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# Ice nucleating particles at a coastal marine boundary layer site: correlations with aerosol type and meteorological conditions

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Ice nucleating  
particles at a coastal  
marine boundary  
layer site

R. H. Mason et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



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# Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.

Title Page

## Abstract

Introduction

## Conclusions

## References

## Tables

## Figures



Back

Close

Full Screen / Esc

[Printer-friendly Version](#)

## Interactive Discussion



## Abstract

Information on what aerosol particle types are the major sources of ice nucleating particles (INPs) in the atmosphere is needed for climate predictions. To determine which aerosol particles are the major sources of immersion-mode INPs at a coastal site in Western Canada, we investigated correlations between INP number concentrations and both concentrations of different atmospheric particles and meteorological conditions. We show that INP number concentrations are strongly correlated with the number concentrations of fluorescent bioparticles between  $-15$  and  $-25$   $^{\circ}\text{C}$ , and that the size distribution of INPs is most consistent with the size distribution of fluorescent bioparticles. We conclude that biological particles were likely the major source of ice nuclei at freezing temperatures between  $-15$  and  $-25$   $^{\circ}\text{C}$  at this site for the time period studied. At  $-30$   $^{\circ}\text{C}$ , INP number concentrations are also well correlated with number concentrations of the total aerosol particles  $\geq 0.5 \mu\text{m}$ , suggesting that non-biological particles may have an important contribution to the population of INPs active at this temperature. As we found that black carbon particles were unlikely to be a major source of ice nuclei during this study, these non-biological INPs may include mineral dust. Furthermore, correlations involving tracers of marine aerosols and marine biological activity indicate that the majority of INPs measured at the coastal site likely originated from terrestrial rather than marine sources. Finally, six existing empirical parameterizations of ice nucleation were tested to determine if they accurately predict the measured INP number concentrations. We found that none of the parameterizations selected are capable of predicting INP number concentrations with high accuracy over the entire temperature range investigated.

## 1 Introduction

At temperatures below approximately  $-37$   $^{\circ}\text{C}$  ice nucleation of solution droplets in the atmosphere occurs by a homogeneous process, but at warmer temperatures ice can

<a href="#">Title Page</a>	
<a href="#">Abstract</a>	<a href="#">Introduction</a>
<a href="#">Conclusions</a>	<a href="#">References</a>
<a href="#">Tables</a>	<a href="#">Figures</a>
<a href="#"></a>	<a href="#"></a>
<a href="#"></a>	<a href="#"></a>
<a href="#">Back</a>	<a href="#">Close</a>
<a href="#">Full Screen / Esc</a>	
<a href="#">Printer-friendly Version</a>	
<a href="#">Interactive Discussion</a>	



also form by heterogeneous nucleation on solid or partially solid aerosol particles called ice nucleating particles (INPs). INPs are a small subset of the total aerosol population (Rogers et al., 1998) whose unique surface properties make them capable of lowering the energy barrier to ice nucleation and hence cause freezing at warmer temperatures or lower supersaturations with respect to ice compared to homogeneous nucleation. Four modes of nucleation have been identified (Vali, 1985): deposition nucleation, where ice forms on the INP directly from the gas phase; condensation freezing, where ice forms during the condensation of water onto the INP; immersion freezing, where ice forms on an INP within a supercooled droplet; and contact freezing, where the impact of a supercooled droplet by an INP initiates freezing. In this study we focus on immersion freezing, which is relevant to ice formation in mixed-phase clouds.

The presence of INPs in the atmosphere can lead to changes in the microphysical properties and lifetime of clouds. As a result, a change in INP concentrations can indirectly modify climate by changing cloud optical properties, lifetime, and cloud extent (e.g. Baker, 1997; Lohmann, 2002; Storelvmo et al., 2011; Creamean et al., 2013). Currently, the role of INPs in climate change is highly uncertain (Boucher et al., 2013). To predict the role of INPs in climate change and precipitation, information on what particle types are the major sources of INPs in the atmosphere is needed. Particle types that we focused on as possible candidates for INPs included primary biological particles, black carbon (BC), and primary marine particles.

Primary biological particles have been identified as a possible source of INPs (e.g. Szyrmer and Zawadzki, 1997; Möhler et al., 2007; Garcia et al., 2012; Hiranuma et al., 2015). Biological particles found to efficiently nucleate ice include bacteria (e.g. Maki et al., 1974; Lindow et al., 1978; Maki and Willoughby, 1978; Kozloff et al., 1983), fungi (e.g. Jayaweera and Flanagan, 1982; Tsumuki et al., 1992; Richard et al., 1996; Ian-none et al., 2011; Haga et al., 2013; Morris et al., 2013), and pollen (e.g. Diehl et al., 2001, 2002; von Blohn et al., 2005; Pummer et al., 2012; Augustin et al., 2013; Hader et al., 2014; O'Sullivan et al., 2015). Strong correlations between number concentrations of INPs and primary biological particles have been found during field studies

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

(Prenni et al., 2009b, 2013; Huffman et al., 2013; Tobo et al., 2013), and biological particles have been observed in ice-crystal residuals of mixed-phase clouds (e.g. Pratt et al., 2009), cloud water (e.g. Joly et al., 2014), and snow samples (e.g. Christner et al., 2008; Morris et al., 2008; Hill et al., 2014). Ice-active biological particles have also been associated with soils (Conen et al., 2011; O'Sullivan et al., 2014; Tobo et al., 2014; Fröhlich-Nowoisky et al., 2015). Model studies have shown that biological particles may not be important for ice nucleation on a global and annual scale (Hoose et al., 2010; Sesartic et al., 2013; Spracklen and Heald, 2014), but may be important on regional and seasonal scales, especially if concentrations of biological particles are high or concentrations of other types of INPs are low (Phillips et al., 2009; Sun et al., 2012; Burrows et al., 2013; Creamean et al., 2013; Yun and Penner, 2013; Costa et al., 2014; Spracklen and Heald, 2014).

Another potential source of INPs in the atmosphere is BC particles (Kärcher et al., 2007). Laboratory studies have given varying results on whether BC particles act as efficient INPs under atmospherically relevant conditions (e.g. Gorbunov et al., 2001; Möhler et al., 2005; Dymarska et al., 2006; Kärcher et al., 2007; DeMott et al., 2009; Friedman et al., 2011; Cziczo et al., 2013; Brooks et al., 2014), although BC is generally considered to be less efficient than mineral dust in the immersion mode (Hoose and Möhler, 2012; Murray et al., 2012; and references therein). Field studies have also produced varying results (e.g. Lin et al., 2006; Cozic et al., 2008; Kamphus et al., 2010; Twohy et al., 2010; Ebert et al., 2011; Corbin et al., 2012; Cziczo et al., 2013; Knopf et al., 2014; McCluskey et al., 2014). Models have suggested that carbonaceous aerosols may have a significant indirect effect on climate if they efficiently nucleate ice (e.g. Lohmann, 2002; Liu et al., 2009; Penner et al., 2009; Yun and Penner, 2013).

Ambient measurements have found INPs in air masses above oceans and coastal sites (e.g. Bigg, 1973; Schnell, 1977; Flyger and Heidam, 1978; Saxena, 1983; Rosinski et al., 1995; Rogers et al., 2001; Prenni et al., 2009a), with some of the INPs identified as biological (e.g. Jayaweera and Flanagan, 1982). Studies have also indicated the potential of marine particles as a source of ice nuclei, including bacteria and

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

<a href="#">Title Page</a>	
<a href="#">Abstract</a>	<a href="#">Introduction</a>
<a href="#">Conclusions</a>	<a href="#">References</a>
<a href="#">Tables</a>	<a href="#">Figures</a>
<a href="#"></a>	<a href="#"></a>
<a href="#"></a>	<a href="#"></a>
<a href="#">Back</a>	<a href="#">Close</a>
<a href="#">Full Screen / Esc</a>	
<a href="#">Printer-friendly Version</a>	
<a href="#">Interactive Discussion</a>	

phytoplankton found in seawater and sea ice (Schnell, 1975, 1977; Schnell and Vali, 1975; Parker et al., 1985; Knopf et al., 2011; Alpert et al., 2011), synthetic sea-salt or sodium chloride particles (Wise et al., 2012; Wagner and Möhler, 2013; Schill and Tolbert, 2014), generated sea spray aerosol using ocean water (DeMott et al., 2013;

Prather et al., 2013), and material from the sea surface microlayer (Wilson et al., 2015). It is therefore possible that the release of biological particles from the oceans may be a source of INPs in the atmosphere. The modeling work of Burrows et al. (2013) indicates that marine INPs may be particularly important in remote regions such as the Southern Ocean.

