

18 November, 2015
Prof. Willy Maenhaut,
Handling Editor of
Atmospheric Chemistry and Physics

Dear Prof. Willy Maenhaut,

We thank your suggestion. We have revised our paper entitled “Annual variation of carbonaceous PM_{2.5} in Malaysia: Influence by Indonesian peatland fires”, which we request you to consider for publication as an *Original Article* in *Atmospheric Chemistry and Physics*.

Our responses to your comments are as follows:

For the main text:

- 1) Page 2, line 22: Replace “gasses and “ by “gases and”.
We replaced “gasses and” by “gases and” (Page 2, line 22).
- 2) Page 4, line 18: Replace “organic compound” by “organic compounds”.
We replaced “organic compound” by “organic compounds” (Page 4, line 18).
- 3) Page 4, line 20: Replace “were quantified” by “was quantified”.
We replaced “were quantified” by “was quantified” (Page 4, line 20).
- 4) Page 5, line 20: Replace “filters range” by “filters ranged”.
We replaced “filters range” by “filters ranged” (Page 5, line 23).
- 5) Page 5, line 31: Replace “acquired on” by “acquired in”.
We replaced “acquired on” by “acquired in” (Page 5, line 31).
- 6) Page 6, line 1: Replace “in the Section” by “in Section”.
We replaced “in the Section” by “in Section” (Page 6, line 1).
- 7) Page 6, line 22: Replace “except August” by “except in August”.
We replaced “except August” by “except in August” (Page 6, line 22).

- 8) Page 6, line 28: Replace “season on September” by “season in September”.
We replaced “season on September” by “season in September” (Page 6, line 28).
- 9) Page 7, line 2: Replace “sites are dominantly downwind regions in” by “site is predominantly downwind of”.
We replaced “sites are dominantly downwind regions in” by “site is predominantly downwind of” (Page 7, line 2).
- 10) Page 7, line 9: Replace “annual concentrations” by “annual average concentrations”.
We replaced “annual concentrations” by “annual average concentrations” (Page 7, line 9).
- 11) Page 7, line 21: Replace “collected on” by “collected in”.
We replaced “collected on” by “collected in” (Page 7, line 21).
- 12) Page 7, line 25: Replace “annual concentrations” by “annual average concentrations”.
We replaced “annual concentrations” by “annual average concentrations” (Page 7, line 26).
- 13) Page 8, line 8: Replace “annual concentrations” by “annual average concentrations”.
We replaced “annual concentrations” by “annual average concentrations” (Page 8, line 9).
- 14) Page 8, line 15: Replace “and generally” by “and is generally”.
We replaced “and generally” by “and is generally” (Page 8, line 16).
- 15) Page 8, line 27: Replace “at IPF” by “at the IPF”.
We replaced “at IPF” by “at the IPF” (Page 8, line 28).
- 16) Page 9, line 26: Replace “annual concentration” by “annual average concentrations”.
We replaced “annual concentration” by “annual average concentrations” (Page 9, line 27).
- 17) Page 9, line 26: Replace “*n*-alkanes concentration” by “*n*-alkanes concentrations”.
We replaced “*n*-alkanes concentration” by “*n*-alkanes concentrations” (Page 9, line 27-28).
- 18) Page 9, line 27: Replace “season is” by “seasons are”.
We replaced “season is” by “seasons are” (Page 9, line 29).
- 19) Page 10, line 21: Replace “at IPF” by “at the IPF”.
We replaced “at IPF” by “at the IPF” (Page 10, line 22).

- 20) Page 10, line 22: Replace “identify IPFs” by “identify IPF”.
We replaced “identify IPFs” by “identify IPF” (Page 10, line 23-24).
- 21) Page 10, line 29: Replace “others should” by “other sources should”.
We replaced “others should” by “other sources should” (Page 10, line 30).
- 22) Page 11, line 25: Replace “galacotsan” by “galactosan”.
We replaced “galacotsan” by “galactosan” (Page 11, line 26).
- 23) Page 12, line 16: Replace “to biomass” by “to a biomass”.
We replaced “to biomass” by “to a biomass” (Page 12, line 18).
- 24) Page 12, line 21: Replace “The differences” by “Differences”.
We replaced “The differences” by “Differences” (Page 12, line 23).
- 25) Page 12, line 22: Replace “of PJ_A” by “of the PJ_A”.
We replaced “of PJ_A” by “of the PJ_A” (Page 12, line 24).
- 26) Page 12, line 23: Replace “to strong influence of IPFs” by “to the strong influence of the IPFs”.
We replaced “to strong influence of IPFs” by “to the strong influence of the IPFs” (Page 12, line 25).
- 27) Page 12, line 24: Replace “for PJ_A” by “for the PJ_A”.
We replaced “for PJ_A” by “for the PJ_A” (Page 12, lines 26-27).
- 28) Page 12, line 25: Replace “influence of IPFs” by “influence of the IPFs”.
We replaced “influence of IPFs” by “influence of the IPFs” (Page 12, line 27).
- 29) Page 13, line 21: Replace “partial overlap” by “partially overlap”.
We replaced “partial overlap” by “partially overlap” (Page 13, line 23).
- 30) Page 13, line 25: Replace “than VA/SA” by “than the VA/SA”.
We replaced “than VA/SA” by “than the VA/SA” (Page 13, line 27).
- 31) Page 14-19: Titles of journal articles should be in lower case instead of in Title Case (e.g., for Betha et al., 2014 and Chow et al., 2007).
We revised titles of journal articles according to the editor’s comment (Page 15-20).

- 32) Page 15, line 10: Replace “vol.62” by “vol. 62”.
We replaced “vol.62” by “vol. 62” (Page 16, line 10).
- 33) Page 17, line 16: Replace “A., Chesworth” by “A., and Chesworth”.
We replaced “A., Chesworth” by “A., and Chesworth” (Page 18, line 16).
- 34) Page 19, line 6: Replace “K. and Kunwar” by “K., and Kunwar”.
We replaced “K. and Kunwar” by “K., and Kunwar” (Page 20, line 6).
- 35) Page 20, line 6 from bottom: “*n*-alkanes” should be left-adjusted.
We revised titles of journal articles according to the editor’s comment (Page 21).

For the Supplemental Information:

- 36) Page 7, caption of Figure S3: Replace “of levoglucosan/mannosan” by “of the levoglucosan/mannosan”.
We replaced “of levoglucosan/mannosan” by “of the levoglucosan/mannosan” (Page 7).

[Others]

- 37) We replaced “to strong influence” by “to the strong influence” (Page 12, line 27).
- 38) We replaced “Fujii, Y., Mahmud, M., Tohno, S., Okuda, T., and Mizohata, A.: Characteristics of PM_{2.5} in Bangi, Selangor, Malaysia during the southwest monsoon season: case study, Aerosol Air Qual. Res., 2015c, accepted” by “Fujii, Y., Mahmud, M., Tohno, S., Okuda, T., and Mizohata, A.: A case study of PM_{2.5} characterization in Bangi, Selangor, Malaysia during the southwest monsoon season, Aerosol Air Qual. Res., 2015c, in press” (see Page 16, lines 21-23).

We look forward to hearing from you at your earliest convenience.

Sincerely,

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1 Annual variations of carbonaceous PM_{2.5} in Malaysia:

2 Influence by Indonesian peatland fires

3
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17 Abstract

18 In this study, we quantified carbonaceous PM_{2.5} in Malaysia through annual observations of
19 PM_{2.5}, focusing on organic compounds derived from biomass burning. We determined organic
20 carbon (OC), elemental carbon (EC) and concentrations of solvent-extractable organic
21 compounds (biomarkers derived from biomass burning sources and *n*-alkanes). We observed
22 seasonal variations in the concentrations of pyrolyzed OC (OP), levoglucosan (LG), mannosan
23 (MN), galactosan, syringaldehyde, vanillic acid (VA) and cholesterol. The average
24 concentrations of OP, LG, MN, galactosan, VA and cholesterol were higher during the
25 southwest monsoon season (June–September) than during the northeast monsoon season
26 (December–March), and these differences were statistically significant. Conversely, the
27 syringaldehyde concentration during the southwest monsoon season was lower. The PM_{2.5}
28 OP/OC₄ mass ratio allowed distinguishing the seven samples, which have been affected by the

1 Indonesian peatland fires (IPFs). In addition, we observed significant differences in the
2 concentrations between the IPF and other samples of many chemical species. Thus, the
3 chemical characteristics of PM_{2.5} in Malaysia appeared to be significantly influenced by IPFs
4 during the southwest monsoon season. Furthermore, we evaluated two indicators, the vanillic
5 acid/syringic acid (VA/SA) and LG/MN mass ratios, which have been suggested as indicators
6 of IPFs. The LG/MN mass ratio ranged from 14 to 22 in the IPF samples and from 11 to 31 in
7 the other samples. Thus, the respective variation ranges partially overlapped. Consequently,
8 this ratio did not satisfactorily reflect the effects of IPFs in Malaysia. In contrast, the VA/SA
9 mass ratio may serve as a good indicator, since it significantly differed between the IPF and
10 other samples. However, the OP/OC₄ mass ratio provided more remarkable differences than
11 the VA/SA mass ratio, offering an even better indicator. Finally, we extracted biomass burning
12 emissions' sources such as IPF, softwood/hardwood burning and meat cooking through
13 varimax-rotated principal component analysis.

14

15 **1 Introduction**

16 Peatland is a terrestrial wetland ecosystem where organic matter production exceeds its
17 decomposition, resulting in net accumulation (Page et al., 2006). Indonesia has the third largest
18 peatland area and the largest tropical peatland area in the world (270,000 km²; Joosten, 2010).
19 Peatland fires occur predominantly in the Sumatra and Kalimantan Islands, Indonesia (Fujii et
20 al., 2014; Page et al., 2002) during the dry season (June–September) mostly due to illegal
21 human activities (Harrison et al., 2009). Because peatland fires are usually underground fires,
22 they are extremely difficult to extinguish. The resulting haze comprises gasses and particulates
23 that are emitted because of biomass burning. It extends beyond Indonesia to the neighbouring
24 countries including Malaysia and Singapore (Betha et al., 2014; Engling et al., 2014; Fujii et
25 al., 2015b; He et al., 2010; See et al., 2006, 2007), limiting visibility and causing health
26 problems to the local population (Emmanuel, 2000; Othman et al., 2014; Pavagadhi, et al.,
27 2013; Sahani et al., 2014). Therefore, Indonesian peatland fires (IPFs) have been recognised as
28 an international problem (Yong and Peh, 2014; Varkkey, 2014).

