

Pesticides in the atmosphere: a comparison of gas-particle partitioning and particle size distribution of legacy and current-use pesticides

C. Degrendele^{1,2}, K. Okonski¹, L. Melymuk¹, L. Landlová¹, P. Kukučka¹, O. Audy¹, J. Kohoutek¹, P. Čupr¹, J. Klánová¹

[1]{Masaryk University, RECETOX (Research Centre for Toxic Compounds in the Environment), Faculty of Science, Kamenice 753/5, 62500 Brno, Czech Republic}

[2]{ Max Planck Institute for Chemistry, Multiphase Chemistry Department, Hahn-Meitner-Weg 1, 55128 Mainz, Germany }

Correspondence to: J. Klánová (klanova@recetox.muni.cz) and L. Melymuk (melymuk@recetox.muni.cz)

Abstract

This study presents a comparison of seasonal variation, gas-particle partitioning and particle-phase size distribution of organochlorine pesticides (OCPs) and current-use pesticides (CUPs) in air. Two years (2012/2013) of weekly air samples were collected at a background site in the Czech Republic using a high-volume air sampler. To study the particle-phase size distribution, air samples were also collected at an urban and rural site in the area of Brno, Czech Republic, using a cascade impactor separating atmospheric particulates according to six size fractions. Major differences were found in the atmospheric distribution of OCPs and CUPs. The atmospheric concentrations of CUPs were driven by agricultural activities while secondary sources such as volatilization from surfaces governed the atmospheric concentrations of OCPs. Moreover, clear differences were observed in gas-particle partitioning; CUP partitioning was influenced by adsorption onto mineral surfaces while OCPs were mainly partitioning to aerosols through absorption. A predictive method for estimating the gas-particle partitioning has been derived and is proposed for polar and non-polar pesticides. Finally, while OCPs and the majority of CUPs were largely found on fine particles, four CUPs (carbendazim, isoproturon, prochloraz and terbuthylazine) had higher

1 concentrations on coarse particles ($>3.0 \mu\text{m}$), which may be related to the pesticide
2 application technique. This finding is particularly important and should be further investigated
3 given that large particles result in lower risks from inhalation (regardless the toxicity of the
4 pesticide) and lower potential for long range atmospheric transport.

5 **1 Introduction**

6 In 1939, dichlorodiphenyltrichloroethane (DDT) was discovered to have insecticidal
7 properties. Since that time, synthetic pesticides have been widely used around the world to
8 control pests in agricultural production (Li and Macdonald, 2005). Legacy organochlorine
9 pesticides (OCPs) are banned for agricultural purposes in most countries, including the Czech
10 Republic, and have been replaced by what are often termed “current-use pesticides” (CUPs).
11 CUPs generally have lower persistence and bioaccumulative potential and higher water
12 solubility, which should result in reduced negative environmental impacts (Kannan et al.,
13 2006). However, given their detection in multiple environmental media, including in remote
14 locations (Koblizková et al., 2012; Zhang et al., 2013), and the relative lack of information
15 regarding their toxic effects, the potential environmental and human risks cannot be
16 neglected. The atmospheric transport of OCPs has been well studied over the last decades
17 (Lammel et al., 2009; Růžicková et al., 2008; Shen et al., 2004), but there is a lack of
18 information on the seasonal trends and partitioning of CUPs which is needed to understand
19 their environmental fate. There are three main processes leading to the presence of pesticides
20 in the air. First, pesticides can enter the atmosphere during application. For example, during
21 spray application, up to 30% of the dosage directly enters the atmosphere (Van den Berg et al.,
22 1999). Another primary emission is wind erosion of soil particles containing sorbed
23 pesticides, which can occur days or weeks after application (Glotfelty et al., 1989). Finally,
24 pesticides are affected by air-surface exchange such as the volatilization from plants and soils,
25 surface waters, and from old industrial sites (Cabrerizo et al., 2011). In the case of pesticides
26 that are not currently authorized for agricultural use (e.g., OCPs), volatilization and wind
27 erosion of soil particles should be the only relevant emission pathways.

28 Once pesticides enter the air, they partition between gas and particulate phases according to
29 their physicochemical properties (vapor pressure, octanol-air partition coefficient K_{oa}), the
30 concentration of total suspended particulate matter (TSP) and meteorological parameters
31 (ambient temperature, relative humidity) (Cousins and Mackay, 2001; Lohmann and Lammel,
32 2004; Pankow, 1987). Knowledge of this gas-particle partitioning is necessary to understand

1 atmospheric residence times, the significance of removal pathways from air (deposition, gas
2 absorption, photodegradation) and the potential for long range atmospheric transport (LRAT)
3 (Bidleman et al., 1986; Eisenreich et al., 1981; Scheyer et al., 2008). Additionally, the
4 atmospheric residence times of particles vary with particle size (Vecchi et al., 2007), further
5 influencing wet/dry deposition and LRAT (Götz et al., 2008). Particle size distribution is also
6 an important factor for human risks from inhalation exposure, as smaller particles penetrate
7 deeper into the respiratory system (Englert, 2004).

8 The gas-particle partitioning of OCPs (Cindoruk, 2011; Scheyer et al., 2008; Sofuoglu et al.,
9 2004) and some CUPs (Borrás et al., 2011; Götz et al., 2007; Sadiki and Poissant, 2008;
10 Sanusi et al., 1999; Sauret et al., 2008; Yao et al., 2008) has been reported. However, the
11 seasonal variation of this partitioning has only been investigated for two CUPs: chlorpyrifos
12 (Li et al., 2014) and alachlor (Sauret et al., 2008). Similarly, knowledge of how CUPs are
13 distributed among different particle sizes is very limited (Coscollà et al., 2014, 2013b), and
14 the seasonality of this particle size distribution has never been investigated for CUPs. To fill
15 these gaps, we assess the seasonal variation of a set of legacy and current-use pesticides
16 (Table 1) in outdoor air, with a focus on the gas-particle partitioning and the particle size
17 distribution. For many of these CUPs, this is the first time that their seasonal gas-particle
18 partitioning and size distributions have been examined.

19

20 **2 Methodology**

21 2.1 Air sampling

22 Air samples were collected in two sampling campaigns. A map of the sampling sites is
23 provided in Fig. S1 in the Supplementary Information (SI). Firstly, to study seasonal trends
24 and gas-particle partitioning, air was sampled at the Košetice observatory (49°34'24"N,
25 15°04'49"E), which is an established background site of the European Monitoring and
26 Evaluation Programme (EMEP) network (Holoubek et al., 2007). The site is located in an
27 agricultural region in central Czech Republic. While the site is located in an agricultural
28 region, it is not directly on cultivated land, therefore the air sampled should not reflect direct
29 emissions from pesticide application (e.g., spray application droplets) but rather the average
30 conditions of a rural air mass. From January 2012 to December 2013, a high volume air

1 sampler (Digitel DH77 with PM₁₀ pre-separator) was used to collect weekly air samples. The
2 sample volume was on average 4310 m³ (~25 m³/h, 7 day sampling duration). Particles were
3 collected on quartz fiber filters (QFFs) (QM-A, 150 mm, Whatman, UK, pore size of 2.2 μm)
4 and gas phase on polyurethane foam (PUF) (two in series, T3037, 110 x 50 mm, 0.030 g/cm³,
5 Molitan a.s., Czech Republic). PUFs were pre-cleaned via Soxhlet-extraction with acetone
6 and dichloromethane for 8 hours each. Fifty-two samples were collected each year. Half of the
7 samples were used for OCPs analysis and half for CUPs analysis (Tables S1 and S2).

8 Secondly, to assess the seasonal variation of the particle size distribution of pesticides,
9 particulate-phase air samples were collected in the area of Brno, the second largest city in the
10 Czech Republic. From October 2009 to October 2010, a high volume air sampler (HV 100-P,
11 Baghirra, CZ) equipped with a multistage cascade impactor (PM₁₀ sampling head and six
12 stage impactor, Tisch Environmental, USA) was used to collect six particle size fractions. The
13 fractions represented particles with aerodynamic diameters of <0.49 μm, 0.49-0.95 μm, 0.95-
14 1.5 μm, 1.5-3.0 μm, 3.0-7.2 μm and 7.2-10 μm and were collected on QFFs (TE-230-QZ, 141
15 x 148 mm, Tisch, Environmental, USA and QM-A, 203 x 254 mm, Whatman, UK, for the
16 backup filters (<0.49 μm)). Sampling was conducted simultaneously at a rural site (Telnice)
17 and at an urban site (Kotlařská). The rural site (49°6'21"N, 16°42'58"E) was located 14 km
18 southeast of the Brno city centre. The main source of pollution at this site is likely agricultural
19 activity, especially from cereals and grapes, which are the main local crops. The urban site
20 (49°12'20"N, 16°35'50"E) was located in a university botanical garden, close to a major
21 traffic junction in the centre of Brno. Only a small amount of pesticides are used within the
22 botanical garden, and do not include any of the target pesticides in the present study. The main
23 sources of pesticides at this site are likely pesticides used in nearby buildings/building
24 materials, and atmospheric transport from the agricultural areas surrounding Brno. Eleven
25 weekly samples were used for CUPs analysis and twelve for OCPs analysis at each site. The
26 remaining samples were analyzed for other SVOCs, presented elsewhere (Degrendele et al.,
27 2014; Okonski et al., 2014). To reach the limit of detection of these compounds, samples were
28 grouped by season (two or three filters) (Tables S3 and S4). The sample volume was on
29 average 9734 m³ (~65 m³/h, 7 day sampling duration).

30 All filters and PUFs were wrapped in aluminum foil, sealed in plastic bags and stored at -
31 18°C until analysis.

32 2.2 Sample preparation and analysis

1 Filters and PUFs were extracted with toluene for OCP analysis and with methanol for CUP
2 analysis, using an automated warm Soxhlet extractor (Büchi Extraction System B-811) for
3 three cycles, each consisting of 60 min of warm Soxhlet and 30 min of solvent rinsing. The
4 extracts were concentrated using a gentle stream of nitrogen. After extraction, OCP extracts
5 were transferred to a glass column (30 mm i.d.) consisting of 0.5 g of activated silica, 30 g of
6 H₂SO₄-modified activated silica and 1 g of non-activated silica and were eluted with 240 mL
7 of DCM:Hexane (1:1 v/v). CUP extracts were passed through syringe filters (nylon
8 membrane, 25 mm diameter, pore size 0.45 µm).

9 OCPs were analyzed by gas chromatography coupled to a tandem mass spectrometer (GC-
10 MS/MS). CUPs were analyzed using an Agilent 1100 high performance liquid chromatograph
11 (HPLC) with a Phenomenex Luna C-18 endcapped analytical column (100 mm x 2.1 mm x 3
12 µm). Analyte detection was performed by tandem mass spectrometry using an AB Sciex Qtrap
13 5500 operating in positive electron spray ionization (ESI+). Further information on all
14 analytical parameters is given in the SI. Identification was based on a comparison of ion ratios
15 and retention times (Table S5) with corresponding isotopically-labeled standards for CUPs
16 and quantification was using internal standards: PCB-121 (Absolute Standards Inc., USA) for
17 OCPs andalachlor-d13, acetochlor-d11, chlorpyrifos d-10, isoproturon d-3, fenitrothion d-6,
18 desisopropylatrazine d-5, dimethoate d-6, diuron d-6, terbuthylazine d-5 and simazine d-10
19 (Toronto Research Chemicals, Canada; Dr. Ehrenstorfer LGC Standards, UK; Chiron AS,
20 Norway; and Neochema, Germany) for CUPs.

