Technical Note: Development of chemoinformatic tools to enumerate functional groups in molecules for organic aerosol characterization

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Abstract. Functional groups (FGs) can be used as a reduced representation of organic aerosol composition in both ambient and environmental controlled chamber studies, as they retain a certain chemical specificity. Furthermore, FG composition has been informative for source apportionment, and various models based on a group contribution framework have been developed to calculate physicochemical properties of organic compounds. In this work, we provide a set of validated chemoinformatic patterns that correspond to: 1) a complete set of functional groups that can entirely describe the molecules comprised in the α -pinene and 1,3,5-trimethylbenzene MCMv3.2 oxidation schemes, 2) FGs that are measurable by Fourier transform infrared spectroscopy (FTIR), 3) groups incorporated in the SIMPOL.1 vapor pressure estimation model, and 4) bonds necessary for the calculation of carbon oxidation state. We also provide example applications for this set of patterns. We compare available aerosol composition reported by chemical speciation measurements and FTIR for different emission sources, and calculate the FG contribution to the O:C ratio of simulated gas phase composition generated from α -pinene photooxidation (using MCMv3.2 oxidation scheme).

1 Introduction

Atmospheric aerosols are complex mixtures of inorganic salts, mineral dust, sea salt, black carbon, metals, organic compounds, and water (Seinfeld and Pandis, 2006). Of these components, the organic fraction can comprise as much as 80% of the aerosol mass (Lim and Turpin, 2002; Zhang et al., 2007), and yet eludes definitive characterization due to the number and diversity of molecule types. There have been many proposals for reducing representations in which a mixture of 10,000+different types of molecules (Hamilton et al., 2004) are represented by some combination of their molecular size, carbon number, polarity, or elemental ratios (Pankow and Barsanti, 2009; Kroll et al., 2011; Daumit et al., 2013; Donahue et al., 2012); many of which are associated with observable quantities [e.g., by aerosol mass spectrometry (AMS; Jayne et al., 2000), gas chromatography mass spectrometry (GC-MS and GCxGC-MS; Rogge et al., 1993; Hamilton et al., 2004)]. Molecular bonds or organic functional groups (FGs), which is the focus of this manuscript, can also be used

to provide reduced representations for mixtures, and has been shown useful for organic mass (OM) quantification, source apportionment, and prediction of hygroscopicity and volatility (e.g., Russell, 2003; Donahue, 2011; Russell et al., 2011; Suda et al., 2014). Examples of property estimation methods include models for pure component vapor pressure (Pankow and Asher, 2008; Compernolle et al., 2011), UNIFAC and its variations for activity coefficients and viscosity (Ming and Russell, 2001; Griffin et al., 2002; Zuend et al., 2008, 2011). The FGs that can be detected or quantified by measurement vary widely by analytical technique, which include Fourier transform infrared spectroscopy (FTIR, Maria et al., 2002), Raman spectroscopy (Craig et al., 2015), spectrophotometry (Aimanant and Ziemann, 2013), nuclear magnetic resonance (NMR, Decesari et al., 2000; Cleveland et al., 2012), and gas chromatography with mass spectrometry and derivatization (Dron et al., 2010).

Projecting specific molecular information available through various forms of mass spectrometry (e.g., Williams et al., 2006; Kalberer et al., 2006; Laskin et al., 2012; Chan et al., 2013; Nguyen et al., 2013; Vogel et al., 2013; Yatavelli et al., 2014; Schilling Fahnestock et al., 2015; Chhabra et al., 2015) or model simulations employing explicit chemical mechanisms (e.g., Jenkin, 2004; Aumont et al., 2005; Herrmann et al., 2005) to a reduced dimensional space represented by some combination of FGs can be useful for measurement intercomparisons, or model-measurement comparisons. For this task, the aerosol community can benefit from developments in the chemoinformatics community. If the structure of a substance is described through its molecular (also referred to as chemical) graph (Balaban, 1985) — which is a set of atoms and their association through bonds — the abundance of arbitrary substructures (also called fragments) can be estimated through pattern matching algorithms called subgraph isomorphisms (Barnard, 1993; Ehrlich and Rarey, 2012; Kerber et al., 2014). Structural information of molecules can be encoded in various representations, including a linear string of ASCII characters denoted as SMILES (Weininger, 1988). A corresponding set of fragments can be specified by SMARTS, which is a superset of the SMILES specification (DAYLIGHT Chemical Information Systems, Inc.). There are many chemoinformatic packages that implement algorithms for pattern matching — for instance, OpenBabel (O'Boyle et al., 2011), Chemistry Development Kit (Steinbeck et al., 2003), OEChem (Openeye Scientific Software, Inc.), RDKit (Landrum), Indigo (GGA Software Services). The concept of using SMILES and SMARTS patterns have been reported for applications in the atmospheric chemistry community (Barley et al., 2011; COBRA, Fooshee et al., 2012). While some sets of SMARTS patterns for substructure matching can additionally be found in literature (Hann et al., 1999; Walters and Murcko, 2002; Olah et al., 2004; Enoch et al., 2008; Barley et al., 2011; Kenny2013) or on web databases — e.g., DAYLIGHT Chemical Information Systems, Inc. (DAYLIGHT Chemical Information Systems, Inc.) — knowledge regarding the extent of specificity and validation of the defined patterns is not available.

In this work, we report specifications for four specific sets of substructures: 1) FGs contained in α -pinene and 1,3,5-trimethylbenzene photooxidation products defined in MCMv3.2 (Jenkin

et al., 1997; Saunders et al., 2003; Jenkin et al., 2003; Bloss et al., 2005), obtained via http: //mcm.leeds.ac.uk/MCM; 2) FGs that are measured or measurable (i.e., have absorption bands) for FTIR analysis (Pavia et al., 2008); 3) molecular fragments used by SIMPOL.1 for estimation of pure organic compound vapor pressures; and 4) bonds used for calculation of carbon oxidation state (\overline{OS}_C) (Kroll et al., 2011, 2015). As there are several ways to define SMARTS patterns for substructure matching, we prescribe a general method for formulating patterns in such a way that permits a user to match and test not only the total number of FGs within a molecule, but to confirm that all atoms within molecule are classified uniquely into a set of FGs (except polyfunctional carbon, which can be associated with many FGs). We present a validation test for the groups defined, and show example applications for mapping molecules onto 2D-VBS space, inter-measurement comparison between OM composition reported by GC-MS and FTIR for several source classes, and discuss implications for further applications. The patterns and software written for this manuscript are provided in a version controlled repository (Appendix C).

2 Methods

In this section, we present a series of patterns corresponding to substructures useful for vapor pressure estimation of FGs in molecules defined by measurements and chemical mechanisms (Section 2.1) as well as the methods and compound sets used for their validation (Section 2.2). We further describe the data set used for constructing a few example applications (Section 2.3).

2.1 Pattern specification for matching substructures

Four groups of patterns are defined: the first group (Table 1, substructures 1-33) corresponding to the complete set of FGs that can be found in the MCMv3.2 α -pinene and 1,3,5-trimethylbenzene oxidation scheme (Jenkin et al., 1997; Saunders et al., 2003), the second group used to study the FG abundance associated with FTIR measurements (FGs not specified before, containing carbon, oxygen and nitrogen atoms; Table 1, substructures 33-57), the third group corresponding to the FGs used to build the SIMPOL.1 model (Pankow and Asher, 2008) to predict pure components vapor pressures that are not present in the first set of patterns (Table 2) and the fourth group used to calculate the oxidation state of carbon atoms (Table 3). The regions of absorption in the IR spectrum associated with FGs patterns are reported in Table 4 as an additional reference. The OpenBabel toolkit (O'Boyle et al., 2011) is called through the pybel library (O'Boyle et al., 2008) in Python to search and enumerate abundances of fragments (most of which are specified by SMARTS) in each molecule (specified by SMILES). A few groups for which SMARTS patterns were difficult to obtain were calculated through algebraic relations specified through the string formatting syntax of the python programming language. In this syntax, values pre-computed through SMARTS matching are combined together to estimate properties for another group. While SMARTS can also describe ring

definitions, ring perception is a difficult task partly due to the varying definitions of a ring, which must consider definition of aromaticity (tautomerism must also be considered) (Berger et al., 2004; May and Steinbeck, 2014). In this work, we use the smallest set of smallest rings (SSSR) (Downs et al., 1989) as defined by OpenBabel and many chemoinformatic software packages to enumerate the number of aromatic rings in this work. Ring enumeration is the only task specific to the software implementation, but otherwise the patterns specified can be ported to other software packages. The full implementation of patterns and scripts described in this manuscript are made available through an online repository (Section C).