29 The main constituent of particulates derived from biomass burning is PM_{2.5} defined as particles
30 having aerodynamic diameters below 2.5 µm, which has been associated with serious health
31 problems (Federal Register, 2006; Schlesinger, 2007). These particulates are primarily
32 composed of organic carbon (OC), which constitutes 50%–60% of the total particle mass (Reid

1 et al., 2005). At present, there are only four papers concerning the PM_{2.5} chemical speciation
2 resulting from IPFs; these papers are based on surface-recorded source-dominated data (Betha
3 et al., 2013; Fujii et al., 2014, 2015a; See et al., 2007). Organic matter is the main component
4 of PM_{2.5} from IPFs as well as from biomass burning in general (Fujii et al., 2014; See et al.,
5 2007). The primary organic compounds such as cellulose and lignin pyrolysis products have
6 been quantified and potential IPF indicators at the receptor site have been suggested by Fujii et
7 al. (2015a). Additional compounds have been discussed by Betha et al. (2013) (metals) and See
8 et al. (2007) (water-soluble ions, metals and polycyclic aromatic hydrocarbons).

9 Several studies exist on the chemical characteristics of haze ambient particulates, which have
10 been potentially affected by IPFs in Malaysia and Singapore (e.g., Abas et al., 2004a, b; Betha
11 et al., 2014; Engling et al., 2014; Fang et al., 1999; Fujii et al., 2015b; He et al., 2010; Keywood
12 et al., 2003; Narukawa et al., 1999; Okuda et al., 2002; See et al., 2006; Yang et al., 2013). In
13 most cases, the field observation periods were short. Even when long-term observations have
14 been obtained, however, only typical chemical species such as ions and metals have been
15 analysed. Nevertheless, organic compounds significantly contribute to the IPF aerosols (Fujii
16 et al., 2014). In Malaysia especially, there are no available quantitative data regarding variations
17 of several organic compound concentrations based on long-term observations of PM_{2.5}.

18 The three major sources of air pollution in Malaysia are mobile, stationary and open burning
19 sources including the burning of solid wastes and forest fires (Afroz, et al., 2003). The annual
20 burned biomass in Malaysia has been estimated to be 23 Tg on average (Streets et al., 2003).
21 Therefore, it is necessary to distinguish the effects of IPFs from those of other sources,
22 particularly local biomass burning. Fujii et al. (2015b) reported the total suspended particulate
23 matter (TSP) concentrations in the different carbon fractions (OC1, OC2, OC3, OC4 and
24 pyrolysed OC (OP)) defined by the IMPROVE_A protocol (Chow et al., 2007) in Malaysia
25 during the haze periods affected by IPFs. They proposed the OP/OC4 mass ratio as a useful
26 indicator of transboundary haze pollution from IPFs at receptor sites even in light haze; the ratio
27 during the haze periods were higher (>4) than during the non-haze periods (<2).

28 In the present study, the carbonaceous PM_{2.5} components are quantitatively characterised using
29 annual PM_{2.5} observations in Malaysia, with special regard to the organic compounds resulting
30 from biomass burning. Furthermore, the OP/OC4 mass ratio is used as an indicator to
31 investigate the effects of IPFs on carbonaceous PM_{2.5} species in this area. In addition, other
32 indicators that potentially record the effects of IPFs are investigated. Finally, possible

1 carbonaceous PM_{2.5} sources are suggested using varimax-rotated principal component analysis
2 (PCA).

3

4 **2 Experimental method**

5 **2.1 Sampling site and period**

6 The sampling site is the Malaysian Meteorological Department (MMD) located in Petaling Jaya
7 (PJ), Selangor, Malaysia (~100 m above sea level, 3° 06' 09" N, 101° 38' 41" E). Eighty-one
8 PM_{2.5} samples were collected on the roof of the MMD's main building (eight stories) from
9 August 2011 to July 2012. A detailed description of the sampling site has been provided by
10 Jamhari et al. (2014). In brief, PJ is located in an industrial area (Department of Environment,
11 2014) ~10 km from Kuala Lumpur. It is predominantly residential and industrial with high-
12 density road traffic.

13

14 **2.2 Sample collection and analysis**

15 PM_{2.5} samples were continuously collected with a Tisch high-volume air sampler (model TE-
16 3070V-2.5-BL) on a quartz-fibre filter for 24 h at a flow rate of 1.13 m³ min⁻¹. Before sampling,
17 the quartz-fibre filters were heated to 500 °C for 3 h. After sampling, OC, elemental carbon
18 (EC) and solvent-extractable organic compounds (SEOC; biomarkers derived from biomass
19 burning sources and *n*-alkanes) were measured.

20 The carbonaceous content ~~were~~was quantified using a DRI model 2001 OC/EC carbon analyser,
21 which employs the thermal optical-reflectance method following the IMPROVE_A protocol.
22 As shown in our former report (Fujii et al., 2014), the IMPROVE_A temperature protocol
23 defines temperature plateaus for thermally-derived carbon fractions as follows: 140 °C for OC1,
24 280 °C for OC2, 480 °C for OC3 and 580 °C for OC4 in helium (He) carrier gas; 580 °C for
25 EC1, 740 °C for EC2 and 840 °C for EC3 in a mixture of 98% He and 2% oxygen (O₂) carrier
26 gas. OC and EC are calculated from the eight carbon fractions as follows:

$$27 \quad \text{OC} = \text{OC1} + \text{OC2} + \text{OC3} + \text{OC4} + \text{OP}, \quad (1)$$

$$28 \quad \text{EC} = \text{EC1} + \text{EC2} + \text{EC3} - \text{OP}, \quad (2)$$

1 where OP is defined as the carbon content measured after the introduction of O₂ until
2 reflectance returns to its initial value at the start of analysis. Blank corrections were performed
3 on the OC and EC data by subtracting the blank filter value from the loaded filter values.

4 SEOC obtained from the quartz-fibre filters were quantified by gas chromatography mass
5 spectrometry (GC/MS). Biomarker organic compound speciation was accomplished following
6 the procedures reported previously (Fujii et al., 2015a, b). To quantify *n*-alkanes, aliquots from
7 the quartz-fibre filter were spiked with internal standards of eicosane-*d*₄₂ and triacontane-*d*₆₂
8 before extraction. Each spiked filter was extracted by ultrasonic agitation for 2 × 20 min periods
9 using 8 mL hexane (Kanto Chemical, purity >96.0%). The combined extracts were filtered
10 through a polytetrafluoroethylene syringe filter (pore size 0.45 μm), dried completely under a
11 gentle stream of nitrogen gas and re-dissolved to 0.1 mL in hexane. Before the GC/MS analysis,
12 ~1.05 μg of tetracosane-*d*₅₀ dissolved in 50 μL of hexane was added as a second internal
13 standard. The *n*-alkanes values were reported in carbon numbers, ranging from 22 to 33 (C₂₂–
14 C₃₃). The extract samples were analysed on a Shimadzu GC/MS system (GCMS-QP2010-Plus,
15 Shimadzu) equipped with a 30 m HP-5MS column (0.25 μm film thickness, 0.25 mm ID). The
16 carrier gas was helium (purity >99.9%) at a pressure of 73.0 kPa (37.2 cm s⁻¹ at 100 °C). The
17 GC oven temperature program was as follows: isothermal at 100 °C for 5 min, 100–300 °C at
18 10 °C min⁻¹ and then 300 °C for 20 min. The injection port and transfer line were maintained
19 at 300 °C. The data for quantitative analysis were acquired in the electron impact mode (70 eV).
20 The mass spectrometer was operated under the selected ion-monitoring scanning mode, and the
21 monitored ions for the quantification of *n*-alkanes were 85 *m/z*. The monitored ions
22 corresponding to the internal standards were 66 *m/z*. The recovery ratios for known amounts of
23 *n*-alkane standards (1 μg addition) on the quartz-fibre filters ranged from 73 to 110% (mean ±
24 standard deviation: 94 ± 6.3%). Blank corrections were performed on the biomarker and *n*-
25 alkane data by subtracting the blank filter value from the loaded filter values.

26

27 **2.3 Source apportionment method**

28 Varimax-rotated PCA was used to identify the possible carbonaceous PM_{2.5} sources at PJ. The
29 following two datasets were considered: (i) PJ_A data, which includes 25 variables (all
30 quantified compounds) and 81 samples (all samples), and (ii) PJ_S data, which includes 25
31 variables and 65 samples (excluded are the samples acquired ~~on~~in September 2011 and June

1 2012, which are influenced by IPFs as shown in ~~the~~ Section 3). PCA results with these datasets
2 are expected to show IPF effects on other sources. It has been suggested that the minimum
3 number of samples (n) for factor analysis should satisfy the following condition (Henry et al.,
4 1984; Karar and Gupta, 2007):

$$5 \quad n > 30 + \frac{V + 3}{2}, \quad (3)$$

6 where V represents the number of variables. Both datasets satisfy this condition.

7 Varimax-rotated PCA followed the procedure proposed by Karar and Gupta (2007) and was
8 accomplished with the R-software (<http://www.R-project.org>). The eigenvalues correspond to
9 the number of factors, which was selected to ensure that the cumulative variance contribution
10 rate is greater than 80%.

12 **3 Results and discussion**

13 **3.1 Air quality and monthly hotspot data**

14 Figure 1 presents the daily variability of the Malaysian Air Pollutant Index (MAPI) and
15 visibility during the sampling periods. The MAPI data were obtained from the Department of
16 Environment Ministry of Natural Resources and Environment website ([http://](http://apims.doe.gov.my/apims/hourly2.php)
17 apims.doe.gov.my/apims/hourly2.php). Hourly visibility data (7:00–17:00) provided by the
18 MMD were used to produce the daily variation in visibility after removing the hourly data
19 corresponding to periods of rainfall. The MAPI values of 0–50, 51–100, 101–200, 201–300 and
20 >300 correspond to good, moderate, unhealthy, very unhealthy and hazardous air quality
21 conditions (Department of Environment, 2014; Fujii et al., 2015b). Good MAPI levels dominate
22 the sampling periods except ~~in~~ August 2011, September 2011 and June 2012. On the other hand,
23 moderate air quality is observed in August 2011, September 2011 and June 2012. The two
24 MAPI values for 15 and 16 June 2012 indicate unhealthy air quality conditions. The average
25 visibility during these two sampling periods (Fig. 1) was below 2.7 km, corresponding to
26 extremely low visibility compared with other intervals.

