1 2	This file includes proposed changes to the original manuscript based on the reviewer comments. The altered sections are highlighted in yellow.
3	Pesticides in the atmosphere: a comparison of gas-particle
4	partitioning and particle size distribution of legacy and
5	current-use pesticides
6	
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16	Abstract

This study presents a comparison of seasonal variation, gas-particle partitioning and particle-phase size distribution of organochlorine pesticides (OCPs) and current-use pesticides (CUPs) in air. Two years (2012/2013) of weekly air samples were collected at a background site in the Czech Republic using a high-volume air sampler. To study the particle-phase size distribution, air samples were also collected at an urban and rural site in the area of Brno, Czech Republic, using a cascade impactor separating atmospheric particulates according to six size fractions. Major differences were found in the atmospheric distribution of OCPs and CUPs. The atmospheric concentrations of CUPs were driven by agricultural activities while secondary sources such as volatilization from surfaces governed the atmospheric concentrations of OCPs. Moreover, clear differences were observed in gas-particle partitioning; CUP partitioning was influenced by adsorption onto mineral surfaces while OCPs were mainly partitioning to aerosols through absorption. A predictive method for estimating the gas-particle partitioning has been derived and is proposed for polar and nonpolar pesticides. Finally, while OCPs and the majority of CUPs were largely found on fine 

particles, four CUPs (carbendazim, isoproturon, prochloraz and terbuthylazine) had higher
 concentrations on coarse particles (>3.0 μm), which may be related to the pesticide
 application technique. This finding is particularly important and should be further investigated
 given that large particles result in lower risks from inhalation (regardless the toxicity of the
 pesticide) and lower potential for long range atmospheric transport.

# 6 1 Introduction

7 In 1939, dichlorodiphenyltrichloroethane (DDT) was discovered to have insecticidal properties. Since that time, synthetic pesticides have been widely used around the world to 8 9 control pests in agricultural production (Li and Macdonald, 2005). Legacy organochlorine pesticides (OCPs) are banned for agricultural purposes in most countries, including the Czech 10 Republic, and have been replaced by what are often termed "current-use pesticides" (CUPs). 11 CUPs generally have lower persistence and bioaccumulative potential and higher water 12 solubility, which should result in reduced negative environmental impacts (Kannan et al., 13 14 2006). However, given their detection in multiple environmental media, including in remote locations (Koblizková et al., 2012; Zhang et al., 2013), and the relative lack of information 15 16 regarding their toxic effects, the potential environmental and human risks cannot be neglected. The atmospheric transport of OCPs has been well studied over the last decades 17 (Lammel et al., 2009; Růzicková et al., 2008; Shen et al., 2004), but there is a lack of 18 19 information on the seasonal trends and partitioning of CUPs which is needed to understand their environmental fate. There are three main processes leading to the presence of pesticides 20 in the air. First, pesticides can enter the atmosphere during application. For example, during 21 spray application, up to 30% of the dosage directly enters the atmosphere (Van den Berg et al., 22 1999). Another primary emission is wind erosion of soil particles containing sorbed 23 pesticides, which can occur days or weeks after application (Glotfelty et al., 1989). Finally, 24 pesticides are affected by air-surface exchange such as the volatilization from plants and soils, 25 surface waters, and from old industrial sites (Cabrerizo et al., 2011). In the case of pesticides 26 that are not currently authorized for agricultural use (e.g., OCPs), volatilization and wind 27 28 erosion of soil particles should be the only relevant emission pathways.

Once pesticides enter the air, they partition between gas and particulate phases according to their physicochemical properties (vapor pressure, octanol-air partition coefficient  $K_{oa}$ ), the concentration of total suspended particulate matter (TSP) and meteorological parameters (ambient temperature, relative humidity) (Cousins and Mackay, 2001; Lohmann and Lammel,

2004; Pankow, 1987). Knowledge of this gas-particle partitioning is necessary to understand 1 atmospheric residence times, the significance of removal pathways from air (deposition, gas 2 absorption, photodegradation) and the potential for long range atmospheric transport (LRAT) 3 (Bidleman et al., 1986; Eisenreich et al., 1981; Schever et al., 2008). Additionally, the 4 atmospheric residence times of particles vary with particle size (Vecchi et al., 2007), further 5 influencing wet/dry deposition and LRAT (Götz et al., 2008). Particle size distribution is also 6 7 an important factor for human risks from inhalation exposure, as smaller particles penetrate deeper into the respiratory system (Englert, 2004). 8

The gas-particle partitioning of OCPs (Cindoruk, 2011; Schever et al., 2008; Sofuoglu et al., 9 2004) and some CUPs (Borrás et al., 2011; Götz et al., 2007; Sadiki and Poissant, 2008; 10 Sanusi et al., 1999; Sauret et al., 2008; Yao et al., 2008) has been reported. However, the 11 seasonal variation of this partitioning has only been investigated for two CUPs: chlorpyrifos 12 (Li et al., 2014) and alachlor (Sauret et al., 2008). Similarly, knowledge of how CUPs are 13 distributed among different particle sizes is very limited (Coscollà et al., 2014, 2013b), and 14 the seasonality of this particle size distribution has never been investigated for CUPs. To fill 15 these gaps, we assess the seasonal variation of a set of legacy and current-use pesticides 16 (Table 1) in outdoor air, with a focus on the gas-particle partitioning and the particle size 17 distribution. For many of these CUPs, this is the first time that their seasonal gas-particle 18 partitioning and size distributions have been examined. 19

20

# 21 2 Methodology

### 22 <u>2.1 Air sampling</u>

Air samples were collected in two sampling campaigns. A map of the sampling sites is 23 provided in Fig. S1 in the Supplementary Information (SI). Firstly, to study seasonal trends 24 and gas-particle partitioning, air was sampled at the Košetice observatory (49°34'24"N, 25 15°04'49"E), which is an established background site of the European Monitoring and 26 27 Evaluation Programme (EMEP) network (Holoubek et al., 2007). The site is located in an agricultural region in central Czech Republic. While the site is located in an agricultural 28 29 region, it is not directly on cultivated land, therefore the air sampled should not reflect direct emissions from pesticide application (e.g., spray application droplets) but rather the average 30

conditions of a rural air mass. From January 2012 to December 2013, a high volume air 1 sampler (Digitel DH77 with PM<sub>10</sub> pre-separator) was used to collect weekly air samples. The 2 sample volume was on average 4310 m<sup>3</sup> ( $\sim 25 \text{ m}^3/\text{h}$ , 7 day sampling duration). Particles were 3 collected on quartz fiber filters (QFFs) (QM-A, 150 mm, Whatman, UK, pore size of 2.2 µm) 4 and gas phase on polyurethane foam (PUF) (two in series, T3037, 110 x 50 mm, 0.030 g/cm<sup>3</sup>, 5 Molitan a.s., Czech Republic). PUFs were pre-cleaned via Soxhlet-extraction with acetone 6 7 and dichloromethane for 8 hours each. Fifty-two samples were collected each year. Half of the samples were used for OCPs analysis and half for CUPs analysis (Tables S1 and S2). 8

Secondly, to assess the seasonal variation of the particle size distribution of pesticides, 9 particulate-phase air samples were collected in the area of Brno, the second largest city in the 10 Czech Republic. From October 2009 to October 2010, a high volume air sampler (HV 100-P, 11 Baghirra, CZ) equipped with a multistage cascade impactor (PM<sub>10</sub> sampling head and six 12 stage impactor, Tisch Environmental, USA) was used to collect six particle size fractions. The 13 fractions represented particles with aerodynamic diameters of <0.49 µm, 0.49-0.95 µm, 0.95-14 1.5 µm, 1.5-3.0 µm, 3.0-7.2 µm and 7.2-10 µm and were collected on QFFs (TE-230-QZ, 141 15 x 148 mm, Tisch, Environmental, USA and OM-A, 203 x 254 mm, Whatman, UK, for the 16 backup filters (<0.49 µm)). Sampling was conducted simultaneously at a rural site (Telnice) 17 and at an urban site (Kotlařská). The rural site (49°6'21"N, 16°42'58"E) was located 14 km 18 southeast of the Brno city centre. The main source of pollution at this site is likely agricultural 19 activity, especially from cereals and grapes, which are the main local crops. The urban site 20 (49°12'20"N, 16°35'50"E) was located in a university botanical garden, close to a major 21 22 traffic junction in the centre of Brno. Only a small amount of pesticides are used within the botanical garden, and do not include any of the target pesticides in the present study. The main 23 sources of pesticides at this site are likely pesticides used in nearby buildings/building 24 materials, and atmospheric transport from the agricultural areas surrounding Brno. Eleven 25 weekly samples were used for CUPs analysis and twelve for OCPs analysis at each site. The 26 remaining samples were analyzed for other SVOCs, presented elsewhere (Degrendele et al., 27 2014; Okonski et al., 2014). To reach the limit of detection of these compounds, samples were 28 grouped by season (two or three filters) (Tables S3 and S4). The sample volume was on 29 average 9734 m<sup>3</sup> ( $\sim$ 65 m<sup>3</sup>/h, 7 day sampling duration). 30

All filters and PUFs were wrapped in aluminum foil, sealed in plastic bags and stored at 18°C until analysis.