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We adapt chemoinformatic tools for use with SIMPOL.1 partly because the portable SMARTS pattern approach is more readily compatible with this model parameterization. We note that EVAP-ORATION vapor pressure model is fitted to more recent diacid measurements and includes positional information and non-linear interactions among FGs (Compernolle et al., 2011). Positional arguments can be included by querying specific structural information from the internal representations of molecular graphs according to implementations in various software packages, or formulating SMARTS patterns which require specificity in the arrangement of neighboring atoms (Barley et al., 2011; Topping et al., 2016). In this work, positional information of FGs are used only for conjugated aldehyde, ketone, and ester with an alkene or benzene ring (Table 1, substructures 40-48). With regards to the use of SIMPOL.1, vapor pressure predictions can also be improved by updating coefficients for the model with new estimates (Yeh and Ziemann, 2015).

SMARTS patterns for tallying the number of FGs can be formulated in many ways. Therefore, we provide an example for the aldehyde FG group to illustrate the development process, with particular attention paid to the description of atoms returned in the matched set and how their bonding environments are defined. We first describe a formulation specific for fulfilling the atom-level validation which requires two patterns to account for all aldehyde groups in the system, and an alternate formulation for only enumerating FGs that requires only a single pattern.

When applied to propionaldehyde, the set of atoms returned by matching the pattern for substructure 9 in Table 1 will be 3, 4, 10 (as labeled in Figure 1a). The first bracket [CX3; \$(C([#1]) (=[0]) [#6])] describes the carbon atom to be matched and returned. CX3 describes a carbon with 3 bonds (effectively sp^2); \$(C([#1]) (=[0]) [#6]) qualifies that it is bonded to hydrogen, oxygen, and another carbon. The expression (=[0; !\$([0][0])]) describes the double-bonded oxygen to this carbon atom; !\$([0][0])] excludes preventing matching of $C=O^+-O^-$ (defined as a separate group, substructure 21 in Table 1) that are present in other molecules (an example is provided in Figure 1b). The last bracket [H] is included to explicitly include the hydrogen atom in the returned set. While the sp^3 carbon attached to the sp^2 is not returned in the set of matched atoms, this additional specificity is necessary to prevent double counting of the same aldehydic group in the formaldehyde molecule, which contains two hydrogen atoms bonded to sp^2 carbon. A separate SMARTS pattern is defined for formaldehyde (Table 1 substructure 15).

(For similar reasons, a SMARTS pattern specific for formic acid has been specified alongside the carboxylic FG.)

In this approach, all atoms in the aldehyde group are matched instead of just the identifying carbon, oxygen, or hydrogen. The advantage of this strict protocol is that we can devise a validation such that each atom in a molecule or chemical system is accounted for by one and only one group—except for polyfunctional carbon—for any proposed set of FGs (Appendix A). Fulfillment of this validation criterion provides a means for interpreting atomic ratios commonly used by the community (e.g., O:C, H:C, and N:C) through contributions of distinctly defined FGs.

Revisiting the aldehyde FG example, an alternative pattern specified only for the purposes of counting FGs for use in SIMPOL.1 is shown in Table 2. We only describe the bonding environment of the sp² carbon and count the number of its occurrence, so a single pattern can be used for both formaldehyde and other aldehyde compounds.

A separate set of SMARTS patterns are defined for estimation of $\overline{\mathrm{OS}}_{\mathrm{C}}$. Instead of FGs, these patterns enumerate the type of bond and atom attached to a carbon atom, and its oxidation state is calculated as the sum of the coefficients corresponding to its bonds.

2.2 Data sets for validation

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The first and the third groups of SMARTS patterns were validated against a set of 99 compounds (Table B1, Appendix B) selected from those used in the development of the SIMPOL.1 method, or occurring in atmospheric aerosol (Section 2.3) (Fraser et al., 2003; Grosjean et al., 1996; Fraser et al., 1998), or from the ChemSpider database (Pence and Williams, 2010) (to test for specific functionalities, eg. secondary amide) or from the MCMv3.2 α-pinene oxidation scheme. The patterns corresponding to the first group were further tested against the complete set of compounds present in the α-pinene and 1,3,5-trimethylbenzene MCMv3.2 oxidation schemes (408 compounds) in order to achieve a complete counting of all the atoms (carbon, oxygen, nitrogen and hydrogen atoms) and to avoid accounting heteroatoms to multiple FGs. The second group (Table 1, substructures 33-57) of SMARTS patterns was tested on a set of 26 compounds (Table B2, Appendix B) selected from the ChemSpider database and the fourth group (Table 3) was tested on a subset of 3 compounds extracted from the set of compounds used for the validation of the first group.

2.3 Data sets for example applications: molecules identified by GC-MS measurements and α -pinene and 1,3,5-TMB photooxidation products specified by the MCMv3.2 mechanism

A classic data set of organic compounds in primary organic aerosol (OA) from automobile exhaust (Rogge et al., 1993) and wood combustion (Rogge et al., 1998) quantified with GC-MS have been analyzed in order to retrieve the FG abundance of the mixture. Each compound, reported by common name in the literature, was converted to its corresponding SMILES string by querying the

ChemSpider database with the Python ChemSpipy package (Swain), which wraps the ChemSpider application programming interface. FG composition, \overline{OS}_{C} and pure component vapor pressure for each compound in the different reported mixture types was estimated using the substructure search algorithm described above. The algorithm previously described was applied to calculate the pure component vapor pressure for each compound i with the SIMPOL.1 model (Pankow and Asher, 2008). The total concentration in both gas phase and particle phase of the compounds reported by Rogge et al. (1993), Rogge et al. (1998), and Hildemann et al. (1991) was used to estimate the OA concentration considering a seed concentration (C_{OA}) in the predilution channel of 10 mg/m³, assuming fresh cooled emissions (Donahue et al., 2006). After diluting the total OA of a factor of 1000 the compounds were partitioned between the two phases based on the partitioning coefficient ξ_i (xi) calculated from the pure component saturation concentration (C_i^0) as described by Donahue et al. (2006).

FG abundance of the set of compounds incorporated in the MCMv3.2 α -pinene and 1,3,5-trimethylbenzene oxidation schemes was analyzed to demonstrate our validation scheme. Furthermore, the gas phase composition generated by α -pinene photooxidation in the presence of NO $_x$ (α -pinene/NO $_x$ ratio of 1.25), with propene as a radical initiator, was simulated using the Kinetic Pre-Processor (KPP, Damian et al., 2002; Sandu and Sander, 2006; Henderson, 2016) incorporating mechanistic information taken from MCMv3.2. Completeness and uniqueness requirements were tested and matched also for the α -pinene and propene MCMv3.2 degradation scheme. Initial concentrations of 240 ppb of α -pinene and 300 ppb of propene, a relative humidity of 61% and a continuous irradiation were chosen as simulation conditions.