To determine which aerosol particles are the major source of INPs in the immersion mode at a coastal site in Western Canada, we investigate correlations between INP number concentrations and both concentrations of different atmospheric particle types and meteorological conditions. Measurements were conducted in August 2013 as part of the Network on Climate and Aerosols: addressing Key Uncertainties in Remote Canadian Environments (NETCARE) project (<http://www.netcare-project.ca/>). A primary goal of the study was to investigate whether primary biological particles, BC particles, or primary marine particles are a major source of INPs at this coastal site. In addition, we also test the ability of parameterizations reported in the literature at predicting the INP number concentrations measured at this coastal site.

## 2 Methods

### 2.1 Site description and instrument location

Measurements were performed at Amphitrite Point (48.92° N, 125.54° W) on the west coast of Vancouver Island in British Columbia, Canada. This was also the location of studies on ozone (McKendry et al., 2014) and cloud condensation nuclei (Yakobi-Hancock et al., 2014). Amphitrite Point (Fig. 1) is located approximately 2.2 km south of the town of Ucluelet (population of 1627 in 2011; Statistics Canada, 2012). The

**Ice nucleating  
particles at a coastal  
marine boundary  
layer site**

R. H. Mason et al.

largest nearby population centers are Nanaimo 120 km to the east, Victoria 170 km to the southeast, and Vancouver 180 km to the east. This region has a temperate maritime climate characterized by warm summers, mild winters, and relatively high levels of cloud cover and precipitation, falling under the Cfb climate type of the Köppen–Geiger classification scheme (Kottek et al., 2006). Local forests contain predominantly coniferous tree species including western hemlock, western redcedar, and Douglas-fir that is characteristic of most low-elevation sites along the west coast of Canada (Austin et al., 2008). The Pacific Ocean is west and south of the site, where the mixing of iron-rich coastal waters with nitrate-rich oceanic waters produces a zone of high primary productivity (Whitney et al., 2005; Ribalet et al., 2010).

Aerosol instrumentation was located in one of two mobile laboratories; one specific to the NETCARE project (labeled 1 in Fig. 1) and one operated by Environment Canada, the British Columbia Ministry of Environment, and Metro Vancouver (labeled 2 in Fig. 1). Aerosols were sampled through louvered total suspended particulate inlets (Mesa Labs Inc., Butler, NJ, USA) or louvered PM<sub>10</sub> inlets (Thermo Scientific, Waltham, MA, USA) atop masts extending 5.5 m a.g.l. The two mobile laboratories were approximately 20 m above mean sea level and 100 m from the high tide line of the Pacific Ocean (McKendry et al., 2014). A row of trees and shrubs approximately 2–10 m in height stood between the laboratories and the rocky shoreline. Adjacent to the laboratories on their seaward side was Amphitrite Lighthouse (labeled 3 in Fig. 1) and Wild Pacific Trail, local tourist attractions and a source of foot traffic during fair weather. Immediately north and east of the site was a station of the Canadian Coast Guard (labeled 4 in Fig. 1).

The meteorological parameters reported in this study were measured at Amphitrite Lighthouse, located approximately halfway between the mobile laboratories and the ocean. Relative humidity and temperature were monitored using an HMP45C probe (Campbell Scientific, Logan, UT, USA) with accuracies of  $\pm 3\%$  and  $\pm 0.2^\circ\text{C}$ , respectively. Wind direction and wind speed were determined by a model 05305L Wind Mon-

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

itor (R. M. Young, Traverse City, Michigan, USA) to a respective accuracy of  $\pm 3^\circ$  and  $\pm 0.2 \text{ m s}^{-1}$ .

## 2.2 Ice nucleating particle measurements

INP number concentrations in the immersion mode were measured using the micro-orifice uniform deposit impactor-droplet freezing technique (MOUDI-DFT; Mason et al., 2015). A Model II 120R MOUDI (MSP Corp., Shoreview, MN, USA) collected size-fractionated aerosol samples by inertial separation (Marple et al., 1991) onto hydrophobic glass cover slips (HR3-215; Hampton Research, Aliso Viejo, CA, USA). To compensate for the thickness of the hydrophobic glass cover slips, spacers were placed between the MOUDI stages. Custom substrate holders were added to the MOUDI impaction plates to maintain consistent positioning of the hydrophobic glass cover slips within the impactor (Mason et al., 2015). Samples from MOUDI stages 2–8 were used in this study, corresponding to a particle size range of 10–0.18  $\mu\text{m}$  (50 % cutoff aerodynamic diameter). Thirty-four sets of MOUDI samples were collected; 18 during the day and 16 at night. The average collection time of a MOUDI sample was 7.8 h. Details of each INP sampling period are available in Table S1 of the Supplement.

The ice-nucleating ability of particles collected by the MOUDI was then determined by the DFT (Koop et al., 2000; Iannone et al., 2011; Mason et al., 2015; Wheeler et al., 2015). Within 24 h of collection, samples were placed in a temperature- and humidity-controlled flow cell that was coupled to an optical microscope (Axiolab; Zeiss, Oberkochen, Germany) with a 5 $\times$  magnification objective. At a sample temperature of 0  $^\circ\text{C}$  a humidified gas flow was introduced, resulting in the formation of water droplets on the sample. Following droplet growth by condensation and coalescence, the droplet size was decreased with a dry gas flow to a final size of approximately 80–160  $\mu\text{m}$  in diameter. On average, more than 99 % of particles on the surface of the hydrophobic glass cover slip were incorporated into droplets by this procedure. Closing valves upstream and downstream of the cell then isolated the flow cell, and the sample temperature was lowered at a constant rate of  $-10 \text{ }^\circ\text{C min}^{-1}$  to  $-40 \text{ }^\circ\text{C}$ . This cooling rate

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



was chosen to minimize the freezing of a liquid droplet by contact with a growing ice crystal. Recent work suggests that changing the cooling rate by an order of magnitude may lead to a shift in freezing temperatures of approximately 0.5–2 °C (Murray et al., 2011; Broadley et al., 2012; Welti et al., 2012; Wright and Petters, 2013; Wright et al., 2013; Wheeler et al., 2015). During droplet growth, evaporation, and cooling, a CCD camera connected to the optical microscope recorded a digital video of the sample. Using the video time stamp and a resistance temperature detector positioned within the flow cell, which was calibrated against the melting point of water droplets approximately 100 µm in diameter, the freezing temperature of each droplet was found by manually noting the increase in droplet opacity immediately following ice nucleation.

Since a small fraction of the sampled particles (less than 1 % on average) was not included in the droplets, there was the possibility of deposition nucleation as well. However, based on an analysis of the videos recorded during the ice nucleation experiments, fewer than 3 % of all freezing events observed were the result of deposition nucleation. Due to the low occurrence of deposition nucleation, only immersion freezing results are reported.