21 2.3 QA/QC

22 Breakthrough of gas phase compounds during air sampling was evaluated by separate
23 quantification of each of the two PUFs placed in series for all the weekly air samples
24 collected at the background site in 2012 (Tables S6 and S7). Based on the results of the
25 breakthrough evaluation, the sampling set-up was deemed appropriate for the quantification
26 of this set of pesticides. Thirteen field blanks and 28 laboratory blanks were analyzed as per
27 samples. Blank levels of individual analytes were below detection (all OCPs and 21 CUPs
28 were below detection in field blanks) or otherwise low (on average <3.5% of sample mass for
29 detected compounds). The concentrations of OCPs and CUPs presented here have been blank
30 corrected by subtracting the average of the field blanks. The OCP analytical method was
31 evaluated using a certified reference material (ASLAB soil standard, Czech Republic)
32 (Lohmann et al., 2012) and recoveries were assessed using spike-recovery tests of air

1 sampling media. Mean OCP recoveries (\pm standard deviation) ranged from $87.2\pm 6.26\%$ to
2 $113\pm 6.10\%$ with an average value of $95.8\pm 8.11\%$ (Table S8). CUP recoveries were
3 determined from spike-recovery tests of air sampling media and ranged from $52.4\pm 21.4\%$ to
4 $115\pm 17.4\%$ (Table S9). The measured concentrations have not been adjusted for recoveries.

5

6 **3 Results and discussion**

7 3.1 Detection frequency at the background site

8 In general, the detection frequency of CUPs related to their legal status, usage amounts and
9 their persistence in the environment, while OCPs were consistently detected ($>57\%$ of
10 samples) throughout the whole sampling period (Table 1). In particular, α -HCH, γ -HCH, p,p' -
11 DDE and p,p' -DDT were detected in every gas phase sample during the two years of
12 sampling, emphasizing the environmental persistence of these OCPs.

13 The CUPs included in this study represent 24% of all pesticides used in agriculture in the
14 Czech Republic (Tables S10 and S11), with acetochlor, chlorpyrifos, chlorotoluron,
15 isoproturon, metamitron, metazachlor, prochloraz and terbuthylazine used in the largest
16 quantities (>90 tonnes of active substance per year) and these CUPs were detected in $>25\%$ of
17 air samples. Isoproturon (detected in 86.5% of samples), metazachlor (86.5%), chlorpyrifos
18 (84.6%), terbuthylazine (78.8%), S-metolachlor (73.1%) and fenpropimorph (65.4%) were the
19 most frequently detected. Acetochlor, atrazine, carbendazim, chlorotoluron, dimethachlor,
20 diuron, metamitron, metribuzin, prochloraz and pyrazon had detection frequencies of 15-55%
21 (Table 1), occurring mostly during periods of agricultural activities. Finally, azinphos methyl
22 and fenitrothion were not detected in any samples and eight CUPs (alachlor, diazinon,
23 dimethoate, disulfoton, fonofos, malathion, simazine, temephos and terbufos) were
24 infrequently detected ($<6\%$). Amongst these infrequently detected pesticides, only dimethoate
25 is authorized for agricultural use in the Czech Republic and is used in very low amounts
26 (Tables S10 and S11). Thus, the infrequent detections of these compounds are likely due to no
27 or limited application in the sampling area.

28 We note that not all the CUPs are in current use in Czech Republic (Table 1); some pesticides,
29 which we have categorized as CUPs to distinguish them from the OCPs, are banned in Czech
30 Republic but remain in use elsewhere. For example, atrazine, a triazine pesticide banned in

1 the European Union since 2003 (European Commission - Health & Consumer Protection
2 Directorate-General, 2003) remains one of the highest use pesticides in USA (U.S.
3 Environmental Protection Agency, 2013). Atrazine was detected in only one sample from May
4 2012 but had more frequent detections between July and November 2013 (Tables S12 and
5 S13).

6 3.2 Total concentrations at the background site

7 Individual OCP and CUP concentrations are presented in Tables 1 and S12-S15.

8 Chlorpyrifos, metazachlor, acetochlor, isoproturon and S-metolachlor were the only CUPs
9 with maximum total (gas+particulate phase) concentrations exceeding 100 pg.m^{-3} , and, except
10 S-metolachlor, these pesticides are all used in quantities >100 tonnes/year in the Czech
11 Republic (Tables S10 and S11). Similarly, carbendazim, chlorotoluron, dimethachlor,
12 fenpropimorph, metamitron and terbuthylazine, which are all authorized for agricultural use
13 and used in quantities >30 tonnes/year (SRS, 2014, 2013), have maximum concentrations
14 higher than 10 pg.m^{-3} . However, beyond this broad categorization, a poor correlation was
15 found between mass used per year and maximum concentration ($r^2=0.362$ and 0.184 in 2012
16 and 2013, respectively). For example, prochloraz, which was used in similar quantities to
17 chlorpyrifos in 2013 (SRS, 2014), had maximum concentrations of only 1.95 pg.m^{-3} (vs. 159
18 pg.m^{-3} for chlorpyrifos). The lack of correlation may be caused by the use of a national
19 pesticide usage database obscuring regional differences, which are of importance given the
20 relatively low atmospheric residence time of CUPs (Coscollà et al., 2013b). Moreover, the
21 pesticide physicochemical properties, their environmental persistence and the pesticide
22 application technique used (e.g., seed treatment vs. spray application) may also influence the
23 atmospheric concentrations of CUPs. Indeed, spray application parameters such as the
24 volatility and viscosity of the pesticide formulation, equipment, weather conditions at the time
25 of application (wind speed and direction, temperature, relative humidity and stability of air at
26 the application site) and operator care, attitude and skill have been identified as factors that
27 influence the emission of pesticide droplets to the air (Gil and Sinfort, 2005), thereby
28 affecting local air concentrations.

29 All of the banned CUPs included in this study had maximum concentrations lower than 2.5
30 pg.m^{-3} (excepting fonofos with a concentration of 8.03 pg.m^{-3} in one sample from August
31 2013), reflecting low current emissions. In particular, atrazine had a maximum concentration

1 of 1.24 pg.m^{-3} in 2012 and lower concentrations ($<0.250 \text{ pg.m}^{-3}$) in 2013. The level of
2 simazine in the single sample in which it was detected was very low ($<0.1 \text{ pg.m}^{-3}$). Similarly,
3 in a recent study, these CUPs were detected in only one sample over the Central North Sea at
4 low concentrations ($<1 \text{ pg.m}^{-3}$) (Mai et al., 2013). In contrast, from 1984-1994 (before the
5 European ban), atrazine and simazine were frequently detected in precipitation (Dubus et al.,
6 2000). These triazines were also routinely detected in atmospheric samples in France during
7 the same period with concentrations up to 51 ng.m^{-3} for atrazine (Sanusi et al., 2000) and 3
8 ng.m^{-3} for simazine (Chevreuil et al., 1996). Thus, the low atmospheric concentrations of
9 atrazine and simazine observed in this study are likely a result of the European ban on use.

10 Of the OCPs, *p,p'*-DDE, γ -HCH and α -HCH had the highest contributions, accounting on
11 average for 56.3%, 15.5% and 11.7% of \sum OCPs. The ratio of *p,p'*-DDT/(*p,p'*-DDE+*p,p'*-
12 DDD) is often used as an indicator of aged technical DDT. A lower ratio is indicative of aged
13 (degraded) DDT, while a value >1 indicates fresh application (Li et al., 2007). In this study,
14 this ratio ranged from 0.0271 to 0.370, suggesting aged DDT.

15 The total concentrations of individual CUPs and OCPs were compared with previous studies
16 (Table S16 and references therein). OCPs levels were comparable to other European
17 background sites (Cabrerizo et al., 2011; Halse et al., 2011). \sum DDT concentrations in this
18 study (1.14-96.3 pg m^{-3}) were considerably lower than those reported in India or in Africa (8-
19 5930 and 8-2178 pg m^{-3} , respectively (Bogdal et al., 2013; Yadav et al., 2015)). The CUPs
20 concentrations reported here were similar to those in the German Bight and North Sea (Mai et
21 al., 2013), but were generally much lower than in Canada (Hayward et al., 2010; Yao et al.,
22 2008), USA (Majewski et al., 2014; Peck and Hornbuckle, 2005) and France (Coscollà et al.,
23 2013c, 2011; Sauret et al., 2008; Scheyer et al., 2008; Schummer et al., 2010).

24 3.3 Seasonal variations at the background site

25 Concentrations of \sum OCPs and \sum CUPs were lowest in January-February and highest in
26 August-September for \sum OCPs and in April-May for \sum CUPs. Individual CUPs and OCPs with
27 consistent detection ($>25\%$ of samples) were generally grouped according to their seasonal
28 trends (Fig. 1). The first group (group A, Fig. 1a) comprises compounds with one growing
29 season concentration peak (April-September). The second group (group B, Fig. 1b) comprises
30 compounds with two peaks, one during the growing season and the second in the plowing
31 season (October-November).

1 Acetochlor, fenpropimorph, S-metolachlor and terbuthylazine are in group A and had
2 maximum concentrations in the April-July period. Dimethachlor and metazachlor are also
3 included in this group but had later peaks, during August-September. These two compounds
4 are used for oil plants and are usually applied later in the summer for weed control of winter
5 grains; this may explain their later maximum concentrations, as has been previously reported
6 for metazachlor (Mai et al., 2013). The peak in concentrations of CUPs in this group is likely
7 associated with the fresh application of pesticides, but also with a contribution from
8 volatilization from soils, plants and surface water at higher temperatures. However, in the case
9 of acetochlor, fenpropimorph and S-metolachlor, which had maximum concentrations during
10 April-May, their total concentrations seemed predominantly influenced by agricultural
11 activity rather than volatilization, as the timing of the peak corresponded with the application
12 season (April-May) rather than with the highest summer temperatures (July-August). A
13 similar pattern of high concentrations during the growing season has been previously reported
14 for acetochlor, alachlor, dimethoate and terbuthylazine (Hayward et al., 2010; Mai et al.,
15 2013; Peck and Hornbuckle, 2005).

