraction) (Table 1) and thus is not considered in our analysis, as it does not truly reflect a "desalted" sample.

4.2   Surface tension depression

The WSOC of the original BB sample contains large amounts of surfactants; at a concentration of 850 mg C L^{-1}, σ was measured to be 59 mN m^{-1} (25°C), 18% lower than the surface tension at infinite dilution with 18-Mohm ultrapure water (71.24±0.53 mN m^{-1}). The σ value of the infinitely diluted solution is very close to pure water, ~2% different from reported literature values for water at room temperature (Yaws, 1999) and the BB is similar in magnitude to the surface tension depression reported by Facchini et al. (2000) for fog water samples and Kiss et al. (2005) for HULIS dissolved in water.

   From our speciated aerosol study, the original sample and hydrophobic component can significantly depress surface tension with increasing carbon concentration; the hydrophilic components do not (Fig. 4). The desalted hydrophobic and desalted hydrophilic components do not depress surface tension as much as their salted counterparts. Of the three salted samples, the HPHOB sample exhibits the largest sensitivity to the presence of salts. The behavior of the HPHOB fraction suggests it is composed of HULIS (Sullivan and Weber, 2006a). The D-HPHIL and HPHIL fractions effectively have the same σ as water, even at very large carbon concentrations (Fig. 4).

Addition of electrolytes to the BB sample (where WSOC concentration is maintained constant at 850 mg L^{-1}) further depresses droplet σ (Fig. 5). This is consistent with behavior seen by Kiss et al. (2005) for HULIS with salts. Greater amounts of NaCl than $(NH_4)_2SO_4$ are required to reduce surface tension by the same amount, as the bivalent SO_4^{2-} is more effective than Cl$^-$ in partitioning the hydrophobic organics (which are anionic surfactants) to the surface layer (Holmberg, 2003; Hunter, 2001). Nonetheless, the increasing presence of an inorganic salt in the BB sample (which could happen e.g., in cloud processing of the aerosol) can further decrease surface tension by more than 20% and hence can have an important impact on CCN activity.

4.3 CCN activity

Figure 6 shows the critical supersaturation, s_c, of particles generated from the BB and fractionated samples, as a function of their d. $(NH_4)_2SO_4$ has been added as a basis of comparison, and based on its van't Hoff factor and molar volume, is expected to be more CCN active than all samples of Table 1 (i.e., its activation spectrum should lie to the left of the samples). Indeed, this is mostly the case; the activation spectra for HPHIL and D-HPHIL are closest to $(NH_4)_2SO_4$. Strong surfactants are not present in the HPHIL, D-HPHIL, as their surface tension is not substantially different from water, even at very high carbon concentrations (Fig. 5). This suggests that the hydrophilic components are composed of soluble, low molecular weight compounds which is consistent with the functional group calibration analysis of Sullivan and Weber (2006a, b) done for similar samples.

Surprisingly, HPHOB (which contains the least hydrophilic component of WSOC) was the most CCN active of all samples, surpassing even $(NH_4)_2SO_4$ (Fig. 6). This seemingly counterintuitive finding can be reconciled when considering the synergism between the salts and organics. HPHOB contains large amounts of salts, so based on that alone, one would expect CCN activity close to pure $(NH_4)_2SO_4$. However, the small amounts of surfactants present in HPHOB, because of the concentrated salts present in the CCN, tend to partition on the CCN surface and depress surface tension,

as was seen in Fig. 5. This "salting out" effect of the organics would then decrease the CCN critical supersaturation, making the HPHOB better CCN than pure $(NH_4)_2SO_4$. "Salting out" of organics can also explain why HPHOB becomes even more CCN active (compared to $(NH_4)_2SO_4$) as the dry CCN diameter decreases (Fig. 6); according to Köhler theory, the concentration of solute (organic and inorganic) increases at the point of activation as dry particle size decreases. Both decrease surface tension as dry size decreases, which implies that the CCN activity of HPHOB and $(NH_4)_2SO_4$ diverge more at small diameters. Conversely, at large d, the concentrations of solute at the point of activation are lower, the surface depression of HPHOB is relatively small, and the CCN activity curve of HPHOB and $(NH_4)_2SO_4$ tend to converge (Fig. 6).

The D-HPHOB fraction has lower CCN activity than $(NH_4)_2SO_4$ and HPHOB. In comparison to $(NH_4)_2SO_4$, lowering of surface tension in D-HPHOB is not compensated by the lack of solute during the desalting. D-HPHOB and BB have similarly low CCN activity and the BB sample is the least active of all samples. As illustrated in Fig. 6, one cannot assume the CCN activity of BB should reside somewhere between HPHOB and HPHIL. Even though the BB sample contains more soluble material (mostly inorganics and some unknown amount of hydrophilics) and did depress surface tension (attributed the presence of electrolytes and hydrophobics) the contribution and interactions with water vapor of all three speciated components: hydrophilics, hydrophobics and inorganics is not additive. Regardless, KTA on all samples should give molar volume estimates that are consistent, i.e., lowest for HPHIL, then BB, and finally, HPHOB.