#### 1 <u>2.2</u> Sample preparation and analysis

Filters and PUFs were extracted with toluene for OCP analysis and with methanol for CUP 2 3 analysis, using an automated warm Soxhlet extractor (Büchi Extraction System B-811) for three cycles, each consisting of 60 min of warm Soxhlet and 30 min of solvent rinsing. The 4 5 extracts were concentrated using a gentle stream of nitrogen. After extraction, OCP extracts 6 were transferred to a glass column (30 mm i.d.) consisting of 0.5 g of activated silica, 30 g of 7 H<sub>2</sub>SO<sub>4</sub>-modified activated silica and 1 g of non-activated silica and were eluted with 240 mL 8 of DCM:Hexane (1:1 v/v). CUP extracts were passed through syringe filters (nylon membrane, 25 mm diameter, pore size 0.45 µm). 9

OCPs were analyzed by gas chromatography coupled to a tandem mass spectrometer (GC-10 MS/MS). CUPs were analyzed using an Agilent 1100 high performance liquid chromatograph 11 (HPLC) with a Phenomenex Luna C-18 endcapped analytical column (100 mm x 2.1 mm x 3 12 μm). Analyte detection was performed by tandem mass spectrometry using an AB Sciex Qtrap 13 5500 operating in positive electron spray ionization (ESI+). Further information on all 14 analytical parameters is given in the SI. Identification was based on a comparison of ion ratios 15 16 and retention times (Table S5) with corresponding isotopically-labeled standards for CUPs and quantification was using internal standards: PCB-121 (Absolute Standards Inc., USA) for 17 18 OCPs and alachlor-d13, acetochlor-d11, chlorpyrifos d-10, isoproturon d-3, fenitrothion d-6, desisopropylatrazine d-5, dimethoate d-6, diuron d-6, terbuthylazine d-5 and simazine d-10 19 (Toronto Research Chemicals, Canada; Dr. Ehrenstorfer LGC Standards, UK; Chiron AS, 20 Norway; and Neochema, Germany) for CUPs. 21

#### 22 <u>2.3 QA/QC</u>

Breakthrough of gas phase compounds during air sampling was evaluated by separate 23 quantification of each of the two PUFs placed in series for all the weekly air samples 24 collected at the background site in 2012 (Tables S6 and S7). Based on the results of the 25 breakthrough evaluation, the sampling set-up was deemed appropriate for the quantification 26 of this set of pesticides. Thirteen field blanks and 28 laboratory blanks were analyzed as per 27 samples. Blank levels of individual analytes were below detection (all OCPs and 21 CUPs 28 were below detection in field blanks) or otherwise low (on average <3.5% of sample mass for 29 detected compounds). The concentrations of OCPs and CUPs presented here have been blank 30 corrected by subtracting the average of the field blanks. The OCP analytical method was 31

evaluated using a certified reference material (ASLAB soil standard, Czech Republic)
(Lohmann et al., 2012) and recoveries were assessed using spike-recovery tests of air
sampling media. Mean OCP recoveries (± standard deviation) ranged from 87.2±6.26% to
113±6.10% with an average value of 95.8±8.11% (Table S8). CUP recoveries were
determined from spike-recovery tests of air sampling media and ranged from 52.4±21.4% to
115±17.4% (Table S9). The measured concentrations have not been adjusted for recoveries.

7

## 8 **3** Results and discussion

#### 9 <u>3.1 Detection frequency at the background site</u>

In general, the detection frequency of CUPs related to their legal status, usage amounts and their persistence in the environment, while OCPs were consistently detected (>57% of samples) throughout the whole sampling period (Table 1). In particular,  $\alpha$ -HCH,  $\gamma$ -HCH, p,p'-DDE and p,p'-DDT were detected in every gas phase sample during the two years of sampling, emphasizing the environmental persistence of these OCPs.

The CUPs included in this study represent 24% of all pesticides used in agriculture in the 15 Czech Republic (Tables S10 and S11), with acetochlor, chlorpyrifos, chlorotoluron, 16 17 isoproturon, metamitron, metazachlor, prochloraz and terbuthylazine used in the largest quantities (>90 tonnes of active substance per year) and these CUPs were detected in >25% of 18 air samples. Isoproturon (detected in 86.5% of samples), metazachlor (86.5%), chlorpyrifos 19 (84.6%), terbuthylazine (78.8%), S-metolachlor (73.1%) and fenpropimorph (65.4%) were the 20 most frequently detected. Acetochlor, atrazine, carbendazim, chlorotoluron, dimethachlor, 21 diuron, metamitron, metribuzin, prochloraz and pyrazon had detection frequencies of 15-55% 22 (Table 1), occurring mostly during periods of agricultural activities. Finally, azinphos methyl 23 and fenitrothion were not detected in any samples and eight CUPs (alachlor, diazinon, 24 dimethoate, disulfoton, fonofos, malathion, simazine, temephos and terbufos) were 25 infrequently detected (<6%). Amongst these infrequently detected pesticides, only dimethoate 26 27 is authorized for agricultural use in the Czech Republic and is used in very low amounts (Tables S10 and S11). Thus, the infrequent detections of these compounds are likely due to no 28 or limited application in the sampling area. 29

We note that not all the CUPs are in current use in Czech Republic (Table 1); some pesticides, 1 which we have categorized as CUPs to distinguish them from the OCPs, are banned in Czech 2 Republic but remain in use elsewhere. For example, atrazine, a triazine pesticide banned in 3 the European Union since 2003 (European Commission - Health & Consumer Protection 4 5 Directorate-General, 2003) remains one of the highest use pesticides in USA (U.S. Environmental Protection Agency, 2013). Atrazine was detected in only one sample from May 6 7 2012 but had more frequent detections between July and November 2013 (Tables S12 and S13). 8

## 9 <u>3.2 Total concentrations at the background site</u>

10 Individual OCP and CUP concentrations are presented in Tables 1 and S12-S15.

11 Chlorpyrifos, metazachlor, acetochlor, isoproturon and S-metolachlor were the only CUPs with maximum total (gas+particulate phase) concentrations exceeding 100 pg.m<sup>-3</sup>, and, except 12 S-metolachlor, these pesticides are all used in quantities >100 tonnes/year in the Czech 13 Republic (Tables S10 and S11). Similarly, carbendazim, chlorotoluron, dimethachlor, 14 fenpropimorph, metamitron and terbuthylazine, which are all authorized for agricultural use 15 and used in quantities >30 tonnes/year (SRS, 2014, 2013), have maximum concentrations 16 higher than 10 pg.m<sup>-3</sup>. However, beyond this broad categorization, a poor correlation was 17 found between mass used per year and maximum concentration ( $r^2=0.362$  and 0.184 in 2012) 18 and 2013, respectively). For example, prochloraz, which was used in similar quantities to 19 chlorpyrifos in 2013 (SRS, 2014), had maximum concentrations of only 1.95 pg.m<sup>-3</sup> (vs. 159 20 pg.m<sup>-3</sup> for chlorpyrifos). The lack of correlation may be caused by the use of a national 21 pesticide usage database obscuring regional differences, which are of importance given the 22 relatively low atmospheric residence time of CUPs (Coscollà et al., 2013b). Moreover, the 23 pesticide physicochemical properties, their environmental persistence and the pesticide 24 application technique used (e.g., seed treatment vs. spray application) may also influence the 25 atmospheric concentrations of CUPs. Indeed, spray application parameters such as the 26 volatility and viscosity of the pesticide formulation, equipment, weather conditions at the time 27 of application (wind speed and direction, temperature, relative humidity and stability of air at 28 the application site) and operator care, attitude and skill have been identified as factors that 29 influence the emission of pesticide droplets to the air (Gil and Sinfort, 2005), thereby 30

31 affecting local air concentrations.

All of the banned CUPs included in this study had maximum concentrations lower than 2.5 1 pg.m<sup>-3</sup> (excepting fonofos with a concentration of 8.03 pg.m<sup>-3</sup> in one sample from August 2 2013), reflecting low current emissions. In particular, atrazine had a maximum concentration 3 of 1.24 pg.m<sup>-3</sup> in 2012 and lower concentrations (<0.250 pg.m<sup>-3</sup>) in 2013. The level of 4 simazine in the single sample in which it was detected was very low ( $<0.1 \text{ pg.m}^{-3}$ ). Similarly, 5 in a recent study, these CUPs were detected in only one sample over the Central North Sea at 6 low concentrations (<1 pg.m<sup>-3</sup>) (Mai et al., 2013). In contrast, from 1984-1994 (before the 7 European ban), atrazine and simazine were frequently detected in precipitation (Dubus et al., 8 2000). These triazines were also routinely detected in atmospheric samples in France during 9 the same period with concentrations up to 51 ng.m<sup>-3</sup> for atrazine (Sanusi et al., 2000) and 3 10 ng.m<sup>-3</sup> for simazine (Chevreuil et al., 1996). Thus, the low atmospheric concentrations of 11 atrazine and simazine observed in this study are likely a result of the European ban on use. 12

13 Of the OCPs, p,p'-DDE,  $\gamma$ -HCH and  $\alpha$ -HCH had the highest contributions, accounting on 14 average for 56.3%, 15.5% and 11.7% of  $\Sigma$ OCPs. The ratio of p,p'-DDT/(p,p'-DDE+p,p'-15 DDD) is often used as an indicator of aged technical DDT. A lower ratio is indicative of aged 16 (degraded) DDT, while a value >1 indicates fresh application (Li et al., 2007). In this study, 17 this ratio ranged from 0.0271 to 0.370, suggesting aged DDT.

The total concentrations of individual CUPs and OCPs were compared with previous studies
(Table S16 and references therein). OCPs levels were comparable to other European

background sites (Cabrerizo et al., 2011; Halse et al., 2011). ΣDDT concentrations in this
 study (1.14-96.3 pg m<sup>-3</sup>) were considerably lower than those reported in India or in Africa (8-

5930 and 8-2178 pg m<sup>-3</sup>, respectively (Bogdal et al., 2013; Yadav et al., 2015)). The CUPs

23 concentrations reported here were similar to those in the German Bight and North Sea (Mai et

al., 2013), but were generally much lower than in Canada (Hayward et al., 2010; Yao et al.,

25 2008), USA (Majewski et al., 2014; Peck and Hornbuckle, 2005) and France (Coscollà et al.,

26 2013c, 2011; Sauret et al., 2008; Scheyer et al., 2008; Schummer et al., 2010).

#### 27 <u>3.3</u> Seasonal variations at the background site

Concentrations of  $\sum OCPs$  and  $\sum CUPs$  were lowest in January-February and highest in August-September for  $\sum OCPs$  and in April-May for  $\sum CUPs$ . Individual CUPs and OCPs with consistent detection (>25% of samples) were generally grouped according to their seasonal trends (Fig. 1). The first group (group A, Fig. 1a) comprises compounds with one growing season concentration peak (April-September). The second group (group B, Fig. 1b) comprises
 compounds with two peaks, one during the growing season and the second in the plowing
 season (October-November).