16 Group B comprises chlorpyrifos, isoproturon, prochloraz, chlorotoluron, diuron, and likely
17 metribuzin, although this is less conclusive due to more limited detection. The first group B
18 peak is attributed to the same factors as described for group A. The off-season (second peak)
19 concentrations are attributed to direct application of pesticides for future cereal crops which
20 usually take place during autumn (Garthwaite et al., 2014). Moreover, volatilization from pre-
21 treated seeds, plants, soils and water and wind erosion facilitated by the plowing of fields,
22 which usually take place during this period, may also contribute to the second peak. In the
23 case of soil volatilization, these compounds, except for isoproturon and metribuzin, are
24 moderately persistent in the soil (Table 1; half-life in soil >45 days) and thus, once they have
25 entered the soil from application or deposition, higher soil concentrations may persist unless
26 anthropogenic soil activity such as plowing occurs. However, terbuthylazine also has
27 moderate persistence in soil and did not have an autumn peak. It is notable that the peak
28 concentrations of chlorpyrifos, isoproturon and chlorotoluron were generally higher (up to
29 4.15 times) in autumn compared to the growing season, suggesting that, for these compounds,
30 autumn emissions are a larger source than emissions during the growing season. October-
31 November peaks of chlorpyrifos have been previously reported in China (Li et al., 2014) and
32 Canada (Hayward et al., 2010).

1 Carbendazim, a fungicide used mostly for oil plants, had a single growing season peak in
2 2012 (in April-June) and two peaks in 2013 (one in May-June and one in September-
3 October). Additionally, this compound had a relative high concentration (12.1 $\text{pg}\cdot\text{m}^{-3}$) during
4 the last sampled week (18-25/12/13). It is unclear what caused these differences between the
5 two study years.

6 Of the OCPs, β -HCH, γ -HCH and *o,p'*-DDD followed the group A seasonal trend, with one
7 peak occurring between May-August. *p,p'*-DDE, *o,p'*-DDT and *p,p'*-DDT behaved as per
8 group B, with two peaks each year. Other OCPs did not have clear seasonal variations. In
9 general, the seasonal trends observed for OCPs were much less pronounced than for CUPs
10 (Fig. S2). For example, the ratio of summer-to-winter concentrations of OCPs ranged from
11 0.758 (*p,p'*-DDD) to 6.54 (*p,p'*-DDT) with an average value of 2.90, while for CUPs, it
12 ranged from 0.188 (diuron) to 167 (metazachlor) with an average value of 28.4.

13 The seasonal variability in pesticides is related to and indicative of the sources of the
14 pesticide. The major cause of the seasonal variability in OCPs is expected to be seasonality in
15 volatilization from soils and other surfaces, thus seasonal variability should be related to
16 temperature variability. Conversely, when seasonality is driven by use/application, as for the
17 CUPs, the relationship with temperature should be weaker and the summer/winter ratios
18 should be greater.

19 An examination of the temperature dependence using the Clausius-Clapeyron equation (see
20 SI) supported this hypothesis. The gaseous pesticide concentrations were expressed as linear
21 regressions of the natural logarithm of partial pressure versus the inverse of temperature (Hoff
22 et al., 1998):

$$23 \quad \ln P = \frac{m}{T} + b \quad (\text{Eq. 1})$$

24 where *m* and *b* are the slope and the intercept of the linear regression, respectively. Partial
25 pressures of individual compounds were calculated for each sample using gas phase
26 concentrations and the ideal gas law.

27 The temperature-dependence of gas-phase concentrations was statistically significant at the
28 99% confidence level for all OCPs except α -HCH, with slopes ranging from -2792 (δ -HCH)
29 to -9802 (*p,p'*-DDT), indicating that OCP concentrations increased with air temperature
30 (Table S17). Generally, a steep slope and high r^2 indicate that temperature-controlled air-

1 surface cycling and short-term transport influenced the ambient gas phase concentrations
2 (Hoff et al., 1998; Wania and Haugen, 1998), while a shallow slope and low r^2 suggest that
3 other factors (i.e., advection, primary sources, atmospheric deposition, degradation) and
4 LRAT influenced concentrations (Lee et al., 2000). Thus, the Clausius-Clapeyron
5 relationships suggest that gas phase concentrations of all OCPs except α -HCH were
6 controlled by re-volatilization from surfaces close to the sampling site. Temperature
7 accounted for 23-84% of the variability in atmospheric concentrations for these compounds.
8 This is in agreement with a previous study showing that atmospheric levels of OCPs at
9 different European background sites were controlled by air-soil exchange (Cabrerizo et al.,
10 2011). The lower temperature dependence of α -HCH suggested that air concentrations were
11 also influenced by LRAT or other confounding factors. For the CUPs which were sufficiently
12 detected in the gas phase, only terbuthylazine and S-metolachlor had a significant temperature
13 dependency (Table S18). For some CUPs, their atmospheric lifetime in relation with OH
14 reaction is relatively small (e.g. about 2 hours for chlorpyrifos (Muñoz et al., 2014)), which
15 may explain the lack of maximum concentrations observed during the warmest periods. These
16 results emphasize the difference in the sources of OCPs and CUPs, with the former being
17 influenced by volatilization while the latter are influenced by temperature-independent local
18 sources (notably pesticide application) or LRAT.

19 3.4 Gas-particle partitioning at the background site

20 It is well known that several sampling artifacts such as blow-on, blow-off, breakthrough and
21 degradation may occur and affect the results about gas-particle partitioning (Melymuk et al.,
22 2014). The reported gas-particle partitioning of pesticides are therefore operationally defined,
23 given the sampling configuration, where gas-phase is defined as the mass of the sample
24 captured on the PUF and particulate-phase is the mass captured on the QFF. Given the large
25 volumes used in this study, breakthrough tests were performed (Table S7) and HCHs were
26 excluded from the discussion of gas-particle partitioning to avoid any bias due to gas-phase
27 breakthrough sampling. Breakthrough is typically the most significant sampling artifact; bias
28 due to filter blow-on/blow-off is not expected to be significant (Melymuk et al., in press).

29 In this study, the pesticides fall into three groups: (1) predominantly particulate phase, (2)
30 predominantly gas phase, and (3) those with significant gas and particulate phase fractions
31 (average measured particulate mass fraction, θ_{meas} , $0.2 < \theta_{\text{meas}} < 0.8$). Six CUPs (carbendazim,
32 chlorotoluron, diuron, fenpropimorph, isoproturon and prochloraz) were predominantly in the

1 particulate phase ($\theta_{\text{meas}} > 0.84$). In particular, prochloraz, diuron and carbendazim (except in
2 one sample in June 2012) were detected only in the particulate phase. A similar dominance of
3 the particulate phase has been reported for carbendazim (Mai et al., 2013) and fenpropimorph
4 (Van Dijk and Guicherit, 1999), but diuron was reported to have an average θ of 0.75
5 (Scheyer et al., 2008), which differs slightly from our results. Three CUPs (chlorpyrifos,
6 acetochlor and dimethachlor) and all the OCPs were predominantly found in the gas phase
7 (average $\theta_{\text{meas}} < 0.20$). In particular, the average particulate-phase mass fractions of
8 chlorpyrifos, *o,p'*-DDE, *p,p'*-DDE and *o,p'*-DDT were < 0.04 (Table 1). Of the OCPs, only
9 *p,p'*-DDD and *p,p'*-DDT had particulate phase fractions > 0.10 . The dominance of the gas
10 phase for chlorpyrifos (Li et al., 2014; Sadiki and Poissant, 2008; Van Dijk and Guicherit,
11 1999) and OCPs (Cindoruk, 2011; Sadiki and Poissant, 2008; Sanusi et al., 1999) is well
12 documented. Finally, four CUPs (atrazine, metazachlor, S-metolachlor and terbuthylazine)
13 were distributed between gas and particulate phases, with average θ_{meas} of 0.63, 0.59, 0.24 and
14 0.45, respectively.

15 Significant correlations ($0.20 < r^2 < 0.94$ and $p < 0.05$) between air temperatures and the gas-
16 particle partitioning coefficient (K_p , in $\text{m}^3 \mu\text{g}^{-1}$, see SI for details) were observed for all OCPs,
17 with higher particulate fractions associated with lower temperatures. Amongst the CUPs, the
18 measured K_p of S-metolachlor and terbuthylazine also correlated with air temperatures
19 ($r^2 = 0.29$ and 0.28 , respectively and $p < 0.05$). The lack of observed relationships in the case of
20 other CUPs suggests that the gas-particle partitioning of the majority of the CUPs is
21 determined by processes which are not or minimally sensitive to temperature.

22 To better understand the mechanisms influencing gas-particle partitioning of pesticides, K_p
23 was compared with K_{oa} and with the soil-air partitioning coefficient (K_{sa} , dimensionless) for
24 four CUPs (chlorpyrifos, isoproturon, metazachlor and terbuthylazine) and three OCPs (*p,p'*-
25 DDD, *p,p'*-DDE and *p,p'*-DDT) (Fig. 2). The temperature-dependency of K_{oa} (Table S19 and
26 Fig. S3) was determined from published relationships (for all OCPs and chlorpyrifos) or from
27 extrapolation (remaining CUPs) based on regression analysis for other compounds and
28 validated for chlorpyrifos (Fig. S4). Details of the calculations can be found in the SI. K_{sa} data
29 used in this study were adopted from the relationship of Davie-Martin et al. (2015), based on
30 laboratory experiments on 22 OCPs and CUPs, as:

$$31 \log K_{\text{sa}} = -26.2 + 0.714 \log K_{\text{oa}, 298.15\text{K}} + \frac{8291}{T_{\text{amb}}} - 0.0128RH + 0.121 \log(100f_{\text{oc}})$$

(Eq. 2)

where, T_{amb} is the ambient temperature (K), RH is the relative humidity (%) and f_{OC} is the organic carbon content of soil. Average monthly RH values and an experimental f_{oc} of 0.03 (Holoubek et al., 2009) were used.

Both K_{oa} and K_{sa} were significantly ($p < 0.05$) correlated with K_p for both OCPs and CUPs ($r^2 = 0.51-0.73$; Fig. 2). However, clear differences were noted between these two classes of compounds. For same K_{oa} (or K_{sa}), the K_p values of CUPs were notably higher than those of the OCPs. This suggests that absorption into organic matter alone, described by K_{oa} , is not sufficient to explain the observed gas-particle partitioning of CUPs and that other types of interactions occur. In other words, while absorption into the organic matter fraction is the dominant process for weakly or non-polar compounds such as OCPs, additionally adsorption to mineral surfaces or soot is significant for more polar compounds such as the CUPs. Indeed, Götz et al., (2007) estimated that the contribution of OM to K_p was 74% for DDT but only 1 and 5% for isoproturon and terbuthylazine, respectively, for which adsorption to mineral surfaces dominated K_p (contributions of 95 and 86%, respectively) and concluded that a predictive model based only on absorptive contribution to organic matter is not recommended for polar compounds such as CUPs. Therefore, assuming absorption to govern sorption, such as in the K_{oa} model (Harner and Bidleman; 1998), will generally lead to high agreement of predicted K_p values with observations for OCPs but low agreement (underestimates) for CUPs (Fig. S5-S6). Thus, a predictive model based only on absorptive contribution to organic matter is not recommended for polar compounds such as CUPs (Götz et al., 2007). However, we note that in Fig. 2a the two slopes intersect around $\log K_{oa}$ of 13. This suggests that for the few polar pesticides with $12 < \log K_{oa} < 13$, the K_{oa} -based approach is still appropriate.