27 Figure 2 presents the monthly hotspot counts in the Sumatra Island detected by the NOAA-18
28 satellite (Indofire). During the southwest monsoon season ~~in~~ September 2011 and June 2012,
29 hotspots exceeded 3,000 on several occasions. The hotspot counts in September 2011 and June

1 2012 mainly derived from the South Sumatra (60% of the hotspot counts) and the Riau (42%)
2 provinces, respectively. The sampling sites ~~are~~ is predominantly downwind ~~regions in~~of the
3 Sumatra Island during the southwest monsoon season. Thus, some samples have probably been
4 affected by IPFs. The three-day backward air trajectories for the sampling periods (Fig. S1)
5 support this conclusion.

6

7 **3.2 PM_{2.5} chemical characteristics and seasonal variations**

8 **3.2.1 OC and EC**

9 The annual average concentrations of OC and EC are 7.0 ± 5.4 and $3.1 \pm 1.1 \mu\text{gC m}^{-3}$,
10 respectively. The OC and EC concentrations' statistical results for each monsoon season appear
11 in Table 1. The average OC concentration during the southwest monsoon season (June–
12 September) is higher than that during other seasons. In particular, an extremely high OC
13 concentration ($>25 \mu\text{g m}^{-3}$) is observed on 12 September 2011 and on 15 and 16 June 2012.
14 There is no statistically significant difference in the EC concentration between the southwest
15 and northeast (December–March) monsoon seasons according to the two-sided Wilcoxon rank
16 sum test (p-value: $p > 0.05$) with R-software. In Bangi (~30 km southeast of the sampling site),
17 the OC concentration was $11 \pm 3.2 \mu\text{gC m}^{-3}$ in September 2013 (Fujii et al., 2015c), in good
18 agreement with the present results for the southwest monsoon season. The OC/EC mass ratios
19 during the southwest monsoon, post-monsoon (October–November), northeast monsoon and
20 pre-monsoon (April–May) season range among 1.2–6.5, 1.4–2.4, 0.99–3.0 and 1.2–2.3,
21 respectively. A high OC/EC mass ratio value (>4) is found only for some samples collected ~~on~~
22 in September 2011 and June 2012. These values have probably been affected by biomass
23 burning, because aerosols emitted from biomass burning usually present higher OC/EC mass
24 ratios (Cong et al., 2015).

25 The daily variations of the OC fractions' mass concentrations during the sampling periods are
26 presented in Fig. 3. The annual average concentrations of OC1, OC2, OC3, OC4 and OP are
27 0.51 ± 0.80 , 1.9 ± 1.1 , 2.3 ± 1.4 , 1.2 ± 0.36 and $1.1 \pm 2.2 \mu\text{g m}^{-3}$, respectively. Statistically
28 significant differences among the OP concentrations during the southwest and northeast
29 monsoon seasons are observed according to the two-sided Wilcoxon rank sum test ($p < 0.001$).
30 In particular, high OP concentrations are clearly observed in September 2011 and June 2012,
31 in addition to the higher OC/EC mass ratios described above. Fujii et al. (2015b) supported that

1 the enhanced OP concentrations in TSP, which are observed in Malaysia during the haze
2 periods, are affected by the IPFs. The enhanced OP concentrations in PM_{2.5} during the
3 southwest monsoon season, which are observed in the present study, are also probably affected
4 by IPFs from the Sumatra Island. The increased number of hotspots recorded (Fig. 2) and
5 backward air trajectories (Fig. S1) further support this conclusion.

6

7 **3.2.2 Biomarkers**

8 Ten biomarkers are identified in this study, which have been suggested as indicators of biomass
9 burning processes such as wood burning and meat cooking. The annual average concentrations
10 of levoglucosan (LG), mannosan (MN), galactosan, *p*-hydroxybenzoic acid, vanillic acid (VA)
11 and syringic acid (SA) are 86 ± 95 , 4.8 ± 5.7 , 1.2 ± 1.6 , 1.1 ± 1.3 , 0.19 ± 0.28 and 0.25 ± 0.28
12 ng m^{-3} , respectively; notably, they exhibit great variability. The annual concentrations of
13 vanillin, syringaldehyde, dehydroabietic acid and cholesterol are 1.2 ± 0.80 , 0.51 ± 0.42 , $1.3 \pm$
14 1.0 and $1.3 \pm 0.72 \text{ ng m}^{-3}$, respectively. The biomarker statistical results for each monsoon
15 season are listed in Table 1.

16 LG is a specific indicator for cellulose burning emissions and is generally formed during
17 cellulose pyrolysis at temperatures above 300 °C (Fujii et al., 2015b; Lin et al., 2010;
18 Shafizadeh, 1984; Simoneit et al., 1999). The MN and galactosan are derived from
19 hemicellulose pyrolysis products; they can also be used as tracers of biomass burning besides
20 LG (e.g., Engling et al., 2014; Fujii et al., 2014, 2015b; Zhu et al., 2015). Statistically significant
21 differences are observed among the concentrations of LG, MN and galactosan obtained during
22 the southwest and northeast monsoon seasons on the basis of the two-sided Wilcoxon rank sum
23 test ($p < 0.001$); high concentrations of these compounds are mostly observed during the
24 southwest monsoon season (especially September 2011 and June 2012; Fig. S2). In Singapore,
25 Engling et al. (2014) suggested that the enhanced concentrations of these compounds during
26 the haze periods were due to the IPFs during the southwest monsoon season. Thus, the presently
27 observed enhanced concentrations of these compounds may also be attributed to the IPFs.

28 In a previous report, PM_{2.5} lignin unit-originating compounds in samples collected at the IPF
29 source were quantified (Fujii et al., 2015a). Lignin is an aromatic polymer consisting of
30 phenylpropane units linked through many ether and C–C linkages. Its aromatic structure varies
31 depending on the species; softwood lignins exclusively contain guaiacyl (G) types, hardwood

1 lignins include both G and syringyl (S) types, whereas herbaceous plants include G, S and *p*-
2 hydroxyphenyl (H) types (Fujii et al., 2015a, b). The composition of these aromatic nuclei
3 within the lignin pyrolysis products resulting from biomass burning may be useful in identifying
4 the biomass type (Fujii et al., 2015a; Simoneit et al., 1993). In the present study, vanillin and
5 VA (compounds derived from G units), syringaldehyde and SA (compounds derived from S
6 units) as well as and *p*-hydroxybenzoic acid (compounds derived from H units or the secondary
7 decomposition of G and S units) (Fujii et al., 2015b) have been quantified. There are significant
8 differences between the concentrations of syringaldehyde and VA derived from lignin pyrolysis
9 during the southwest and northeast monsoon seasons on the basis of the two-sided Wilcoxon
10 rank sum test ($p < 0.001$), corresponding to seasonal variations. The average VA concentration
11 during the southwest monsoon season is 5.3 times greater than that during the northeast
12 monsoon season. In contrast, the average concentration of syringaldehyde during the northeast
13 monsoon season is 2.6 times greater than that during the southwest monsoon season. This may
14 be due to the transboundary pollution by prevailing winds from the Chinese region including
15 Thailand and Vietnam during the northeast monsoon season (Fig. S1; Khan et al., 2015).

16 Dehydroabietic acid and cholesterol are quantified as indicators of softwood burning and meat
17 cooking, respectively (Fujii et al., 2015b; Lin et al., 2010). The two-sided Wilcoxon rank sum
18 test indicates that the difference between the cholesterol concentration during the southwest and
19 northeast monsoon seasons is statistically significant ($p < 0.001$). The dehydroabietic acid and
20 cholesterol concentrations recorded in the interval between June and July 2014 in Bangi, which
21 is located ~30 km southeast of the sampling site, range between 2.6–8.7 and 1.5–5.7 ng m⁻³,
22 respectively (Fujii et al., 2015b). The PJ industrial area's concentrations of these compounds
23 are lower than those in the Bangi suburban area owing to the decreased impact of softwood
24 burning and meat cooking in PJ.

25

26 3.2.3 *N*-alkanes

27 The total annual average concentrations of *n*-alkanes is 79 ± 63 ng m⁻³. The total *n*-alkanes
28 concentrations during the southwest monsoon, post-monsoon, northeast monsoon and pre-
29 monsoon seasons ~~is~~ are 110 ± 93 , 57 ± 20 , 67 ± 18 and 55 ± 41 ng m⁻³, respectively. The highest
30 concentration is observed during the southwest monsoon season. Figure 4 illustrates the
31 molecular distribution of *n*-alkanes during the southwest and northeast monsoon seasons. There
32 are no significant differences among the concentrations of C₂₂–C₂₆, C₂₉, C₃₀ and C₃₂ in the two

1 seasons ($p > 0.05$). High concentrations of $>C_{24}$ are mainly observed in September 2011 and
2 June 2012 when many hotspots are detected in the Sumatra Island (Fig. 2). Fujii et al. (2015a)
3 suggested that IPFs increase the C_{27} , C_{28} and C_{29} concentrations in $PM_{2.5}$ at the receptor site
4 relative to other sources such as vehicle and biomass burning. Thus, the enhanced n -alkanes
5 concentrations in $PM_{2.5}$ during the southwest monsoon season may be mainly attributed to IPFs.

6 The carbon number maximum (C_{max}) in n -alkanes during the southwest and northeast monsoon
7 seasons is C_{27} (in 83% of the samples) and C_{26} (89%), respectively (Fig. 5). Reported C_{max}
8 values range from 27 to 33, characteristic of biogenic sources (higher plant-wax), whereas
9 lower C_{max} values may indicate major petrogenic input (Abas et al., 2004a; Gogou et al., 1996;
10 He et al., 2010). The C_{max} during the southwest monsoon season (C_{27}) suggests primarily
11 biogenic sources and is in perfect agreement with the measured value for the IPF source (Fujii
12 et al., 2015b).

13 The carbon preference index (CPI) has been widely used to roughly estimate the effects of
14 anthropogenic or biogenic sources (e.g., Bray and Evans, 1961; Chen et al., 2014; He et al.,
15 2010; Yamamoto et al., 2013). The CPI values are calculated by the following equation based
16 on the suggestion by Bray and Evans (1961).

$$17 \quad CPI = 0.5 \times \left(\frac{C_{25} + C_{27} + C_{29} + C_{31}}{C_{26} + C_{28} + C_{30} + C_{32}} + \frac{C_{25} + C_{27} + C_{29} + C_{31}}{C_{24} + C_{26} + C_{28} + C_{30}} \right) \quad (4)$$

18 The CPI values are generally high ($CPI > 5$) when there is no serious input from fossil fuel
19 hydrocarbons ($CPI = 1$) (Yamamoto et al., 2013, and references therein). The CPI values during
20 the southwest and northeast monsoon seasons are 1.3 ± 0.12 and 1.0 ± 0.14 , respectively; these
21 values are close to one for both seasons, indicating an anthropogenic n -alkane source. Thus, the
22 CPI value is not susceptible to IPF influence, since the CPI value at [the](#) IPF source is 1.6 ± 0.13
23 (Fujii et al., 2015a), which is not high. Consequently, the CPI cannot be used to identify IPFs
24 sources at a receptor site.