4 Acetochlor, fenpropimorph, S-metolachlor and terbuthylazine are in group A and had 5 maximum concentrations in the April-July period. Dimethachlor and metazachlor are also 6 included in this group but had later peaks, during August-September. These two compounds 7 are used for oil plants and are usually applied later in the summer for weed control of winter 8 grains; this may explain their later maximum concentrations, as has been previously reported for metazachlor (Mai et al., 2013). The peak in concentrations of CUPs in this group is likely 9 associated with the fresh application of pesticides, but also with a contribution from 10 volatilization from soils, plants and surface water at higher temperatures. However, in the case 11 12 of acetochlor, fenpropimorph and S-metolachlor, which had maximum concentrations during April-May, their total concentrations seemed predominantly influenced by agricultural 13 activity rather than volatilization, as the timing of the peak corresponded with the application 14 season (April-May) rather than with the highest summer temperatures (July-August). A 15 similar pattern of high concentrations during the growing season has been previously reported 16 for acetochlor, alachlor, dimethoate and terbuthylazine (Hayward et al., 2010; Mai et al., 17 18 2013; Peck and Hornbuckle, 2005).

Group B comprises chlorpyrifos, isoproturon, prochloraz, chlorotoluron, diuron, and likely 19 metribuzin, although this is less conclusive due to more limited detection. The first group B 20 peak is attributed to the same factors as described for group A. The off-season (second peak) 21 22 concentrations are attributed to direct application of pesticides for future cereal crops which usually take place during autumn (Garthwaite et al., 2014). Moreover, volatilization from pre-23 treated seeds, plants, soils and water and wind erosion facilitated by the plowing of fields, 24 which usually take place during this period, may also contribute to the second peak. In the 25 case of soil volatilization, these compounds, except for isoproturon and metribuzin, are 26 moderately persistent in the soil (Table 1; half-life in soil>45 days) and thus, once they have 27 entered the soil from application or deposition, higher soil concentrations may persist unless 28 anthropogenic soil activity such as plowing occurs. However, terbuthylazine also has 29 moderate persistence in soil and did not have an autumn peak. It is notable that the peak 30 concentrations of chlorpyrifos, isoproturon and chlorotoluron were generally higher (up to 31 4.15 times) in autumn compared to the growing season, suggesting that, for these compounds, 32

autumn emissions are a larger source than emissions during the growing season. October November peaks of chlorpyrifos have been previously reported in China (Li et al., 2014) and
 Canada (Hayward et al., 2010).

Carbendazim, a fungicide used mostly for oil plants, had a single growing season peak in
2012 (in April-June) and two peaks in 2013 (one in May-June and one in SeptemberOctober). Additionally, this compound had a relative high concentration (12.1 pg.m<sup>-3</sup>) during
the last sampled week (18-25/12/13). It is unclear what caused these differences between the
two study years.

9 Of the OCPs,  $\beta$ -HCH,  $\gamma$ -HCH and o,p'-DDD followed the group A seasonal trend, with one 10 peak occurring between May-August. p,p'-DDE, o,p'-DDT and p,p'-DDT behaved as per 11 group B, with two peaks each year. Other OCPs did not have clear seasonal variations. In 12 general, the seasonal trends observed for OCPs were much less pronounced than for CUPs 13 (Fig. S2). For example, the ratio of summer-to-winter concentrations of OCPs ranged from 14 0.758 (p,p'-DDD) to 6.54 (p,p'-DDT) with an average value of 2.90, while for CUPs, it 15 ranged from 0.188 (diuron) to 167 (metazachlor) with an average value of 28.4.

The seasonal variability in pesticides is related to and indicative of the sources of the pesticide. The major cause of the seasonal variability in OCPs is expected to be seasonality in volatilization from soils and other surfaces, thus seasonal variability should be related to temperature variability. Conversely, when seasonality is driven by use/application, as for the CUPs, the relationship with temperature should be weaker and the summer/winter ratios should be greater.

An examination of the temperature dependence using the Clausius-Clapeyron equation (see SI) supported this hypothesis. The gaseous pesticide concentrations were expressed as linear regressions of the natural logarithm of partial pressure versus the inverse of temperature (Hoff et al., 1998):

$$lnP = \frac{m}{T} + b$$
 (Eq. 1)

where m and b are the slope and the intercept of the linear regression, respectively. Partial
pressures of individual compounds were calculated for each sample using gas phase
concentrations and the ideal gas law.

The temperature-dependence of gas-phase concentrations was statistically significant at the 1 99% confidence level for all OCPs except  $\alpha$ -HCH, with slopes ranging from -2792 ( $\delta$ -HCH) 2 to -9802 (*p*,*p*'-DDT), indicating that OCP concentrations increased with air temperature 3 (Table S17). Generally, a steep slope and high  $r^2$  indicate that temperature-controlled air-4 surface cycling and short-term transport influenced the ambient gas phase concentrations 5 (Hoff et al., 1998; Wania and Haugen, 1998), while a shallow slope and low  $r^2$  suggest that 6 7 other factors (i.e., advection, primary sources, atmospheric deposition, degradation) and LRAT influenced concentrations (Lee et al., 2000). Thus, the Clausius-Clapeyron 8 9 relationships suggest that gas phase concentrations of all OCPs except  $\alpha$ -HCH were controlled by re-volatilization from surfaces close to the sampling site. Temperature 10 accounted for 23-84% of the variability in atmospheric concentrations for these compounds. 11 12 This is in agreement with a previous study showing that atmospheric levels of OCPs at different European background sites were controlled by air-soil exchange (Cabrerizo et al., 13 2011). The lower temperature dependence of  $\alpha$ -HCH suggested that air concentrations were 14 15 also influenced by LRAT or other confounding factors. For the CUPs which were sufficiently detected in the gas phase, only terbuthylazine and S-metolachlor had a significant temperature 16 17 dependency (Table S18). For some CUPs, their atmospheric lifetime in relation with OH reaction is relatively small (e.g. about 2 hours for chlorpyrifos (Muñoz et al., 2014)), which 18 may explain the lack of maximum concentrations observed during the warmest periods. These 19 results emphasize the difference in the sources of OCPs and CUPs, with the former being 20 influenced by volatilization while the latter are influenced by temperature-independent local 21 22 sources (notably pesticide application) or LRAT.

## 23 <u>3.4 Gas-particle partitioning at the background site</u>

It is well known that several sampling artifacts such as blow-on, blow-off, breakthrough and 24 degradation may occur and affect the results about gas-particle partitioning (Melymuk et al., 25 2014). The reported gas-particle partitioning of pesticides are therefore operationally defined, 26 given the sampling configuration, where gas-phase is defined as the mass of the sample 27 captured on the PUF and particulate-phase is the mass captured on the QFF. Given the large 28 volumes used in this study, breakthrough tests were performed (Table S7) and HCHs were 29 excluded from the discussion of gas-particle partitioning to avoid any bias due to gas-phase 30 breakthrough sampling. Breakthrough is typically the most significant sampling artifact; bias 31 due to filter blow-on/blow-off is not expected to be significant (Melymuk et al., in press). 32

In this study, the pesticides fall into three groups: (1) predominantly particulate phase, (2) 1 predominantly gas phase, and (3) those with significant gas and particulate phase fractions 2 (average measured particulate mass fraction,  $\theta_{\text{meas}}$ , 0.2<  $\theta_{\text{meas}}$ <0.8). Six CUPs (carbendazim, 3 4 chlorotoluron, diuron, fenpropimorph, isoproturon and prochloraz) were predominantly in the 5 particulate phase ( $\theta_{\text{meas}} > 0.84$ ). In particular, prochloraz, diuron and carbendazim (except in one sample in June 2012) were detected only in the particulate phase. A similar dominance of 6 7 the particulate phase has been reported for carbendazim (Mai et al., 2013) and fenpropimorph (Van Dijk and Guicherit, 1999), but diuron was reported to have an average  $\theta$  of 0.75 8 9 (Scheyer et al., 2008), which differs slightly from our results. Three CUPs (chlorpyrifos, acetochlor and dimethachlor) and all the OCPs were predominantly found in the gas phase 10 (average  $\theta_{\text{meas}}$  < 0.20). In particular, the average particulate-phase mass fractions of 11 chlorpyrifos, o,p'-DDE, p,p'-DDE and o,p'-DDT were <0.04 (Table 1). Of the OCPs, only 12 13 p,p'-DDD and p,p'-DDT had particulate phase fractions >0.10. The dominance of the gas phase for chlorpyrifos (Li et al., 2014; Sadiki and Poissant, 2008; Van Dijk and Guicherit, 14 1999) and OCPs (Cindoruk, 2011; Sadiki and Poissant, 2008; Sanusi et al., 1999) is well 15 documented. Finally, four CUPs (atrazine, metazachlor, S-metolachlor and terbuthylazine) 16 17 were distributed between gas and particulate phases, with average  $\theta_{\text{meas}}$  of 0.63, 0.59, 0.24 and 18 0.45, respectively. Significant correlations  $(0.20 < r^2 < 0.94$  and p<0.05) between air temperatures and the gas-19 particle partitioning coefficient ( $K_{p}$  in m<sup>3</sup>µg<sup>-1</sup>, see SI for details) were observed for all OCPs, 20 with higher particulate fractions associated with lower temperatures. Amongst the CUPs, the 21 measured  $K_{\rm p}$  of S-metolachlor and terbuthylazine also correlated with air temperatures 22  $(r^2=0.29 \text{ and } 0.28, \text{ respectively and } p<0.05)$ . The lack of observed relationships in the case of 23

(1 - 0.2) and (0.20), respectively and p(0.03). The lack of observed relationships in the case of

24 other CUPs suggests that the gas-particle partitioning of the majority of the CUPs is

25 determined by processes which are not or minimally sensitive to temperature.

26 To better understand the mechanisms influencing gas-particle partitioning of pesticides,  $K_p$ 

was compared with  $K_{oa}$  and with the soil-air partitioning coefficient ( $K_{sa}$ , dimensionless) for

four CUPs (chlorpyrifos, isoproturon, metazachlor and terbuthylazine) and three OCPs (p,p')-

29 DDD, p,p'-DDE and p,p'-DDT) (Fig. 2). The temperature-dependency of  $K_{oa}$  (Table S19 and

Fig. S3) was determined from published relationships (for all OCPs and chlorpyrifos) or from

- 31 extrapolation (remaining CUPs) based on regression analysis for other compounds and
- 32 validated for chlorpyrifos (Fig. S4). Details of the calculations can be found in the SI.  $K_{sa}$  data

- 1 used in this study were adopted from the relationship of Davie-Martin et al. (2015), based on
- 2 laboratory experiments on 22 OCPs and CUPs, as:

3 
$$\log K_{sa} = -26.2 + 0.714 \log K_{oa,298.15K} + \frac{8291}{T_{amb}} - 0.0128RH + 0.121 \log(100f_{oc})$$

4

(Eq. 2)

5 where,  $T_{amb}$  is the ambient temperature (K), RH is the relative humidity (%) and  $f_{OC}$  is the

6 organic carbon content of soil. Average monthly RH values and an experimental  $f_{oc}$  of 0.03

7 (Holoubek et al., 2009) were used.