190 3 Results

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3.1 Validation

Figures 2 shows that the enumerated FGs used by the SIMPOL.1 method (Table 2) are identical to the values enumerated manually. Matched FTIR FGs in Table 1 (substructures 33-57) are also identical to the true number of FGs in the set of compounds used for evaluation (Table B2), but are not shown as each group except alkane CH is matched at most once and a similar plot is uninformative. Figure 3 shows the completeness condition met, and Figure 4 shows the specificity criterion fulfilled of the first set of chemoinformatic patterns (Table 1, substructures 1-33). The carbon atoms can be accounted by multiple FGs if polyfunctional: methylene and methyl groups are matched 2 and 3 times respectively by alkane CH group (substructure 1 in Table 1), while the carbon atoms in small molecules included in the test set have only 1 carbon atom that is matched 4 times (e.g. methanol, which has 3 alkane CH and 1 alcohol substructures).

3.2 Example applications

Mapping composition in 2-D volatility basis set space. The algorithm described has been used to project molecular composition of GC-MS and MCM compounds to 2D-VBS space delineated by carbon oxidation and pure component saturation concentration (C⁰) (Figure 5). The properties of vehicle-related primary OA and wood combustion compounds measured by GC-MS are generally consistent with those reported for hydrocarbon-like OA and biomass burning OA, respectively, derived from PMF analysis of AMS spectra (Donahue et al., 2012). The low oxidation state is observed on account of more than 60% of carbon atoms being associated with methylene groups (-CH₂-, oxidation state of -2) in long-chain hydrocarbon compounds, and an association to lesser degree with CH groups in aromatic rings (oxidation state of -1) and methyl groups (-CH₃, oxidation state of -3). Most compounds in the MCMv3.2 system correspond to intermediate volatility organic compounds (IVOC), with only a small fraction with the semivolatile organic compound (SVOC) regime. When using of MCMv3.2 for simulation of secondary OA formation, additional mechanisms (e.g.,

pounds (IVOC), with only a small fraction with the semivolatile organic compound (SVOC) regime. When using of MCMv3.2 for simulation of secondary OA formation, additional mechanisms (e.g., in the condensed phase) are necessary to introduce low volatility organic compounds (LVOC) as observed in atmospheric and environmental controlled chamber observations (Ehn et al., 2014; Shiraiwa et al., 2014). Higher oxidation states than for compounds in the GC-MS set are observed on account of the larger number of functional groups containing electronegative atoms (oxygen and nitrogen) bonded to carbon.

Source apportionment In Figure 6, the FG distributions of aerosol collected during wood-burning and vehicle emission studies (Rogge et al., 1993; Rogge et al., 1998) have been compared to estimates from FTIR measurements of ambient samples separated by factor analytic decomposition (Positive Matrix Factorization or PMF; Paatero and Tapper, 1994) during September 2008 study period in California (Hawkins and Russell, 2010). The studies by Rogge et al. (1993, 1998) have been chosen as they have been used as a reference in the study of composition of organic aerosol from combustion sources (Heringa et al., 2012). The FTIR factor components from this study are consistent with similarly labeled factors from other field campaigns (Russell et al., 2011). The GC-MS reports approximately 20% of the OA mass (Fine et al., 2002), while the FTIR quantifies around 90% (Maria et al., 2003); these fractions form the bases for comparisons. For the study using FTIR, the biomass burning fraction was approximately 50% of the total OA during intensive fire periods, and the fossil fuel combustion comprised 95% of the overall OA during the campaign (Hawkins and Russell, 2010).

From this comparison, we find that the oxidized fraction is much higher in the biomass burning aerosol composition estimated by FTIR. The high abundance of alkane CH bonds in the compounds reported by GC-MS can be explained by the preference of this analytical method to characterize the least oxidized fraction of the collected aerosol. While high abundance of carbonyl groups are reported in FTIR measurements of biomass burning aerosol (Liu et al., 2009; Russell et al., 2009; Hawkins and Russell, 2010), more recent methods including advanced derivatization (Dron et al.,

2010) are necessary for quantification of carbonyl containing compounds by GC-MS. In addition, neither amine compounds nor levogucosan were reported in this GC-MS study. Levoglucosan is a polysaccharide compound often used as a tracer for burning and decomposition of cellulose reported in modern GC-MS measurements (Simoneit, 1999). However, FTIR does not report high fraction of alcohol COH as levoglucosan near particular fuel sources may be found mostly in supermicron diameter particles (Radzi bin Abas et al., 2004) (submicron OA was analyzed by Hawkins and Russell, 2010), its degradation in the atmosphere is rapid (Hennigan et al., 2010; Cubison et al., 2011; Lai et al., 2014), and the overall mass contribution to biomass burning OA is small (less than 2% by mass, Leithead et al., 2006).

Both estimation methods agree that more than 90% of OM mass is composed of alkane-CH for vehicle sources The fraction characterized by GC-MS and FTIR with PMF have associated uncertainties from derivatization and thermal separation in the chromatography column or in statistical separation, respectively, and lead to different fractions of mass reported. However, the approximate consistency in FG abundances estimated by the two methods, suggest that the fraction not analyzed by the GC-MS may not vary significantly from the measured fraction by FTIR in these aerosol types.

Oxygenated FG contribution to O:C ratio Using the first set of SMARTS patterns we are able to match all the oxygen atoms, accounting them to specific FGs, in the α -pinene and 1,3,5-trimethylbenzene MCMv3.2 oxidation mechanisms. We can therefore calculate the contribution of each FG to the total O:C ratio of the gas phase mixture. In Figure 7, contributions of FGs to the O:C ratio of the gas phase mixture generated by α -pinene photooxidation in low NO_x conditions (Section 2.3) is reported as a function of irradiation time. A singular peroxyacyl nitrate compound (peroxyacetyl nitrate) accounts for 26% of the total gas phase mass. The peroxyacyl nitrate functional group furthermore accounts for the greatest fraction of the total O:C ratio after 20 hours of simulation (53% of the total O:C), as it contains five oxygen atoms per FG. A full analysis on oxidation products with gas/particle partitioning is discussed by Ruggeri et al. (2016). This type of analysis can provide intermediate information that is useful to suggest constraints on the form of oxygenation (and resulting change in organic mixture vapor pressure) assumed by simplified models such as the Statistical Oxidation Model (Cappa and Wilson, 2012).

4 Conclusions

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We introduced the application of chemoinformatic tools that allow us to perform substructure matching in molecules to enumerate FGs present in compounds relevant for organic aerosol chemistry. We developed 50+ substructure patterns and validated them over a list of 125 compounds that were selected in order to account for all the functional groups (FGs) represented. We demonstrate how these tools can facilitate intercomparisons between GC-MS and FTIR measurements, and mapping of compounds onto the VBS space described by pure component vapor pressure and oxidation state.

We further introduce a novel approach for defining a set of patterns which accounts for each atom in a chemical system once and only once (except for polyfunctional carbon atoms associated with multiple FGs). This condition is confirmed by an atomic-level validation scheme applied to chemically explicit α -pinene and 1,3,5-TMB degradation mechanisms. This validation scheme provides an intermediate resolution between molecular speciation and atomic composition, and permits apportionment of conventionally aggregated quantities such as O:C, H:C, and N:C to contributions from individual FGs. We illustrate its application to the photochemical degradation of α -pinene from speciated simulations using MCMv3.2.

These applications can be further adapted for other methods developed to match substructures for other measurements, or enumerate groups used in group contribution methods for estimation of vapor pressures, activity coefficients, and Henry's law constants (Raventos-Duran et al., 2010; Compernolle et al., 2011; Zuend et al., 2011). The proposed validation approach can also be followed to define FG patterns containing sulfur and halide bonds that absorb in the infrared region presently not included in this work.

Appendix A: Group validation

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Let us consider a set of atoms A in molecule k and a set of FGs G. $\{a :\in A_k, a \in g\}$ denotes the set of atoms in molecule k which also is a member of group g, where $g \in G$. Completeness of G is defined by the condition that the combination of atoms matched by all groups in G comprises the full set of atoms A_k for every molecule:

$$\bigcup_{g \in G} \{a : a \in A_k, a \in g\} = A_k \quad \forall k$$

Specificity or minimal redundancy in G is defined by the condition that the intersection of atoms from all groups, excluding the set of polyfunctional carbon atoms $C_k^p \subset A_k$, comprises the empty set:

$$\bigcap_{g \in G} \{a : a \in A_k, a \in g\} \setminus C_k^{\mathsf{p}} = \varnothing \quad \forall k$$

Appendix B: Compounds used for testing the chemoinformatic patterns

Table B1: List of the compounds used to test the chemoinformatic patterns used in the SIMPOL.1 (Pankow and Asher, 2008) group contribution method to calculate pure component vapor pressure (Table 2).