The atmospheric number concentration of INPs within the size cut of each MOUDI stage, [INPs( $T$ )], was evaluated using the following equation:

$$[\text{INPs}(T)] = -\ln \left( \frac{N_u(T)}{N_o} \right) N_o \left( \frac{A_{\text{deposit}}}{A_{\text{DFT}} V} \right) f_{nu} f_{ne} \quad (1)$$

where  $N_u(T)$  is the number of unfrozen droplets at temperature  $T$ ,  $N_o$  is the total number of droplets,  $A_{\text{deposit}}$  is the total area of the sample deposit on the hydrophobic glass cover slips,  $A_{\text{DFT}}$  is the area of the sample analyzed by the DFT,  $V$  is the volume of air sampled by the MOUDI,  $f_{nu}$  is a correction factor to account for changes in particle concentration across each MOUDI sample (because the DFT analyzes only a fraction of the entire sample), and  $f_{ne}$  is a correction factor to account for the uncertainty associated with the number of nucleation events in each experiment following Koop et al. (1997). Additional details are available in Mason et al. (2015). Equation (1) takes

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

into account the possibility of multiple INPs being contained in a single droplet using the method of Vali (1971). The total INP number concentration was found by summing the INP number concentrations over all analyzed MOUDI stages. INP number concentrations have been adjusted to standard temperature and pressure.

### 5 2.3 Total and fluorescent aerosol measurements with sizes $\geq 0.5 \mu\text{m}$

A model-4A waveband integrated bioaerosol sensor (WIBS-4A; Droplet Measurement Technologies, Boulder, CO, USA) was used to find both the total and fluorescent aerosol number concentrations with sizes  $\geq 0.5 \mu\text{m}$ . Particles that enter the WIBS-4A first transect a continuous-wave 635 nm diode laser. The forward-scattered light from 10 the continuous-wave laser is detected with a quadrant photomultiplier tube for the determination of particle size and asymmetry factor based on the signal intensity and asymmetry, respectively. The detected forward-scattered light also triggers excitation pulses from xenon lamps, the first at a wavelength of 280 nm and the second at 370 nm. The excitation pulses may lead to fluorescent emission from the particle, which is then 15 collected in two wavelength ranges: 310–400 nm (short wavelength region) and 420–650 nm (long wavelength region). This results in sample information provided for each particle in three fluorescence channels: excitation at 280 nm, emission in the short wavelength region (FL1); excitation at 280 nm, emission in the long wavelength region (FL2); and excitation at 370 nm, emission in the long wavelength region (FL3). Detailed 20 descriptions of the instrument can be found in Kaye et al. (2005), Gabey et al. (2010), and Healy et al. (2012a). The sample and total flow rates of the WIBS-4A were 0.63 and  $2.3 \text{ L min}^{-1}$ , respectively, and number concentrations have been adjusted to standard temperature and pressure.

The fluorescent channels used in the WIBS-4A allow for the detection of fluorophores characteristic of biological activity. These fluorophores include the amino acid tryptophan, the cofactor NAD(P)H, and the micronutrient riboflavin. While some non-biological species can produce a fluorescent signal (Pan et al., 1999; Sivaprakasam et al., 2004; Bones et al., 2010; Pöhlker et al., 2012; Lee et al., 2013), the number of 25

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)

◀

▶

[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)



[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

fluorescent particles is generally considered to be a lower limit to the number of primary biological particles (Huffman et al., 2010, 2012; Pöhlker et al., 2012). In addition, fluorescence microscopy measurements of samples collected during this field study show high concentrations of fluorescent biological particles (see below). Therefore, fluorescent particles detected using the WIBS-4A are hereafter referred to as fluorescent bioparticles.

## 2.4 Fluorescence microscopy

Aerosol samples were collected onto glass cover slips using a custom single-stage impactor operating at a flow rate of  $1.2 \text{ L min}^{-1}$  with a 50 % cutoff aerodynamic diameter of  $0.5 \mu\text{m}$ . Prior to sample collection, the substrates were coated with a thin layer of high viscosity grease (Baysilone grease, Bayer, Germany) to reduce particle bounce.

Fluorescence microscopy images were taken on a BZ-9000 fluorescence microscope (Keyence, Inc., Osaka, Japan) equipped with a 120 W super high-compression mercury lamp and a 1.5 megapixel monochrome CCD camera. Images were obtained using the following fluorescence filters: OP-66834 DAPI-BP ( $\lambda_{\text{ex}} = 360/20 \text{ nm}$ ,  $\lambda_{\text{dichroic}} = 400 \text{ nm}$ ,  $\lambda_{\text{abs}} = 460/25 \text{ nm}$ ), OP-66836 GFP-BP ( $\lambda_{\text{ex}} = 470/20 \text{ nm}$ ,  $\lambda_{\text{dichroic}} = 495 \text{ nm}$ ,  $\lambda_{\text{abs}} = 535/25 \text{ nm}$ ), and OP-66838 TexasRed ( $\lambda_{\text{ex}} = 560/20 \text{ nm}$ ,  $\lambda_{\text{dichroic}} = 595 \text{ nm}$ ,  $\lambda_{\text{abs}} = 630/30 \text{ nm}$ ). Filter specifications are given as wavelength of maximum absorbance or excitation and full width at half maximum ( $\lambda/\text{FWHM}$ ).

## 2.5 Black carbon (BC) measurements

BC mass concentrations were measured using a multi-angle absorption photometer (MAAP model 5012; Thermo Scientific, Franklin, MA, USA). Detailed descriptions of the MAAP are available in Petzold et al. (2002), Petzold and Schönlinner (2004), and Petzold et al. (2005). Within the MAAP, particles are continuously collected on a glass fiber filter. The intensity of transmitted and forward-scattered light through the aerosol particle layer and filter matrix is measured by a photodetector located beneath the filter

at a frequency of 1 Hz. The signal strength is attenuated by the presence of both light-absorbing particles and particles that cause backscattering. As the angular distribution of back-scattered light is related to the fraction of non-absorbing particles (Petzold and Schönlinner, 2004), four additional photodetectors located above the filter are used to quantify the non-absorbing component of the sample. The absorbance by the collected aerosol is then related to a mass of BC using a mass-specific absorption coefficient of  $6.6 \text{ m}^2 \text{ g}^{-1}$ . Mass concentrations have been adjusted to standard temperature and pressure.

Non-BC material such as mineral dusts and brown carbon can also absorb 670 nm wavelength light used in the MAAP, albeit with smaller absorption coefficients than BC (Yang et al., 2009). We follow the recommendation of Petzold et al. (2013) for BC data derived from optical absorption methods and hereafter refer to MAAP data as measurements of equivalent black carbon (eBC).

## 2.6 Ion measurements

Size-resolved aerosol samples were collected on Teflon® filters (Pall Corporation, Port Washington, NY, USA) using a second MOUDI (model 110R). Samples were collected on the inlet, stage 1, and stages 7–10 of the MOUDI with stages 2–6 being removed prior to collection. The flow rate through the MOUDI was on average  $24 \text{ L min}^{-1}$ , resulting in a collected size range of 0.068 to  $> 20 \mu\text{m}$  (50 % cutoff aerodynamic diameter). Collection times ranged from approximately 45–49 h and samples were stored at  $4^\circ\text{C}$  for a period of one month before analysis.

Mass concentrations of sodium and methanesulfonic acid (MSA) were found using cationic and anionic chromatography following the method of Phinney et al. (2006). Briefly, filters were extracted with sonication in 10 mL of deionized water for 1 h, and samples were analyzed with a Dionex DX600 ion chromatograph using an AS11-HC column and a CS12 column for anions and cations, respectively. Filter blanks were measured to be below the limit of detection for both analytes. Mass concentrations were adjusted to standard temperature and pressure.

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

## 2.7 Back trajectories

Back trajectories spanning a period of 72 h were calculated for each sampling period using the Hybrid Single-Particle Lagrangian Integrated Trajectory (HYSPLIT4) model of the National Oceanographic and Atmospheric Administration and the GDAS1 meteorological data archive (Draxler and Rolph, 2014). To determine if the air mass changed during a sampling period, back trajectories were initiated at the beginning of the sampling period and every 2 h until the end of the sampling period. Back trajectories were used to assign each sampling period to one of four general air mass categories: (i) *coastal NW* where boundary layer air (defined here as an altitude below 1000 m) has traversed land northwest of the sampling site during its approach; (ii) *coastal SE* where boundary layer air has traversed land southeast of the sampling site during its approach; (iii) *Pacific Ocean* where boundary layer air has approached directly from the ocean and has not encountered land prior to arrival at the sampling site; and (iv) *free troposphere* where the air mass has spent more than 50 % of the 72 h back trajectory in the free troposphere. In four sampling periods, back trajectories initiated at different times in the sampling period indicated that the air mass changed during sampling, such as a change in the predominant altitude of the air mass from the free troposphere to the marine boundary layer. In these situations, the air mass category to which the majority of the back trajectories belonged was selected as the air mass category of the sample.