Interestingly, the predicted method derived for soil-air partitioning which takes into account both absorption (K_{oa}) and adsorption (T, RH) (Davie-Martin et al., 2015) was a better predictor for gas-particle partitioning, given that similar slopes (0.857 ± 0.0332) were observed for OCPs and CUPs (Fig. 2b). The difference between the intercepts is 1.25. Based on this relationship, we propose an improved method for prediction of gas-particle partitioning of pesticides:

$$\log K_p = 0.857 \cdot \left(-26.2 + 0.714 \log K_{oa,298.15 K} + \frac{8291}{T_{amb}} - 0.0128RH + 0.121 \log(100f_{oc}) \right) - 10.5 + p$$

(Eq. 3)

1 with p being a factor accounting for polarity, $p = 1.25$ for CUPs and $p = 0$ for OCPs. By using
2 eq. 3, good agreements between predicted and measured $\log K_p$ of CUPs and OCPs are found
3 (rmse = 0.61, Fig. 3), with few exceptions for some CUPs.

4 The relationship between K_p and K_{sa} suggest similar partitioning from the gas phase to
5 aerosols and to soils including absorption to organic matter and adsorption to mineral
6 surfaces. In fact, the role of adsorption to mineral surfaces in air-soil exchange studies has
7 been rather neglected over the last decades (Mackay, 2001; Odabasi and Cetin, 2012a; Wang
8 et al., 2012; Wei et al., 2014) and should be further considered (Davie-Martin et al., 2015;
9 Goss et al., 2004).

10 The seasonal variation of the relationship of K_p with K_{oa} and K_{sa} was also examined (Fig. S7).
11 Given that many CUPs were mainly detected in spring and autumn, we choose these two
12 seasons (spring and autumn were defined from March to June and from September to
13 November, respectively). Interestingly, better correlations were obtained between K_p and K_{oa}
14 for CUPs in autumn compared to spring ($r^2 = 0.71$ and 0.49 , respectively, $p < 0.05$) while there
15 was no variation for OCPs ($r^2 = 0.63$). This suggests that a process other than absorption in
16 organic matter gains significance for CUPs in spring but less in autumn. This could be related
17 to a higher concentration (specific surface area) of mineral dust during the spring sampling
18 period (adsorption, see above). Pesticide application technique could potentially affect gas-
19 particle partitioning through mass transport kinetics limitations (non-equilibrium). Indeed,
20 $10\times$ higher particulate fractions were found for chlorpyrifos for two samples in spring 2012
21 ($\theta_{meas} = 0.19$ and 0.33), suggesting a potential influence of application or agricultural activities.
22 However, there is limited knowledge in this area. The differences in RH in spring and autumn
23 (73.1 vs. 83.5 %, respectively) may also have contributed to the observed differences, as
24 water layers and deliquescence may affect OM accessibility (higher in spring). Götz et al.,
25 (2007) estimated that the influence of absorption on K_p for polar pesticides was negligible for
26 RH between 40 and 80% while it was dominant for $RH > 80\%$ (contributing to 30-90%).
27 Similarly, Davie-Martin et al., (2015) found that RH had a negligible effect on K_{sa} for
28 $RH > 80\%$ for semi-arid soils. The influence of RH on K_p has generally not been considered in
29 predictive methods except with polyparameter linear free energy relationships (pp-LFER,
30 (Goss, 1997)), which considers all types of molecular interactions. Unfortunately, many input
31 parameters needed for this method are not available for CUPs or are associated with large
32 uncertainties (Davie-Martin et al., 2015; Götz et al., 2007). The new predictive K_p model

1 proposed in this study and based on easily accessible parameters allows consideration of both
2 meteorological variables (RH, T), compound-specific properties (K_{oa} , factor accounting for
3 the polarity, p) and soil composition (f_{oc}).

4

5 3.5 Particle size distribution at the urban and rural sites

6 The particle size distribution of CUPs and OCPs was determined at the rural (Telnice) and
7 urban (Kotlářská) site. Only the pesticides with significant particle fractions (average
8 $\theta_{meas} > 10\%$) and detection will be discussed in this section. However, because chlorpyrifos is
9 one of the most widely used insecticides in the world (Solomon et al., 2014), we also include
10 it in the further discussion. Amongst the pesticides with sufficient detection (Tables S20-S23),
11 nine pesticides had highest concentrations on particles $< 0.95 \mu\text{m}$, four pesticides had highest
12 concentrations in the $> 1.5 \mu\text{m}$ fraction and one pesticide showed no size distribution pattern.
13 The seasonal size distributions of fenpropimorph and isoproturon are shown in Fig. 4 as
14 representative of the pesticides dominated by the fine and coarse fractions, respectively.

15 Particulate phase concentrations of $\sum\text{CUPs}$ at the rural site ranged from 110 to 408 $\text{pg}\cdot\text{m}^{-3}$ and
16 were higher than at the urban site ($\sum\text{CUPs} = 30.3 - 112 \text{ pg}\cdot\text{m}^{-3}$). In contrast, similar
17 concentrations were observed for $\sum\text{OCPs}$ at the both rural (14.4 – 50.1 $\text{pg}\cdot\text{m}^{-3}$) and urban
18 (18.2 – 42.2 $\text{pg}\cdot\text{m}^{-3}$) sites. As suggested by the seasonal trends at the background site, this
19 indicates that current agricultural emissions are driving CUP concentrations, while OCPs are
20 the result of diffuse pollution and thus do not have a strong urban-rural gradient. In general,
21 seasonal variations of particulate OCPs and CUPs were similar to those observed at the
22 background site. However, in these samples, the second autumn peak was observed only for
23 diuron, isoproturon and chlorotoluron at lower concentrations than during the growing season.

24 One CUP (alachlor) had sporadic detection outside of the growing season and no clear trend
25 in particle size distributions at either site (Tables S20 and S22). Nine CUPs (acetochlor,
26 atrazine, chlorpyrifos, diuron, fenpropimorph, metazachlor, S-metolachlor, simazine and
27 terbufos) had higher concentrations on fine particles and were on average 35-76% associated
28 with particles $< 0.95 \mu\text{m}$. This distribution did not shift significantly when concentrations were
29 normalized by particle mass in each size fraction (Table S22). To the best of our knowledge,
30 only one study has reported the particle size distribution of CUPs (Coscollà et al., 2013b) and
31 this included acetochlor and fenpropimorph in common with our study, also found largely on

1 fine particles. Similarly, *p,p'*-DDD and *p,p'*-DDT also had highest concentrations on fine
2 particles (<0.95 μm), which accounted for 43-63% and 50-91% of the total particulate phase
3 mass, respectively. It is interesting to note that the size distribution of diuron, fenpropimorph
4 and *p,p'*-DDD and *p,p'*-DDT did not show any variation by season or site. The presence of
5 these compounds in the fine fraction (per air volume and per particle mass) is attributed to the
6 sorption of gas phase pesticides to fine particles due to their higher surface area and the
7 coagulation of ultrafine to fine particles (Coscollà et al., 2013b). Moreover, as the
8 mechanisms of wet and dry deposition are less efficient for removing particles in the 0.1-1 μm
9 and 0.05-2 μm size range respectively (Zhang and Vet, 2006), these compounds are expected
10 to have higher atmospheric residence times compared to compounds which are mostly present
11 on coarse particles.

12 Four pesticides (carbendazim, isoproturon, prochloraz and terbuthylazine) were found
13 predominantly on coarse particles (>3.0 μm) in all seasons at both sites. Indeed, when the
14 maximum total concentration occurred (i.e. in spring or summer), 45-70% of the total
15 particulate phase mass of these compounds was on particles >3.0 μm . Similar size
16 distributions were observed when the concentrations were normalized by mass (Table S22). In
17 general, coarse particles are the result of mechanical processes such as wind erosion of soil
18 particles and most of these pesticides are moderately persistent in the soil (DT_{50} =40-120 days)
19 and thus might be subject to wind erosion. The presence of pesticides on coarse particles
20 could also be related to the pesticide application technique, as it has been shown that the type
21 and amount of emissions during application (either drift or airborne residues) are strongly
22 related to the application technique, and independent of the physicochemical properties of the
23 compound applied (FOCUS, 2008). A very wide range of application techniques are used; for
24 example, prochloraz exists as an emulsifiable concentrate, while carbendazim, isoproturon
25 and terbuthylazine mostly exist as soluble concentrates, and chlorpyrifos can be applied as
26 either a soluble concentrate or as solid particles directly to soil (PPDB, 2013). The f_{om} , not
27 measured in this study, may influence observed particle distributions, particularly given that
28 fine particles may contain a higher carbonaceous fraction (Putaud et al., 2004). The lack of f_{om}
29 data is a limitation in understanding the particle size distributions, however, we note that
30 individual samples (therefore with the same f_{om} values) had some CUPs predominantly found
31 on coarse particles and others predominantly found on fine particles, suggesting that factors
32 other than f_{om} are controlling their particle size distribution. We hypothesize that differences
33 in type of application (emulsifiable vs. soluble concentrates, type of spray application,

1 application to plants vs. soil vs. seeds) may lead to differences in the particle size distribution
2 of pesticides, yet very little specific information is available on how particle size distribution
3 relates to application techniques.

4 Coarse particles have a shorter residence time in the atmosphere because they settle rapidly
5 and are efficiently removed by wet and dry deposition. Moreover, these particles are less
6 likely to penetrate deeply into the human respiratory system (Englert, 2004). Thus, should
7 these distributions apply on a wider scale, carbendazim, isoproturon, prochloraz and
8 terbuthylazine could be considered as pollutants with low risks of human inhalation exposure
9 (discarding the potential toxicity of individual substances) and LRAT potential. Additional
10 research on the link between pesticide application techniques and local/regional atmospheric
11 concentrations and distributions are needed in order to reduce inhalation exposure of
12 agricultural workers.

13

14 **4 Conclusions**

15 Although OCPs have been banned for agricultural use decades ago, this study highlights the
16 fact that they are still frequently detected in atmospheric samples at a background site in
17 Central Europe due to their persistence in environmental matrices. Presently, more than 270
18 plant protection products are registered for agricultural use in the Czech Republic (SRS,
19 2014) with limited knowledge on potential environmental and human risks. This study
20 improves knowledge of the characterization of atmospheric behavior of 27 CUPs,
21 representing about 24% of the national market and found three major differences than what is
22 observed for OCPs. Firstly, regarding their seasonal variations, atmospheric concentrations of
23 CUPs were largely driven by agricultural practices while secondary sources such as
24 volatilization from surfaces governed atmospheric concentrations of OCPs. Secondly, clear
25 differences were observed in gas-particle partitioning, with an influence of adsorption onto
26 mineral surfaces for CUPs while OCPs were mainly partitioning to aerosols through
27 absorption. Based on the recent work of Davie-Martin et al., (2015), a basic predictive method
28 for K_p is proposed for polar and non-polar pesticides, which relies on easily accessible
29 parameters. This method should be tested for other sampling sites and aerosol composition to
30 determine its broader utility for polar pesticides in cases when parameters needed for pp-
31 LFER are not available. Finally, while OCPs and the majority of CUPs were largely found on

1 fine particles, four CUPs (carbendazim, isoproturon, prochloraz and terbuthylazine) had
2 higher concentrations on coarse particles ($>3.0\ \mu\text{m}$) which may be caused by the pesticide
3 application technique. This finding is particularly important and should be further investigated
4 given that large particles results in lower risks from inhalation (regardless the toxicity of the
5 pesticide) and lower potential for long range atmospheric transport.