5 Köhler theory analysis of samples

5.1 Molar volume estimations

We compute average organic molar volumes for the original and desalted hydrophobic and hydrophilic samples (i.e., BB, D-HPHIL, D-HPHOB), where the organic volume fraction dominates (Table 4) and KTA is subject to less uncertainty (Padró et al., 2007[1]).

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The inorganic compounds present in the sample are assumed to be a mixture of ammonium sulfate, ammonium chloride, and ammonium nitrate in proportion to their ionic concentrations (Table 1).

In computing the volume fraction of each constituent in the dry aerosol, Eq. (4), we multiplied the WSOC concentration with 1.4 to obtain the total dissolved organic carbon mass concentration (Turpin and Lim, 2001); the relative amount of each component (OC and inorganic salts) in solution is then used to determine their mass fraction, m_i, in the dry aerosol (Table 1). In applying Eq. (4), the density of $(NH_4)_2SO_4$, NH_4Cl, NH_4NO_3 was taken to be 1760, 1800, and 1500 kg m^{-3}, respectively (Perry et al., 1997). OC density was the most uncertain, and here is assumed to be 1400 kg m^{-3} (Turpin and Lim, 2001). In the case of hydrophobic molar volume analysis, the WSOC concentration exceeds the critical micelle concentration (which is not predicted by the Szyszkowski-Langmuir model) beyond which surface tension depression asymptotes. Because of this, the surface tension of hydrophobics is set to 50% of pure water, corresponding to the lower limit observed for HULIS (Kiss et al., 2005).

The effective van't Hoff factor, v, (defined as the number of ions released into the solution times the osmotic coefficient) expresses the impact of dissociation on water activity. For multicomponent electrolyte solutions, v can be accurately computed with existing thermodynamic models (Nenes et al., 1998) and has been successfully applied for activation of inorganic CCN (e.g., Raymond and Pandis, 2002). However, in solutions of WSOC with electrolytes (which characterize all the samples of this study), v is quite uncertain as speciation in solution is not known, and modeling complex organic-water-inorganic interactions is challenging. For this reason, we approximate v as follows: *i*) $v=2$ for NH_4Cl and NH_4NO_3, *ii*) $v=3$ for $(NH_4)_2SO_4$, *iii*) $v=1$ for organics in the BB sample, as the sample is acidic and organics are not expected to substantially dissociate, *iv*) $v=1$ for organics in the HPHOB and D-HPHOB samples, as they are very weakly dissociating and *v*) $v=2.5$ for D-HPHIL, as we assume the sample to be composed of dicarboxylic acids neutralized with ammonium during the extraction process; the ammonium salts are then assumed to dissociate as effectively as $(NH_4)_2SO_4$.

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The molar volumes of the organic aerosol components are estimated employing KTA (presented in Sect. 2) and the above effective van't Hoff assumptions, the results of which are shown in Table 4. The molar volume of the hydrophilic organic (6.2×10^{-5} m^3 mol^{-1}) is comparable to that of inorganic salts (7.46×10^{-5} m^3 mol^{-1} and 2.97×10^{-5} m^3 mol^{-1} for $(NH_4)_2SO_4$ and $(NH_4)Cl$, respectively) and low molecular weight mono- and di-carboxylic chains (e.g., formic and acetic acid, and succinic, glutaric, and oxalic acid with 7.54×10^{-5} m^3 mol^{-1}, 9.34×10^{-5} m^3 mol^{-1}, and 6.4×10^{-5} m^3 mol^{-1}, respectively) typical of those identified in the XAD-8 calibration studies (Sullivan and Weber, 2006a). The significantly larger estimated molar volume of the hydrophobic fraction (5.6×10^{-4} m^3 mol^{-1} using method $\mathbf{b_1}$), 6.1×10^{-4} m^3 mol^{-1} using method $\mathbf{b_2}$) is consistent of the existence of HULIS within the sample which has been previously shown by mass spectrometery methods to have molecular weights up to 1000 amu (Graber and Rudich, 2006). Nevertheless, we speculate that the average organic molar mass inferred from KTA could be smaller for several reasons: i) The KTA value is a number averaged property, which weighs the average towards a smaller number, when compared to mass-based averaging, and, ii) HULIS may have a higher ν than unity, as they are comprised of several polyfunctional groups that may contain polar groups like carbonyls, carboxyls, and hydroxls (Graber and Rudich, 2006; Kiss et al., 2002). Therefore, when KTA is applied assuming $\nu = 1$, the inferred molar volume decreases to compensate.