Both  $K_{oa}$  and  $K_{sa}$  were significantly (p<0.05) correlated with  $K_p$  for both OCPs and CUPs (r<sup>2</sup> = 8 9 0.51-0.73; Fig. 2). However, clear differences were noted between these two classes of 10 compounds. For same  $K_{oa}$  (or  $K_{sa}$ ), the  $K_{p}$  values of CUPs were notably higher than those of the OCPs. This suggests that absorption into organic matter alone, described by  $K_{oa}$ , is not 11 sufficient to explain the observed gas-particle partitioning of CUPs and that other types of 12 13 interactions occur. In other words, while absorption into the organic matter fraction is the 14 dominant process for weakly or non-polar compounds such as OCPs, additionally adsorption to mineral surfaces or soot is significant for more polar compounds such as the CUPs. Indeed, 15 Götz et al., (2007) estimated that the contribution of OM to  $K_p$  was 74% for DDT but only 1 16 and 5% for isoproturon and terbuthylazine, respectively, for which adsorption to mineral 17 surfaces dominated  $K_p$  (contributions of 95 and 86%, respectively) and concluded that a 18 19 predictive model based only on absorptive contribution to organic matter is not recommended for polar compounds such as CUPs. Therefore, assuming absorption to govern sorption, such 20 as in the  $K_{oa}$  model (Harner and Bidleman; 1998), will generally lead to high agreement of 21 predicted  $K_p$  values with observations for OCPs but low agreement (underestimates) for CUPs 22 (Fig. S5-S6). Thus, a predictive model based only on absorptive contribution to organic 23 matter is not recommended for polar compounds such as CUPs (Götz et al., 2007). However, 24 we note that in Fig. 2a the two slopes intersect around  $\log K_{oa}$  of 13. This suggests that for the 25 few polar pesticides with  $12 < \log K_{oa} < 13$ , the  $K_{oa}$ -based approach is still appropriate. 26 Interestingly, the predicted method derived for soil-air partitioning which takes into account 27

both absorption ( $K_{oa}$ ) and adsorption (T, RH) (Davie-Martin et al., 2015) was a better predictor for gas-particle partitioning, given that similar slopes (0.857 ± 0.0332) were observed for OCPs and CUPs (Fig. 2b). The difference between the intercepts is 1.25. Based on this relationship, we propose an improved method for prediction of gas-particle
partitioning of pesticides:

$$\log K_p = 0.857 \cdot \left(-26.2 + 0.714 \log K_{oa,298.15 K} + \frac{8291}{T_{amb}} - 0.0128RH + 0.121 \log(100f_{oc})\right) - 10.5 + p$$

3

4 with p being a factor accounting for polarity, p = 1.25 for CUPs and p = 0 for OCPs. By using 5 eq. 3, good agreements between predicted and measured  $\log K_p$  of CUPs and OCPs are found

6 (rmse = 0.61, Fig. 3), with few exceptions for some CUPs.

7 The relationship between  $K_p$  and  $K_{sa}$  suggest similar partitioning from the gas phase to 8 aerosols and to soils including absorption to organic matter and adsorption to mineral 9 surfaces. In fact, the role of adsorption to mineral surfaces in air-soil exchange studies has 10 been rather neglected over the last decades (Mackay, 2001; Odabasi and Cetin, 2012a; Wang 11 et al., 2012; Wei et al., 2014) and should be further considered (Davie-Martin et al., 2015; 12 Goss et al., 2004).

The seasonal variation of the relationship of  $K_p$  with  $K_{oa}$  and  $K_{sa}$  was also examined (Fig. S7). 13 Given that many CUPs were mainly detected in spring and autumn, we choose these two 14 seasons (spring and autumn were defined from March to June and from September to 15 November, respectively). Interestingly, better correlations were obtained between  $K_{p}$  and  $K_{oa}$ 16 for CUPs in autumn compared to spring ( $r^2 = 0.71$  and 0.49, respectively, p<0.05) while there 17 was no variation for OCPs ( $r^2 = 0.63$ ). This suggests that a process other than absorption in 18 organic matter gains significance for CUPs in spring but less in autumn. This could be related 19 20 to a higher concentration (specific surface area) of mineral dust during the spring sampling period (adsorption, see above). Pesticide application technique could potentially affect gas-21 22 particle partitioning through mass transport kinetics limitations (non-equilibrium). Indeed, 10× higher particulate fractions were found for chlorpyrifos for two samples in spring 2012 23 24  $(\theta_{\text{meas}}=0.19 \text{ and } 0.33)$ , suggesting a potential influence of application or agricultural activities. However, there is limited knowledge in this area. The differences in RH in spring and autumn 25 (73.1 vs. 83.5 %, respectively) may also have contributed to the observed differences, as 26 water layers and deliquescence may affect OM accessibility (higher in spring). Götz et al., 27 (2007) estimated that the influence of absorption on  $K_{\rm p}$  for polar pesticides was negligible for 28 29 RH between 40 and 80% while it was dominant for RH>80% (contributing to 30-90%).

(Eq. 3)

Similarly, Davie-Martin et al., (2015) found that RH had a negligible effect on  $K_{sa}$  for 1 RH>80% for semi-arid soils. The influence of RH on  $K_p$  has generally not been considered in 2 predictive methods except with polyparameter linear free energy relationships (pp-LFER, 3 (Goss, 1997)), which considers all types of molecular interactions. Unfortunately, many input 4 parameters needed for this method are not available for CUPs or are associated with large 5 uncertainties (Davie-Martin et al., 2015; Götz et al., 2007). The new predictive  $K_p$  model 6 7 proposed in this study and based on easily accessible parameters allows consideration of both meteorological variables (RH, T), compound-specific properties ( $K_{oa}$ , factor accounting for 8 the polarity, p) and soil composition  $(f_{oc})$ . 9

10

#### 11 <u>3.5</u> Particle size distribution at the urban and rural sites

The particle size distribution of CUPs and OCPs was determined at the rural (Telnice) and 12 urban (Kotlářská) site. Only the pesticides with significant particle fractions (average 13  $\theta_{\text{meas}}$  >10%) and detection will be discussed in this section. However, because chlorpyrifos is 14 one of the most widely used insecticides in the world (Solomon et al., 2014), we also include 15 16 it in the further discussion. Amongst the pesticides with sufficient detection (Tables S20-S23), nine pesticides had highest concentrations on particles <0.95 µm, four pesticides had highest 17 18 concentrations in the  $>1.5 \mu m$  fraction and one pesticide showed no size distribution pattern. The seasonal size distributions of fenpropimorph and isoproturon are shown in Fig. 4 as 19 representative of the pesticides dominated by the fine and coarse fractions, respectively. 20

Particulate phase concentrations of  $\Sigma$ CUPs at the rural site ranged from 110 to 408 pg.m<sup>-3</sup> and 21 were higher than at the urban site ( $\Sigma CUPs = 30.3 - 112 \text{ pg.m}^{-3}$ ). In contrast, similar 22 concentrations were observed for  $\Sigma$ OCPs at the both rural (14.4 - 50.1 pg.m<sup>-3</sup>) and urban 23  $(18.2 - 42.2 \text{ pg.m}^{-3})$  sites. As suggested by the seasonal trends at the background site, this 24 25 indicates that current agricultural emissions are driving CUP concentrations, while OCPs are the result of diffuse pollution and thus do not have a strong urban-rural gradient. In general, 26 27 seasonal variations of particulate OCPs and CUPs were similar to those observed at the background site. However, in these samples, the second autumn peak was observed only for 28 29 diuron, isoproturon and chlorotoluron at lower concentrations than during the growing season.

30 One CUP (alachlor) had sporadic detection outside of the growing season and no clear trend 31 in particle size distributions at either site (Tables S20 and S22). Nine CUPs (acetochlor,

atrazine, chlorpyrifos, diuron, fenpropimorph, metazachlor, S-metolachlor, simazine and 1 terbufos) had higher concentrations on fine particles and were on average 35-76% associated 2 with particles <0.95 µm. This distribution did not shift significantly when concentrations were 3 normalized by particle mass in each size fraction (Table S22). To the best of our knowledge, 4 only one study has reported the particle size distribution of CUPs (Coscollà et al., 2013b) and 5 this included acetochlor and fenpropimorph in common with our study, also found largely on 6 7 fine particles. Similarly, p,p'-DDD and p,p'-DDT also had highest concentrations on fine particles (<0.95 µm), which accounted for 43-63% and 50-91% of the total particulate phase 8 9 mass, respectively. It is interesting to note that the size distribution of diuron, fenpropimorph and *p*,*p*'-DDD and *p*,*p*'-DDT did not show any variation by season or site. The presence of 10 these compounds in the fine fraction (per air volume and per particle mass) is attributed to the 11 12 sorption of gas phase pesticides to fine particles due to their higher surface area and the 13 coagulation of ultrafine to fine particles (Coscollà et al., 2013b). Moreover, as the mechanisms of wet and dry deposition are less efficient for removing particles in the 0.1-1 µm 14 15 and 0.05-2 µm size range respectively (Zhang and Vet, 2006), these compounds are expected to have higher atmospheric residence times compared to compounds which are mostly present 16 17 on coarse particles.