## 20 3 Results and discussion

### 3.1 Back trajectories and the dependence of INP concentrations on air mass classification

Back trajectories initiated at the midpoint of each INP sampling period are shown in Fig. 2. The back trajectories indicate that 88 % of the air masses sampled spent the majority of their 72 h prior to reaching the site over the Pacific Ocean within the marine

<a href="#">Title Page</a>	
<a href="#">Abstract</a>	<a href="#">Introduction</a>
<a href="#">Conclusions</a>	<a href="#">References</a>
<a href="#">Tables</a>	<a href="#">Figures</a>
<a href="#"></a>	<a href="#"></a>
<a href="#"></a>	<a href="#"></a>
<a href="#">Back</a>	<a href="#">Close</a>
<a href="#">Full Screen / Esc</a>	
<a href="#">Printer-friendly Version</a>	
<a href="#">Interactive Discussion</a>	



boundary layer (est. < 1000 m). Furthermore, air masses approached the sampling site from an onshore direction with minimal flow over land apart from coastal regions. Average local wind directions of 89 to 297° during INP sampling support this finding.

Shown in Fig. 3 is the number concentration of INPs as a function of time, color-coded by the classification of the air mass. At ice-activation temperatures between –15 and –25 °C there is no obvious trend with the type of the air mass. At –30 °C, INP number concentrations associated with air masses from the coastal SE (red points) appear to be higher than INP number concentrations associated with other air masses, but the statistics are low for the coastal SE air masses, especially at –30 °C. Figure 4 shows that the mean values for the different air mass types vary by less than a factor of 2.6. We conclude that INP number concentrations did not exhibit a strong dependence on the type of air mass sampled. The correlation analysis presented in Sects. 3.2–3.5 uses the entire dataset (i.e. the data were not differentiated based on air mass type). We further explore the dependence on air mass type in Sect. 3.6.

### 3.2 Are biological particles a major source of ice nuclei?

To investigate if biological particles are an important source of INPs at the coastal site, we determined correlations between INPs and fluorescent bioparticles. In the following correlation analysis, WIBS-4A data are limited to particle sizes of 10 µm or smaller to better match the size range of the MOUDI-DFT. The correlation coefficients ( $R$ ) of linear fits to the data are presented in Table 1 with correlation plots at a freezing temperature of –25 °C shown in Fig. 5 and plots at –15, –20, and –30 °C given in the Supplement. Here we use the scheme of Dancey and Reidy (2011) where correlations with an  $R$  value of 0.1–0.3, 0.4–0.6, and 0.7–0.9 are classified as weak, moderate, and strong, respectively. Only correlations with statistical significance ( $P$  value < 0.05) are discussed.

With  $R$  values between 0.74 and 0.83, INP number concentrations are strongly correlated with the number concentrations of fluorescent bioparticles for INPs active between –15 and –25 °C (Figs. 5a and S1 in the Supplement, Table 1). At these tem-

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

peratures, fluorescent bioparticles have the largest correlation coefficients with INPs compared to all of the other parameters investigated. This suggests that biological particles are an important component of the INP population. Using similar fluorescence techniques, others have also noted strong correlations between INPs and primary biological particles during ambient measurements (Prenni et al., 2009b, 2013; Huffman et al., 2013; Tobo et al., 2013).

To further investigate the relationship between biological particles and INPs, we compared the size distributions of INPs with the size distributions of total particles and fluorescent bioparticles, using samples where all three measurements were available.

Shown in Fig. 6a–d are the average number concentrations of INPs as a function of particle size for droplet freezing temperatures ranging from –15 to –30 °C. The shapes of all four INP size distributions were nearly identical with a single mode at an aerodynamic diameter of 3.2–5.6 µm.

Also shown in Fig. 6 are the average size distributions of total particles and fluorescent bioparticles as measured with the WIBS-4A over the size range of 0.5–10 µm. Due to the decrease in WIBS counting efficiency at particle sizes below approximately 0.7 µm (Healy et al., 2012b), the number concentration of particles sized 0.5–1.0 µm should be considered a lower limit. The size distribution of total particles (Fig. 6e) was found to be unimodal with the mode at 0.5–1.0 µm. Fluorescent bioparticles were bimodally distributed (Fig. 6f) with one mode at 1.8–3.2 µm and another at 0.5–1.0 µm. Figure 6 illustrates that the size distributions of INPs are more closely related to the size distribution of fluorescent bioparticles than total particles, suggesting that biological particles may have had a greater contribution to the INP population than non-biological particles.

In addition to the WIBS-4A, the presence of biological material in sampled air was verified by fluorescence microscopy. Images of a sample collected on 11 August 2013 are shown in Fig. 7 as an example. The fraction of particles exhibiting fluorescence on this day based on the WIBS-4A was close to the campaign average value; 7.1 % vs. an average of 7.8 %. The image here shows a sample containing many biological parti-

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)

◀

▶

[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)



cles, identified by their blue color which is characteristic of biological fluorophores such as proteins and coenzymes (Pöhlker et al., 2012). Most of these biological particles had a similar morphology with an ellipsoidal shape, approximately 11.9 µm in length × 4.1 µm in width, and multi-nucleation with three septa. Morphologically, many of these  
5 appear to be fungal macroconidia, consistent with the physical attributes of ascospores (Carlile et al., 2001; Leslie and Summerell, 2006; Webster and Weber, 2007; Maheshwari, 2011). Fungal spores can be ice-active at the temperatures used here (Jayaweera and Flanagan, 1982; Pouleur et al., 1992; Tsumuki et al., 1992; Richard et al., 1996; Iannone et al., 2011; Haga et al., 2013, 2014; Fröhlich-Nowoisky et al., 2015), and  
10 the size of the bioparticles observed in Fig. 7 (an estimated aerodynamic diameter of 4.8 µm assuming a prolate spheroid shape and unit density) matches the mode in the INP size distributions of Fig. 6. Predicting the optical diameter that the WIBS-4A would measure for such a particle is difficult, but it is reasonable that they could be detected as slightly larger or smaller depending on the axis on which the incident light impinges.

### 15 3.3 Is black carbon a major source of ice nuclei?

Sources of BC at the sampling site include local marine ship traffic. Atmospheric size distributions obtained at other locations demonstrate that most BC particles are smaller than 1 µm (Schwarz et al., 2008, 2013; Schroder et al., 2015). As shown in Fig. 6, the majority of INPs identified here were larger than 1 µm at all of the temperatures  
20 studied. It is therefore likely that BC particles were not the major source of INPs at the sampling site. As correlations between INPs and eBC are moderate at –15 to –25 °C ( $R = 0.47\text{--}0.60$ , Table 1), we also investigated correlations between INPs and CO, NO<sub>x</sub>, and SO<sub>2</sub> because their sources, like BC, include fossil fuel combustion and biomass burning (Galanter et al., 2000; Gadi et al., 2003; United States Environmental  
25 Protection Agency, 2014). The correlations between INPs and CO, NO<sub>x</sub>, and SO<sub>2</sub> are not statistically significant (see Table S2 of the Supplement), further suggesting that BC was not a major INP source.