The average molar mass for the original BB samples was found to be 248 g mol^{-1}. Assuming that the organics in the original BB sample is a mixture of D-HPHIL and D-HPOB, we can infer the relative proportion of HPHIL and HPOB via the following equation

$$M_{BB} = \chi M_{HPHIL} + (1 - \chi) M_{HPHOB} \tag{10}$$

where χ is the mol fraction of hydrophilics in the original BB sample and M_{BB}, M_{HPHIL}, M_{HPHOB} are the inferred molar masses from the BB, HPHIL and HPHOB samples.

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Equation (10) can be solved for χ to yield

$$\chi = \left(\frac{M_{\mathrm{BB}} - M_{\mathrm{HPHIL}}}{M_{\mathrm{HPHOB}} - M_{\mathrm{HPHIL}}} \right) \qquad (11)$$

We determine that $\chi \sim 0.25$, i.e., there are approximately 3 hydrophilic compounds for every 1 hydrophobic macromolecule in the original WSOC mixture.

5.2   Molar volume sensitivity and uncertainty analysis

Application of the sensitivity analysis requires quantification of the uncertainty for all parameters that affect $\left(\frac{M_j}{\rho_j} \right)$. $\Delta\sigma/\sigma$ is 2% (Sect. 4.2), $\Delta\omega$ is the standard deviation derived from the fit of s and d experimental data to the Köhler curve, $\Delta\upsilon_i$ is 0.5, $\Delta\upsilon_i$ is 0.2, and $\Delta\varepsilon_i$ and $\Delta\varepsilon_j$ are uncertainties associated with assuming organic aerosol density of 1.4 to 1.6 g mol^{-1} (Turpin and Lim, 2001).

The sensitivity analysis suggests that one of the largest source of uncertainty for molar volume estimates arises from ω (Tables 6, 7, and 8), which is not surprising, given that it cumulatively expresses CCN activity. Furthermore, ω may vary significantly in the presence of strong surfactants, as the concentration of solute at the critical diameter (which varies considerably over a range of s_c) controls σ. CCN with low s_c (large d) tend to have low WSOC concentration at activation and do not affect σ depression at the droplet layer as much as CCN of smaller d and higher s_c. For theses reasons, σ may contribute considerable uncertainty in organic molar volume estimates (up to 10% as shown) but can be the second largest source of uncertainty for samples containing strong surfactants (e.g., hydrophobics). In the case of hydrophobics, uncertainty from exact knowledge of σ may introduce even larger uncertainty in molar volume estimates. The uncertainty in the van't Hoff factor may also be an important source of uncertainty in estimated organic molar volume ($\sim$24%) even though inorganic fractions and uncertainties are small (especially for the BB and D-HPHOB samples).

Using an average organic mass density of 1.4 g cm^{-3} (Turpin and Lim, 2001) we find the organic component in the original sample (BB) to have an average molar mass of

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248±117 g mol^{-1}, the hydrophobics in D-HPHOB to be on average 780±231 g mol^{-1} and the hydrophilic component of D-HPHIL to be the lightest, with an average of 87±26 g mol^{-1} (Table 4).

6 Implications and summary

5   This study is focused on characterizing the properties of water-soluble organics found in fresh biomass burning aerosol. The aerosol, after collection upon filters, is dissolved in water and fractionated into hydrophobic and hydrophilic components using XAD-8 solid phase extraction, and subsequently desalted. The original, fractionated and desalted samples are then characterized for their surfactant properties and average

10 thermodynamic properties relevant for CCN activation (i.e., surface tension depression, solubility and average molar volume). Characterization of solubility and molar volume is done by combining CCN activity measurements with Köhler Theory using the method of "Köhler Theory Analysis" (KTA) (first introduced by Padró et al., 2007[1], and further developed here). The surface tension and CCN activity of these different samples are

15 measured with a KSV CAM 200 goniometer and a DMT Streamwise Thermal Gradient CCN Counter, respectively.

It was found that the less hygroscopic soluble hydrophobic fractions can readily activate at high supersaturations and depending on the presence of inorganic species, can exhibit larger CCN activity than pure $(NH_4)_2SO_4$. This phenomenon can be attributed

20 to "salting-out" of organics to the CCN surface layer from the presence of electrolytes; this would decrease surface tension and critical supersaturation. This hypothesis is supported by direct measurements of surface tension, as well as from measurements of CCN activity.

Surfactant behavior in the samples is attributed to the hydrophobic fraction that exhibit

25 common properties to HULIS. The presence of inorganic salts may enhance surface tension depression to an extent in which insoluble hydrophobic aerosol may be better CCN than their pure inorganic counterparts. This supports the suggestion by

Kiss et al. (2005), to include the interaction of inorganic and organic species effect on surface tension, whenever appropriate, in aerosol-cloud interaction studies.