Compound or MCMv3.2 internal name Smiles

2,2-dimethyl pentaneCCCC(C)(C)C1,1-dimethyl cyclohexaneCC1(CCCCC1)CcyclobutanolC1CC(C1)O1,2-pentanediolCCCC(CO)ObutanalCCCC=O

2-octanone CCCCCC(=O)C
heptanal CCCCCCC=O
ethanoic acid CC(=O)O
butanoic acid CCCC(=O)O
4-oxo-pentanoic acid CC(=O)CCC(=O)O
2,4-hexadienal C/C=C/C=C/C=O
3-butenoic-acid C=CCC(=O)O

2-phenyl-propane CC(C)C1=CC=CC=C1
2-phenyl-ethanol C1=CC=C(C=C1)CCO
2-hydroxy-1-methyl-benzene CC1=CC=CC1O

3-methyl-benzoic acid CC1=CC(=CC=C1)C(=O)O

4-amino-3-methylbenzoic acid CC1=C(C=CC(=C1)C(=O)O)N

1-butoxy-2-ethoxyethane O(CCCC)CCOCC

cis-2,4-dimethyl-1,3-dioxane C[C@H]1OCC[C@@H](C)O13-methylbutyl nitrate CC(C)CCO[N+](=O)[O-]

 $\begin{array}{lll} \text{2-methyl-propyl ethanoate} & & & & & & & & \\ \text{1-methyl-propyl butanoate} & & & & & & \\ \text{1-methyl-propyl butanoate} & & & & & \\ \text{2-nitro-1-propanol} & & & & & \\ \text{2-nitro-1-propanol} & & & & & \\ \text{CC(C)[N+](=O)[O-]} \\ \text{ethyl nitroacetate} & & & & \\ \text{CCOC(=O)C[N+](=O)[O-]} \\ \text{di-n-butyl peroxide} & & & \\ \text{CC(C)(C)OOC(C)(C)C} \\ \text{peroxyacetylnitrate} & & & \\ \text{CC(=O)OO[N+](=O)[O-]} \\ \end{array}$

ethyl-hydroperoxide CCOO butyl-hydroperoxide CCCCOO

butanedioic acid C(CC(=O)O)C(=O)O

methylbutanedioic acid CC(CC(=O)O)C(=O)O
benzoic acid C1=CC=C(C=C1)C(=O)O

1,3,5-benzenetricarboxylic acid C1=C(C=C1C(=O)O)C(=O)O)C(=O)O

1,2,4,5-benzenetetracarboxylic acid C1=C(C(=C1C(=O)O)C(=O)O)C(=O)O)C(=O)O2,6-naphthalenedicarboxylic acid C1=CC2=C(C=C2)C(=O)O)C=C1C(=O)O

dehydroabietic acid CC(C)C1=CC2=C(C=C1)[C@]3(CCC[C@@]([C@@H]3CC2)(C)C(=O)O)C

dinitrophenol C1=CC(=C(C(=C1)O)[N+](=O)[O-])[N+](=O)[O-]

perylene C1=CC2=C3C(=C1)C4=CC=CC5=C4C(=CC=C5)C3=CC=C2

benzo[ghi]perylene C1=CC2=C3C(=C1)C4=CC=CC5=C4C6=C(C=C5)C=CC(=C36)C=C2

benzo[ghi]fluoranthene C1=CC2=C3C(=C1)C4=CC=CC5=C4C3=C(C=C2)C=C5

anthracene-9,10-dione C1=CC=C2C(=C1)C(=O)C3=CC=CC=C3C2=O

trans-2-butene C/C=C/C

peroxyacetyl nitrate CC(=O)OO[N+](=O)[O-]

acetone CC(=O)C
glyoxal C(=O)C=O
crotonaldehyde C/C=C/C=O
cyclohexanone C1CCC(=O)CC1
cyclohex-2-eneone C1CC=CC(=O)C1
1-(4-methyl-phenyl)-ethanone Cc1ccc(cc1)C(=O)C
1-phenyl-1-butanone CCCC(=O)c1ccccc1

2,4-dimethyl-benzaldehyde CC1=CC(=C(C=C1)C=O)C

cyclohexane C1CCCCC1

1,1-dimethyl cyclopentane CC1(CCCC1)C

3-ethyl-phenol CCc1ccc(c1)O

p-hydroxybiphenyl C1=CC=C(C=C1)C2=CC=C(C=C2)O

 cis-2-butene-1,4-diol
 C(/C=C/CO)O

 oct-2-en-4-ol
 OC(/C=C/C)CCCC

 1,7-heptanediol
 C(CCCO)CCCO

pinic acid CC1(C(CC1C(=O)O)CC(=O)O)C
norpinic acid CC1(C(CC1C(=O)O)C(=O)O)C

octadeca-9-enoic acid CCCCCCC/C=C/CCCCCCC(=O)O pentamethyl benzoic acid Cc1c(c(c(c(c1C)C)C(=O)O)C)C

heptanamide CCCCCC(=O)N diethyl-butanamide CCC(CC)(CC)C(=O)N

n-ethyl-n-phenylamine CCNc1cccc1

triethanolamine C(CO)N(CCO)CCO
methyl dimethoxyethanoate COC(C(=O)OC)OC
methyl benzoate COC(=O)c1ccccc1
2-methyl-propyl benzoate CC(C)COC(=O)c1ccccc1

1,3-dioxolan C1COCO1

2-phenyl-1,3-dioxolane c1ccc(cc1)C2OCCO2
2,4-dimethoxybenzoic acid COc1ccc(c(c1)OC)C(=O)O

phenylmethyl nitrate C1=CC=C(C=C1)CO[N+](=O)[O-]2,4-dinitrophenol c1cc(c(c1[N+](=O)[O-])[N+](=O)[O-])O

4-nitrophenol c1cc(ccc1[N+](=O)[O-])O

2-methyl-6-nitrobenzoic acid Cc1cccc(c1C(=O)O)[N+](=O)[O-]

di-(1-methyl-propyl) peroxide CCC(C)OOC(C)CC ethylbutanamide CCCC(=O)NCC

 $\begin{array}{ccc} \text{C811CO3} & & & & & & & \\ \text{O]OC(=O)CC1CC(C(=O)O)C1(C)C} \\ \text{APINBOO} & & & & & & \\ \text{C0-][O+]=CCC1CC(C(=O)C)C1(C)C} \\ \text{C106O2} & & & & & & \\ \text{O=CCC(=O)CC(C(=O)C)C(C)(C)O[O]} \\ \end{array}$

C721O OC(=O)C1CC([O])C1(C)C

2,2-Dimethylpropaneperoxoic acid OOC(=O)C(C)(C)C

APINCO CC1=CCC(CC10)C(C)(C)[O]
C89CO2 O=CCC1CC(C(=0)[O])C1(C)C

 $\begin{array}{ccc} \text{C10PAN2} & \text{O=N(=O)OOC(=O)CC1CC(C(=O)C)C1(C)C} \\ \text{Pinanol} & \text{O=N(=O)OC1(C)C(O)CC2CC1C2(C)C} \\ \text{C811CO3H} & \text{OOC(=O)CC1CC(C(=O)O)C1(C)C} \\ \text{C106OOH} & \text{O=CCC(=O)CC(C(=O)C)C(C)(C)OO} \end{array}$

Ethyl sulfate CCOS(=O)(=O)O

Toluene Cc1cccc1
Nitroperoxymethane COON(=O)=O
Diethylamine CCNCC
Dimethylamine CNC

Table B2. List of compounds used to test the substructures 33-57 in Table 1.