25

26 **3.3 Indonesian peatland fire effect**

27 The hotspot data and backward air trajectories suggest that IPFs strongly modify many chemical
28 species concentrations mostly during the southwest monsoon season. However, IPFs do not
29 always occur during the southwest monsoon season. Therefore, significant differences in some
30 chemical species concentrations among samples affected by IPF and other [sources](#) should be

1 observed. To distinguish IPF samples from other samples obtained during the southwest
2 monsoon season, the OP/OC4 mass ratio is used, which is a useful indicator for IPF (Fujii et
3 al., 2015b). The ratio value is >4 for seven samples (11–13 September 2011 and 14–17 June
4 2012); these samples are regarded as the IPF samples. The OP/OC4 mass ratio for the IPF and
5 other samples is 7.4 ± 3.4 and 0.44 ± 0.49 , respectively, exhibiting significant differences
6 among them according to the two-sided Wilcoxon rank sum test ($p < 0.001$). Figure 6 shows the
7 p-values used to determine the statistical significance in a hypothesis test of the differences
8 between the IPF and other samples for all the quantified species. Significant differences (p
9 < 0.001) are recorded for many chemical species. Thus, the chemical characteristics of $PM_{2.5}$ in
10 Malaysia are significantly influenced by IPFs.

11 Furthermore, the VA/SA and LG/MN mass ratios in the IPF source are investigated as potential
12 indicators, as suggested in previous studies (Fujii et al., 2014, 2015a). The VA/SA mass ratio
13 for IPF and other samples is 1.7 ± 0.36 and 0.59 ± 0.27 , respectively, providing a good indicator
14 ($p < 0.001$). Although the VA/SA mass ratio at the IPF source is 1.1 ± 0.16 (Fujii et al., 2015a),
15 the ratios for IPF samples are higher. Opsahl and Benner (1998) reported photochemical
16 reactivity of VA and SA in the Mississippi River water. They demonstrated that the early
17 degradation of SA in the water is mostly due to its higher photochemical reactivity compared
18 with VA. Even though there are no reports of such degradations in air, SA is considered to be
19 less stable than VA in air as well as in water, which leads to an increased VA/SA ratio after
20 long-range transportation. On the other hand, the LG/MN mass ratio for the IPF and other
21 samples ranges from 14 to 22 and 11 to 31, respectively (Fig. S3). Therefore, the LG/MN mass
22 ratio is inappropriate to extract the effects of IPF in Malaysia, because its value's ranges in the
23 IPF and other samples partially overlap.

24 The daily variability of the C_{27} and LG concentration as well as the VA/SA and OP/OC4 mass
25 ratios are presented in Fig. 7; similar trends are observed in all cases. However, the
26 concentrations of LG, MN and galactosan (Fig. S2) increase abruptly on 10 August 2011,
27 although this sample is not categorised as an IPF sample. We hypothesised that this increase
28 results from local biomass burning, since LG emissions are produced by several different
29 biomass burning sources (Oros and Simoneit, 2001a,b; Oros et al., 2006). Therefore, LG levels
30 are not directly indicative of the IPF contribution in Malaysia; instead, C_{27} may be a useful
31 indicator (Fig. 7). Although the VA/SA mass ratio can be used as an IPF indicator, as we

1 mentioned before, the OP/OC4 mass ratio highlights the differences between the IPF and other
2 samples better than the VA/SA mass ratio (Fig. 7).

3

4 **3.4 Carbonaceous PM_{2.5} contributions**

5 The possible sources of carbonaceous PM_{2.5} are investigated through varimax-rotated PCA of
6 the PJ_A and PJ_S datasets. Over 80% of the cumulative variance in the PJ_A and PJ_S datasets
7 is explained by three and five factors, respectively (Table 2). For the PJ_A data (Table 2a), the
8 total variance explained by the three factors is 80%. Factor A1, which explains 60% of the
9 variance, is heavily loaded (loading factor: >0.65) with OC, LG, MN, galactosan, *p*-
10 hydroxybenzoic acid, VA and C₂₅–C₃₃, which direct towards an IPF source. Factor A2, which
11 corresponds to 12% of the variance, is heavily loaded with C₂₂–C₂₄, suggesting a petrogenic
12 source (Abas et al., 2004a; Gogou et al., 1996; He et al., 2010). Factor A3, which explains 8.0%
13 of the variance in the data set, is heavily loaded with SA and dehydroabietic acid, indicating
14 mixed (softwood and hardwood) biomass burning sources. For the PJ_S dataset (Table 2b), the
15 total variance explained by five factors is 82%. Factor S1 explains 43% of the data's variance
16 and is heavily loaded with C₂₇–C₃₃, which suggests tire wear emission (Rogge et al., 1993).
17 Factor S2 explains 19% of the variance and is heavily loaded with LG, MN, galactosan, VA
18 and SA, which correspond to a biomass burning source. Factor S3, which explains 11% of the
19 variance, is heavily loaded with C₂₂–C₂₆, which indicate a petrogenic source, similar to factor
20 A2. Although heavy loading with only syringaldehyde is found in factor S4 (5.0% of the
21 variance), its source could not be identified. Finally, factor S5 explains 4.5% of the variance
22 and is heavily loaded with EC and cholesterol, which are produced when cooking meat.

23 ~~The d~~Differences of the factor loadings between PJ_A and PJ_S data are observed. For the PCA
24 result of the PJ_A dataset, the factors such as tire wear (factor S1) and cooking (factor S5) as
25 shown in Table 2b are not extracted due to the strong influence of the IPFs. Although a
26 petrogenic source is identified from both results, C25 and C26 are not heavily loaded for the
27 PJ_A dataset. This is also considered to be due to the strong influence of the IPFs.

28 Wahid et al. (2013) reported varimax-rotated PCA results on the distribution of inorganic ions
29 within fine-mode aerosols (<1.5 μm) at Kuala Lumpur, which is close to the present study's
30 sampling site (~10 km). They extracted three principal components from this analysis: (1) motor
31 vehicles, (2) soil and earth's crust and (3) sea spray. Jamhari et al. (2014) applied varimax-

1 rotated PCA on polycyclic aromatic hydrocarbon data in PM₁₀ at Kuala Lumpur. They extracted
2 two factors, which were attributed to (1) natural gas emission and coal combustion and (2)
3 vehicles and gasoline emissions. In the present study, only biomass burning could be identified
4 as a factor through comparison with these previous analyses. Factors such as soil, sea spray and
5 coal combustion could not be identified, because the key inorganic compounds produced from
6 these sources were not determined.

7

8 **4 Conclusions**

9 Annual PM_{2.5} observations in Malaysia have been conducted to quantitatively characterise
10 carbonaceous PM_{2.5}, especially focusing on organic compounds derived from biomass burning
11 for the first time. The main conclusions are summarised as follows:

12 Concentrations of OP, LG, MN, galactosan, syringaldehyde, VA and cholesterol exhibit
13 seasonal variability. The average concentrations of OP, LG, MN, galactosan, VA and
14 cholesterol during the southwest monsoon season are higher than those during the northeast
15 monsoon season, and the differences are statistically significant. In contrast, the syringaldehyde
16 concentration during the southwest monsoon season is lower.

17 Seven IPF samples are distinguished on the basis of the PM_{2.5} OP/OC₄ mass ratio. In addition,
18 significant differences are observed for the concentrations of many chemical species between
19 the IPF and other samples. Thus, the PM_{2.5} chemical characteristics in Malaysia are clearly
20 influenced by IPFs during the southwest monsoon season. Furthermore, two previously
21 suggested indicators of IPF sources have been evaluated, the VA/SA and LG/MN mass ratio.
22 The LG/MN mass ratio ranges from 14 to 22 in the IPF samples and from 11 to 31 in the other
23 samples. The two ratio distributions partially overlap. Thus, the LG/MN mass ratio is not
24 considered appropriate for extracting the effects of IPFs in Malaysia. In contrast, significant
25 differences among the VA/SA mass ratios in the IPF and other samples suggest that it may
26 serve as a good indicator. However, the OP/OC₄ mass ratio differentiates the IPF samples better
27 than the VA/SA mass ratio. Consequently, the OP/OC₄ mass ratio is proposed as a better
28 indicator than the VA/SA mass ratio. Finally, varimax-rotated PCA enabled to discriminate
29 biomass burning components such as IPFs, softwood/hardwood burning and meat cooking.

30

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2

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12

1 Table 1. Statistical results of chemical species concentrations. Av = Average. Sd = Standard deviation.

Compounds	Southwest monsoon (June–September)		Post-monsoon (October–November)		Northeast monsoon (December–March)		Pre-monsoon (April–May)	
	Av ± Sd	Range	Av ± Sd	Range	Av ± Sd	Range	Av ± Sd	Range
OC and EC [$\mu\text{g m}^{-3}$]								
OC	10 ± 7.8	3.6–36	5.6 ± 2.4	2.5–11	5.2 ± 1.4	2.7–8.2	4.2 ± 1.4	2.8–7.3
EC	3.0 ± 0.95	1.0–5.6	3.2 ± 1.3	1.1–5.9	3.4 ± 1.1	1.6–6.1	2.6 ± 1.2	1.4–4.5
Biomarkers [ng m^{-3}]								
levoglucosan	160 ± 130	32–490	64 ± 39	19–130	40 ± 14	17–64	49 ± 21	23–86
mannosan	8.4 ± 8.2	1.5–30	3.4 ± 2.6	0.95–9.1	2.6 ± 1.2	0.84–5.3	2.5 ± 1.2	1.2–5.3
galactosan	2.3 ± 2.3	0.38–8.3	0.86 ± 0.72	0.29–2.8	0.60 ± 0.35	0.13–1.3	0.62 ± 0.34	0.33–1.5
<i>p</i> -hydroxybenzoic acid	1.9 ± 1.9	0.18–7.5	0.79 ± 0.67	0.036–2.2	0.64 ± 0.30	0.20–1.2	0.50 ± 0.25	0.24–1.0
vanillin	1.6 ± 1.1	0.54–5.5	1.2 ± 0.66	0.45–2.2	1.0 ± 0.38	0.21–1.7	0.96 ± 0.42	0.30–1.7
syringaldehyde	0.29 ± 0.22	0.085–1.0	0.59 ± 0.22	0.26–1.2	0.77 ± 0.54	0.074–2.2	0.36 ± 0.22	0.093–0.77
vanillic acid	0.39 ± 0.39	0.074–1.9	0.11 ± 0.070	0.031–0.22	0.073 ± 0.057	0.013–0.26	0.066 ± 0.027	0.034–0.12
syringic acid	0.35 ± 0.41	0.075–2.4	0.26 ± 0.21	0.058–0.59	0.17 ± 0.13	0.029–0.64	0.16 ± 0.084	0.049–0.28
dehydroabietic acid	1.7 ± 1.1	0.10–5.4	1.1 ± 0.69	0.31–2.4	1.1 ± 1.1	0.14–4.6	0.67 ± 0.24	0.16–0.98
cholesterol	1.8 ± 0.82	0.50–3.7	1.2 ± 0.51	0.57–2.0	0.98 ± 0.51	0.026–2.0	1.3 ± 0.56	0.51–2.0
<i>n</i> -alkanes [ng m^{-3}]								
docosane	3.2 ± 0.82	1.8–5.0	2.9 ± 0.61	2.0–4.0	3.0 ± 0.53	1.9–4.2	4.0 ± 4.8	2.1–19
tricosane	3.6 ± 1.2	2.0–7.2	3.2 ± 0.91	2.0–4.8	3.2 ± 0.65	1.8–4.4	5.0 ± 7.6	2.1–29
tetracosane	5.8 ± 3.2	2.5–19	5.7 ± 1.7	3.3–8.7	6.1 ± 2.3	2.9–15	6.3 ± 8.5	2.7–33
pentacosane	8.9 ± 6.7	3.5–34	5.7 ± 2.3	3.1–11	6.0 ± 1.6	3.7–9.2	5.8 ± 5.5	3.2–23
hexacosane	13 ± 9.8	4.3–49	8.6 ± 3.7	3.6–18	9.7 ± 2.8	5.0–16	7.1 ± 5.3	3.5–23