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

### 3.4 Are particles from the ocean a major source of ice nuclei?

Situated in a region of high oceanic primary productivity (Whitney et al., 2005; Ribalet et al., 2010) with onshore winds, particles of marine origin are a potential source of INPs at the sampling site. Therefore, correlations between INP number concentrations and tracers of marine aerosols and marine biological activity were explored. Since primary marine aerosols are ejected from the ocean by the bursting of entrained bubbles (Blanchard and Woodcock, 1957; Blanchard, 1963, 1989; Andreas, 1998), sodium was used as a tracer of primary particles from the ocean. The strength of correlations between INPs and sodium are given in Table 1. Although the correlations range from weakly-to-moderately negative to strongly positive, the large  $P$  values (0.20 or greater) indicate that the results are not statistically significant. Due in part to the long sampling times required for the sodium measurements, only three to six data points were available for the sodium correlation analysis.

MSA is often used as a marker for marine biological productivity (Saltzman et al., 1986; Savoie et al., 1994; Sorooshian et al., 2009; Gaston et al., 2010; Becagli et al., 2013) because it is chemically stable and its precursor, dimethylsulfide, is produced by primary biological activity in the ocean (Andreae et al., 1985; Charlson et al., 1987; Keller, 1989; Bates et al., 1992; Kettle et al., 1999). As INP number concentrations are closely correlated to bioparticles at warmer droplet freezing temperatures, one may expect correlations of a similar magnitude between INPs and MSA if the marine environment was indeed acting as an important source of biological INPs. As is shown in Table 1, no statistically significant correlations are found as  $P$  values are large (0.15–0.50).

Finally, correlations between wind speed and INP number concentration were investigated. As the dominant source of bubble entrainment in the ocean is breaking waves (O'Dowd and de Leeuw, 2007), the rate of sea-spray production is dependent in part on wind speed. For this correlation, wind speed was first raised to the power of 3.41 using the power law of Monahan and Muircheartaigh (1980) that relates whitecap cov-

<a href="#">Title Page</a>	
<a href="#">Abstract</a>	<a href="#">Introduction</a>
<a href="#">Conclusions</a>	<a href="#">References</a>
<a href="#">Tables</a>	<a href="#">Figures</a>
<a href="#"></a>	<a href="#"></a>
<a href="#"></a>	<a href="#"></a>
<a href="#">Back</a>	<a href="#">Close</a>
<a href="#">Full Screen / Esc</a>	
<a href="#">Printer-friendly Version</a>	
<a href="#">Interactive Discussion</a>	

verage to wind speed. The correlation found at  $-30^{\circ}\text{C}$  is statistically significant ( $P$  value  $< 0.05$ ), but the magnitude of the correlation coefficient is only moderate ( $R = 0.48$ ; see Table 1). The average wind speed during INP sampling exceeded the onset speed for whitecap formation, approximately  $4 \text{ m s}^{-1}$  (O'Dowd and de Leeuw, 2007), in only 47% of samples and daily observations at the site noted infrequent wave activity. Furthermore, some of the highest INP concentrations were found when the wind speed was less than  $4 \text{ m s}^{-1}$ .

All correlations between INPs and parameters indicative of marine aerosols and marine biological activity are either moderate at best or not statistically significant. For these reasons, correlations involving sodium, MSA, and wind speed do not provide strong evidence that marine particles were a major contributor to the INP population. Recent measurements have shown the presence of INPs in the sea surface microlayer (Wilson et al., 2015). Our measurements do not contradict these findings, since we do not rule out the ocean as a source of INPs. One possibility is that biological INPs released by local vegetation were present in sufficient numbers at this site to overwhelm the presence of any INPs from the ocean.

### 3.5 What is the major source of ice nuclei active at $-30^{\circ}\text{C}$ ?

At warmer droplet freezing temperatures ( $-15$  to  $-25^{\circ}\text{C}$ ), the strongest correlations are observed between number concentrations of fluorescent bioparticles and INPs. In contrast, at  $-30^{\circ}\text{C}$  the strength of correlations between INPs and fluorescent bioparticles and INPs and total particles  $> 0.5 \mu\text{m}$  in diameter are equal ( $R = 0.66$ ; Table 1). It is therefore likely that both biological and non-biological particles were important sources of INPs active at  $-30^{\circ}\text{C}$ . Good correlations between INPs and total particles  $> 0.5 \mu\text{m}$  have also been observed in several other field studies (e.g. DeMott et al., 2010; Chou et al., 2011; Field et al., 2012; Prenni et al., 2013; Tobo et al., 2013; Jiang et al., 2015).

Since the INP size distributions of Fig. 6 and the correlations of Table 1 do not provide strong evidence of BC particles or the ocean being a major source of INPs active at  $-30^{\circ}\text{C}$ , it is possible that mineral dust was a major source of INPs as mineral dust

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

particles are known to efficiently nucleate ice at this temperature (e.g. DeMott et al., 2003; Cziczo et al., 2004; Field et al., 2006; Möhler et al., 2006; Marcolli et al., 2007; Zimmermann et al., 2008; Klein et al., 2010; Niedermeier et al., 2010; Chou et al., 2011; Atkinson et al., 2013; Yakobi-Hancock et al., 2013; Wheeler et al., 2015). The size distribution of INPs did not drastically change between  $-25$  and  $-30^{\circ}\text{C}$  (Fig. 6c and d), and the dominant mode in the surface area distribution of airborne mineral dust (Maring et al., 2003) can occur at approximately the same size range as biological INPs (Després et al., 2012). While in a very different ecosystem and climatic region, Prenni et al. (2009b) noted that the relative contribution of mineral dust particles to the total number of INPs in the Amazon region increased as ice nucleation temperature decreased. Only below  $-27^{\circ}\text{C}$  did the amount of mineral dust significantly influence the number of INPs, while above this temperature most INPs were biological (Prenni et al., 2009b).

### 3.6 Do the potential sources of ice nuclei change with air mass classification?

In the preceding sections we did not differentiate data based on air mass classification. Here we present correlations within each of the four air mass categories introduced in Sect. 2.7 to investigate if the major sources of INPs vary with air mass type. The correlations for each air mass type are given in Table 2. Correlations involving sodium and MSA are not included due to insufficient data, and only statistically significant correlations will be discussed ( $P < 0.05$ ).

The general trends presented in Table 1 for the undifferentiated data are also found in Table 2 for the various air mass categories. In coastal NW, Pacific Ocean, and free tropospheric air masses, INP number concentrations are well correlated to those of fluorescent bioparticles at temperatures between  $-15$  and  $-25^{\circ}\text{C}$  with  $R$  values ranging from 0.64 to 0.99 (an average of 0.89), and in free tropospheric air masses a very strong correlation is also found at  $-30^{\circ}\text{C}$  ( $R = 1.00$ ). In most cases, these are the strongest correlations noted at a given temperature. This again suggests that many INPs may have been biological.

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)  
[Back](#)[▶](#)  
[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

In coastal NW and free tropospheric air masses, INPs and total particles are also closely correlated. These correlations are strong in the case of coastal NW air masses at ice activation temperatures of  $-15$  to  $-25$  °C ( $R = 0.70\text{--}0.85$ ) and very strong in air masses from the free troposphere between  $-20$  and  $-30$  °C ( $R = 0.98\text{--}0.99$ ). The correlation coefficients are significantly greater than those found in the undifferentiated data of Table 1. With the average fraction of particles that exhibited fluorescence in these air masses being close to the campaign average, the good correlations with total particles suggest that non-biological INPs such as mineral dust may have also contributed to the INP population.

Correlations of INPs with eBC are strong ( $R = 0.71\text{--}0.84$ ) at  $-25$  °C and above in coastal NW air masses and very strong ( $R = 0.99$ ) at  $-15$  °C in air masses from the free troposphere. Correlations of INPs with CO and  $\text{SO}_2$  in these air masses are also moderate to very strong in some cases (see Table S3 of the Supplement). However, more than 84 and 100 % of INPs active at these temperatures were larger than  $1 \mu\text{m}$  in size in air masses from the coastal NW and the free troposphere, respectively. Given the dominance of supermicron INPs in these two air mass types, it is unlikely that BC was an important source of INPs.