Using an average organic mass density of $1.4\,\mathrm{g\,cm^{-3}}$ we infer the hydrophobics to be on average $780\pm231\,\mathrm{g\,mol^{-1}}$ and the hydrophilics to be on average $87\pm26\,\mathrm{g\,mol^{-1}}$. We also estimate the relative molar ratio of hydrophilics to hydrophobics to being 3:1. Solubility limits, seen as an abrupt change in the scale dependence of s_c from $d^{-1.5}$ to d^{-a} ($a<-1.5$), were not observed (Padró et al., 2007[1]).

The inferred molar volumes for the samples considered in this study are on average subject to 36% uncertainty. Most of the uncertainty arises from the slope of the $s_c - d$ fit and σ (Padró et al., 2007[1]), and v in the case of aggregate organics. Nevertheless, the molar volume estimates are in agreement with expected ranges for these compounds and suggest that KTA can be applied effectively to characterize CCN activity of water-soluble organic aerosol.

The study shown here presents a novel method to describe the complex detailed inorganic, organic and water vapor interactions within a Köhler theory framework appropriate for GCM parameterizations of aerosol-cloud interactions. We have demonstrated that this novel method can successfully be applied to a very complex aerosol and provide aggregate properties that comprehensively characterize its CCN activity. In future work, the properties from other sources of carbonaceous aerosol should be characterized as presented here. Over time, such efforts will provide a comprehensive set of constraints for physically-based assessments of the indirect effects of carbonaceous aerosol.

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Table 1. Composition of each sample considered in this study.

Sample	[a]WSOC (mg C L^{-1})	[b]Cl^- ($\mu g\, L^{-1}$)	[b]SO_4^{2-} ($\mu g\, L^{-1}$)	[b]NO_3^- ($\mu g\, L^{-1}$)
BB	850	17 510	19 698	20 834
HPHIL	40	399 031	7700	0
HPHOB	375	4058	27	0
D-BB	600	2438	4056	3310
D-HPHIL	130	7.5	15.3	0
D-HPHOB	450	15.6	53.8	0

[a] The uncertainty of WSOC measurements is 3–5%
[b] The uncertainty is 40, 200, and 133 $\mu g\, L^{-1}$ for Cl^-, SO_4^{2-}, and NO_3^- measurements, respectively.

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Table 2. Average α and β parameters of the Szyszkowski-Langmuir model for all samples considered.

Sample	α $(\text{mN m}^{-1}\,\text{K}^{-1})$	β^c (L mg^{-1})
BB	2.78	1.72×10^{-6}
HPHIL	1.00	5.78×10^{-7}
HPHOB	9.18	6.02×10^{-6}
D-HPHIL	1.54×10^{-1}	8.23×10^{-6}
D-HPHOB	6.98	4.62×10^{-6}

[c]Measurements obtained between 296 and 299°K.

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Table 3. Properties used for Köhler Theory Analysis of all samples.

Property (units)	BB	D-HPHIL	D-HPHOB
σ $(N\,m^{-1})$	6.83×10^{-2}	6.85×10^{-2}	3.53×10^{-2}
ω $(m^{-1.5})^{d}$	7.78×10^{-14}	3.86×10^{-14}	6.26×10^{-14}
$\upsilon_{(NH_4)Cl}$	2	2	2
$\upsilon_{(NH_4)_2SO_4}$	3	3	3
$\upsilon_{NH_4NO_3}$	2	2	2
$\varepsilon_{(NH_4)Cl}$	0.018	0	0
$\varepsilon_{(NH_4)_2SO_4}$	0.019	0	0
$\varepsilon_{NH_4NO_3}$	0.019	0	0
$\varepsilon_{organic}$	0.94	1	1
$\nu_{organic}$	1	2.5	1

[d] obtained only for the data with $s_c \leq 0.6\%$

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Table 4. Results of Köhler Theory Analysis.

Sample	$\frac{M_j}{\rho_j}$ (m^3 mol^{-1})	M_j [e] (g mol^{-1})
BB	1.6×10^{-4}	248
D-HPHIL	6.2×10^{-5}	87
D-HPHOB	5.6×10^{-4} (method $\mathbf{b_1}$)[f]	780 (method $\mathbf{b_1}$)[f]
	6.1×10^{-4} (method $\mathbf{b_2}$)	859 (method $\mathbf{b_2}$)

[e] Assuming an average density of 1400 kg m^{-3} (Turpin and Lim, 2001)
[f] Determined from data with s_c between 0.4% and 0.6%.