書式変更: 左揃え

heptacosane	16 ± 14	4.7–64	7.2 ± 2.6	3.6–12	8.2 ± 2.4	3.7–14	5.8 ± 3.4	3.3–16
octacosane	12 ± 12	2.6–54	4.3 ± 1.8	1.7–7.9	5.9 ± 3.0	2.3–17	3.6 ± 1.7	2.3–8.2
nonacosane	13 ± 13	3.0–55	4.9 ± 2.1	1.5–8.7	6.3 ± 2.2	3.3–13	4.5 ± 1.4	2.6–7.8
triacontane	7.9 ± 7.8	2.0–36	3.8 ± 2.0	1.6–9.0	5.2 ± 2.7	2.0–16	3.3 ± 1.7	1.7–8.3
hentriacontane	14 ± 14	2.8–59	4.8 ± 1.9	1.8–8.4	5.7 ± 2.0	3.3–11	4.3 ± 1.2	2.9–6.9
dotriacontane	6.7 ± 5.5	1.6–27	3.4 ± 0.72	2.4–4.5	4.6 ± 1.3	2.8–7.8	3.1 ± 0.88	1.8–4.4
tritriacontane	6.8 ± 7.1	1.2–33	2.5 ± 0.97	1.1–4.2	2.8 ± 0.92	1.2–5.0	2.1 ± 0.72	1.5–3.8

1

1 Table 2a. Factor loadings from varimax-rotated PCA of PJ_A data. A1–A3 indicate factors.

	A1	A2	A3
OC	<u>0.97</u>	0.10	0.16
EC	0.29	0.37	0.51
levoglucosan	<u>0.81</u>	-0.05	0.17
mannosan	<u>0.89</u>	0.00	0.11
galactosan	<u>0.90</u>	0.02	0.08
<i>p</i> -hydroxybenzoic acid	<u>0.94</u>	0.04	0.22
vanillin	0.61	0.15	0.25
syringaldehyde	-0.17	0.12	0.40
vanillic acid	<u>0.65</u>	-0.10	0.55
syringic acid	0.28	-0.11	<u>0.81</u>
dehydroabietic acid	0.15	-0.01	<u>0.86</u>
cholesterol	0.36	0.14	0.39
C ₂₂	0.03	<u>0.95</u>	0.05
C ₂₃	0.07	<u>0.95</u>	0.05
C ₂₄	0.30	<u>0.92</u>	0.06
C ₂₅	<u>0.81</u>	0.54	0.14
C ₂₆	<u>0.86</u>	0.43	0.13
C ₂₇	<u>0.95</u>	0.23	0.13
C ₂₈	<u>0.96</u>	0.18	0.07
C ₂₉	<u>0.97</u>	0.13	0.12
C ₃₀	<u>0.92</u>	0.25	0.05
C ₃₁	<u>0.97</u>	0.10	0.13
C ₃₂	<u>0.93</u>	0.15	0.11
C ₃₃	<u>0.97</u>	0.10	0.13
% variance	60	12	8.0
% cumulative	60	72	80

2

1 Table 2b. Factor loadings from varimax-rotated PCA of PJ_S data. S1–S5 indicate factors.

	S1	S2	S3	S4	S5	2
OC	0.47	0.47	0.10	0.08	0.57	
EC	0.39	0.20	0.25	0.26	<u>0.65</u>	
levoglucosan	0.09	<u>0.71</u>	-0.03	-0.52	0.19	
mannosan	0.19	<u>0.84</u>	0.02	-0.26	0.28	
galactosan	0.17	<u>0.83</u>	0.06	-0.09	0.41	
<i>p</i> -hydroxybenzoic acid	0.26	0.62	0.08	0.23	0.42	
vanillin	0.22	0.32	0.07	0.05	0.61	
syringaldehyde	0.24	0.13	0.01	<u>0.74</u>	0.07	
vanillic acid	-0.12	<u>0.81</u>	-0.04	0.22	-0.01	
syringic acid	0.02	<u>0.81</u>	0.00	0.37	0.26	
dehydroabietic acid	0.18	0.44	0.04	0.12	0.60	
cholesterol	0.01	0.17	0.15	-0.21	<u>0.77</u>	
C ₂₂	0.05	-0.02	<u>0.97</u>	-0.04	0.05	
C ₂₃	0.05	0.00	<u>0.97</u>	-0.04	0.04	
C ₂₄	0.28	-0.03	<u>0.94</u>	0.04	-0.01	
C ₂₅	0.33	0.10	<u>0.85</u>	0.05	0.35	
C ₂₆	0.61	0.05	<u>0.68</u>	0.14	0.24	
C ₂₇	<u>0.67</u>	0.08	0.53	0.10	0.35	
C ₂₈	<u>0.86</u>	0.06	0.27	-0.01	0.01	
C ₂₉	<u>0.89</u>	0.14	0.18	0.08	0.29	
C ₃₀	<u>0.84</u>	0.03	0.33	0.04	-0.12	
C ₃₁	<u>0.77</u>	0.24	0.07	0.10	0.47	
C ₃₂	<u>0.88</u>	-0.04	0.02	0.10	0.16	
C ₃₃	<u>0.72</u>	0.28	-0.03	0.14	0.49	
% variance	43	19	11	5.0	4.5	
% cumulative	43	62	72	77	82	

1 **Figure Captions**

2

3 Figure 1. Daily variability of the MAPI and visibility during the sampling periods.

4 Figure 2. Monthly hotspot counts in the Sumatra Island.

5 Figure 3. Daily variation of the OC fractions' mass concentrations during the sampling periods.

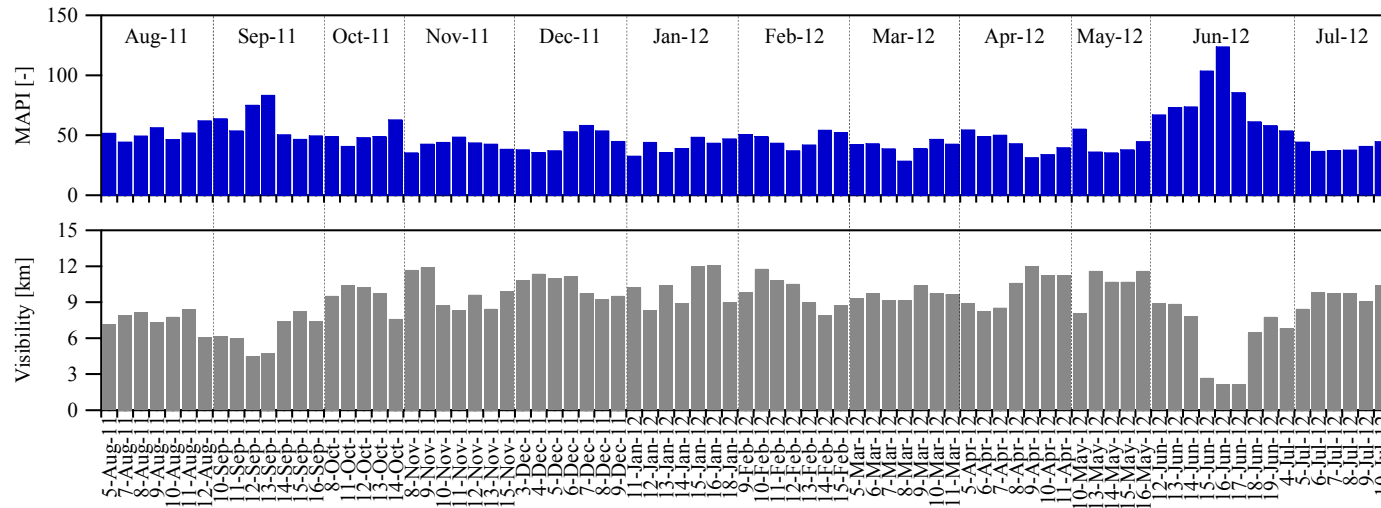
6 Figure 4. Box-whisker plots of molecular distributions of *n*-alkanes during the (a) southwest
7 and (b) northeast monsoon seasons. The horizontal lines in the box represent the 25th, 50th, and
8 75th percentiles. The whiskers represent the 10th and 90th percentiles.

9 Figure 5. Number fraction of C_{max} in the $PM_{2.5}$ samples for each monsoon season.

10 Figure 6. P-values to determine significance in the two-sided Wilcoxon rank sum test between
11 the IPF and other samples.

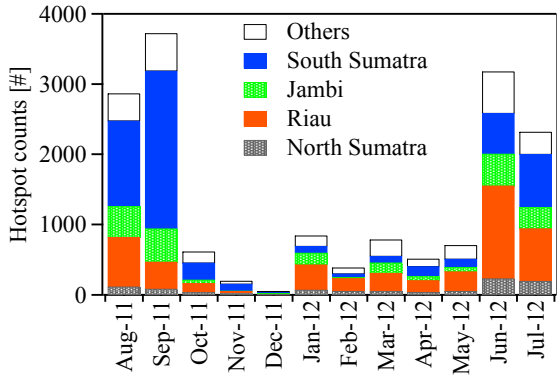
12 Figure 7. Daily variability of the C_{27} and LG concentration as well as the VA/SA and OP/OC4
13 mass ratios during the sampling periods.