Compound name	Smiles
propane	CCC
pentyne	CCCC#C
benzene	c1ccccc1
pentenal	CC/C=C/C=O
benzaldehyde	c1ccc(cc1)C=O
cinnamaldehyde	c1ccc(cc1)C=CC=O
mesityloxide	CC(=CC(=O)C)C
acetophenone	CC(=O)c1ccccc1
benzophenone	c1ccc(cc1)C(=O)c2cccc2
cyclopentanone	C1CCC(=O)C1
biacetyl	CC(=O)C(=O)C
pentadione	CC(=O)CC(=O)C
methylmethacrylate	CC(=C)C(=O)OC
methylbenzoate	COC(=O)c1ccccc1
vinylacetate	CC(=O)OC=C
butyrolactone	C1CC(=O)OC1
ethanoic anhydride	CC(=O)OC(=O)C
acetyl chloride	CC(=O)Cl
propionitrile	CCC#N
methyl isocyanate	CN=C=O
methyl isothiocyanate	CN=C=S
ethanimine	CC=N
acetone oxime	CC(=NO)C
nitrobenzene	c1ccc(cc1)[N+](=O)[O-]
nitropropane	CCC[N+](=O)[O-]

Appendix C: Software program

ASCII tables of the SMARTS patterns and the python program assembled for this work is released as Python program, APRL-SSP (<u>APRL Substructure Search Program</u>; Takahama, 2015), licensed under the GNU Public License version 3.0. In this program, series of scripts allow users to access the functionality of pybel and ChemSpiPy through input and output files defined as CSV-formatted tables.

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Tables

Table 1a. Substructures matched in order to account for the complete set of carbons and oxygen atoms in the set of compounds constituting the α -pinene and 1,3,5-trimethylbenzene degradation scheme in MCM v3.2 (substructures 1-33) and extra molecular substructures measurable with FTIR (substructures 33-57). For space constraints the SMARTS patterns have been reported on multiple lines, even if the SMARTS notation requires unique lines.

Ν°	Substructure	Definition	Chemoinformatic definition	Matched pattern
1	Quaternary carbon	A carbon atom bonded to four carbon atoms. ¹	[\$([C]([#6])([#6])([#6])[#6])]	C-C-C C
2	Alkane CH	Hydrogen atom attached to a sp3 carbon atom.	[CX4][H]	C C C
3	Alkene CH	Hydrogen atom attached to a non aromatic sp ² carbon atom.	[CX3;\$(C=C)][H]	H, C,
4	Aromatic CH	Hydrogen atom attached to an aromatic sp ² carbon atom.	[c][H]	С-Н
5	C sp ² non quaternary	A non aromatic sp ² carbon atom bonded to three carbons.	[CX3;\$([C]([#6])(=[#6])[C])]	C-CH ₃
6	C sp ² aromatic non quaternary	An aromatic sp ² carbon atom bonded to three carbon atoms.	[c;\$(c(c)[C])]	C-CH ₃
7	Alcohol OH	A compound containing an -OH (hydroxyl) group bonded to a tetrahedralcarbon atom. 1	[C;!\$(C=O)][OX2H][H]	_с-о-н
8	Ketone	A compound containing a carbonyl group bonded to two carbon atoms. 1	[CX3;\$(C([#6])(=[0])[#6])] (=[0;!\$([0][0])]))) 0=0 , 0=0
9	Aldehyde	A compound containing a -CHO group. ¹ (excludes formaldehyde)	[CX3;\$(C([#1])(=[0])[#6])] (=[0;!\$([0][0])])[H]	H, C, C,

¹Brown et al. (2012)

 $^{^2\}mathrm{Miloslav}$ et al.

Table 1b. Continued

Ν°	Substructure	Definition	Chemoinformatic definition	Matched pattern
10	Carboxylic acid	A compound containing a carboxyl, -COOH, group. 1 [CX3; !\$([CX3][H])] (=0) [OX2H][H]		-\c-c',
11	Formic acid	Formic acid compound.	[CX3] (=O) ([H]) [OX2H] [H]	H-Ć OH
12	Acyloxy radical	Oxygen-centered radicals consisting of an acyl radical bonded to an oxygen atom. ²	[C;\$(C=O)](=O)[OX2; !\$([OX2][H]);!\$([OX2][O]); !\$([OX2][N]);!\$([OX2]([#6]) [#6])]	-ć′ ••••••••••••••••••••••••••••••••••••
13	Ester	A derivative of a carboxylic acid in which H of the carboxyl group is replaced by a carbon. ¹	[CX3H1,CX3](=O) [OX2H0][#6;!\$([C]=[O])]	-c, -c, -
14	Ether	An -OR group, where R is an alkyl group. ¹	[OD2]([#6;!\$(C=O)]) [#6;!\$(C=O)]	- <u>c-o-c</u>
15	Formaldehyde	Formaldehyde compound.	[CX3;\$(C(=[0])([#1])[#1])] (=[0;!\$([0][0])])([H])[H]	0:C H
16	Phenol OH	Compounds having one or more hydroxy groups attached to a benzene or other arene ring. ²	[c;!\$(C=O)][OX2H][H]	С-ОН
17	Oxy radical (alkoxy)	Oxygen centered radical consisting of an oxygen bonded to an alkyl.	[#6;!\$(C=0)][OX2;!\$([OX2][H]); !\$([OX2][O]);!\$([OX2][N]); !\$([OX2]([#6])[#6]); !\$([OX2][S])]	- <u>\</u> c-o •
18	Carboxylic amide (primary, secondary and tertiary)	A derivative of a carboxylic acid in which the -OH is replaced by an amine. ¹	[CX3] (=0) [NX3; !\$ (N=0)] ([#6,#1]) [#6,#1]	O C N C(H)
19	Peroxide	Compounds of structure ROOR in which R may be any organyl group. ²	[#6][OD2][OD2,OD1][#6]	>c_o_o_c <u>/</u>
20	Peroxy radical	Oxygen centered radical derived from an hydroperoxyde.	[0;!\$([0][#6]);!\$([0][H]); !\$([0X2][N]);!\$(0=C)][0] [#6;!\$([C](=O)~OO)]	-c-o-o•
21	C=O ⁺ -O ⁻ group	Group of the type C=O ⁺ -O ⁻	[O;!\$([O][#6]);!\$([O][H]); !\$([OX2][N]);!\$(O=C)] [O]=[#6;!\$([C](=O)~OO)] ([#6,#1])[#6,#1]	C=O ⁺ -O ⁻
22	C-nitro	Compounds having the nitrogroup, -NO ₂ (free valence on nitrogen), which is attached to a carbon. ²	[#6][\$([NX3](=0)=0), \$([NX3+](=0)[0-])](~[0]) (~[0])	,O C(Ar)-N, O
23	Organonitrate	Compounds having the nitrogroup, -NO ₂ (free valence on nitrogen), which is attached to an oxygen. ²	[#6][0][\$([NX3](=[OX1]) (=[OX1])0),\$([NX3+]([OX1-]) (=[OX1])0)](~[O])(~[O])	
24	Peroxyacyl nitrate	Functional group containing a -COOONO ₂ .	[C] (=0)00[N] (~0)~[0])C-0-0-N,
25	Peroxy acid	Acids in which an acidic -OH group has been replaced by an -OOH group. ²	C(=0)0[0][H]	O, C-0-0H