Table 5. Sensitivity of Molar Volume to its dependant parameters.

x	$\Phi_x = \frac{\partial}{\partial x}\left(\frac{M_j}{\rho_j}\right)$
σ	$\left(\frac{3\times256}{27}\left(\frac{M_w}{\rho_w}\right)^2\left(\frac{1}{RT}\right)^3\frac{\sigma^2\omega^{-2}}{\varepsilon_j\upsilon_j}\right)\left(\frac{M_j}{\rho_j}\right)^2$
ω	$\left(\frac{2\times256}{27}\left(\frac{M_w}{\rho_w}\right)^2\left(\frac{1}{RT}\right)^3\frac{\sigma^3\omega^{-3}}{\varepsilon_j\upsilon_j}\right)\left(\frac{M_j}{\rho_j}\right)^2$
$\upsilon_{(NH_4)Cl}$	$\left(\frac{\frac{\rho_{NH_4Cl}}{M_{NH_4Cl}}\varepsilon_{(NH_4)Cl}}{\varepsilon_j\upsilon_j}\right)\left(\frac{M_j}{\rho_j}\right)^2$
$\upsilon_{(NH_4)_2SO_4}$	$\left(\frac{\frac{\rho_{(NH_4)_2SO_4}}{M_{(NH_4)_2SO_4}}\varepsilon_{(NH_4)_2SO_4}}{\varepsilon_j\upsilon_j}\right)\left(\frac{M_j}{\rho_j}\right)^2$
$\upsilon_{(NH_4)NO_3}$	$\left(\frac{\frac{\rho_{(NH_4)NO_3}}{M_{(NH_4)NO_3}}\varepsilon_{(NH_4)NO_3}}{\varepsilon_j\upsilon_j}\right)\left(\frac{M_j}{\rho_j}\right)^2$
$\varepsilon_{(NH_4)Cl}$	$\frac{1}{\varepsilon_j\upsilon_j}\left(\frac{\rho_{NH_4Cl}}{M_{NH_4Cl}}\varepsilon_{NH_4Cl}\upsilon_{NH_4Cl}+\frac{\rho_{NH_4NO_3}}{M_{NH_4NO_3}}\upsilon_{NH_4NO_3}\right)\left(\frac{M_j}{\rho_j}\right)^2$
$\varepsilon_{(NH_4)_2SO_4}$	$\frac{1}{\varepsilon_j\upsilon_j}\left(\frac{\rho_{(NH_4)_2SO_4}}{M_{(NH_4)_2SO_4}}\varepsilon_{(NH_4)_2SO_4}\upsilon_{(NH_4)_2SO_4}+\frac{\rho_{NH_4NO_3}}{M_{NH_4NO_3}}\upsilon_{NH_4NO_3}\right)\left(\frac{M_j}{\rho_j}\right)^2$
v_j	$\frac{256}{27}\left(\frac{M_w}{\rho_w}\right)^2\left(\frac{1}{RT}\right)^3\frac{-\sigma^3\omega^{-3}\upsilon_j^{-2}}{\varepsilon_j}\left(\frac{M_j}{\rho_j}\right)^2+\left(\sum_{i\neq j}\frac{\frac{\rho_j}{M_i}\varepsilon_i\upsilon_i}{\varepsilon_j}\right)\upsilon_j^{-2}\left(\frac{M_j}{\rho_j}\right)^2$
ε_j	$\frac{256}{27}\left(\frac{M_w}{\rho_w}\right)^2\left(\frac{1}{RT}\right)^3\frac{-\sigma^3\omega^{-2}\varepsilon_j^{-2}}{\upsilon_j}\left(\frac{M_j}{\rho_j}\right)^2-\left(\begin{array}{l}\frac{\rho_{NH_4Cl}}{M_{NH_4Cl}}\frac{\varepsilon_{NH_4Cl}\upsilon_{NH_4Cl}}{\upsilon_j}(-\varepsilon_j^{-2})\\+\frac{\rho_{(NH_4)_2SO_4}}{M_{(NH_4)_2SO_4}}\frac{\varepsilon_{(NH_4)_2SO_4}\upsilon_{(NH_4)_2SO_4}}{\upsilon_j}(-\varepsilon_J^{-2})\\+\frac{\rho_{(NH_4)NO_3}}{M_{(NH_4)NO_3}}\frac{\upsilon_{(NH_4)NO_3}}{\varepsilon_j\upsilon_j}\\+\frac{\rho_{(NH_4)NO_3}}{M_{(NH_4)NO_3}}\frac{(1-\varepsilon_{NH_4Cl}-\varepsilon_{(NH_4)_2SO_4}-\varepsilon_j)\upsilon_{(NH_4)NO_3}}{\upsilon_j}(-\varepsilon_j^{-2})\end{array}\right)\left(\frac{M_j}{\rho_j}\right)^2$

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Table 6. Molar Volume Sensitivity Analysis for BB Sample.