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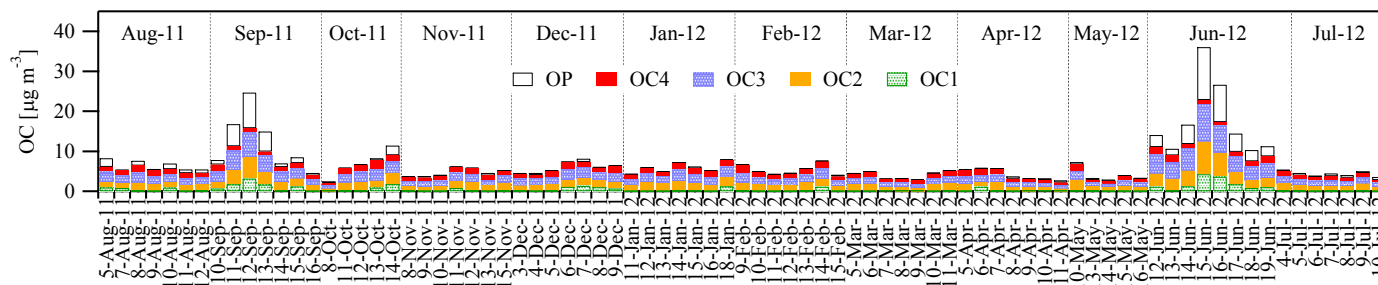
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Figure 1. Daily variability of the MAPI and visibility during the sampling periods.



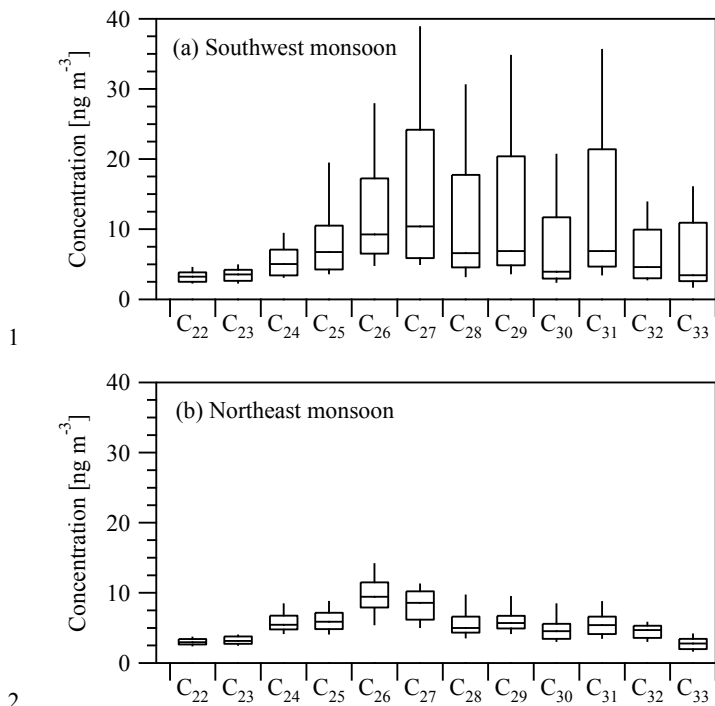
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Figure 2. Monthly hotspot counts in the Sumatra Island.



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Figure 3. Daily variation of the OC fractions' mass concentrations during the sampling periods.

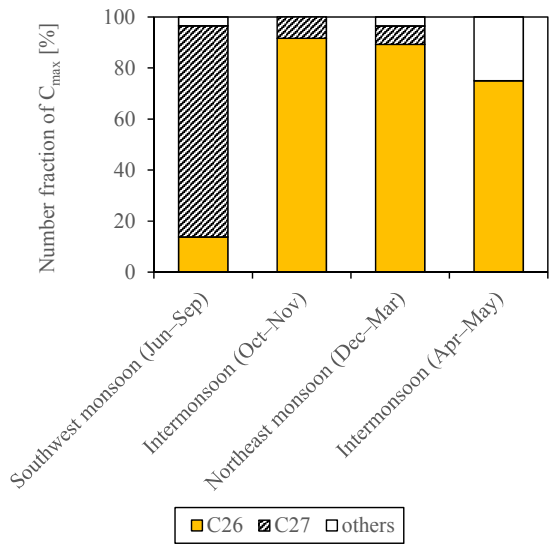


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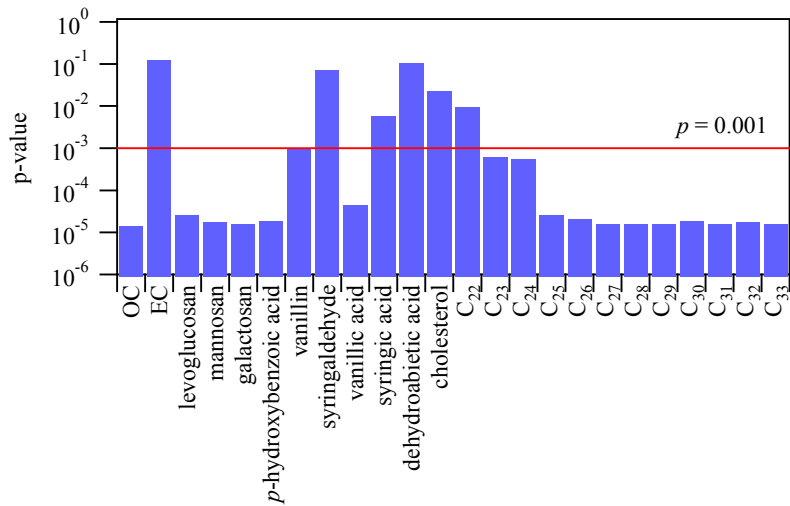
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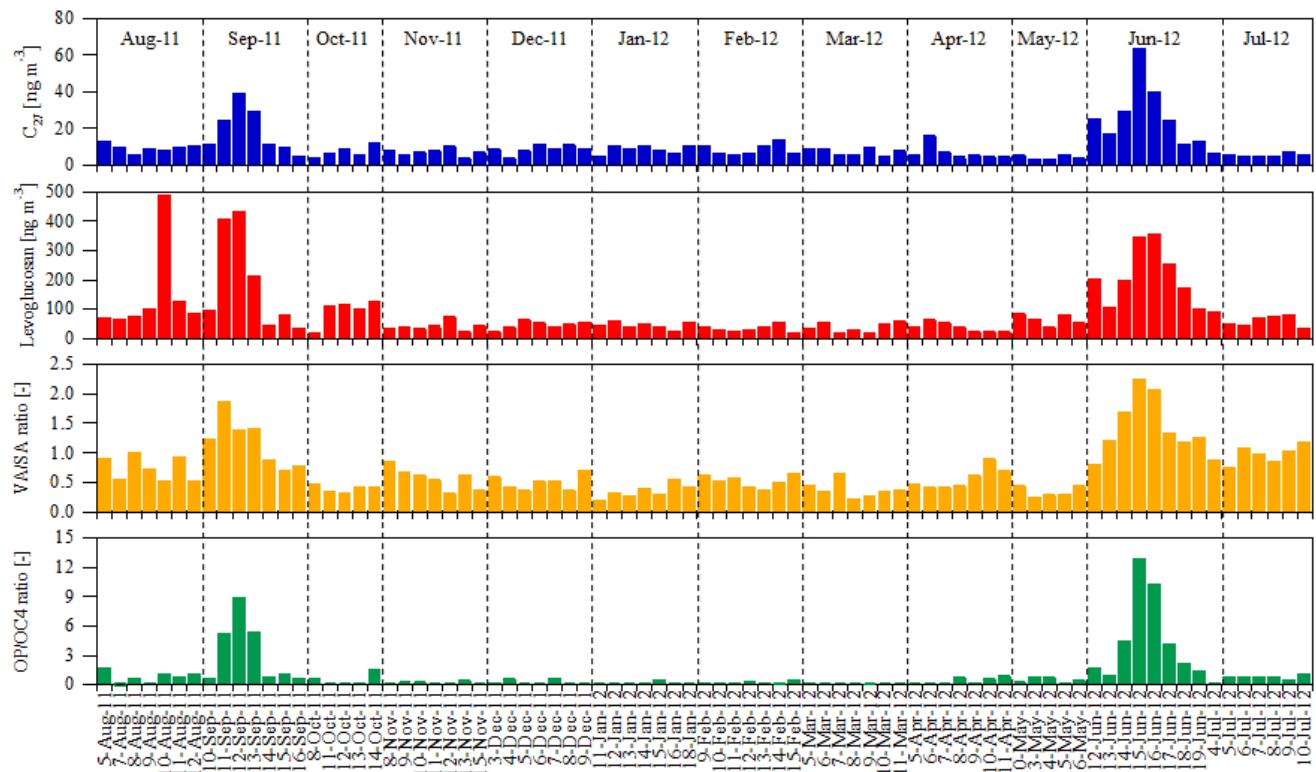
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Figure 5. Number fraction of C_{max} in the $PM_{2.5}$ samples for each monsoon season.



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Figure 6. P-values to determine significance in the two-sided Wilcoxon rank sum test between the IPF and other samples.



1
 2 Figure 7. Daily variability of the C_{27} and LG concentration as well as the VA/SA and OP/OC4 mass ratios during the sampling periods.