6

7 **Supporting Information**

8 Description of samples collected, analytical methods for CUPs and OCPs, usage of pesticides
9 in the Czech Republic, Clausius-Clapeyron plots, description of calculation for predicted
10 particulate fractions and atmospheric concentrations of individual CUPs and OCPs are
11 provided.

12

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22

23 **References**

- 24 Bidleman, T.F., Billings, W.N., Foreman, W.T., 1986. Vapor-particle partitioning of
25 semivolatile organic compounds: estimates from field collections. *Environ. Sci. Technol.*
26 20, 1038–43.
- 27 Bogdal, C., Scheringer, M., Abad, E., Abalos, M., van Bavel, B., Hagberg, J., Fiedler, H.,
28 2013. Worldwide distribution of persistent organic pollutants in air, including results of

- 1 air monitoring by passive air sampling in five continents. *TrAC Trends Anal. Chem.* 46,
2 150–161. doi:10.1016/j.trac.2012.05.011
- 3 Borrás, E., Sánchez, P., Muñoz, A., Tortajada-Genaro, L.A., 2011. Development of a gas
4 chromatography-mass spectrometry method for the determination of pesticides in
5 gaseous and particulate phases in the atmosphere. *Anal. Chim. Acta* 699, 57–65.
6 doi:10.1016/j.aca.2011.05.009
- 7 Cabrerizo, A., Dachs, J., Jones, K.C., Barceló, D., 2011. Soil-Air exchange controls on
8 background atmospheric concentrations of organochlorine pesticides. *Atmos. Chem.*
9 *Phys.* 11, 12799–12811. doi:10.5194/acp-11-12799-2011
- 10 Chevreuil, M., Garmouma, M., Teil, M.-J., Chesterikoff, A., 1996. Occurrence of
11 organochlorines (PCBs, pesticides) and herbicides (triazines, phenylureas) in the
12 atmosphere and in the fallout from urban and rural stations of the Paris area. *Sci. Total*
13 *Environ.* 182, 25–37. doi:10.1016/0048-9697(95)05032-9
- 14 Cindoruk, S.S., 2011. Atmospheric organochlorine pesticide (OCP) levels in a metropolitan
15 city in Turkey. *Chemosphere* 82, 78–87. doi:10.1016/j.chemosphere.2010.10.003
- 16 Coscollà, C., Castillo, M., Pastor, A., Yusa, V., 2011. Determination of 40 currently used
17 pesticides in airborne particulate matter (PM 10) by microwave-assisted extraction and
18 gas chromatography coupled to triple quadrupole mass spectrometry. *Anal. Chim. Acta*
19 693, 72–81. doi:10.1016/j.aca.2011.03.017
- 20 Coscollà, C., Hart, E., Pastor, A., Yusà, V., 2013a. LC-MS characterization of contemporary
21 pesticides in PM10 of Valencia Region, Spain. *Atmos. Environ.* 77, 394–403.
22 doi:http://dx.doi.org/10.1016/j.atmosenv.2013.05.022
- 23 Coscollà, C., Munoz, A., Borrás, E., Vera, T., Rodenas, M., Yusa, V., 2014. Particle size
24 distributions of currently used pesticides in ambient air of an agricultural Mediterranean
25 area. *Atmos. Environ.* 95, 29–35. doi:10.1016/j.atmosenv.2014.06.022
- 26 Coscollà, C., Yahyaoui, A., Colin, P., Robin, C., Martinon, L., Val, S., Baeza-Squiban, A.,
27 Mellouki, A., Yusà, V., 2013b. Particle size distributions of currently used pesticides in a
28 rural atmosphere of France. *Atmos. Environ.* 81, 32–38.
29 doi:10.1016/j.atmosenv.2013.08.057
- 30 Coscollà, C., Yahyaoui, A., Colin, P., Robin, C., Martinon, L., Val, S., Baeza-Squiban, A.,
31 Mellouki, A., Yusà, V., 2013c. Particle size distributions of currently used pesticides in a
32 rural atmosphere of France. *Atmos. Environ.* 81, 32–38.
33 doi:http://dx.doi.org/10.1016/j.atmosenv.2013.08.057
- 34 Cousins, I.T., Mackay, D., 2001. Gas-particle partitioning of organic compounds and its
35 interpretation using relative solubilities. *Environ. Sci. Technol.* 35, 643–647.
36 doi:10.1021/es001123m
- 37 Davie-Martin, C.L., Hageman, K.J., Chin, Y.-P., Rougé, V., Fujita, Y., 2015. Influence of
38 temperature, relative humidity, and soil properties on the soil-air partitioning of

- 1 semivolatile pesticides: Laboratory measurements and predictive models. *Environ. Sci.*
2 *Technol.* 49, 10431–10439. doi:10.1021/acs.est.5b02525
- 3 Degrendele, C., Okonski, K., Melymuk, L., Landlová, L., Kukučka, P., Čupr, P., Klánová, J.,
4 2014. Size specific distribution of the atmospheric particulate PCDD/Fs, dl-PCBs and
5 PAHs on a seasonal scale: Implications for cancer risks from inhalation. *Atmos. Environ.*
6 98, 410–416. doi:http://dx.doi.org/10.1016/j.atmosenv.2014.09.001
- 7 Dubus, I.G., Hollis, J.M., Brown, C.D., 2000. Pesticides in rainfall in Europe. *Environ. Pollut.*
8 110, 331–344. doi:10.1016/S0269-7491(99)00295-X
- 9 Eisenreich, S.J., Looney, B.B., Thornton, J.D., 1981. Airborne organic contaminants in the
10 Great Lakes ecosystem. *Environ. Sci. Technol.* 15, 30–38. doi:10.1021/es00083a002
- 11 Englert, N., 2004. Fine particles and human health--a review of epidemiological studies.
12 *Toxicol. Lett.* 149, 235–42. doi:10.1016/j.toxlet.2003.12.035
- 13 European Commission - Health & Consumer Protection Directorate-General, 2003. Review
14 report for the active substance atrazine [WWW Document].
- 15 FOCUS, 2008. Pesticides in Air: Considerations for Exposure Assessment. Rep. Focus Work.
16 Gr. Pestic. Air (FOCUS Air Group).
- 17 Garthwaite, D.G., Hudson, S., Barker, I., Parrish, G., Smith, L., Pietravalle, S., 2014.
18 Pesticide usage survey report 250. Arable crops in the United Kingdom 2012 (including
19 aerial applications 2012). Department for Environment, Food & Rural Affairs.
- 20 Gil, Y., Sinfort, C., 2005. Emission of pesticides to the air during sprayer application: A
21 bibliographic review. *Atmos. Environ.* 39, 5183–5193.
22 doi:http://dx.doi.org/10.1016/j.atmosenv.2005.05.019
- 23 Glotfelty, D.E., Leech, M.M., Jersey, J., Taylor, A.W., 1989. Volatilization and Wind Erosion
24 of Soil Surface Applied Atrazine ., *J. Agric. Food Chem.* 546–551.
- 25 Goss, K.-U., 1997. Conceptual Model for the Adsorption of Organic Compounds from the
26 Gas Phase to Liquid and Solid Surfaces. *Envir Sci Technol* 31, 3600–3605.
27 doi:10.1021/es970361n
- 28 Goss, K.U., Buschmann, J., Schwarzenbach, R., 2004. Adsorption of organic vapors to air-dry
29 soils: model predictions and experimental validation. *Environ. Sci. Technol.* 38, 3667–
30 3673.
- 31 Götz, C.W., Scheringer, M., MacLeod, M., Roth, C.M., Hungerbühler, K., 2007. Alternative
32 Approaches for Modeling Gas–Particle Partitioning of Semivolatile Organic Chemicals:
33 Model Development and Comparison. *Environ. Sci. Technol.* 41, 1272–1278.
34 doi:10.1021/es060583y
- 35 Götz, C.W., Scheringer, M., MacLeod, M., Wegmann, F., Hungerbühler, K., 2008. Regional
36 differences in gas–particle partitioning and deposition of semivolatile organic

- 1 compounds on a global scale. *Atmos. Environ.* 42, 554–567.
2 doi:<http://dx.doi.org/10.1016/j.atmosenv.2007.08.033>
- 3 Halse, a. K., Schlabach, M., Eckhardt, S., Sweetman, a., Jones, K.C., Breivik, K., 2011.
4 Spatial variability of POPs in European background air. *Atmos. Chem. Phys.* 11, 1549–
5 1564. doi:10.5194/acp-11-1549-2011
- 6 Hayward, S.J., Gouin, T., Wania, F., 2010. Levels and Seasonal Variability of Pesticides in the
7 Rural Atmosphere of Southern Ontario. *J. Agric. Food Chem.* 58, 1077–1084.
8 doi:10.1021/jf902898f
- 9 Hoff, R.M., Brice, K.A., Halsall, C.J., 1998. Nonlinearity in the slopes of Clausius-Clapeyron
10 plots for SVOCs. *Environ. Sci. Technol.* 32, 1793–1798.
- 11 Holoubek, I., Dušek, L., Sáňka, M., Hofman, J., Čupr, P., Jarkovský, J., Zbiral, J., Klánová, J.,
12 2009. Soil burdens of persistent organic pollutants - Their levels, fate and risk. Part I.
13 Variation of concentration ranges according to different soil uses and locations. *Environ.*
14 *Pollut.* 157, 3207–3217. doi:10.1016/j.envpol.2009.05.031
- 15 Holoubek, I., Klánová, J., Jarkovský, J., Kohoutek, J., 2007. Trends in background levels of
16 persistent organic pollutants at Kosetice observatory, Czech Republic. Part I. Ambient air
17 and wet deposition 1996-2005. *J. Environ. Monit.* 9, 557–63.
- 18 Kannan, K., Ridal, J., Struger, J., 2006. Pesticides in the Great Lakes, in: Hites, R. (Ed.),
19 Persistent Organic Pollutants in the Great Lakes, The Handbook of Environmental
20 Chemistry. Springer Berlin Heidelberg, pp. 151–199. doi:10.1007/698_5_041
- 21 Koblizková, M., Lee, S.C., Harner, T., 2012. Sorbent impregnated polyurethane foam disk
22 passive air samplers for investigating current-use pesticides at the global scale. *Atmos.*
23 *Pollut. Res.* 3, 456–462.
- 24 Lammel, G., Klánová, J., Kohoutek, J., Prokeš, R., Ries, L., Stohl, A., 2009. Observation and
25 origin of organochlorine compounds and polycyclic aromatic hydrocarbons in the free
26 troposphere over central Europe. *Environ. Pollut.* 157, 3264–3271.
27 doi:10.1016/j.envpol.2009.05.028
- 28 Lee, R.G.M., Burnett, V., Harner, T., Jones, K.C., 2000. Short-term temperature-dependent
29 air-surface exchange and atmospheric concentrations of polychlorinated naphthalenes
30 and organochlorine pesticides. *Environ. Sci. Technol.* 34, 393–398.
31 doi:10.1021/es9907414
- 32 Li, H., Ma, H., Lydy, M.J., You, J., 2014. Occurrence, seasonal variation and inhalation
33 exposure of atmospheric organophosphate and pyrethroid pesticides in an urban
34 community in South China. *Chemosphere* 95, 363–369.
35 doi:<http://dx.doi.org/10.1016/j.chemosphere.2013.09.046>
- 36 Li, J., Zhang, G., Guo, L., Xu, W., Li, X., Lee, C.S.L., Ding, A., Wang, T., 2007.
37 Organochlorine pesticides in the atmosphere of Guangzhou and Hong Kong: Regional
38 sources and long-range atmospheric transport. *Atmos. Environ.* 41, 3889–3903.
39 doi:10.1016/j.atmosenv.2006.12.052