Property x (units)	Δx	Φ_x (m^3 mol^{-1} x^{-1})	Molar volume uncertainty (%)
σ (N m^{-1})	1.37×10^{-3}	1.063×10^{-2}	9.7
ω (m$^{-1.5}$)	9.49×10^{-15}	6.25×10^{9}	39.3
υ_{NH_4Cl}	0.5	1.49×10^{-5}	4.9
$\upsilon_{(NH_4)_2SO_4}$	0.5	7.43×10^{-6}	2.5
$\upsilon_{NH_4NO_3}$	0.5	6.08×10^{-6}	2.0
ε_{NH_4Cl}	1.95×10^{-3}	9.34×10^{-4}	1.2
$\varepsilon_{(NH_4)_2SO_4}$	2.03×10^{-3}	9.22×10^{-4}	1.2
$\varepsilon_{organic}$	6.36×10^{-3}	1.09×10^{-3}	4.6
$v_{organic}$	0.20	1.78×10^{-4}	23.6
Total uncertainty			47.5

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Table 7. Molar Volume Sensitivity Analysis for D-HPHIL Sample.

Property x (units)	Δx	Φ_x ($m^3\ mol^{-1}\ x^{-1}$)	Molar volume uncertainty (%)
σ ($N\,m^{-1}$)	1.37×10^{-3}	3.90×10^{-3}	7.2
ω ($m^{-1.5}$)	4.43×10^{-15}	4.62×10^{9}	27.6
υ_{NH_4Cl}	0.5	3.58×10^{-8}	0.0
$\upsilon_{(NH_4)_2SO_4}$	0.5	2.70×10^{-9}	0.0
ε_{NH_4Cl}	6.89×10^{-5}	8.27×10^{-5}	0.0
$\varepsilon_{(NH_4)_2SO_4}$	1.31×10^{-5}	8.26×10^{-5}	0.0
$\varepsilon_{organic}$	8.2×10^{-5}	1.72×10^{-4}	0.0
$v_{organic}$	0.20	3.56×10^{-5}	9.6
Total uncertainty			30.1

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Table 8. Molar Volume Sensitivity Analysis for D-HPHOB Sample.

Property x (units)	Δx	Φ_x (m^3 mol^{-1} x^{-1})	Molar volume uncertainty (%)
σ (N m^{-1})	7.06×10^{-4}	5.53×10^{-2}	7.0
ω ($m^{-1.5}$)	4.43×10^{-15}	2.08×10^{10}	16.5
υ_{NH_4Cl}	0.5	2.65×10^{-7}	0.0
$\upsilon_{(NH_4)_2SO_4}$	0.5	3.38×10^{-7}	0.0
ε_{NH_4Cl}	3.63×10^{-6}	1.16×10^{-2}	0.0
$\varepsilon_{(NH_4)_2SO_4}$	0	1.16×10^{-2}	0.0
$\varepsilon_{organic}$	1.16×10^{-5}	1.23×10^{-2}	0.0
$v_{organic}$	0.20	6.50×10^{-4}	23.3
Total uncertainty			29.5

Table 9. Molar Volume Sensitivity Analysis for D-HPHOB at each supersaturation measured.