### 3.7 Can existing parameterizations accurately predict measured INP concentrations?

Empirical parameterizations have been developed to predict ice nucleation in atmospheric models. Here we investigate whether or not these parameterizations are consistent with the current measurements. In total we tested six different parameterizations: those of Fletcher (1962), hereafter F62; Cooper (1986), hereafter C86; Meyers et al. (1992), hereafter M92; DeMott et al. (2010), hereafter D10; and two from Tobo et al. (2013), hereafter T13<sub>total</sub> and T13<sub>fluorescent</sub>. Details on these parameterizations are given in the Supplement.

In Fig. 8 we compare measured INP number concentrations with predicted INP number concentrations based on the parameterizations discussed above. The parameter-

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

izations of D10, T13<sub>total</sub> and T13<sub>fluorescent</sub> require knowledge of either total particle or fluorescent bioparticle number concentrations with sizes  $> 0.5 \mu\text{m}$ . Here we use the data from the WIBS-4A over its full size range ( $0.5\text{--}23.7 \mu\text{m}$ ). Note the parameterization of T13<sub>fluorescent</sub> based on fluorescent bioparticle number concentrations was formulated using measurements from an ultraviolet aerodynamic particle sizer (UV-APS), whereas this study uses a WIBS-4A. As noted in Healy et al. (2014), there may be discrepancies between the number concentrations of fluorescent bioparticles detected by the UV-APS and WIBS-4A. With more fluorescent channels and more sensitive electronics, the WIBS-4A may probe different fluorophores than the UV-APS, thus detecting greater concentrations of fluorescent bioparticles and in turn leading to greater predicted INP number concentrations. Also, the INP number concentrations measured by the MOUDI-DFT are for particle sizes of  $0.18\text{--}10 \mu\text{m}$ , whereas the INP measurements used to formulate the parameterizations of M92, D10, and T13 were for particles  $\leq 3$ ,  $\leq 1.6$ , and  $\leq 2.4 \mu\text{m}$ , respectively. As a result, when reporting measured INP number concentrations in Fig. 8 we limit the MOUDI-DFT data to particle sizes that overlap with those used to formulate the parameterizations (see the Supplement for details).

It is evident in Fig. 8 that none of the parameterizations are able to consistently predict the measured INP number concentrations within a factor of 5 over the entire temperature range investigated. The most accurate parameterization is that of C86 (Fig. 8b), predicting 25 and 57 % of the INP number concentrations within a factor of 2 and 5, respectively, of the solid 1 : 1 line. While the C86 parameterization works reasonably well at temperatures of  $-15$  to  $-25^\circ\text{C}$ , at lower temperatures it becomes increasingly inaccurate, possibly due to it being applied outside the temperature range over which it was developed ( $-5$  to  $-25^\circ\text{C}$ ).

The parameterizations of D10, T13<sub>total</sub> and T13<sub>fluorescent</sub> incorporate measurements of total particles or fluorescent bioparticles, but are found to be poor predictors of the values measured in this study as on average only 41 % of INP number concentrations are predicted within a factor of 5 (Fig. 8d–f). A number of datasets from diverse locations were used in the development of the D10 parameterization, but those with

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

a strong marine influence were not included because sea salt is not known to be an efficient ice nucleus. Given the proximity of our sampling site to the Pacific Ocean (Fig. 1) and the back trajectories of the sampled air masses (Fig. 2), a marine influence in our samples may contribute to the somewhat poor performance of the D10 parameterization and the over-estimation of INPs shown in Fig. 8d. The T13<sub>total</sub> and T13<sub>fluorescent</sub> parameterizations were developed using data from a forested site in Colorado. Differences in the composition, concentration, and ice nucleating ability of both biological and non-biological particles between the continental forest of T13 and the coastal site of this study may have contributed to the inaccuracy of the T13<sub>total</sub> and T13<sub>fluorescent</sub> parameterizations (Fig. 8e and f).

## 4 Summary and conclusions

The number concentrations of 0.18–10 µm INPs active in the immersion mode were determined at a coastal site in Western Canada during the summer of 2013 as part of the NETCARE project. We investigated the strength of linear correlations between these INP values and measurements of total particles, fluorescent bioparticles, eBC, sodium, MSA, and wind speed and also compared their size distributions where these measurements were available. We found that (1) biological particles, possibly from local vegetation, were likely the major source of ice nuclei at freezing temperatures between –15 and –25 °C; (2) non-biological particles such as mineral dust may also have had an important contribution to the population of INPs active at –30 °C; (3) the prevalence of supermicron INPs makes BC particles an unlikely source of ice nuclei; and (4) there was no evidence of marine particles being a significant source of ice nuclei, although the ocean as a source of INPs cannot be ruled out. One possibility is that biological INPs released by local vegetation were present in sufficient numbers at this site to overwhelm the presence of any INPs from the ocean.

Six empirical parameterizations of ice nucleation for use in atmospheric models were tested to determine the accuracy with which they predict INP number concentrations

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

at this coastal site. Overall, none of the parameterizations were found to be suitable, predicting only 1 to 57 % of INPs within a factor of 5 of the measured value. This highlights the need for the development of INP parameterizations that are appropriate for this complex environment.

- 5    **The Supplement related to this article is available online at  
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Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



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## Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)

◀

▶

◀

▶

[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)



**Ice nucleating  
particles at a coastal  
marine boundary  
layer site**

R. H. Mason et al.

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

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## Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)



[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)



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R. H. Mason et al.

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)

◀

▶

[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)



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Discussion Paper | Discussion Paper

Discussion Paper | Discussion Paper

Discussion Paper | Discussion Paper

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)



[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)



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## Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)

◀

▶

◀

▶

[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)



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## Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)

◀

▶

[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)



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Ice nucleating  
particles at a coastal  
marine boundary  
layer site

R. H. Mason et al.

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)

◀

▶

◀

▶

[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)



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Ice nucleating  
particles at a coastal  
marine boundary  
layer site

R. H. Mason et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures



◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



**Ice nucleating  
particles at a coastal  
marine boundary  
layer site**

R. H. Mason et al.

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Discussion Paper | Discussion Paper

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

## Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.

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Discussion Paper

Discussion Paper

Discussion Paper

Discussion Paper

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)



[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)



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## Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)

◀

▶

[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)



## Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)

◀

▶

◀

▶

[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)



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[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

## Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.

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Discussion Paper

Discussion Paper

Discussion Paper

Discussion Paper

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)



[◀](#)

[▶](#)

[◀](#)

[▶](#)

[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)



## Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.

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Title Page	
Abstract	Introduction
Conclusions	References
Tables	Figures
 	 
Back	Close
Full Screen / Esc	
Printer-friendly Version	
Interactive Discussion	

## Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.

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Discussion Paper | Discussion Paper

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)



[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)



---

Ice nucleating  
particles at a coastal  
marine boundary  
layer site

R. H. Mason et al.

---

<a href="#">Title Page</a>	
<a href="#">Abstract</a>	<a href="#">Introduction</a>
<a href="#">Conclusions</a>	<a href="#">References</a>
<a href="#">Tables</a>	<a href="#">Figures</a>
<a href="#"></a>	<a href="#"></a>
<a href="#"></a>	<a href="#"></a>
<a href="#">Full Screen / Esc</a>	
<a href="#">Printer-friendly Version</a>	
<a href="#">Interactive Discussion</a>	

## Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.

**Table 1.** Correlation coefficients ( $R$ ) for linear regression analyses of INPs vs. fluorescent bioparticles, total aerosol particles, eBC, sodium, MSA, and wind speed<sup>a</sup>. Correlations with statistical significance ( $P < 0.05$ ) are shown in bold.