Four pesticides (carbendazim, isoproturon, prochloraz and terbuthylazine) were found 18 predominantly on coarse particles (>3.0  $\mu$ m) in all seasons at both sites. Indeed, when the 19 20 maximum total concentration occurred (i.e. in spring or summer), 45-70% of the total particulate phase mass of these compounds was on particles >3.0 µm. Similar size 21 22 distributions were observed when the concentrations were normalized by mass (Table S22). In general, coarse particles are the result of mechanical processes such as wind erosion of soil 23 particles and most of these pesticides are moderately persistent in the soil ( $DT_{50}$ =40-120 days) 24 and thus might be subject to wind erosion. The presence of pesticides on coarse particles 25 26 could also be related to the pesticide application technique, as it has been shown that the type 27 and amount of emissions during application (either drift or airborne residues) are strongly related to the application technique, and independent of the physicochemical properties of the 28 compound applied (FOCUS, 2008). A very wide range of application techniques are used; for 29 example, prochloraz exists as an emulsifiable concentrate, while carbendazim, isoproturon 30 and terbuthylazine mostly exist as soluble concentrates, and chlorpyrifos can be applied as 31 32 either a soluble concentrate or as solid particles directly to soil (PPDB, 2013). The fom, not measured in this study, may influence observed particle distributions, particularly given that 33

fine particles may contain a higher carbonaceous fraction (Putaud et al., 2004). The lack of form 1 data is a limitation in understanding the particle size distributions, however, we note that 2 individual samples (therefore with the same for values) had some CUPs predominantly found 3 4 on coarse particles and others predominantly found on fine particles, suggesting that factors other than f<sub>om</sub> are controlling their particle size distribution. We hypothesize that differences 5 in type of application (emulsifiable vs. soluble concentrates, type of spray application, 6 7 application to plants vs. soil vs. seeds) may lead to differences in the particle size distribution of pesticides, yet very little specific information is available on how particle size distribution 8 9 relates to application techniques.

Coarse particles have a shorter residence time in the atmosphere because they settle rapidly 10 and are efficiently removed by wet and dry deposition. Moreover, these particles are less 11 likely to penetrate deeply into the human respiratory system (Englert, 2004). Thus, should 12 these distributions apply on a wider scale, carbendazim, isoproturon, prochloraz and 13 terbuthylazine could be considered as pollutants with low risks of human inhalation exposure 14 (discarding the potential toxicity of individual substances) and LRAT potential. Additional 15 research on the link between pesticide application techniques and local/regional atmospheric 16 concentrations and distributions are needed in order to reduce inhalation exposure of 17 18 agricultural workers.

19

#### 20 4 Conclusions

21 Although OCPs have been banned for agricultural use decades ago, this study highlights the fact that they are still frequently detected in atmospheric samples at a background site in 22 Central Europe due to their persistence in environmental matrices. Presently, more than 270 23 24 plant protection products are registered for agricultural use in the Czech Republic (SRS, 2014) with limited knowledge on potential environmental and human risks. This study 25 improves knowledge of the characterization of atmospheric behavior of 27 CUPs, 26 27 representing about 24% of the national market and found three major differences than what is observed for OCPs. Firstly, regarding their seasonal variations, atmospheric concentrations of 28 29 CUPs were largely driven by agricultural practices while secondary sources such as volatilization from surfaces governed atmospheric concentrations of OCPs. Secondly, clear 30 differences were observed in gas-particle partitioning, with an influence of adsorption onto 31

mineral surfaces for CUPs while OCPs were mainly partitioning to aerosols through 1 absorption. Based on the recent work of Davie-Martin et al., (2015), a basic predictive method 2 for  $K_p$  is proposed for polar and non-polar pesticides, which relies on easily accessible 3 parameters. This method should be tested for other sampling sites and aerosol composition to 4 5 determine its broader utility for polar pesticides in cases when parameters needed for pp-LFER are not available. Finally, while OCPs and the majority of CUPs were largely found on 6 7 fine particles, four CUPs (carbendazim, isoproturon, prochloraz and terbuthylazine) had higher concentrations on coarse particles (>3.0  $\mu$ m) which may be caused by the pesticide 8 application technique. This finding is particularly important and should be further investigated 9 given that large particles results in lower risks from inhalation (regardless the toxicity of the 10 pesticide) and lower potential for long range atmospheric transport. 11

12

#### **13 Supporting Information**

Description of samples collected, analytical methods for CUPs and OCPs, usage of pesticides in the Czech Republic, Clausius-Clapeyron plots, description of calculation for predicted particulate fractions and atmospheric concentrations of individual CUPs and OCPs are provided.

18

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# 1 References

- Bidleman, T.F., Billings, W.N., Foreman, W.T., 1986. Vapor-particle partitioning of
   semivolatile organic compounds: estimates from field collections. Environ. Sci. Technol.
   20, 1038–43.
- Bogdal, C., Scheringer, M., Abad, E., Abalos, M., van Bavel, B., Hagberg, J., Fiedler, H.,
  2013. Worldwide distribution of persistent organic pollutants in air, including results of
  air monitoring by passive air sampling in five continents. TrAC Trends Anal. Chem. 46,
  150–161. doi:10.1016/j.trac.2012.05.011
- Borrás, E., Sánchez, P., Muñoz, A., Tortajada-Genaro, L.A., 2011. Development of a gas
  chromatography-mass spectrometry method for the determination of pesticides in
  gaseous and particulate phases in the atmosphere. Anal. Chim. Acta 699, 57–65.
  doi:10.1016/j.aca.2011.05.009
- Cabrerizo, A., Dachs, J., Jones, K.C., Barceló, D., 2011. Soil-Air exchange controls on
   background atmospheric concentrations of organochlorine pesticides. Atmos. Chem.
   Phys. 11, 12799–12811. doi:10.5194/acp-11-12799-2011
- Chevreuil, M., Garmouma, M., Teil, M.-J., Chesterikoff, A., 1996. Occurrence of
  organochlorines (PCBs, pesticides) and herbicides (triazines, phenylureas) in the
  atmosphere and in the fallout from urban and rural stations of the Paris area. Sci. Total
  Environ. 182, 25–37. doi:10.1016/0048-9697(95)05032-9
- Cindoruk, S.S., 2011. Atmospheric organochlorine pesticide (OCP) levels in a metropolitan
   city in Turkey. Chemosphere 82, 78–87. doi:10.1016/j.chemosphere.2010.10.003

Coscollà, C., Castillo, M., Pastor, A., Yusa, V., 2011. Determination of 40 currently used
 pesticides in airborne particulate matter (PM 10) by microwave-assisted extraction and
 gas chromatography coupled to triple quadrupole mass spectrometry. Anal. Chim. Acta
 693, 72–81. doi:10.1016/j.aca.2011.03.017

- Coscollà, C., Hart, E., Pastor, A., Yusà, V., 2013a. LC-MS characterization of contemporary
   pesticides in PM10 of Valencia Region, Spain. Atmos. Environ. 77, 394–403.
   doi:http://dx.doi.org/10.1016/j.atmosenv.2013.05.022
- Coscollà, C., Munoz, A., Borras, E., Vera, T., Rodenas, M., Yusa, V., 2014. Particle size
   distributions of currently used pesticides in ambient air of an agricultural Mediterranean
   area. Atmos. Environ. 95, 29–35. doi:10.1016/j.atmosenv.2014.06.022
- Coscollà, C., Yahyaoui, A., Colin, P., Robin, C., Martinon, L., Val, S., Baeza-Squiban, A.,
   Mellouki, A., Yusà, V., 2013b. Particle size distributions of currently used pesticides in a
   rural atmosphere of France. Atmos. Environ. 81, 32–38.
   doi:10.1016/j.atmosenv.2013.08.057
- Coscollà, C., Yahyaoui, A., Colin, P., Robin, C., Martinon, L., Val, S., Baeza-Squiban, A.,
   Mellouki, A., Yusà, V., 2013c. Particle size distributions of currently used pesticides in a
   rural atmosphere of France. Atmos. Environ. 81, 32–38.
- 39 doi:http://dx.doi.org/10.1016/j.atmosenv.2013.08.057

- Cousins, I.T., Mackay, D., 2001. Gas--particle partitioning of organic compounds and its
   interpretation using relative solubilities. Environ. Sci. Technol. 35, 643–647.
   doi:10.1021/es001123m
- Davie-Martin, C.L., Hageman, K.J., Chin, Y.-P., Rougé, V., Fujita, Y., 2015. Influence of
  temperature, relative humidity, and soil properties on the soil–air partitioning of
  semivolatile pesticides: Laboratory measurements and predictive models. Environ. Sci.
  Technol. 49, 10431–10439. doi:10.1021/acs.est.5b02525
- Begrendele, C., Okonski, K., Melymuk, L., Landlová, L., Kukučka, P., Čupr, P., Klánová, J.,
  2014. Size specific distribution of the atmospheric particulate PCDD/Fs, dl-PCBs and
  PAHs on a seasonal scale: Implications for cancer risks from inhalation. Atmos. Environ.
  98, 410–416. doi:http://dx.doi.org/10.1016/j.atmosenv.2014.09.001
- Dubus, I.G., Hollis, J.M., Brown, C.D., 2000. Pesticides in rainfall in Europe. Environ. Pollut.
   110, 331–344. doi:10.1016/S0269-7491(99)00295-X
- Eisenreich, S.J., Looney, B.B., Thornton, J.D., 1981. Airborne organic contaminants in the
- Eisenreich, S.J., Looney, B.B., Thornton, J.D., 1981. Airborne organic contaminants in the
   Great Lakes ecosystem. Environ. Sci. Technol. 15, 30–38. doi:10.1021/es00083a002
- Englert, N., 2004. Fine particles and human health--a review of epidemiological studies.
   Toxicol. Lett. 149, 235–42. doi:10.1016/j.toxlet.2003.12.035
- European Commission Health & Consumer Protection Directorate-General, 2003. Review
   report for the active substance atrazine [WWW Document].
- FOCUS, 2008. Pesticides in Air: Considerations for Exposure Assessment. Rep. Focus Work.
   Gr. Pestic. Air (FOCUS Air Group).
- Garthwaite, D.G., Hudson, S., Barker, I., Parrish, G., Smith, L., Pietravalle, S., 2014.
   Pesticide usage survey report 250. Arable crops in the United Kingdom 2012 (including aerial applications 2012). Department for Environment, Food & Rural Affairs.
- Gil, Y., Sinfort, C., 2005. Emission of pesticides to the air during sprayer application: A
  bibliographic review. Atmos. Environ. 39, 5183–5193.
  doi:http://dx.doi.org/10.1016/j.atmosenv.2005.05.019
- Glotfelty, D.E., Leech, M.M., Jersey, J., Taylor, A.W., 1989. Volatilization and Wind Erosion
   of Soil Surface Applied Atrazine ,. J. Agric. Food Chem. 546–551.
- Goss, K.-U., 1997. Conceptual Model for the Adsorption of Organic Compounds from the
   Gas Phase to Liquid and Solid Surfaces. Envir Sci Technol 31, 3600–3605.
   doi:10.1021/es970361n
- Goss, K.U., Buschmann, J., Schwarzenbach, R., 2004. Adsorption of organic vapors to air-dry
   soils: model predictions and experimental validation. Environ. Sci. Technol. 38, 3667–
   3673.
- Götz, C.W., Scheringer, M., MacLeod, M., Roth, C.M., Hungerbühler, K., 2007. Alternative
   Approaches for Modeling Gas–Particle Partitioning of Semivolatile Organic Chemicals:

- Model Development and Comparison. Environ. Sci. Technol. 41, 1272–1278.
   doi:10.1021/es060583y
- Götz, C.W., Scheringer, M., MacLeod, M., Wegmann, F., Hungerbühler, K., 2008. Regional
   differences in gas-particle partitioning and deposition of semivolatile organic
   compounds on a global scale. Atmos. Environ. 42, 554–567.
   doi:http://dx.doi.org/10.1016/j.atmoscapy.2007.08.033
- 6 doi:http://dx.doi.org/10.1016/j.atmosenv.2007.08.033
- Halse, a. K., Schlabach, M., Eckhardt, S., Sweetman, a., Jones, K.C., Breivik, K., 2011.
  Spatial variability of POPs in European background air. Atmos. Chem. Phys. 11, 1549– 1564. doi:10.5194/acp-11-1549-2011
- Hayward, S.J., Gouin, T., Wania, F., 2010. Levels and Seasonal Variability of Pesticides in the
   Rural Atmosphere of Southern Ontario. J. Agric. Food Chem. 58, 1077–1084.
   doi:10.1021/jf902898f
- Hoff, R.M., Brice, K.A., Halsall, C.J., 1998. Nonlinearity in the slopes of Clausius-Clapeyron
   plots for SVOCs. Environ. Sci. Technol. 32, 1793–1798.
- Holoubek, I., Dušek, L., Sáňka, M., Hofman, J., Čupr, P., Jarkovský, J., Zbíral, J., Klánová, J.,
  2009. Soil burdens of persistent organic pollutants Their levels, fate and risk. Part I.
  Variation of concentration ranges according to different soil uses and locations. Environ.
  Pollut. 157, 3207–3217. doi:10.1016/j.envpol.2009.05.031
- Holoubek, I., Klánová, J., Jarkovský, J., Kohoutek, J., 2007. Trends in background levels of
  persistent organic pollutants at Kosetice observatory, Czech Republic. Part I. Ambient air
  and wet deposition 1996-2005. J. Environ. Monit. 9, 557–63.
- Kannan, K., Ridal, J., Struger, J., 2006. Pesticides in the Great Lakes, in: Hites, R. (Ed.),
   Persistent Organic Pollutants in the Great Lakes, The Handbook of Environmental
   Chemistry. Springer Berlin Heidelberg, pp. 151–199. doi:10.1007/698 5 041
- Koblizková, M., Lee, S.C., Harner, T., 2012. Sorbent impregnated polyurethane foam disk
   passive air samplers for investigating current-use pesticides at the global scale. Atmos.
   Pollut. Res. 3, 456–462.
- Lammel, G., Klánová, J., Kohoutek, J., Prokeš, R., Ries, L., Stohl, A., 2009. Observation and
   origin of organochlorine compounds and polycyclic aromatic hydrocarbons in the free
   troposphere over central Europe. Environ. Pollut. 157, 3264–3271.
- 31 doi:10.1016/j.envpol.2009.05.028
- Lee, R.G.M., Burnett, V., Harner, T., Jones, K.C., 2000. Short-term temperature-dependent
   air-surface exchange and atmospheric concentrations of polychlorinated naphthalenes
   and organochlorine pesticides. Environ. Sci. Technol. 34, 393–398.
   doi:10.1021/es9907414
- Li, H., Ma, H., Lydy, M.J., You, J., 2014. Occurrence, seasonal variation and inhalation
  exposure of atmospheric organophosphate and pyrethroid pesticides in an urban
  community in South China. Chemosphere 95, 363–369.
- 39 doi:http://dx.doi.org/10.1016/j.chemosphere.2013.09.046

- Li, J., Zhang, G., Guo, L., Xu, W., Li, X., Lee, C.S.L., Ding, A., Wang, T., 2007.
   Organochlorine pesticides in the atmosphere of Guangzhou and Hong Kong: Regional sources and long-range atmospheric transport. Atmos. Environ. 41, 3889–3903. doi:10.1016/j.atmosenv.2006.12.052
- Li, Y.F., Macdonald, R.W., 2005. Sources and pathways of selected organochlorine pesticides
  to the Arctic and the effect of pathway divergence on HCH trends in biota: a review. Sci.
  Total Environ. 342, 87–106. doi:http://dx.doi.org/10.1016/j.scitotenv.2004.12.027
- Lohmann, R., Klanova, J., Kukucka, P., Yonis, S., Bollinger, K., 2012. PCBs and OCPs on a
  East-to-West Transect: The Importance of Major Currents and Net Volatilization for
  PCBs in the Atlantic Ocean. Environ. Sci. Technol. 46, 10471–10479.
- 11 doi:10.1021/es203459e
- Lohmann, R., Lammel, G., 2004. Adsorptive and absorptive contributions to the gas-particle
   partitioning of polycyclic aromatic hydrocarbons: State of knowledge and recommended
   parametrization for modeling. Environ. Sci. Technol. 38, 3793–3803.
- 15 doi:10.1021/es035337q
- Mackay, D., 2001. Multimedia Environmental Models The Fugacity Approach Second
   Edition, Second Edi. ed. Lewis.
- Mai, C., Theobald, N., Lammel, G., Hühnerfuss, H., 2013. Spatial, seasonal and vertical distributions of currently-used pesticides in the marine boundary layer of the North Sea. Atmos. Environ. 75, 92–102. doi:http://dx.doi.org/10.1016/j.atmosenv.2013.04.027
- Majewski, M.S., Coupe, R.H., Foreman, W.T., Capel, P.D., 2014. Pesticides in Mississippi air
   and rain: A comparison between 1995 and 2007. Environ. Toxicol. Chem. 33, 1283–
   1293. doi:10.1002/etc.2550
- Melymuk, L., Bohlin, P., Sáňka, O., Pozo, K., Klánová, J., 2014. Current challenges in air
   sampling of semi-volatile organic contaminants: sampling artifacts and their influence on
   data comparability. Environ. Sci. Technol. doi:10.1021/es502164r
- Melymuk, L., Bohlin-Nizzetto, P., Prokeš, R., Kukučka, P., Klánová, J., n.d. Sampling
  artifacts in active air sampling of semivolatile organic contaminants: Comparing
  theoretical and measured artifacts and evaluating implications for monitoring networks.
  Environ. Pollut. doi:doi:10.1016/j.envpol.2015.12.015
- Muñoz, A., Ródenas, M., Borrás, E., Vázquez, M., Vera, T., 2014. The gas-phase degradation
   of chlorpyrifos and chlorpyrifos-oxon towards OH radical under atmospheric conditions.
- 33 Chemosphere 111, 522–528. doi:10.1016/j.chemosphere.2014.04.087
- Odabasi, M., Cetin, B., 2012a. Determination of octanol-air partition coefficients of
  organochlorine pesticides (OCPs) as a function of temperature: Application to air-soil
  exchange. J. Environ. Manage. 113, 432–439. doi:10.1016/j.jenvman.2012.10.010
- Odabasi, M., Cetin, B., 2012b. Determination of octanol-air partition coefficients of
   organochlorine pesticides (OCPs) as a function of temperature: application to air-soil
   exchange. J. Environ. Manage. 113, 432–9.

- Okonski, K., Degrendele, C., Melymuk, L., Landlová, L., Kukučka, P., Vojta, Š., Jiri, K., 1 Čupr, P., Klánová, J., 2014. Particle size distribution of halogenated flame retardants and 2 3 implications for atmospheric deposition and transport. Env. Sci Technol 48, 14426-4 14434.
- 5 Pankow, J.F., 1987. Review and comparative analysis of the theories on partitioning between the gas and aerosol particulate phases in the atmosphere. Atmos. Environ. 21, 2275-6 2283. doi:http://dx.doi.org/10.1016/0004-6981(87)90363-5 7
- Peck, A.M., Hornbuckle, K.C., 2005. Gas-Phase Concentrations of Current-Use Pesticides in 8 Iowa. Environ. Sci. Technol. 39, 2952–2959. doi:10.1021/es0486418 9
- PPDB, 2013. The Pesticide Properties DataBase (PPDB) [WWW Document]. Dev. by Agric. 10 Environ. Res. Unit (AERU), Univ. Hertfordshire, 2006-2013. 11
- Putaud, J.-P., Raes, F., Van Dingenen, R., Brüggemann, E., Facchini, M.C., Decesari, S., 12 Fuzzi, S., Gehrig, R., Hüglin, C., Laj, P., Lorbeer, G., Maenhaut, W., Mihalopoulos, N., 13 Müller, K., Querol, X., Rodriguez, S., Schneider, J., Spindler, G., Brink, H. ten, Tørseth,
- 14
- 15 K., Wiedensohler, A., 2004. A European aerosol phenomenology-2: chemical
- characteristics of particulate matter at kerbside, urban, rural and background sites in 16 17 Europe. Atmos. Environ. 38, 2579–2595.
- doi:http://dx.doi.org/10.1016/j.atmosenv.2004.01.041 18
- Růzicková, P., Klánová, J., Cupr, P., Lammel, G., Holoubek, I., 2008. An assessment of air-19 soil exchange of polychlorinated biphenyls and organochlorine pesticides across central 20 21 and southern Europe. Environ. Sci. Technol. 42, 179-85.
- Sadiki, M., Poissant, L., 2008. Atmospheric concentrations and gas-particle partitions of 22
- 23 pesticides: Comparisons between measured and gas-particle partitioning models from
- source and receptor sites. Atmos. Environ. 42, 8288-8299. 24
- doi:http://dx.doi.org/10.1016/j.atmosenv.2008.07.041 25
- Sanusi, A., Millet, M., Mirabel, P., Wortham, H., 2000. Comparison of atmospheric pesticide 26 concentrations measured at three sampling sites: local, regional and long-range transport. 27 Sci. Total Environ. 263, 263–277. doi:http://dx.doi.org/10.1016/S0048-9697(00)00714-2 28
- Sanusi, A., Millet, M., Mirabel, P., Wortham, H., 1999. Gas-particle partitioning of pesticides 29 in atmospheric samples. Atmos. Environ. 33, 4941–4951. 30
- 31 doi:http://dx.doi.org/10.1016/S1352-2310(99)00275-7
- Sauret, N., Wortham, H., Putaud, J.-P., Mirabel, P., 2008. Study of the effects of 32
- environmental parameters on the gas/particle partitioning of current-use pesticides in 33 urban air. Atmos. Environ. 42, 544-553. 34
- doi:http://dx.doi.org/10.1016/j.atmosenv.2007.09.012 35
- Scheyer, A., Morville, S., Mirabel, P., Millet, M., 2008. Gas/particle partitioning of lindane 36 and current-used pesticides and their relationship with temperature in urban and rural air 37
- in Alsace region (east of France). Atmos. Environ. 42, 7695–7705. 38
- doi:http://dx.doi.org/10.1016/j.atmosenv.2008.05.029 39