1 **Supplemental Information**

2 **Annual variations of carbonaceous PM_{2.5} in Malaysia:**
3 **Influence of Indonesian peatland fire**

4

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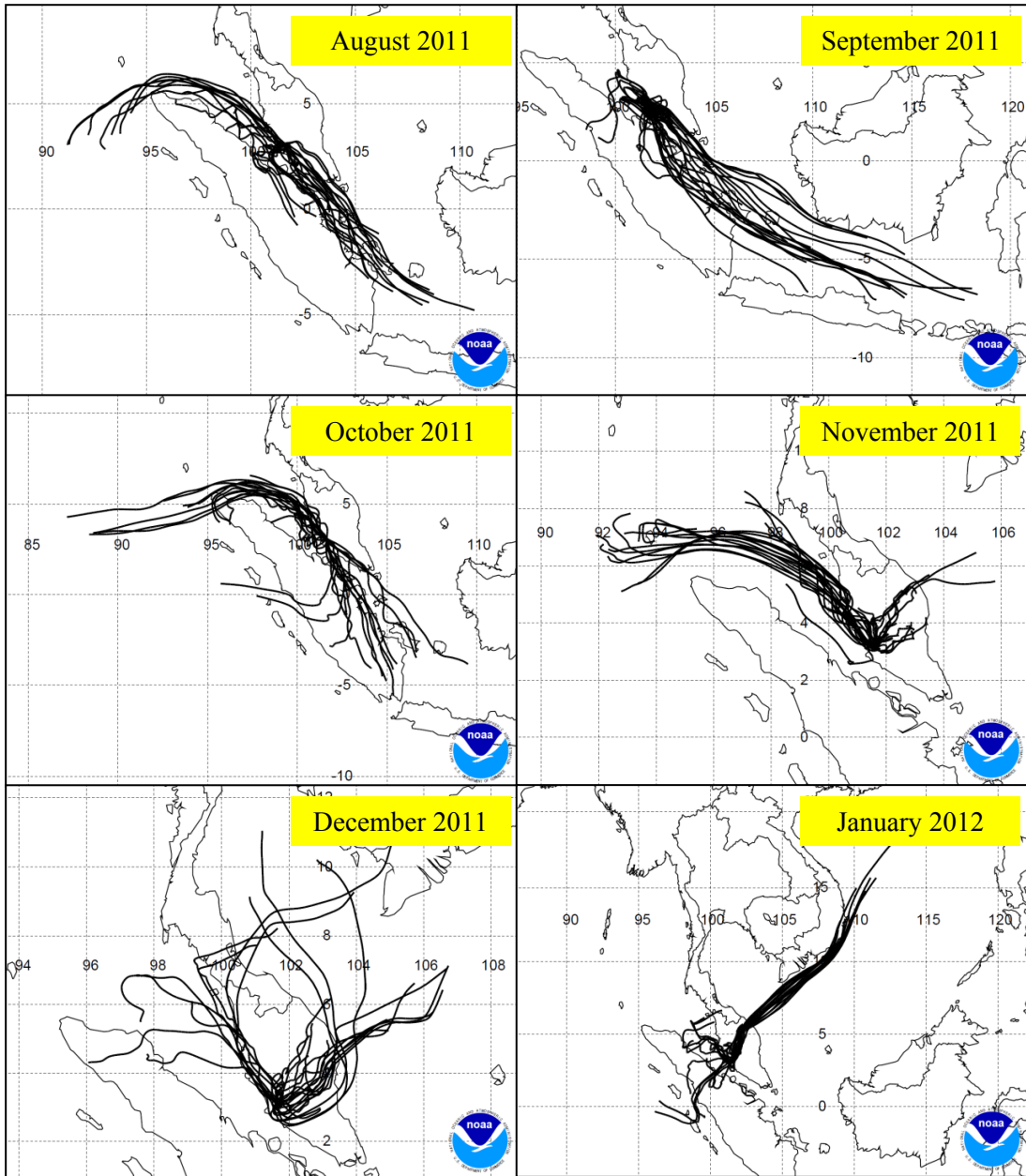
12 [5]{Institute for Environment and Development, Universiti Kebangsaan Malaysia, Bangi,
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14 [6]{Research Organization for University-Community Collaborations, Osaka Prefecture
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16 Correspondence to: Y. Fujii (fujii.yusuke.86n@st.kyoto-u.ac.jp)

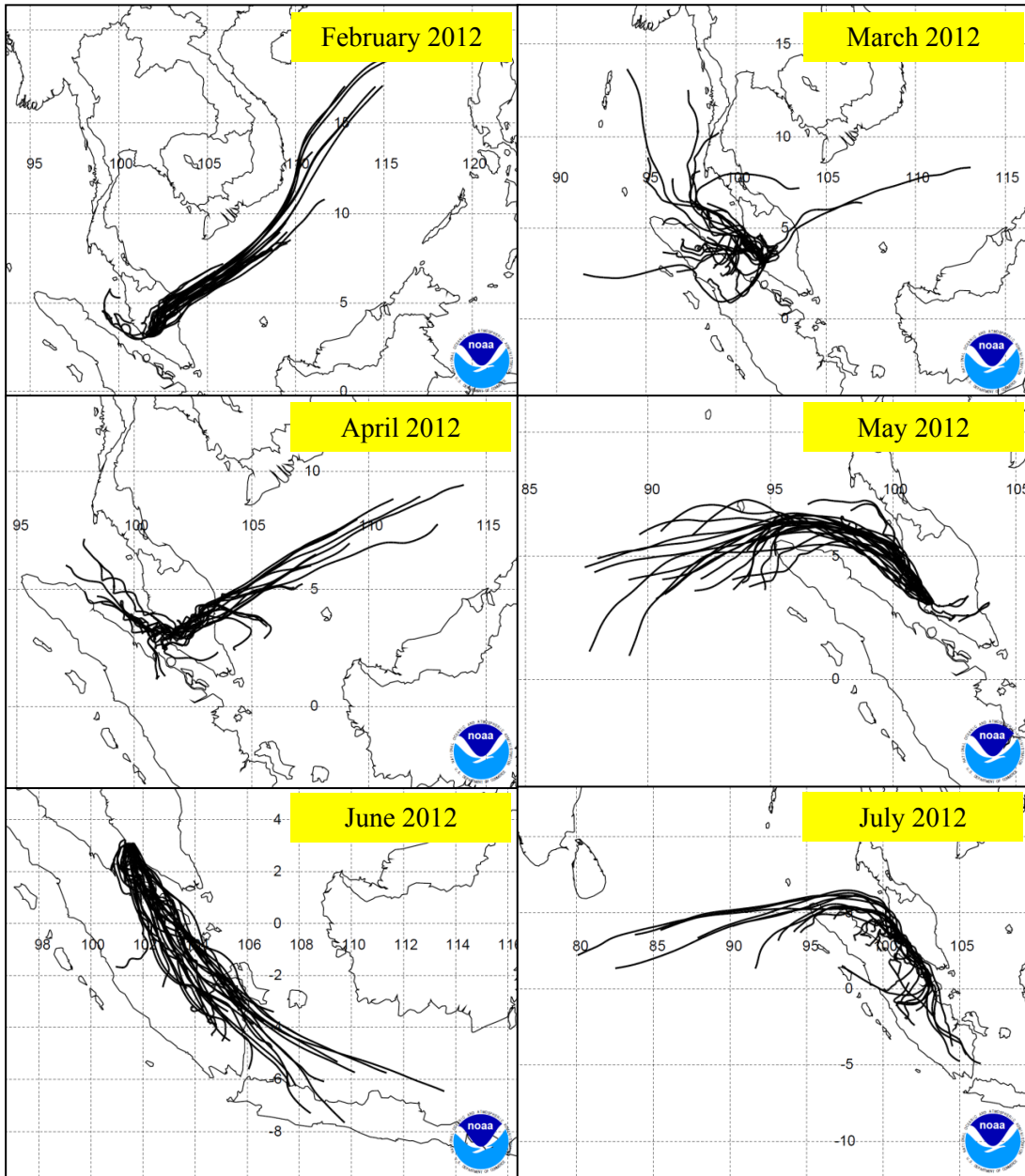
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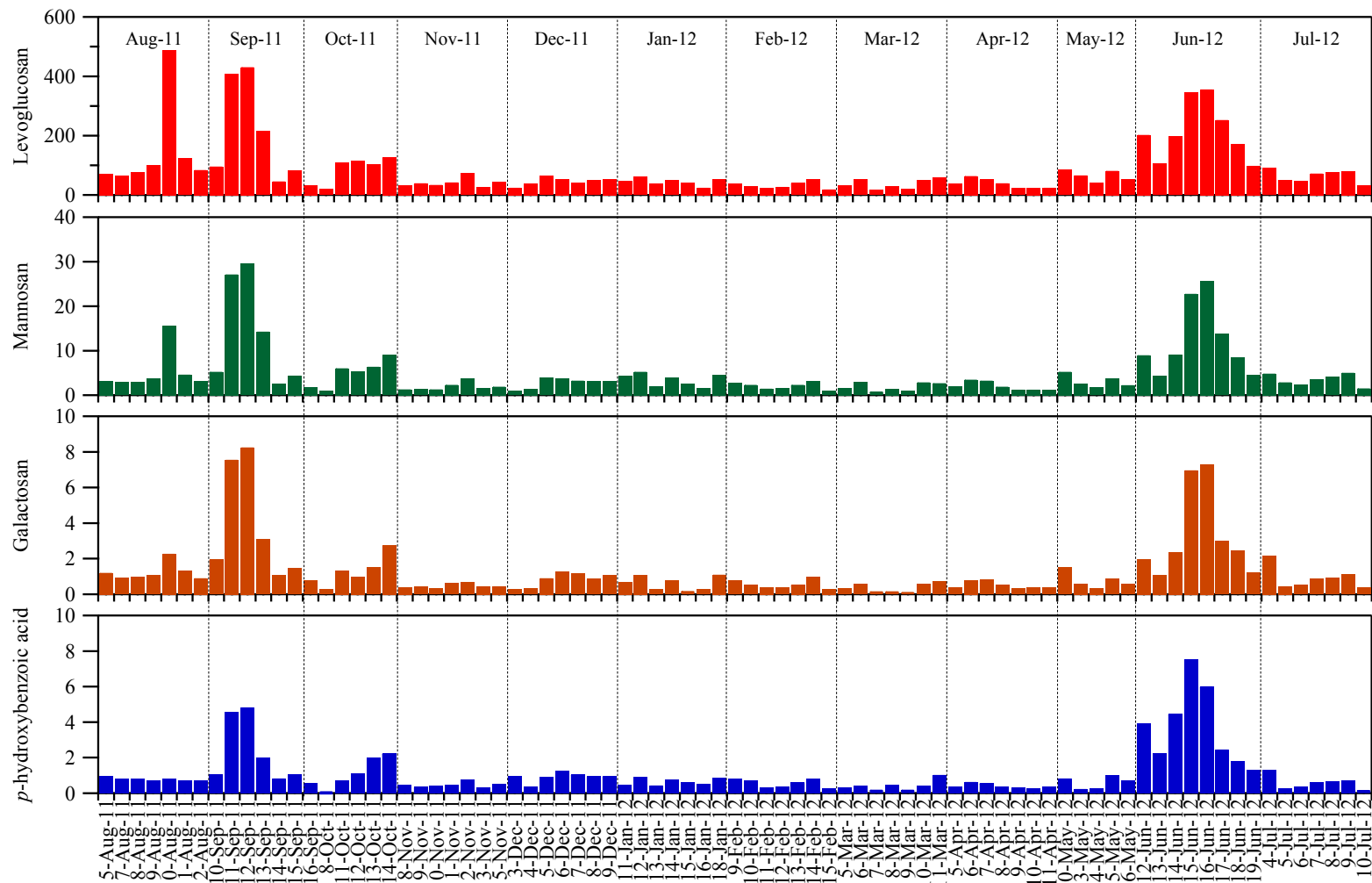
Figure S1. Backward air trajectories during the sampling periods. The 3-days backward air trajectories every 6 hours with 500 m above ground level arriving at the sampling site in local time were calculated during the sampling periods by the Hybrid Single Particle Lagrangian Integrated Trajectory model (Draxler and Hess, 2004) based on meteorological data obtained from the Global Data Assimilation.