- 1 Li, Y.F., Macdonald, R.W., 2005. Sources and pathways of selected organochlorine pesticides
2 to the Arctic and the effect of pathway divergence on HCH trends in biota: a review. *Sci.*
3 *Total Environ.* 342, 87–106. doi:<http://dx.doi.org/10.1016/j.scitotenv.2004.12.027>
- 4 Lohmann, R., Klanova, J., Kukucka, P., Yonis, S., Bollinger, K., 2012. PCBs and OCPs on a
5 East-to-West Transect: The Importance of Major Currents and Net Volatilization for
6 PCBs in the Atlantic Ocean. *Environ. Sci. Technol.* 46, 10471–10479.
7 doi:10.1021/es203459e
- 8 Lohmann, R., Lammel, G., 2004. Adsorptive and absorptive contributions to the gas-particle
9 partitioning of polycyclic aromatic hydrocarbons: State of knowledge and recommended
10 parametrization for modeling. *Environ. Sci. Technol.* 38, 3793–3803.
11 doi:10.1021/es035337q
- 12 Mackay, D., 2001. *Multimedia Environmental Models The Fugacity Approach Second*
13 *Edition, Second Edi.* ed. Lewis.
- 14 Mai, C., Theobald, N., Lammel, G., Hühnerfuss, H., 2013. Spatial, seasonal and vertical
15 distributions of currently-used pesticides in the marine boundary layer of the North Sea.
16 *Atmos. Environ.* 75, 92–102. doi:<http://dx.doi.org/10.1016/j.atmosenv.2013.04.027>
- 17 Majewski, M.S., Coupe, R.H., Foreman, W.T., Capel, P.D., 2014. Pesticides in Mississippi air
18 and rain: A comparison between 1995 and 2007. *Environ. Toxicol. Chem.* 33, 1283–
19 1293. doi:10.1002/etc.2550
- 20 Melymuk, L., Bohlin, P., Sářka, O., Pozo, K., Klánová, J., 2014. Current challenges in air
21 sampling of semi-volatile organic contaminants: sampling artifacts and their influence on
22 data comparability. *Environ. Sci. Technol.* doi:10.1021/es502164r
- 23 Melymuk, L., Bohlin-Nizzetto, P., Prokeš, R., Kukučka, P., Klánová, J., n.d. Sampling
24 artifacts in active air sampling of semivolatile organic contaminants: Comparing
25 theoretical and measured artifacts and evaluating implications for monitoring networks.
26 *Environ. Pollut.* doi:10.1016/j.envpol.2015.12.015
- 27 Muñoz, A., Ródenas, M., Borrás, E., Vázquez, M., Vera, T., 2014. The gas-phase degradation
28 of chlorpyrifos and chlorpyrifos-oxon towards OH radical under atmospheric conditions.
29 *Chemosphere* 111, 522–528. doi:10.1016/j.chemosphere.2014.04.087
- 30 Odabasi, M., Cetin, B., 2012a. Determination of octanol-air partition coefficients of
31 organochlorine pesticides (OCPs) as a function of temperature: Application to air-soil
32 exchange. *J. Environ. Manage.* 113, 432–439. doi:10.1016/j.jenvman.2012.10.010
- 33 Odabasi, M., Cetin, B., 2012b. Determination of octanol-air partition coefficients of
34 organochlorine pesticides (OCPs) as a function of temperature: application to air-soil
35 exchange. *J. Environ. Manage.* 113, 432–9.
- 36 Okonski, K., Degrendele, C., Melymuk, L., Landlová, L., Kukučka, P., Vojta, Š., Jiri, K.,
37 Čupr, P., Klánová, J., 2014. Particle size distribution of halogenated flame retardants and
38 implications for atmospheric deposition and transport. *Env. Sci Technol* 48, 14426–
39 14434.

- 1 Pankow, J.F., 1987. Review and comparative analysis of the theories on partitioning between
2 the gas and aerosol particulate phases in the atmosphere. *Atmos. Environ.* 21, 2275–
3 2283. doi:[http://dx.doi.org/10.1016/0004-6981\(87\)90363-5](http://dx.doi.org/10.1016/0004-6981(87)90363-5)
- 4 Peck, A.M., Hornbuckle, K.C., 2005. Gas-Phase Concentrations of Current-Use Pesticides in
5 Iowa. *Environ. Sci. Technol.* 39, 2952–2959. doi:10.1021/es0486418
- 6 PPDB, 2013. The Pesticide Properties DataBase (PPDB) [WWW Document]. Dev. by Agric.
7 Environ. Res. Unit (AERU), Univ. Hertfordshire, 2006-2013.
- 8 Putaud, J.-P., Raes, F., Van Dingenen, R., Brüggemann, E., Facchini, M.C., Decesari, S.,
9 Fuzzi, S., Gehrig, R., Hüglin, C., Laj, P., Lorbeer, G., Maenhaut, W., Mihalopoulos, N.,
10 Müller, K., Querol, X., Rodriguez, S., Schneider, J., Spindler, G., Brink, H. ten, Tørseth,
11 K., Wiedensohler, A., 2004. A European aerosol phenomenology—2: chemical
12 characteristics of particulate matter at kerbside, urban, rural and background sites in
13 Europe. *Atmos. Environ.* 38, 2579–2595.
14 doi:<http://dx.doi.org/10.1016/j.atmosenv.2004.01.041>
- 15 Růžicková, P., Klánová, J., Cupr, P., Lammel, G., Holoubek, I., 2008. An assessment of air-
16 soil exchange of polychlorinated biphenyls and organochlorine pesticides across central
17 and southern Europe. *Environ. Sci. Technol.* 42, 179–85.
- 18 Sadiki, M., Poissant, L., 2008. Atmospheric concentrations and gas-particle partitions of
19 pesticides: Comparisons between measured and gas-particle partitioning models from
20 source and receptor sites. *Atmos. Environ.* 42, 8288–8299.
21 doi:<http://dx.doi.org/10.1016/j.atmosenv.2008.07.041>
- 22 Sanusi, A., Millet, M., Mirabel, P., Wortham, H., 2000. Comparison of atmospheric pesticide
23 concentrations measured at three sampling sites: local, regional and long-range transport.
24 *Sci. Total Environ.* 263, 263–277. doi:[http://dx.doi.org/10.1016/S0048-9697\(00\)00714-2](http://dx.doi.org/10.1016/S0048-9697(00)00714-2)
- 25 Sanusi, A., Millet, M., Mirabel, P., Wortham, H., 1999. Gas–particle partitioning of pesticides
26 in atmospheric samples. *Atmos. Environ.* 33, 4941–4951.
27 doi:[http://dx.doi.org/10.1016/S1352-2310\(99\)00275-7](http://dx.doi.org/10.1016/S1352-2310(99)00275-7)
- 28 Sauret, N., Wortham, H., Putaud, J.-P., Mirabel, P., 2008. Study of the effects of
29 environmental parameters on the gas/particle partitioning of current-use pesticides in
30 urban air. *Atmos. Environ.* 42, 544–553.
31 doi:<http://dx.doi.org/10.1016/j.atmosenv.2007.09.012>
- 32 Scheyer, A., Morville, S., Mirabel, P., Millet, M., 2008. Gas/particle partitioning of lindane
33 and current-used pesticides and their relationship with temperature in urban and rural air
34 in Alsace region (east of France). *Atmos. Environ.* 42, 7695–7705.
35 doi:<http://dx.doi.org/10.1016/j.atmosenv.2008.05.029>
- 36 Schummer, C., Mothiron, E., Appenzeller, B.M.R., Rizet, A.-L., Wennig, R., Millet, M., 2010.
37 Temporal variations of concentrations of currently used pesticides in the atmosphere of
38 Strasbourg, France. *Environ. Pollut.* 158, 576–584.
39 doi:<http://dx.doi.org/10.1016/j.envpol.2009.08.019>

- 1 Shen, L., Wania, F., Lei, Y.D., Teixeira, C., Muir, D.C.G., Bidleman, T.F., 2004. Atmospheric
2 distribution and long-range transport behavior of organochlorine pesticides in north
3 America. *Environ. Sci. Technol.* 39, 409–420. doi:10.1021/es049489c
- 4 Shoeib, M., Harner, T., 2002. Using measured octanol-air partition coefficients to explain
5 environmental partitioning of organochlorine pesticides. *Environ. Toxicol. Chem.* 21,
6 984–990. doi:10.1897/1551-5028(2002)021<0984:umoapc>2.0.co;2
- 7 Sofuoglu, A., Cetin, E., Bozacioglu, S.S., Sener, G.D., Odabasi, M., 2004. Short-term
8 variation in ambient concentrations and gas/particle partitioning of organochlorine
9 pesticides in Izmir, Turkey. *Atmos. Environ.* 38, 4483–4493.
10 doi:10.1016/j.atmosenv.2004.05.036
- 11 Solomon, K.R., Williams, W.M., Mackay, D., Purdy, J., Giddings, J., Giesy, J., 2014.
12 Properties and Uses of Chlorpyrifos in the United States, in: Giesy, J.P., Solomon, K.R.
13 (Eds.), *Ecological Risk Assessment for Chlorpyrifos in Terrestrial and Aquatic Systems*
14 *in the United States, Reviews of Environmental Contamination and Toxicology*. Springer
15 International Publishing, pp. 13–34. doi:10.1007/978-3-319-03865-0_2
- 16 SRS, 2014. State Phytosanitary Administration. Usage of active substances in 2013, Czech
17 Republic. http://eagri.cz/public/web/file/439536/celek_2013_EN.pdf.
- 18 SRS, 2013. State Phytosanitary Administration. Usage of active substances in 2012, Czech
19 Republic. http://eagri.cz/public/web/file/439598/celek_2012_EN.pdf.
- 20 U.S. Environmental Protection Agency, 2013. Atrazine Updates [WWW Document]. Pestic.
21 Reregistration.
- 22 University of Hertfordshire, 2013. The Pesticide Properties DataBase (PPDB) [WWW
23 Document]. Dev. by Agric. Environ. Res. Unit (AERU), Univ. Hertfordshire, 2006-2013.
- 24 US EPA, 2014. Estimation Programs Interface Suite™ for Microsoft® Windows, v 4.11.
25 United States Environmental Protection Agency, Washington, DC, USA.
- 26 Van den Berg, F., Kubiak, R., Benjey, W.G., Majewski, M.S., Yates, S.R., Reeves, G.L.,
27 Smelt, J.H., van der Linden, A.M.A., 1999. Emission of Pesticides into the Air. *Water*.
28 *Air. Soil Pollut.* 115, 195–218. doi:10.1023/A:1005234329622
- 29 Van Dijk, H.F.G., Guicherit, R., 1999. Atmospheric dispersion of current-use pesticides: A
30 review of the evidence from monitoring studies. *Water. Air. Soil Pollut.*
31 doi:10.1023/A:1005293020536
- 32 Vecchi, R., Marcazzan, G., Valli, G., 2007. A study on nighttime-daytime PM10 concentration
33 and elemental composition in relation to atmospheric dispersion in the urban area of
34 Milan (Italy). *Atmos. Environ.* 41, 2136–2144. doi:10.1016/j.atmosenv.2006.10.069
- 35 Wang, X.P., Sheng, J.J., Gong, P., Xue, Y.G., Yao, T.D., Jones, K.C., 2012. Persistent organic
36 pollutants in the Tibetan surface soil: Spatial distribution, air-soil exchange and
37 implications for global cycling. *Environ. Pollut.* 170, 145–151.
38 doi:10.1016/j.envpol.2012.06.012