- 1 Schummer, C., Mothiron, E., Appenzeller, B.M.R., Rizet, A.-L., Wennig, R., Millet, M., 2010. Temporal variations of concentrations of currently used pesticides in the atmosphere of 2 3 Strasbourg, France. Environ. Pollut. 158, 576-584.
- doi:http://dx.doi.org/10.1016/j.envpol.2009.08.019 4
- 5 Shen, L., Wania, F., Lei, Y.D., Teixeira, C., Muir, D.C.G., Bidleman, T.F., 2004. Atmospheric distribution and long-range transport behavior of organochlorine pesticides in north 6 America. Environ. Sci. Technol. 39, 409-420. doi:10.1021/es049489c 7
- Shoeib, M., Harner, T., 2002. Using measured octanol-air partition coefficients to explain 8 9 environmental partitioning of organochlorine pesticides. Environ. Toxicol. Chem. 21, 984–990. doi:10.1897/1551-5028(2002)021<0984:umoapc>2.0.co;2 10
- Sofuoglu, A., Cetin, E., Bozacioglu, S.S., Sener, G.D., Odabasi, M., 2004. Short-term 11 variation in ambient concentrations and gas/particle partitioning of organochlorine 12 13 pesticides in Izmir, Turkey. Atmos. Environ. 38, 4483-4493.
- doi:10.1016/j.atmosenv.2004.05.036 14
- Solomon, K.R., Williams, W.M., Mackay, D., Purdy, J., Giddings, J., Giesy, J., 2014. 15 Properties and Uses of Chlorpyrifos in the United States, in: Giesy, J.P., Solomon, K.R. 16 17 (Eds.), Ecological Risk Assessment for Chlorpyrifos in Terrestrial and Aquatic Systems 18 in the United States, Reviews of Environmental Contamination and Toxicology. Springer
- International Publishing, pp. 13-34. doi:10.1007/978-3-319-03865-0 2 19
- SRS, 2014. State Phytosanitary Administration. Usage of active substances in 2013, Czech 20 21 Republic. http://eagri.cz/public/web/file/439536/celek 2013 EN.pdf.
- SRS, 2013. State Phytosanitary Administration. Usage of active substances in 2012, Czech 22 23 Republic. http://eagri.cz/public/web/file/439598/celek 2012 EN.pdf.
- U.S. Environmental Protection Agency, 2013. Atrazine Updates [WWW Document]. Pestic. 24 Reregistration. 25
- University of Hertfordshire, 2013. The Pesticide Properties DataBase (PPDB) [WWW 26 Document]. Dev. by Agric. Environ. Res. Unit (AERU), Univ. Hertfordshire, 2006-2013. 27
- US EPA, 2014. Estimation Programs Interface Suite<sup>TM</sup> for Microsoft® Windows, v 4.11. 28 United States Environmental Protection Agency, Washington, DC, USA. 29
- Van den Berg, F., Kubiak, R., Benjey, W.G., Majewski, M.S., Yates, S.R., Reeves, G.L., 30 Smelt, J.H., van der Linden, A.M.A., 1999. Emission of Pesticides into the Air. Water. 31 Air. Soil Pollut. 115, 195–218. doi:10.1023/A:1005234329622 32
- Van Dijk, H.F.G., Guicherit, R., 1999. Atmospheric dispersion of current-use pesticides: A 33 review of the evidence from monitoring studies. Water. Air. Soil Pollut. 34 doi:10.1023/A:1005293020536 35

#### Vecchi, R., Marcazzan, G., Valli, G., 2007. A study on nighttime-daytime PM10 concentration 36 37 and elemental composition in relation to atmospheric dispersion in the urban area of Milan (Italy). Atmos. Environ. 41, 2136–2144. doi:10.1016/j.atmosenv.2006.10.069 38

- Wang, X.P., Sheng, J.J., Gong, P., Xue, Y.G., Yao, T.D., Jones, K.C., 2012. Persistent organic
  pollutants in the Tibetan surface soil: Spatial distribution, air-soil exchange and
  implications for global cycling. Environ. Pollut. 170, 145–151.
- 4 doi:10.1016/j.envpol.2012.06.012
- Wania, F., Haugen, J., 1998. Temperature dependence of atmospheric concentrations of
   semivolatile organic compounds. Environ. Sci. ... 1013–1021. doi:10.1021/es970856c
- Wei, Y.L., Bao, L.J., Wu, C.C., He, Z.C., Zeng, E.Y., 2014. Association of soil polycyclic aromatic hydrocarbon levels and anthropogenic impacts in a rapidly urbanizing region:
  Spatial distribution, soil-air exchange and ecological risk. Sci. Total Environ. 473-474, 676–684. doi:10.1016/j.scitotenv.2013.12.106
- Yadav, I.C., Devi, N.L., Syed, J.H., Cheng, Z., Li, J., Zhang, G., Jones, K.C., 2015. Current
  status of persistent organic pesticides residues in air, water, and soil, and their possible
  effect on neighboring countries: A comprehensive review of India. Sci. Total Environ.
  511, 123–137. doi:10.1016/j.scitotenv.2014.12.041
- Yao, Y., Harner, T., Blanchard, P., Tuduri, L., Waite, D., Poissant, L., Murphy, C., Belzer, W.,
  Aulagnier, F., Sverko, E., 2008. Pesticides in the Atmosphere Across Canadian
  Agricultural Regions. Environ. Sci. Technol. 42, 5931–5937. doi:10.1021/es800878r
- Zhang, L., Vet, R., 2006. A review of current knowledge concerning size-dependent aerosol
   removal. China Particuology 4, 272–282. doi:http://dx.doi.org/10.1016/S1672 2515(07)60276-0
- Zhang, N., Yang, Y., Liu, Y., Tao, S., 2009. Determination of octanol-air partition coefficients
   and supercooled liquid vapor pressures of organochlorine pesticides. J. Environ. Sci.
   Heal. Part B 44, 649–656.
- Zhang, X.M., Meyer, T., Muir, D.C.G., Teixeira, C., Wang, X.W., Wania, F., 2013.
- Atmospheric deposition of current use pesticides in the Arctic: Snow core records from
  the Devon Island Ice Cap, Nunavut, Canada. Environ. Sci. Impacts 15, 2304–2311.
  doi:10.1039/c3em00433c

# 29 TABLE

### 30 31

Table 1: Physicochemical properties and atmospheric concentrations (in pg.m<sup>-3</sup>) of individual OCPs and CUPs at background site. ND indicates "not detected"

Compound	Type of pesticide <sup>a</sup>	In use <sup>b</sup>	Half-life in soil (days) (PPDB, 2013)	Vapor pressure (Pa)	$\log K_{oa}$	Detection frequency (%)	Range of total concentrations (pg.m <sup>-3</sup> )	Range of gas phase concentrations (pg.m <sup>-3</sup> )	Range of particle phase concentrations (pg.m <sup>-3</sup> )	Average measured particulate fraction $(\theta_{meas})$
Acetochlor	Н	Y	14	2.20E-05 <sup>c</sup>	9.07 <sup>e</sup>	50.0	ND-181	ND-158	ND-23.2	$0.14\pm0.32$
Alachlor	Н	Ν	14	2.90E-03 <sup>c</sup>	9.98 <sup>e</sup>	5.77	ND-0.82	ND-0.23	ND-0.82	$0.85\pm0.26$
Atrazine	Η	Ν	75	3.90E-05 <sup>c</sup>	9.62 <sup>g</sup>	21.2	ND-1.24	ND-0.76	ND-0.49	$0.63\pm0.46$
Azinphos Methyl	Ι	Ν	10	5.00E-07 <sup>c</sup>	8.76 <sup>f</sup>	0.00	ND	ND	ND	ND
Carbendazim	F	Y	40	9.00E-05 <sup>c</sup>	10.6 <sup>f</sup>	42.3	ND-12.5	ND-0.22	ND-12.5	$0.98\pm0.10$
Chlorotoluron	Н	Y	45	5.00E-06 <sup>c</sup>	10.6 <sup>g</sup>	48.1	ND-25.1	ND-0.48	ND-24.7	$0.95\pm0.20$
Chlorpyrifos	Ι	Y	50	1.43E-03 <sup>c</sup>	8.41 <sup>h</sup>	84.6	ND-159	ND-158	ND-9.43	$0.037\pm0.064$
Diazinon	Ι	Ν	9.1	1.20E-02 <sup>c</sup>	9.14 <sup>e</sup>	1.92	ND-0.18	ND-0.18	ND	0.0
Dimethachlor	Н	Y	7.0	6.40E-04 <sup>c</sup>	9.34 <sup>d</sup>	40.4	ND-71.3	ND-70.8	ND-9.36	$0.18\pm0.37$
Dimethoate	Ι	Y	2.6	2.47E-04 <sup>c</sup>	9.15 <sup>f</sup>	3.85	ND-0.08	ND	ND-0.08	$1.0 \pm 0.00$
Disulfoton	Ι	Ν	30	7.20E-03 <sup>c</sup>	8.07 <sup>d</sup>	1.92	ND-2.22	ND	ND-2.22	1
Diuron	Н	Ν	76	1.15E-06 <sup>c</sup>	10.4 <sup>f</sup>	32.7	ND-1.23	ND	ND-1.23	$1.0 \pm 0.00$
Fenitrothion	Ι	Ν	2.7	6.76E-04 <sup>c</sup>	7.72 <sup>d</sup>	0.00	ND	ND	ND	ND
Fenpropimorph	F	Y	35	3.90E-03 <sup>c</sup>	8.93 <sup>e</sup>	65.4	ND-73.8	ND-1.27	ND-73.8	$0.91\pm0.28$
Fonofos	Ι	Ν	99	2.70E-02 <sup>c</sup>	7.48 <sup>d</sup>	5.76	ND-8.03	ND	ND-8.03	$1.0 \pm 0.00$
Isoproturon	Н	Y	12	5.50E-06 <sup>c</sup>	11.2 <sup>g</sup>	86.5	ND-413	ND-122	ND-291	$0.84\pm0.29$
Malathion	Ι	Ν	0.17	3.10E-03 <sup>c</sup>	9.06 <sup>e</sup>	3.85	ND-0.30	ND-0.30	ND-0.13	$0.50 \ \pm 0.70$
Metamitron	Н	Y	30	7.44E-07 <sup>c</sup>	11.2 <sup>d</sup>	25.0	ND-16.5	ND-16.5	ND-6.41	$0.23\pm0.44$
Metazachlor	Н	Y	8.6	9.30E-05 <sup>c</sup>	9.76 <sup>e</sup>	86.5	ND-344	ND-262	ND-275	$0.59\pm0.38$
Metribuzin	Н	Y	11.5	1.21E-04 <sup>c</sup>	10.0 <sup>d</sup>	15.4	ND-5.46	ND-5.46	ND-1.83	$0.22 \pm 0.41$