Table 1c. Continued

Ν°	Substructure	Definition	Chemoinformatic definition	Matched pattern
26	Acylperoxy radical	Oxygen centered radical c(=0)0[0;!\$([0][H]); edited from a peroxy acid.)C-0-0 •
27	Organosulfate	Esters compounds derived from alcohol and sulfuric acids functional groups.	[#6][0][SX4; \$([SX4](=0)(=0)(0)0), \$([SX4+2]([0-])([0-])(0)0)] (~[0])(~[0])(~[0])	-C-O-S-O-
28	Hydroperoxide	A compound containing an -OOH group. ¹	[#6;!\$(C=O)][OD2] [OX2H,OD1][#1]	_C-O-O-H
29	Primary amine	An amine in which nitrogen is bonded to one carbon and two hydrogens. ¹	[#6][NX3;H2;!\$(NC=O)] ([H])[H]	C-N-H
30	Secondary amine	An amine in which nitrogen is bonded to two carbons and one hydrogen. ¹	[#6][NX3;H;!\$(NC=O)] ([#6])[H]	
31	Tertiary amine	An amine in which nitrogen is bonded to three carbons. ¹	[#6][NX3;H0;!\$(NC=O); !\$(N=O)]([#6])[#6]	
32	Peroxy nitrate	Functional group containing a COONO ₂ .	[#6][0;!\$(OOC(=0))] [0;!\$(OOC(=0))][N](~0)~[0]	-c-o-o-v, o
33	Anhydride	Two acyl groups bonded to an oxygen atom. 1	[CX3] (=0) [0] [CX3] (=0)	C,C:0 C,C:0
34	Alcohol O-H and Phenol O-H	Alcohol and phenol O-H.	[OX2H;\$([O]([#6])[H]); !\$([O](C=O)[H])][H]	O H and
35	Alkane C-H in -CH ₃	C-H bonds in CH ₃ group.	[CX4;\$(C([H])([H])[H])][H]	H -C-H
36	Alkane C-H in -CH ₂	C-H bonds in CH ₂ group.	[CX4;\$(C([H])([H]) ([!#1])[!#1])][H]	-C-C-C \
37	Alkynes C-H	Hydrogen bonded to a sp carbon in an alkyne group.	[C;\$(C#C)][H]	—C≣C−H
38	Alkynes C≡C	Two carbons that are triple bonded.	[C]#[C]	_c <u>=</u> c_
39	Aromatic C=C	Two aromatic carbons bonded with an aromatic bond.	c:c	CC
40	Conjugated aldehyde C=O and α,β C=C	An aldehyde C=O conjugated with an alkene C=C in α and β positions.	[CX3;\$(C(=[0])([#1])[C]=[C])] ([C]=[C;!\$(Cc)]) (=[0;!\$([0][0]))[H]	С=С-С-Н
41	Conjugated aldehyde C=O and phenyl	An aldehyde C=O conjugated with a phenyl group.	[CX3;\$(C(=[0])([#1]) [c;\$(c1cc[c]cc1)])]([#6,#1]) (=[0;!\$([0][0])])[H]	Ar-C-H O
42	Conjugated aldehyde C=O and α,β C=C and phenyl	An aldehyde C=O conjugated with alkene C=C in α and β positions and a phenyl group.	[CX3; \$ (C(=[0]) ([#1]) [C]=[C] [c; \$ (c1cc[c]cc1)])] ([C]) (=[0;!\$([0][0])]) [H]	Ar-C=C-C-H

Table 1d. Continued

Ν°	Substructure	Definition	Chemoinformatic definition	Matched pattern
43	Conjugated ketone C=O and α , α C=C	A ketone C=O conjugated with an alkene C=C in α and β positions.	[CX3; \$ (C([#6]) (=[0]) [C]=[C])]([C]) (=[0;!\$([0][0])])[C]	C=C-C-C
44	Conjugated ketone C=O and phenyl	A ketone C=O conjugated with a phenyl group.	[CX3;\$(C([C])(=[0]) [c;\$(c1cc[c]cc1)])]([C]) (=[0;!\$([0][0])])[c]	Ar-C-C
45	Conjugated ketone C=O and two phenyl	A ketone C=O conjugated with two phenyl groups.	[CX3; \$ (C([c, \$ (clcc[c]ccl)]) (=[0]) [c; \$ (clcc[c]ccl)])] ([c]) (=[0; !\$ ([0][0])]) [c]	Ar-C-Ar O
46	Conjugated ester C=O and α,β C=C	An ester C=O conjugated with alkene C=C in α and β positions .	[C;!\$(Cc)]=[C] [CX3;\$([C]([O][C]) (=[O])[C]=[C])]([O][C]) (=[O;!\$([O][O])])	c=c-c-o-c
47	Conjugated ester C=O and phenyl	A ester C=O conjugated with a phenyl group.	[CX3;\$([C]([O][C])(=[O]) [c,\$(c1cc[c]cc1)])]([O][C]) (=[O;!\$([O][O])])	Ar-C-O-C
48	Conjugated ester C-O with C=C or phenyl	An ester C=O conjugated with alkene C=C in α and β positions and a phenyl group.	[CX3; \$([C]([#6])(=[0])[0] [C]=[C]), \$([C]([#6])(=[0]) [O][c; \$(clcc[c]ccl)])] (=[0;!\$([0][0])])[0] [#6; \$(C=C), \$(clcc[c]ccl)]	
49	Nonacid carbonyl	Carbonyl group in ketones and aldehydes.	[CX3;\$(C([#6,#1])(=[0]) [#6,#1])](=[0;!\$([0][0])])	C(H) C=O
50	Acyl Chloride	An acyl group bonded to a chloride atom.	[C,\$([C]([#6])(=[O]))] (=O)[C1])c C Cl
51	Isocyanate	An -N=C=O group.	[N; \$([N]([#6])=[C]=[O])] =[C]=[O]	_C-N=C=O
52	Isothiocyanate	An -N=C=S group.	[N; \$([N]([#6])=[C]=[S])] =[C]=[S]	-)c-N=C=S
53	Imine	A carbon- nitrogen double bond, $R_2C=NR$.	[C;\$(C([#6,#1])([#6,#1])=[N])] =[N][#1,#6]	C(H) $C(H)$ $C(H)$
54	Oxime	A carbon-nitrogen double bond, $R_2C=NOH$.	[C;\$(C([#6,#1])([#6,#1]) =[N][O][H])]=[N][O][H]	C(H) C=N OH C(H)
55	Aliphatic nitro	Compounds having the nitro group, -NO ₂ (free valence on nitrogen), which is attached to an alifatic carbon.	[C][\$([NX3](=0)=0), \$([NX3](=0)[O-])]+ (~[O])(~[O])	
56	Aromatic nitro	Compounds having the nitro group, -NO ₂ (free valence on nitrogen), which is attached to an aromatic carbon.	[c][\$([NX3](=0)=0), \$([NX3](=0)[0-])]+ (~[0])(~[0])	Ar – N,
57	Nitrile	A carbon atom bonded to a nitrogen atom with a triple bond.	[C;\$([C]#[N])]#[N]	-}C-C≣N

Table 2: Chemical substructures required by SIMPOL.1 model (Pankow and Asher, 2008). The column denoted by k corresponds to the group number of Pankow and Asher (2008), Table 5. For the calculation of the ester (SIMPOL.1), the generic ester specified in Table 1 (substructure 13) is specified. The group named 'Carbon number on the OH side of an amide' is used in the calculation of the 'carbon number on the acid side of an amide' but is not present in the SIMPOL.1 groups indicated by Pankow and Asher (2008).