- 1 Wania, F., Haugen, J., 1998. Temperature dependence of atmospheric concentrations of
2 semivolatile organic compounds. *Environ. Sci. ...* 1013–1021. doi:10.1021/es970856c
- 3 Wei, Y.L., Bao, L.J., Wu, C.C., He, Z.C., Zeng, E.Y., 2014. Association of soil polycyclic
4 aromatic hydrocarbon levels and anthropogenic impacts in a rapidly urbanizing region:
5 Spatial distribution, soil-air exchange and ecological risk. *Sci. Total Environ.* 473-474,
6 676–684. doi:10.1016/j.scitotenv.2013.12.106
- 7 Yadav, I.C., Devi, N.L., Syed, J.H., Cheng, Z., Li, J., Zhang, G., Jones, K.C., 2015. Current
8 status of persistent organic pesticides residues in air, water, and soil, and their possible
9 effect on neighboring countries: A comprehensive review of India. *Sci. Total Environ.*
10 511, 123–137. doi:10.1016/j.scitotenv.2014.12.041
- 11 Yao, Y., Harner, T., Blanchard, P., Tuduri, L., Waite, D., Poissant, L., Murphy, C., Belzer, W.,
12 Aulagnier, F., Sverko, E., 2008. Pesticides in the Atmosphere Across Canadian
13 Agricultural Regions. *Environ. Sci. Technol.* 42, 5931–5937. doi:10.1021/es800878r
- 14 Zhang, L., Vet, R., 2006. A review of current knowledge concerning size-dependent aerosol
15 removal. *China Particuology* 4, 272–282. doi:http://dx.doi.org/10.1016/S1672-
16 2515(07)60276-0
- 17 Zhang, N., Yang, Y., Liu, Y., Tao, S., 2009. Determination of octanol-air partition coefficients
18 and supercooled liquid vapor pressures of organochlorine pesticides. *J. Environ. Sci.*
19 *Heal. Part B* 44, 649–656.
- 20 Zhang, X.M., Meyer, T., Muir, D.C.G., Teixeira, C., Wang, X.W., Wania, F., 2013.
21 Atmospheric deposition of current use pesticides in the Arctic: Snow core records from
22 the Devon Island Ice Cap, Nunavut, Canada. *Environ. Sci. Impacts* 15, 2304–2311.
23 doi:10.1039/c3em00433c
- 24

1 **TABLE**

2

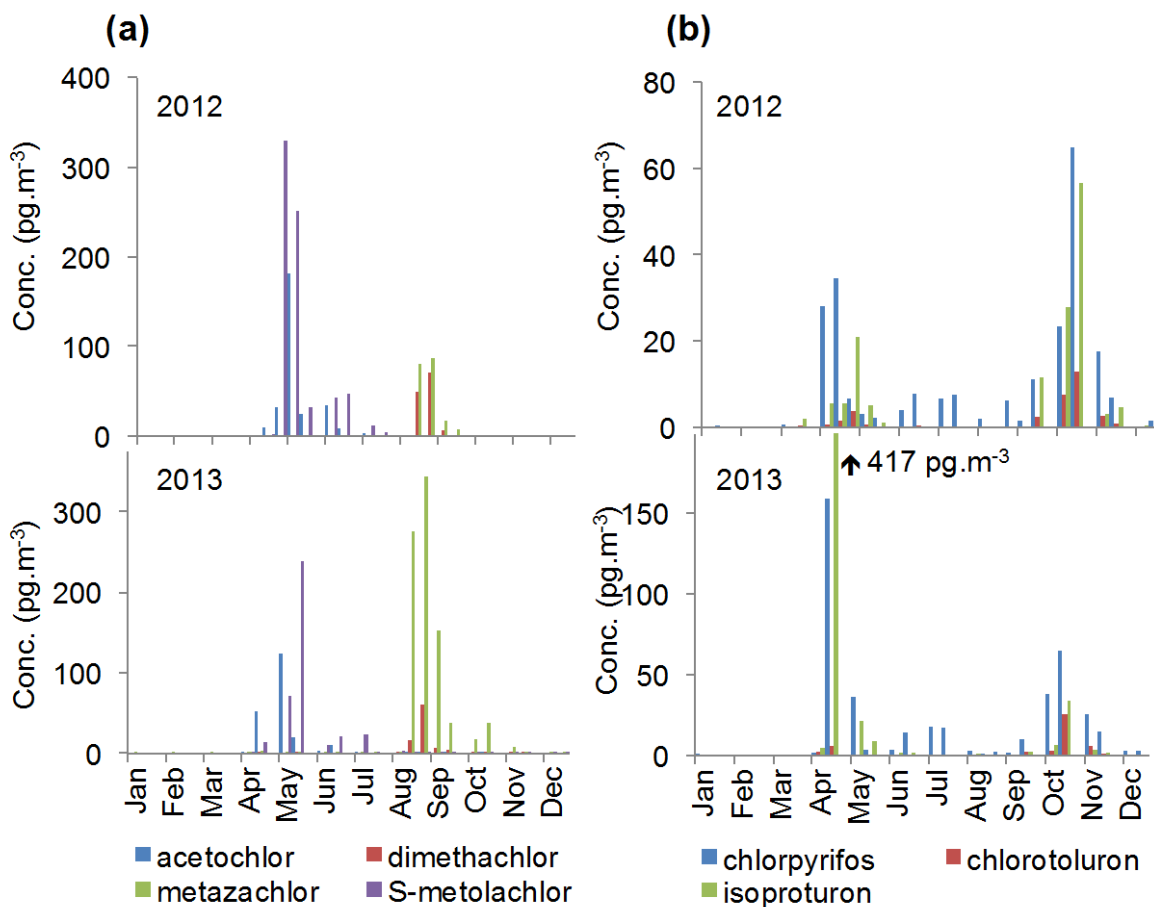
3 Table 1: Physicochemical properties and atmospheric concentrations (in $\text{pg}\cdot\text{m}^{-3}$) of individual OCPs and CUPs at background site. ND indicates “not detected”

Compound	Type of pesticide ^a		In use ^b	Half-life in soil (days) (PPDB, 2013)	Vapor pressure (Pa)	$\log K_{\text{oa}}$	Detection frequency (%)	Range of total concentrations ($\text{pg}\cdot\text{m}^{-3}$)	Range of gas phase concentrations ($\text{pg}\cdot\text{m}^{-3}$)	Range of particle phase concentrations ($\text{pg}\cdot\text{m}^{-3}$)	Average measured particulate fraction (θ_{meas})
Acetochlor	H	Y	14	2.20E-05 ^c	9.07 ^e	50.0	ND-181	ND-158	ND-23.2	0.14 ± 0.32	
Alachlor	H	N	14	2.90E-03 ^c	9.98 ^e	5.77	ND-0.82	ND-0.23	ND-0.82	0.85 ± 0.26	
Atrazine	H	N	75	3.90E-05 ^c	9.62 ^g	21.2	ND-1.24	ND-0.76	ND-0.49	0.63 ± 0.46	
Azinphos Methyl	I	N	10	5.00E-07 ^c	8.76 ^f	0.00	ND	ND	ND	ND	
Carbendazim	F	Y	40	9.00E-05 ^c	10.6 ^f	42.3	ND-12.5	ND-0.22	ND-12.5	0.98 ± 0.10	
Chlorotoluron	H	Y	45	5.00E-06 ^c	10.6 ^g	48.1	ND-25.1	ND-0.48	ND-24.7	0.95 ± 0.20	
Chlorpyrifos	I	Y	50	1.43E-03 ^c	8.41 ^h	84.6	ND-159	ND-158	ND-9.43	0.037 ± 0.064	
Diazinon	I	N	9.1	1.20E-02 ^c	9.14 ^e	1.92	ND-0.18	ND-0.18	ND	0.0	
Dimethachlor	H	Y	7.0	6.40E-04 ^c	9.34 ^d	40.4	ND-71.3	ND-70.8	ND-9.36	0.18 ± 0.37	
Dimethoate	I	Y	2.6	2.47E-04 ^c	9.15 ^f	3.85	ND-0.08	ND	ND-0.08	1.0 ± 0.00	
Disulfoton	I	N	30	7.20E-03 ^c	8.07 ^d	1.92	ND-2.22	ND	ND-2.22	1	
Diuron	H	N	76	1.15E-06 ^c	10.4 ^f	32.7	ND-1.23	ND	ND-1.23	1.0 ± 0.00	
Fenitrothion	I	N	2.7	6.76E-04 ^c	7.72 ^d	0.00	ND	ND	ND	ND	
Fenpropimorph	F	Y	35	3.90E-03 ^c	8.93 ^e	65.4	ND-73.8	ND-1.27	ND-73.8	0.91 ± 0.28	
Fonofos	I	N	99	2.70E-02 ^c	7.48 ^d	5.76	ND-8.03	ND	ND-8.03	1.0 ± 0.00	
Isoproturon	H	Y	12	5.50E-06 ^c	11.2 ^g	86.5	ND-413	ND-122	ND-291	0.84 ± 0.29	
Malathion	I	N	0.17	3.10E-03 ^c	9.06 ^e	3.85	ND-0.30	ND-0.30	ND-0.13	0.50 ± 0.70	
Metamitron	H	Y	30	7.44E-07 ^c	11.2 ^d	25.0	ND-16.5	ND-16.5	ND-6.41	0.23 ± 0.44	
Metazachlor	H	Y	8.6	9.30E-05 ^c	9.76 ^e	86.5	ND-344	ND-262	ND-275	0.59 ± 0.38	
Metribuzin	H	Y	11.5	1.21E-04 ^c	10.0 ^d	15.4	ND-5.46	ND-5.46	ND-1.83	0.22 ± 0.41	