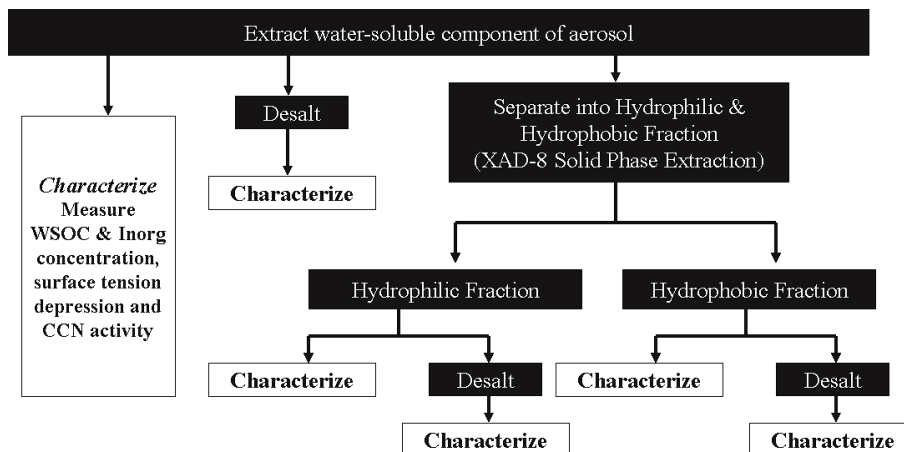| Quantity (units)	s=1.00%	0.80%	0.60%	0.50%	0.40%	0.30%	0.20%
$\left(\frac{M_j}{\rho_j}\right)$ ($m^3\,mol^{-1}$)	5.97×10^{-4}	5.80×10^{-4}	5.57×10^{-4}	4.45×10^{-4}	6.51×10^{-4}	5.98×10^{-4}	8.68×10^{-4}
σ ($N\,m^{-1}$)	3.53×10^{-2}	3.53×10^{-2}	3.53×10^{-2}	3.53×10^{-2}	3.53×10^{-2}	3.53×10^{-2}	3.53×10^{-2}
$\Delta\sigma$	7.06×10^{-4}	7.06×10^{-4}	7.06×10^{-4}	7.06×10^{-4}	7.06×10^{-4}	7.06×10^{-4}	7.06×10^{-4}
Φ_σ ($m^4\,mol^{-1}\,N^{-1}$)	5.93×10^{-2}	5.76×10^{-2}	5.42×10^{-2}	4.43×10^{-2}	6.47×10^{-2}	5.96×10^{-2}	8.65×10^{-2}
$\Phi_\sigma\Delta\sigma$	7.01%	7.02%	6.87%	7.02%	7.02%	7.03%	7.03%
ω ($m^{-1.5}$)	6.48×10^{-14}	6.39×10^{-14}	6.32×10^{-14}	5.60×10^{-14}	6.77×10^{-14}	6.48×10^{-14}	7.81×10^{-14}
$\Delta\omega$	4.43×10^{-15}	4.43×10^{-15}	4.43×10^{-15}	4.43×10^{-15}	4.43×10^{-15}	4.43×10^{-15}	4.43×10^{-15}
Φ_ω	2.15×10^{10}	2.12×10^{10}	2.02×10^{10}	1.86×10^{10}	2.25×10^{10}	2.16×10^{10}	2.60×10^{10}
$\Phi_\omega\Delta\omega$	15.96%	16.21%	16.04%	18.50%	15.31%	16.00%	13.28%
$\varepsilon_{organic}$	1.00	1.00	1.00	1.00	1.00	1.00	1.00
$\Delta\varepsilon_{organic}$	8.47×10^{-4}	8.47×10^{-4}	8.47×10^{-4}	8.47×10^{-4}	8.47×10^{-4}	8.47×10^{-4}	8.03×10^{-19}
$\Phi_{\varepsilon organic}$ ($m^3\,mol^{-1}$)	0	1.33×10^{-2}	1.22×10^{-2}	7.83×10^{-3}	1.70×10^{-2}	1.41×10^{-2}	2.91×10^{-2}
$\Phi_{organic}\Delta\varepsilon_{organic}$	0.00%	1.94%	1.86%	1.49%	2.21%	1.99%	0.00%
$\upsilon_{organic}$	1.00	1.00	1.00	1.00	1.00	1.00	1.00
$\Delta\upsilon_{organic}$	0.2	0.2	0.2	0.2	0.2	0.2	0.2
$\Phi_{\upsilon organic}$ ($m^3\,mol^{-1}$)	0	6.57×10^{-4}	6.24×10^{-4}	4.07×10^{-4}	1.11×10^{-3}	6.41×10^{-4}	8.35×10^{-4}
$\Phi_{\upsilon organic}\Delta\upsilon_{organic}$	0.00%	22.65%	22.40%	18.27%	34.14%	21.44%	19.23%

Fig. 1. Procedure used for sample analysis. The water-soluble component of the aerosol sample is extracted and characterized for carbon and inorganic content, surface tension and CCN properties. The sample is then desalted, fractionated into hydrophilic (HPHIL), hydrophobic (HPHOB), desalted hydrophilic (D-HPHIL) and desalted hydrophobic (D-HPHOB) components, all of which are characterized.

Fig. 2. Experimental setup for measuring CCN activity. Atomized sample is dried, charged and classified with a DMA. The classified aerosol stream is split into the CPC and the CCN instrument.

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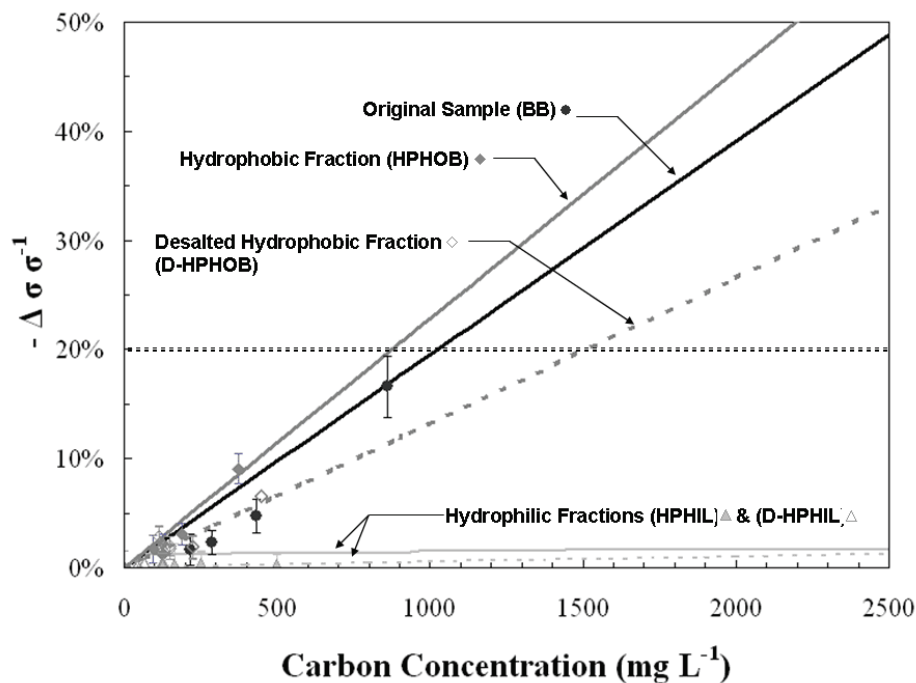**Fig. 3.** Example of CCN/CN data (D-HPHIL) with their corresponding sigmoidal fits.