Groups	Chemoinformatic definition	
Groups	or reference to Table 1	k
Carbon number	[#6]	1
	{Carbon number}-	
Carlo an arranda a card	{Carbon number on the OH side of an amide}-1	
Carbon number on the acid side of an amide*†	<pre>if ({Amide, primary}+{Amide, secondary}</pre>	2
side of an amide	+{Amide, tertiary}> 0)	
	else 0	
Aromatic ring [‡]	count_aromatic_rings(molecule)	3
Non aromatic ring [‡]	count_nonaromatic_rings(molecule)	4
C=C (non aromatic)	C=C	5
C=C-C=O in non-aromatic ring	[\$(C=CC=O);A;R]	6
Hydroxyl (alkyl)	Table 1, number 7	7
Aldehyde	[CX3; \$ (C([#1]) (=[0]) [#6, #1])] (=[0;!\$([0][0])])	8
Ketone	Table 1, number 8	9
Carboxylic acid	[CX3] (=0) [OX2H] [H]	10
Ester (SIMPOL.1) [†]	{Ester}-{Nitroester}	11
Ether (SIMPOL.1)	[OD2]([C;!R;!\$(C=O)])[C;!R;!\$(C=O)]	12
Ether, alicyclic	[OD2;R]([C;!\$(C=0);R])[C;!\$(C=0);R]	13
Ether, aromatic	c~[O,o]~[c,C&!\$(C=O)]	14
Nitrate	Table 1, number 23	15
Nitro	Table 1, number 22	16
Aromatic hydroxyl (e.g. phenol)	Table 1, number 16	17
Amine, primary	[C][NX3;H2;!\$(NC=O)]([H])[H]	18
Amine, secondary	[C][NX3;H;!\$(NC=O)]([C])[H]	19
Amine, tertiary	[C][NX3;H0;!\$(NC=O);!\$(N=O)]([C])[C]	20
Amine, aromatic	[N;!\$(NC=O);!\$(N=O);\$(Na)]	21
Amide, primary	[CX3;\$(C(=[0])[NX3;!\$(N=0)])](=[0])[N]([#1])[#1]	22
A!JJ	[CX3; \$ (C(=[0]) [NX3; !\$ (N=0)] ([#6]) [#1])] (=[0])	22
Amide, secondary	[N][#1]	23
Amida tartiary	[CX3;\$(C(=[0])[NX3;!\$(N=0)]([#6])[#6])](=[0])	24
Amide, tertiary	[N]	24

Carbonylperoxynitrate	Table 1, number 24	
Peroxide	Table 1, number 19	
Hydroperoxide	Table 1, number 28	
Carbonylperoxyacid	Table 1, number 25	28
Nitrophenol [‡]	<pre>count_nitrophenols(molecule,'{phenol},'{nitro})</pre>	29
	[#6][OX2H0][CX3,CX3H1](=O)[C;\$(C[N](~[O])~[O]),	
Nitroester*	\$(CC[N](~[0])~[0]),\$(CCC[N](~[0])~[0]),	30
Muocstei	\$(CCCC[N](~[0])~[0]),	
	\$(CCCCC[N](~[0])~[0])]	
	[C;\$(C[NX3][CH,CC](=O)),\$(CC[NX3][CH,CC](=O)),	
Carbon number on the OH side of an amide	\$(CCC[NX3][CH,CC](=0)),\$(CCCC[NX3][CH,CC](=0)),	
	\$(CCCCC[NX3[CH,CC](=0))]	

^{*}In the case of the calculations of the number of carbons on the acid side of an amide and for nitroester is this table, these patterns provide correct counting for compounds with a maximum of 5 carbon atoms on the acid side of an amide or in between the ester and the nitro group respectively. To match cases with higher number of carbon atoms, it is necessary to repeat the specified pattern with an augmented number of carbons specified in the code.

[‡]User-defined functions which access additional molecular structure information for ring structures. molecule is a reserved name indicating an object of the Molecule class defined by the pybel library for our implementation, and entries in quoted braces '{} passed as arguments correspond to the matched substructure prior to enumeration. These functions are provided as part of the companion program (Appendix C). This functional interface abstracts the calculation such that the patterns above can be used with any chemoinformatic software package provided that the implementation of ring enumeration functions are changed accordingly.

[†]Quantities are calculated from other groups; the code shown is executable string formatting syntax of the Python programming language. Entries in braces {} are replaced by the number of matched groups designated by name.

Table 3. List of SMARTS patterns and coefficients associated with each bond type, used to calculate the carbon oxidation state as described in the Section 2.

Bond	SMARTS pattern	Coefficient
С-Н	[#6][H]	-1
C-C	[#6]-[#6]	0
C=C	[#6]=[#6]	0
C≡C	[#6]#[#6]	0
C-O	[#6]-[#8]	1
C=O	[#6]=[#8]	2
C-N	[#6]-[#7]	1
C=N	[#6]=[#7]	2
C≡N	[#6]#[#7]	2
C-S	[#6]-[#16]	1
C=S	[#6]=[#16] 2	
C≡S	[#6]#[#16]	3

Table 4: Absorption bands in the infrared region of different FGs and the correspondence in Table 1.

N°	Functional group and	Wayanan kan (am =1)
1	functional groups pattern	Wavenumber (cm ⁻¹)
2, 35, 36		2900 (C-H stretch),
	Alkane C-H	1450 and 1375 (bend in CH ₃),
		1465 (bend in CH ₂)
		3100 (C-H stretch),
3	Alkene C-H	720 (Bend, rocking),
		100-650 (Out of plane bend)
37	Alkyne C-H	3300 (Stretch)
4	Aromatic C-H	3000 (C-H stretch),
4	Afolhatic C-H	900-690 (Out of plane bend)
38	Alkyne C≡C	2150 (CC stretch)
39	Aromatic C=C	1600 and 1475 (Stretch)
		3400 (O-H stretch),
7, 16, 34	Alcohol and phenol	1440-1220 (C-O-H bend),
		1260-1000 (C-O stretch),
	Carboxylic acid COOH	3400 - 2400 (O-H stretch),
10, 11		1730-1700 (C=O stretch),
		1320-1210 (stretch)
	Aldehyde and ketone	1740 (aldehyde C=O stretch),
8, 9, 15, 49		1720-1708 (ketone C=O stretch),
0, 9, 13, 49		1300-1100 (ketone C(C=O)C bend),
		2860-2800 and 2760-1200 (aldehyde C-H stretch)
		1640 - 1560 (N-H bend, in primary amines),
	Amines	3500-3300 (secondary and primary amines N-H stretch),
29, 30, 31		1500 (secondary amines N-H bend),
		800 (secondary and primary amines N-H out of plane bend),
		1350-1000 (C-N stretch)
14	Ether	1300-1000 (C-O stretch)
13	Ester	1750-1735 (C=O stretch),
13	Ester	1300-1000 (C-O stretch)
		1680-1630 (C=O stretch),
18, (SIMPOL.1 groups)	Amide	3350 and 3180 (primary amide N-H stretch),
10, (ontil OL.1 groups)		3300 (secondary amide N-H stretch),
		1640-1550 (primary and secondary amide N-H bend)

27	Organosulfate	876 (C-O-S stretch)
23	Organonitrate	1280 (symmetric NO ₂ stretch)
50	Acid Chloride	1850-1775 (C=O stretch),
30	Acid Cilionae	730-550 (C-Cl stretch)
		1600-1640 (aliphatic nitro -NO ₂ asymmetric stretch),
22, 55, 56	Nitro	1390-1315 (aliphatic nitro -NO ₂ symmetric stretch),
22, 33, 30	Nitio	1550-1490 (aromatic nitro -NO ₂ asymmetric stretch),
		1355-1315 (aromatic nitro -NO ₂ symmetric stretch)
57	Nitrile	2250 (stretch, if conjugated 1780-1760)
51	Isocyanate	2270 (stretch)
52	Isothiocyanate	2125 (stretch)
53	Imine	1690-1640 (stretch)
33	Anhydride	1830-1800 (C=O stretch),
33	Aimydride	1775-1740 (C-O stretch)
		1700-1680 and 1640 (conjugated aldehyde C=O
		with C=C in α and β),
40, 41, 42	Conjugated aldehyde	1700-1660 and 1600-1450 (conjugated aldehyde C=O
		with phenyl),
		1680 (conjugated aldehyde C=O with C=C and phenyl),
		1700-1675 and 1644-1617 (conjugated ketone C=O
43, 44, 45	Conjugated ketone	and α, β C=C),
13, 11, 13	Conjugated Retone	1700-1680 and 1600-1450 (conjugated ketone C=O with phenyl),
		1670-1600 (conjugated ketone and two phenyl)
		1740-1715 and 1640-1625 (conjugated ester C=O and α , β C=C),
46, 47, 48	Conjugated ester	1740-1715 and 1600-1450 (conjugated ester C=O and phenyl),
		1765-1762 (conjugated ester C-O with C=C or phenyl)

Figures

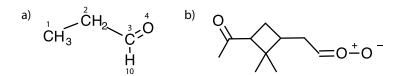


Figure 1. Propionaldehyde (a, SMILES code CCC=O) and compound named APINOOB in MCMv3.2 scheme (b, SMILES code [O-][O+]=CCC1CC(C(=O)C)C1(C)C). The carbon and oxygen atoms are enumerated, together with the hydrogen of the aldehyde group in compound a.