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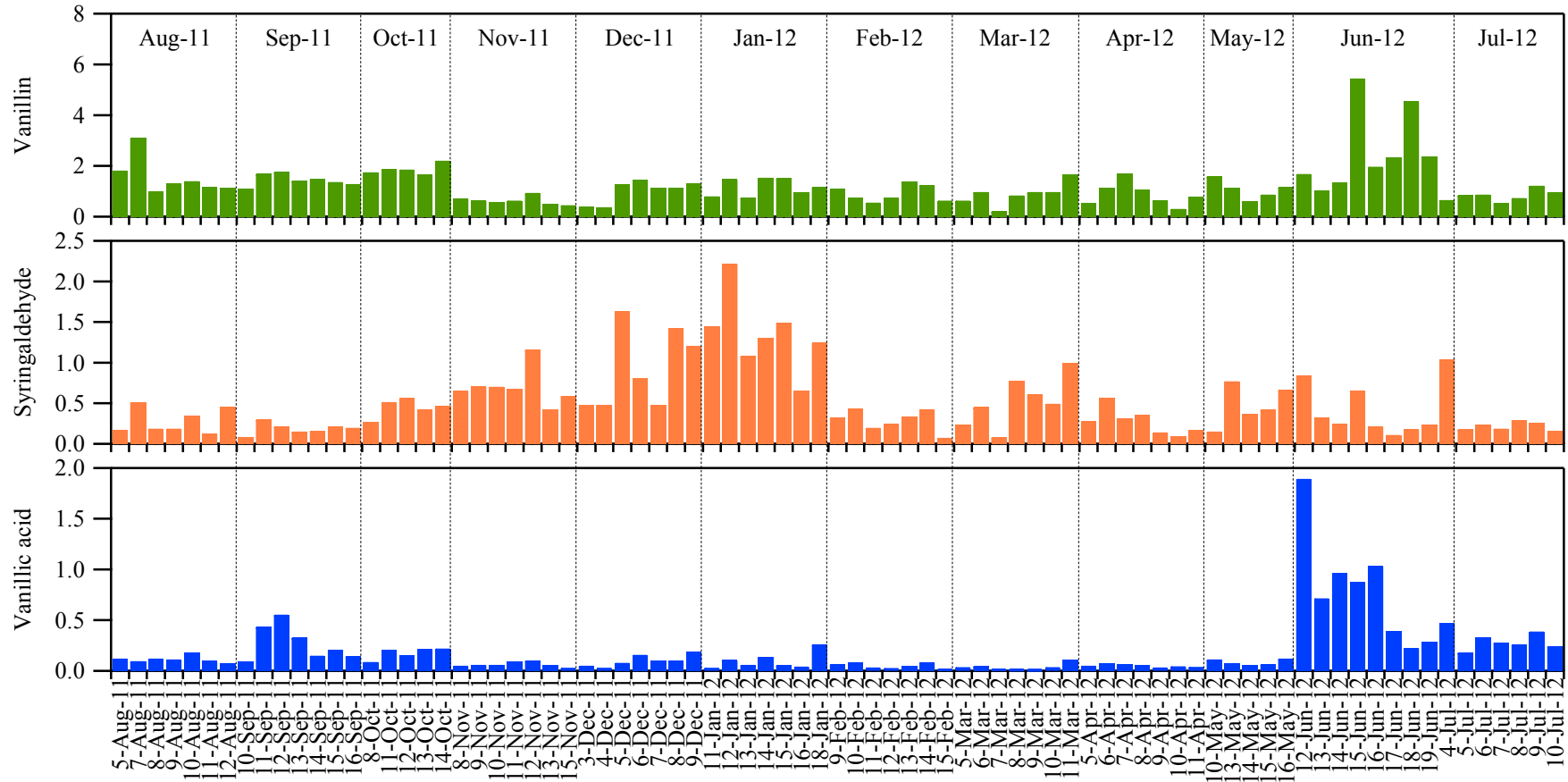
3 Figure S1. Continued.



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2 Figure S2. Daily variation of biomarker concentrations [ng m⁻³] during the sampling periods.

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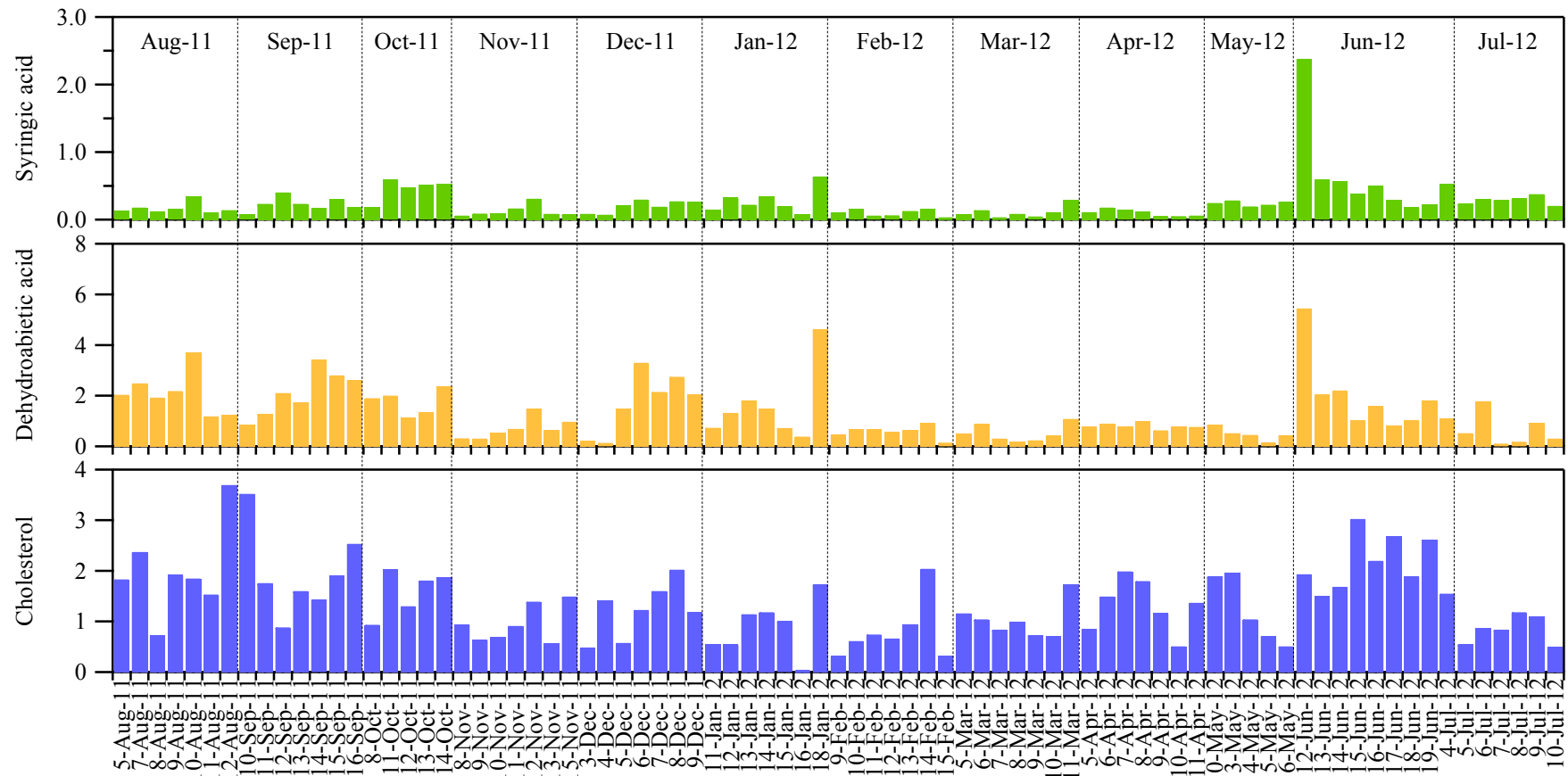


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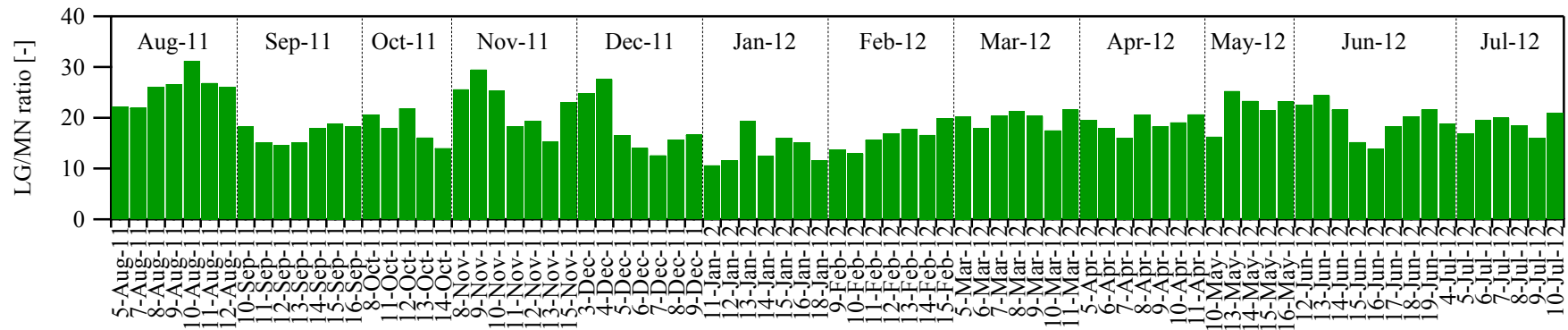
4 Figure S2. Continued.

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Figure S2. Continued.



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3 Figure S3. Daily variation of the levoglucosan/mannosan ratio (LG/MN) during the sampling periods.

1 **References**

- 2 Draxler, R.R. and Hess, G.D.: Description of the HYSPLIT 4 Modeling System, NOAA
- 3 Technical Memorandum ERL ARL-224, 2004.