Prochloraz	F	Y	120	1.50E-04 <sup>c</sup>	13.6 <sup>d</sup>	55.8	ND-1.95	ND	ND-1.95	$1.0 \pm 0.00$
Pyrazon	Н	Y	31	1.00E-09 <sup>c</sup>	9.01 <sup>d</sup>	15.4	ND-2.25	ND-0.80	ND-2.25	$0.91 \pm 0.26$
Simazine	Н	Ν	60	8.10E-07 <sup>c</sup>	9.59 <sup>g</sup>	1.92	ND-0.087	ND	ND-0.087	1
S-metolachlor	Н	Y	15	3.70E-03 <sup>c</sup>	9.33 <sup>d</sup>	73.1	ND-329	ND-309	ND-91.0	$0.24 \pm 0.34$
Temephos	Ι	Ν	2	9.50E-06 <sup>c</sup>	13.1 <sup>d</sup>	5.77	ND-0.21	ND-0.21	ND-0.11	$0.67\pm0.58$
Terbufos	Ι	Ν	8	3.46E-02 <sup>c</sup>	7.49 <sup>d</sup>	1.92	ND-0.80	ND	ND-0.61	1
Terbuthylazine	Н	Y	75.1	1.20E-04 <sup>c</sup>	$9.03^{\mathrm{f}}$	78.8	ND-53.8	ND-33.8	ND-31.6	$0.45 \pm 0.35$
∑CUPs							ND-662	ND-365	ND-323	
-										
α-HCH		Ν	175	3.44E-02 <sup>d</sup>	7.61 <sup>i</sup>	100	1.09-9.79	1.08-9.78	ND-0.031	< 0.01
β-НСН		Ν		3.44E-02 <sup>d</sup>	8.88 <sup>i</sup>	69.2	ND-0.59	ND-0.59	ND-0.074	$0.033 \pm 0.051$
γ-HCH	Ι	Ν		3.44E-02 <sup>d</sup>	7.85 <sup>i</sup>	100	0.488-21.8	0.470-21.8	ND-0.043	< 0.01
δ-НСН		Ν		3.44E-02 <sup>d</sup>	8.84 <sup>i</sup>	57.7	ND-0.42	ND-0.42	ND-0.065	$0.055\pm0.097$
<i>o,p</i> '-DDE		Ν		5.99E-03 <sup>d</sup>	9.26 <sup>j</sup>	96.2	ND-1.42	ND-1.42	ND-0.054	$0.018\pm0.071$
<i>p,p</i> '-DDE		Ν		3.44E-03 <sup>d</sup>	9.68 <sup>i</sup>	100	1.14-71.4	0.612-71.4	ND-0.96	$0.037\pm0.074$
<i>o,p</i> '-DDD		Ν		8.45E-04 <sup>d</sup>	9.57 <sup>j</sup>	73.1	ND-1.30	ND-1.28	ND-0.11	$0.065 \pm 0.16$
<i>p,p</i> '-DDD		Ν		1.23E-03 <sup>d</sup>	10.1 <sup>i</sup>	75.0	ND-2.61	ND-2.56	ND-0.40	$0.11 \pm 0.18$
<i>o,p</i> '-DDT	Ι	Ν	6200	1.68E-03 <sup>d</sup>	9.45 <sup>i</sup>	92.3	ND-9.18	ND-9.18	ND-0.11	$0.033\pm0.08$
<i>p,p</i> '-DDT	Ι	Ν	6200	1.43E-04 <sup>d</sup>	9.82 <sup>i</sup>	100	0.414-9.99	0.13-9.99	ND-0.50	$0.13 \pm 0.19$
∑OCPs		Ν					4.51-122	2.87-122	ND-1.96	

<sup>a</sup> H: Herbicide, I: Insecticide and F: Fungicide. <sup>b</sup> Y: Authorized for agricultural use in Czech Republic during the sampling period and N: Not authorized for agricultural purposes in Czech Republic during the sampling period. <sup>c</sup> (University of Hertfordshire, 2013). <sup>d</sup> (US EPA, 2014). <sup>e</sup> (Coscollà et al., 2013b). <sup>f</sup> (Coscollà et al., 2013a). <sup>g</sup> (Götz et al., 2007). <sup>h</sup> (Odabasi and Cetin, 2012b). <sup>i</sup> (Shoeib and Harner, 2002). <sup>j</sup> (Zhang et al., 2009).

**35 FIGURES** 

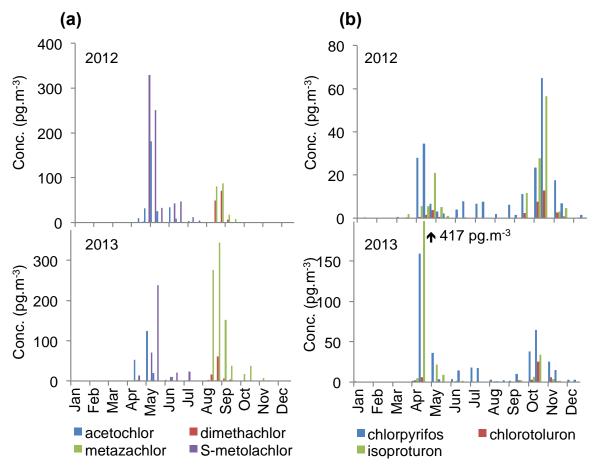
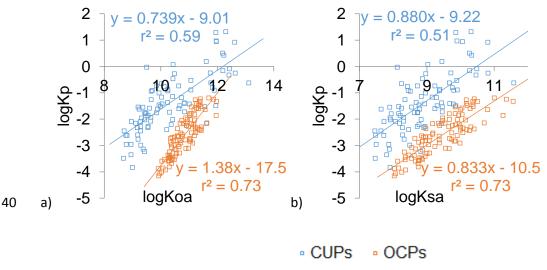
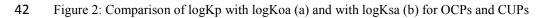
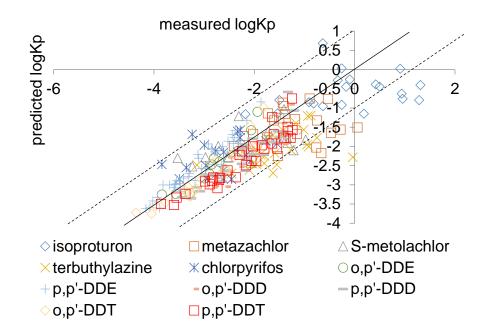


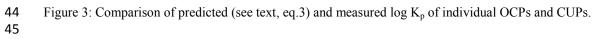
Figure 1: Seasonal variation of selected CUPs with (a) one peak per year during the growing season and (b) two
peaks per year, in April-July and October-November

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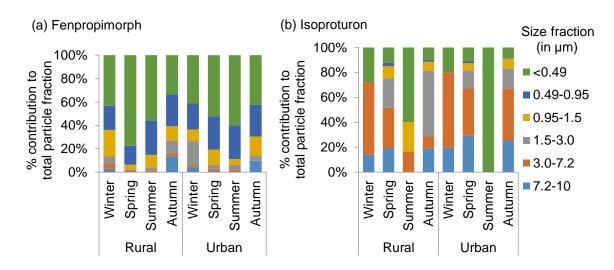


Figure 4: Seasonal particle size distribution of (a) fenpropimorph and (b) isoproturon at the rural and urban sites.
 Fenpropimorph represents the group of pesticides predominantly found on fine particles and isoproturon for the coarse particles

# 50 CAPTIONS

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- 52 Table 2: Physicochemical properties and atmospheric concentrations (in pg.m<sup>-3</sup>) of individual
- 53 OCPs and CUPs at background site. ND indicates "not detected"
- 54 Figure 1: Seasonal variation of selected CUPs with (a) one peak per year during the growing
- season and (b) two peaks per year, in April-July and October-November

- 56 Figure 2: Comparison of logKp with logKoa (a) and with logKsa (b) for OCPs and CUPs
- Figure 3: Comparison of predicted (see text, eq.3) and measured log K<sub>p</sub> of individual OCPs
  and CUPs.
- 59 Figure 4: Seasonal particle size distribution of (a) fenpropimorph and (b) isoproturon at the
- 60 rural and urban sites. Fenpropimorph represents the group of pesticides predominantly found
- 61 on fine particles and isoproturon for the coarse particles

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