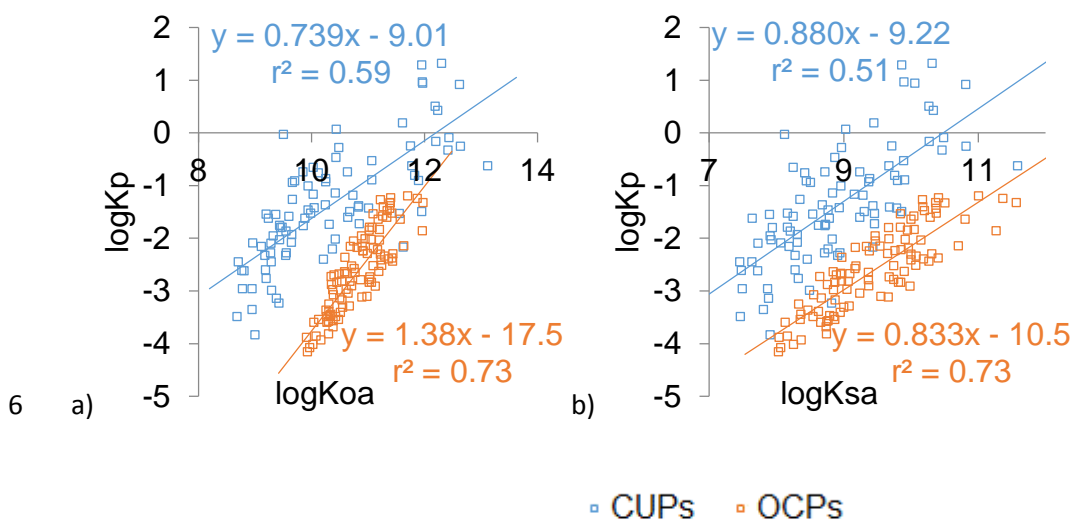
Prochloraz	F	Y	120	1.50E-04 ^c	13.6 ^d	55.8	ND-1.95	ND	ND-1.95	1.0 ± 0.00
Pyrazon	H	Y	31	1.00E-09 ^c	9.01 ^d	15.4	ND-2.25	ND-0.80	ND-2.25	0.91 ± 0.26
Simazine	H	N	60	8.10E-07 ^c	9.59 ^g	1.92	ND-0.087	ND	ND-0.087	1
S-metolachlor	H	Y	15	3.70E-03 ^c	9.33 ^d	73.1	ND-329	ND-309	ND-91.0	0.24 ± 0.34
Temephos	I	N	2	9.50E-06 ^c	13.1 ^d	5.77	ND-0.21	ND-0.21	ND-0.11	0.67 ± 0.58
Terbufos	I	N	8	3.46E-02 ^c	7.49 ^d	1.92	ND-0.80	ND	ND-0.61	1
Terbutylazine	H	Y	75.1	1.20E-04 ^c	9.03 ^f	78.8	ND-53.8	ND-33.8	ND-31.6	0.45 ± 0.35
ΣCUPs							ND-662	ND-365	ND-323	
<i>α</i> -HCH		N	175	3.44E-02 ^d	7.61 ⁱ	100	1.09-9.79	1.08-9.78	ND-0.031	<0.01
<i>β</i> -HCH		N		3.44E-02 ^d	8.88 ⁱ	69.2	ND-0.59	ND-0.59	ND-0.074	0.033 ± 0.051
<i>γ</i> -HCH	I	N		3.44E-02 ^d	7.85 ⁱ	100	0.488-21.8	0.470-21.8	ND-0.043	<0.01
<i>δ</i> -HCH		N		3.44E-02 ^d	8.84 ⁱ	57.7	ND-0.42	ND-0.42	ND-0.065	0.055 ± 0.097
<i>o,p'</i> -DDE		N		5.99E-03 ^d	9.26 ^j	96.2	ND-1.42	ND-1.42	ND-0.054	0.018 ± 0.071
<i>p,p'</i> -DDE		N		3.44E-03 ^d	9.68 ⁱ	100	1.14-71.4	0.612-71.4	ND-0.96	0.037 ± 0.074
<i>o,p'</i> -DDD		N		8.45E-04 ^d	9.57 ^j	73.1	ND-1.30	ND-1.28	ND-0.11	0.065 ± 0.16
<i>p,p'</i> -DDD		N		1.23E-03 ^d	10.1 ⁱ	75.0	ND-2.61	ND-2.56	ND-0.40	0.11 ± 0.18
<i>o,p'</i> -DDT	I	N	6200	1.68E-03 ^d	9.45 ⁱ	92.3	ND-9.18	ND-9.18	ND-0.11	0.033 ± 0.08
<i>p,p'</i> -DDT	I	N	6200	1.43E-04 ^d	9.82 ⁱ	100	0.414-9.99	0.13-9.99	ND-0.50	0.13 ± 0.19
ΣOCPs		N					4.51-122	2.87-122	ND-1.96	

1 ^a H: Herbicide, I: Insecticide and F: Fungicide. ^b Y: Authorized for agricultural use in Czech Republic during the sampling period and N: Not
2 authorized for agricultural purposes in Czech Republic during the sampling period. ^c (University of Hertfordshire, 2013). ^d (US EPA, 2014). ^e (Coscollà
3 et al., 2013b). ^f (Coscollà et al., 2013a). ^g (Götz et al., 2007). ^h (Odabasi and Cetin, 2012b). ⁱ (Shoeib and Harner, 2002). ^j (Zhang et al., 2009).

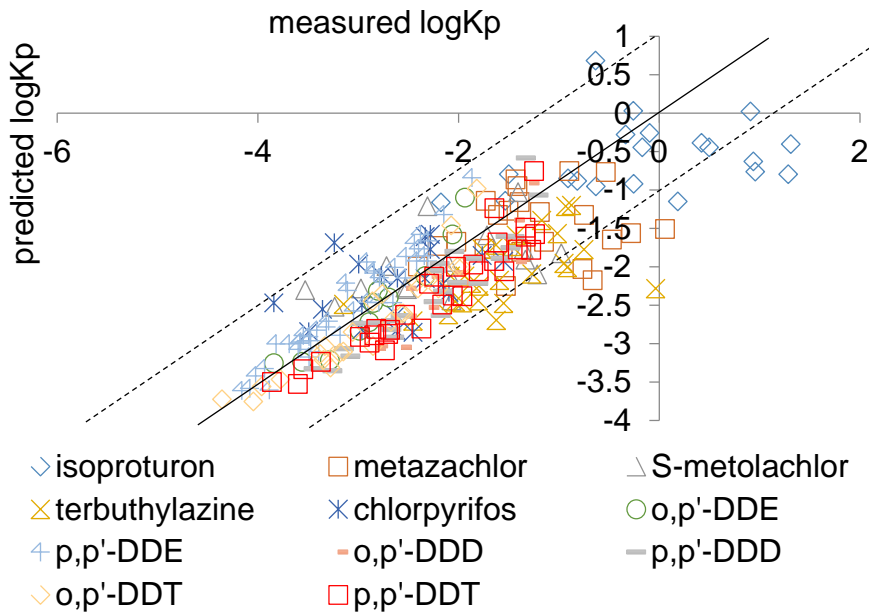
1 **FIGURES**



2
3 Figure 1: Seasonal variation of selected CUPs with (a) one peak per year during the growing season and (b) two
4 peaks per year, in April-July and October-November
5



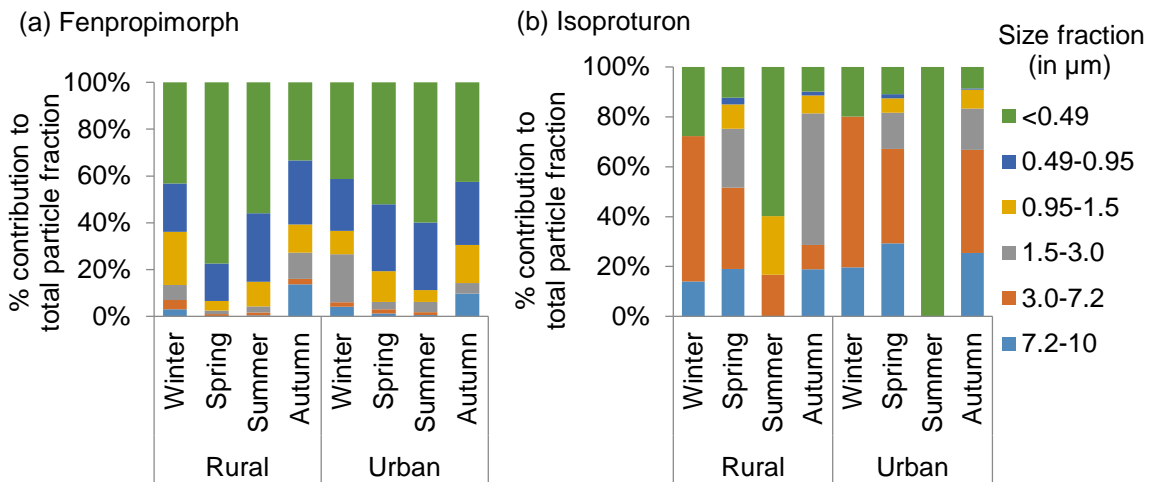
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7
8 Figure 2: Comparison of $\log K_p$ with $\log K_{oa}$ (a) and with $\log K_{sa}$ (b) for OCPs and CUPs



1

2 Figure 3: Comparison of predicted (see text, eq.3) and measured log K_p of individual OCPs and CUPs.

3



4

5 Figure 4: Seasonal particle size distribution of (a) fenpropimorph and (b) isoproturon at the rural and urban sites.
 6 Fenpropimorph represents the group of pesticides predominantly found on fine particles and isoproturon for the
 7 coarse particles

8 **CAPTIONS**

9

10 Table 2: Physicochemical properties and atmospheric concentrations (in $\text{pg}\cdot\text{m}^{-3}$) of individual
 11 OCPs and CUPs at background site. ND indicates “not detected”

12 Figure 1: Seasonal variation of selected CUPs with (a) one peak per year during the growing
 13 season and (b) two peaks per year, in April-July and October-November

- 1 Figure 2: Comparison of $\log K_p$ with $\log K_{oa}$ (a) and with $\log K_{sa}$ (b) for OCPs and CUPs
- 2 Figure 3: Comparison of predicted (see text, eq.3) and measured $\log K_p$ of individual OCPs
- 3 and CUPs.
- 4 Figure 4: Seasonal particle size distribution of (a) fenpropimorph and (b) isoproturon at the
- 5 rural and urban sites. Fenpropimorph represents the group of pesticides predominantly found
- 6 on fine particles and isoproturon for the coarse particles