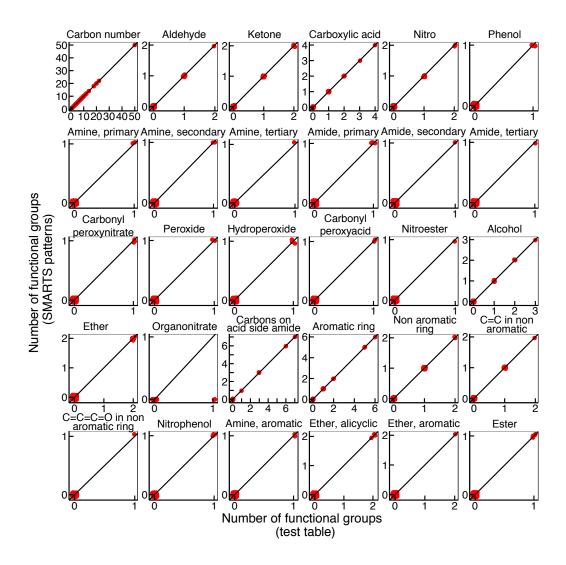


Figure 2. Validation of the developed chemoinformatic patterns for the chemical substructures required in the SIMPOL.1 model (Pankow and Asher, 2008). This validation set includes 99 compounds as described in Section 2.

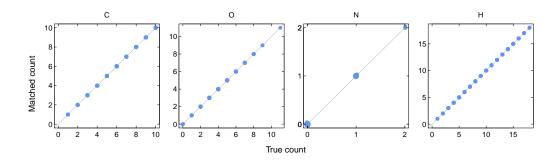


Figure 3. Test of the completeness of matching of all the atoms in the α -pinene and 1,3,5-trimethylbenzene degradation scheme in MCMv3.2 by the SMARTS patterns in Table 1, substructures 1-33.

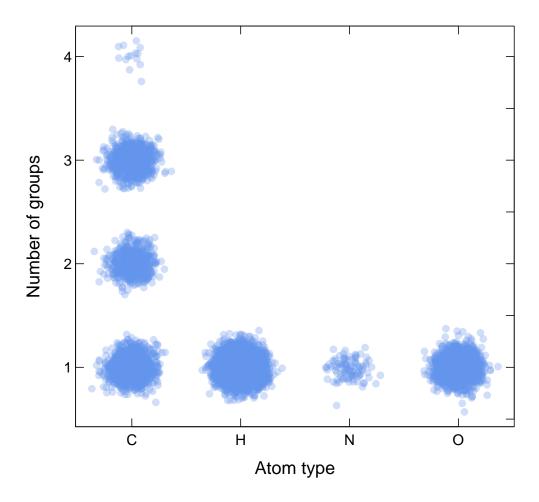


Figure 4. Test for the uniqueness of matching for each atom. Number of times a specific atom has been matched, in the α -pinene and 1,3,5-trimethylbenzene degradation scheme in MCMv3.2 by the SMARTS patterns in Table 1, substructures 1-33. Oxygen, nitrogen and hydrogen atoms are matched only once. The carbon atoms are matched multiple times when multifunctional.

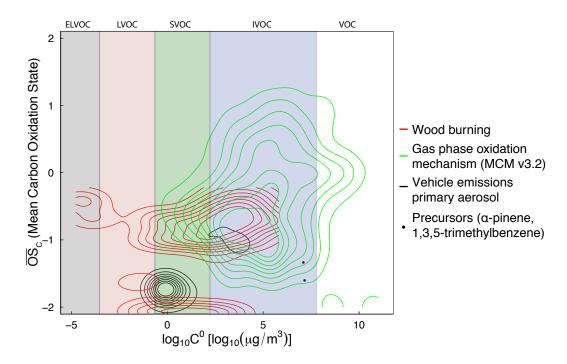


Figure 5. Logarithm of the pure component saturation concentration ($\log_{10}C^0$) and mean carbon oxidation state of each compound ($\overline{\rm OS}_{\rm C}$) measured by Rogge et al. (1993) and Rogge et al. (1998) for biomass burning and vehicle emissions sources (green and blue lines), and of each molecule constituting the MCMv3.2 gas phase oxidation mechanism of α -pinene and 1,3,5-trimethylbenzene. The lines in the plot denote isolines (0,0.1,...,0.9) of the maximum density estimate for the different compound sets. The black dots indicate the position of α -pinene and 1,3,5-trimethylbenzene. The area of the plot is divided in volatility regions according to the classification of Donahue et al. (2012).

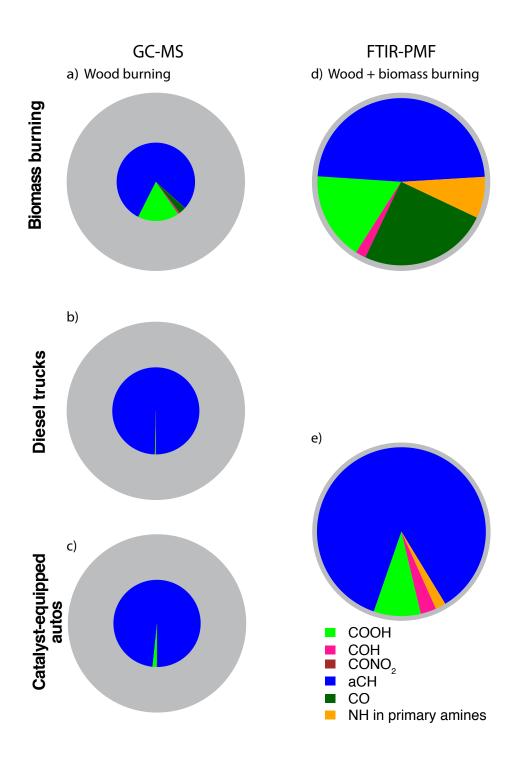


Figure 6. Comparison of the FG distribution of the quantified fraction measured by GC-MS (a,b and c; Rogge et al., 1998; Rogge et al., 1993) and FTIR-PMF (d and e; Hawkins and Russell, 2010) in aerosol emitted by biomass burning (a and d) and vehicle emission (b,c and e) sources. The gray area is the unresolved OA fraction by the two different analytical techniques used (around 80% for GC-MS and around 10% for FTIR). The type of biomass burning is specified in the pie charts a and d.

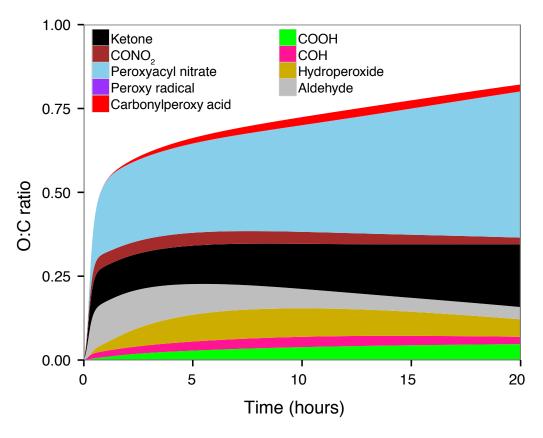


Figure 7. Time series of FG contributions to the total O:C of the gas phase generated by photooxidation of α -pinene in low-NO_x regime, simulated using the MCMv3.2 degradation scheme.