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Fig. 4. Surface tension of fractionated biomass burning samples. Effective surface tension curves are based on Szyszkowski-Langmuir fit to data at room temperatures. The original sample (solid black), hydrophobic (dark grey), and hydrophilic fractions (light grey) are shown. The desalted components are represented by dashed lines. The double dashed line indicates a surface tension depression of 20%.

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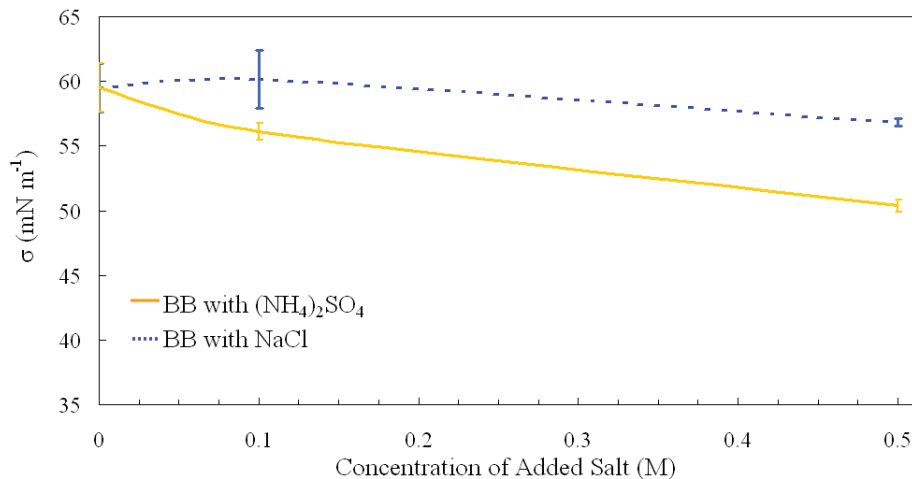Back | Close

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Fig. 5. Surface Tension of BB sample with the addition of $(NH_4)_2SO_4$ and NaCl. WSOC concentration is constant at $850\,\mathrm{mg\,L^{-1}}$).

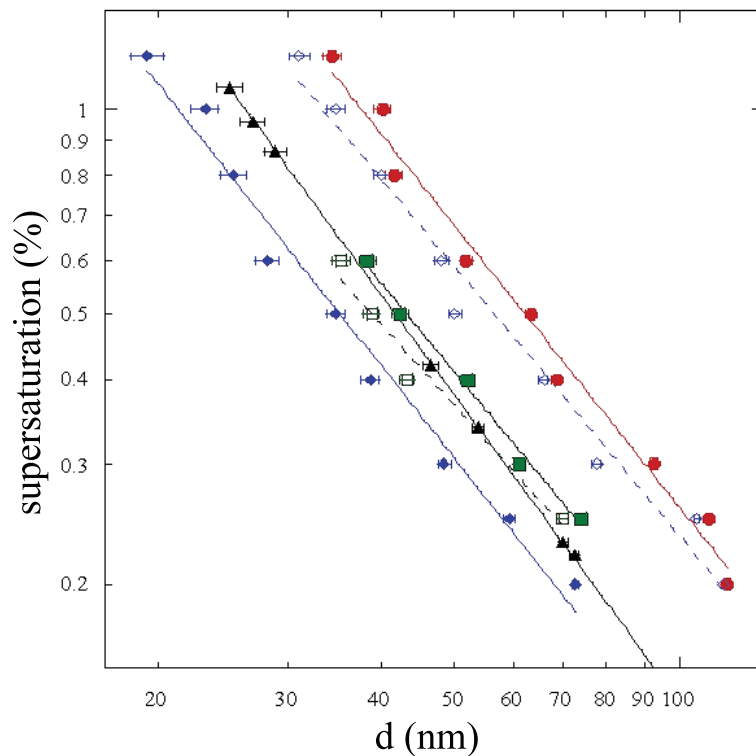**Fig. 6.** Critical supersaturation vs. dry particle diameter for BB Fractions and $(NH_4)_2SO_4$. The original sample (solid red circles), hydrophobic (blue diamonds), hydrophilic (green squares) and $(NH_4)_2SO_4$ (black triangles) are shown. The desalted samples are represented by dashed lines and open symbols.

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