Measurement	Relation to the INP number concentration											
	−15 °C			−20 °C			−25 °C			−30 °C		
	$R$	$P$ <sup>b</sup>	$n$ <sup>c</sup>	$R$	$P$	$n$	$R$	$P$	$n$	$R$	$P$	$n$
Fluorescent bioparticles [0.5–10 µm]	<b>0.74</b>	< 0.01	28	<b>0.77</b>	< 0.01	28	<b>0.83</b>	< 0.01	28	<b>0.66</b>	< 0.01	23
Total particles [0.5–10 µm]	<b>0.33</b>	0.04	28	<b>0.36</b>	0.03	28	<b>0.49</b>	< 0.01	28	<b>0.66</b>	< 0.01	23
eBC	<b>0.47</b>	< 0.01	34	<b>0.59</b>	< 0.01	34	<b>0.60</b>	< 0.01	34	0.25	0.11	27
Sodium	−0.35	0.25	6	0.13	0.40	6	0.32	0.27	6	0.82	0.20	3
MSA	0.17	0.38	6	0.51	0.15	6	0.27	0.30	6	0.00	0.50	3
(Wind speed) <sup>3.41</sup>	0.05	0.39	34	0.01	0.48	34	0.15	0.19	34	<b>0.48</b>	< 0.01	27

<sup>a</sup> Using the power law dependence of whitecap coverage on wind speed found by Monahan and Muircheartaigh (1980), wind speed was raised to the power of 3.41.

<sup>b</sup> The  $P$  value is a conditional probability that is the probability of obtaining an  $R$  value equal to or greater than the given  $R$  value if there is no correlation between INPs and the given parameter.

<sup>c</sup>  $n$  represents the number of data points used in determining the correlation.

- [Title Page](#)
- [Abstract](#) | [Introduction](#)
- [Conclusions](#) | [References](#)
- [Tables](#) | [Figures](#)
- [◀](#) | [▶](#)
- [◀](#) | [▶](#)
- [Back](#) | [Close](#)
- [Full Screen / Esc](#)
- [Printer-friendly Version](#)
- [Interactive Discussion](#)



## Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.

Title Page	
Abstract	Introduction
Conclusions	References
Tables	Figures
Back	Close
Full Screen / Esc	

Printer-friendly Version

Interactive Discussion



**Table 2.** Correlation coefficients ( $R$ ) for linear regression analyses of INPs vs. fluorescent bioparticles, total aerosol particles, eBC, and wind speed<sup>a</sup> within each category of air mass. Correlations with statistical significance ( $P < 0.05$ ) are shown in bold.

Air Mass	Measurement	Relation to the INP number concentration											
		−15 °C			−20 °C			−25 °C			−30 °C		
		$R$	$P$ <sup>b</sup>	$n$ <sup>c</sup>		$R$	$P$	$n$		$R$	$P$	$n$	
Coastal NW	Fluorescent bioparticles [0.5–10 µm]	<b>0.94</b>	< 0.01	9	<b>0.94</b>	< 0.01	9	<b>0.96</b>	< 0.01	9	0.65	0.08	6
	Total particles [0.5–10 µm]	<b>0.85</b>	< 0.01	9	<b>0.70</b>	0.02	9	<b>0.71</b>	0.02	9	0.67	0.07	6
	eBC	<b>0.71</b>	< 0.01	11	<b>0.80</b>	< 0.01	11	<b>0.84</b>	< 0.01	11	0.53	0.11	7
	(Wind speed) <sup>3.41</sup>	-0.38	0.12	11	-0.39	0.12	11	-0.22	0.26	11	0.26	0.29	7
Coastal SE	Fluorescent bioparticles [0.5–10 µm]	-0.07	0.48	3	-0.53	0.32	3	-0.85	0.17	3	NA <sup>d</sup>		
	Total particles [0.5–10 µm]	-0.17	0.45	3	-0.61	0.29	3	-0.90	0.14	3	NA		
	eBC	0.07	0.46	5	0.28	0.32	5	0.67	0.11	5	0.96	0.09	3
	(Wind speed) <sup>3.41</sup>	-0.34	0.29	5	-0.27	0.33	5	-0.14	0.41	5	0.21	0.43	3
Pacific Ocean	Fluorescent bioparticles [0.5–10 µm]	<b>0.80</b>	< 0.01	12	<b>0.74</b>	< 0.01	12	<b>0.64</b>	0.01	12	0.23	0.24	12
	Total particles [0.5–10 µm]	0.13	0.34	12	0.30	0.17	12	0.21	0.25	12	0.25	0.22	12
	eBC	0.24	0.21	14	0.37	0.10	14	0.26	0.19	14	0.06	0.42	13
	(Wind speed) <sup>3.41</sup>	-0.10	0.37	14	-0.26	0.18	14	-0.26	0.19	14	-0.21	0.25	13
Free troposphere	Fluorescent bioparticles [0.5–10 µm]	<b>0.97</b>	0.02	4	<b>0.99</b>	< 0.01	4	<b>0.99</b>	< 0.01	4	<b>1.00</b>	< 0.01	4
	Total particles [0.5–10 µm]	0.86	0.07	4	<b>0.98</b>	0.01	4	<b>0.99</b>	< 0.01	4	<b>0.98</b>	0.01	4
	eBC	<b>0.99</b>	< 0.01	4	0.89	0.05	4	0.88	0.06	4	0.89	0.06	4
	(Wind speed) <sup>3.41</sup>	-0.89	0.05	4	-0.70	0.15	4	-0.67	0.17	4	-0.68	0.16	4

<sup>a</sup> Using the power law dependence of whitecap coverage on wind speed found by Monahan and Muircheartaigh (1980), wind speed was raised to the power of 3.41.

<sup>b</sup> The  $P$  value is a conditional probability that is the probability of obtaining an  $R$  value equal to or greater than the given  $R$  value if there is no correlation between INPs and the given parameter.

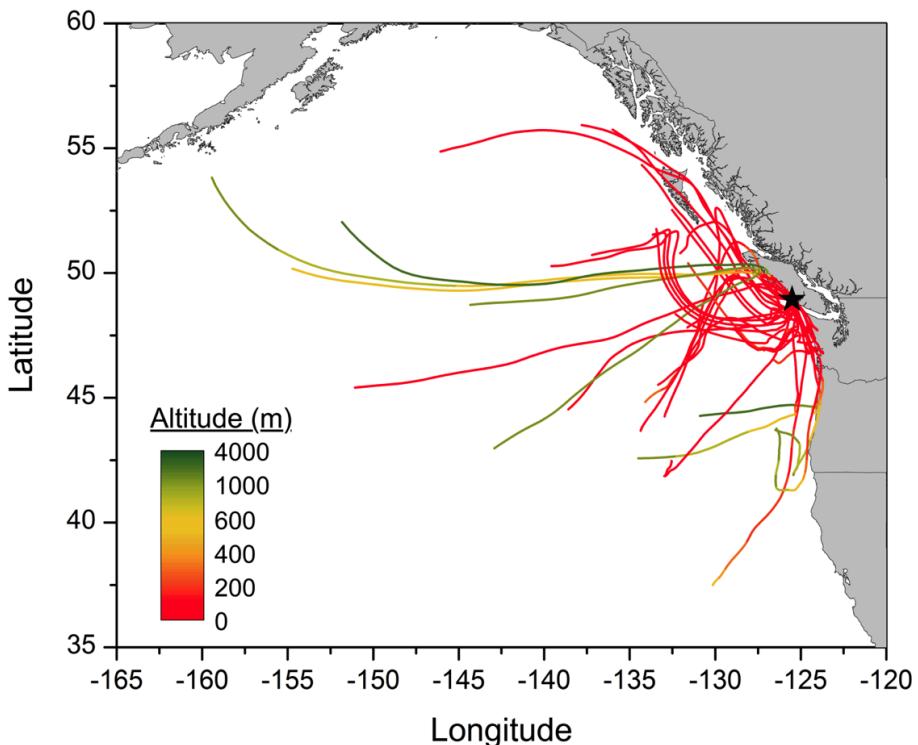
<sup>c</sup>  $n$  represents the number of data points used in determining the correlation.

<sup>d</sup> NA = not available due to insufficient data.

**Figure 1.** A satellite image of the sampling site: (1) location of the MOUDIs and the WIBS-4A; (2) location of the MAAP; and (3) Amphitrite Lighthouse where most meteorological data was collected; and (4) a station of the Canadian Coast Guard with supporting infrastructure. The image was modified from Bing Maps, 2014 (<http://www.bing.com/maps/>). Inset: the location of the sampling site in British Columbia, Canada.

## Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.



**Figure 2.** Seventy-two hour HYSPLIT4 back trajectories of the air masses analyzed at the coastal site (black star) during INP sampling periods. Each back trajectory was initiated from a height of 5.5 m a.g.l. and at the midpoint of the sampling period.

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)

[◀](#)

[▶](#)

[◀](#)

[▶](#)

[Back](#)

[Close](#)

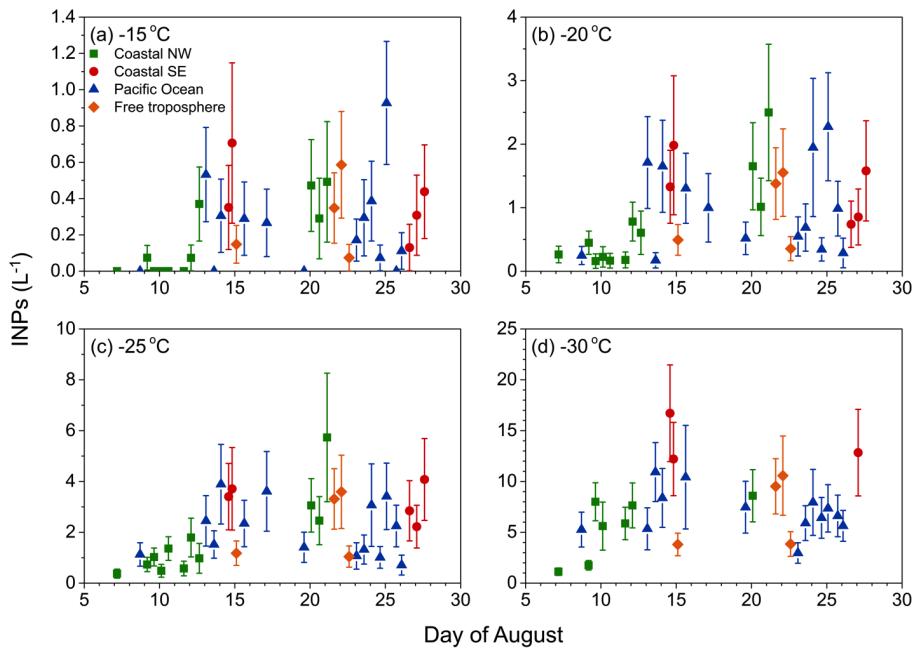
[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)

## Ice nucleating particles at a coastal marine boundary layer site

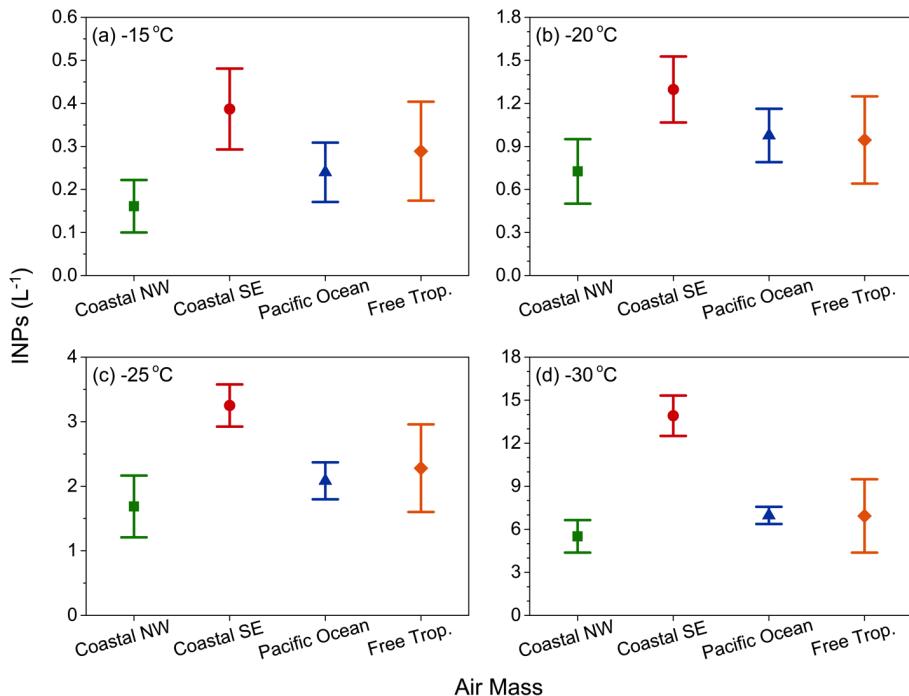
R. H. Mason et al.



**Figure 3.** INP number concentrations as a function of date determined at ice-activation temperatures of **(a)**  $-15$ , **(b)**  $-20$ , **(c)**  $-25$ , and **(d)**  $-30$   $^{\circ}\text{C}$ . The symbols are color coded by air mass category (see Sect. 2.7 for details). Fewer data points are available at  $-30$   $^{\circ}\text{C}$  as INP number concentrations can only be determined to the temperature where all droplets are frozen and Eq. (1) becomes undefined.

## Ice nucleating particles at a coastal marine boundary layer site

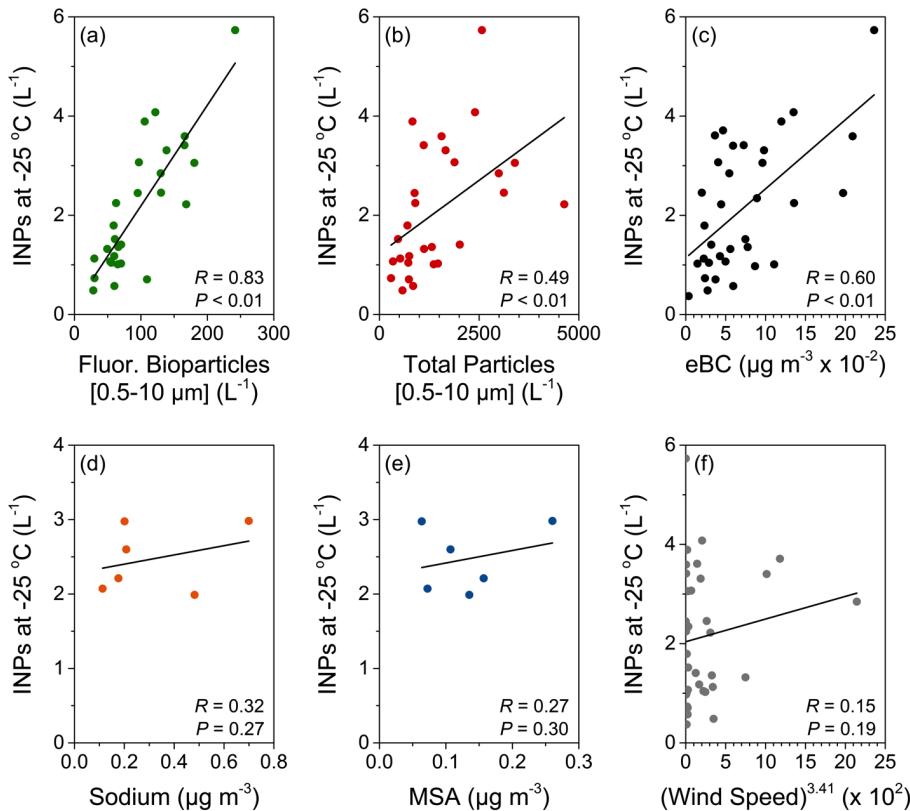
R. H. Mason et al.



**Figure 4.** Mean INP number concentrations found in each of the four categories of air masses sampled at **(a)**  $-15$ , **(b)**  $-20$ , **(c)**  $-25$ , and **(d)**  $-30$   $^{\circ}\text{C}$ . The scheme for air mass classification is given in Sect. 2.7. Uncertainties are given as the standard error of the mean.

## Ice nucleating particles at a coastal marine boundary layer site

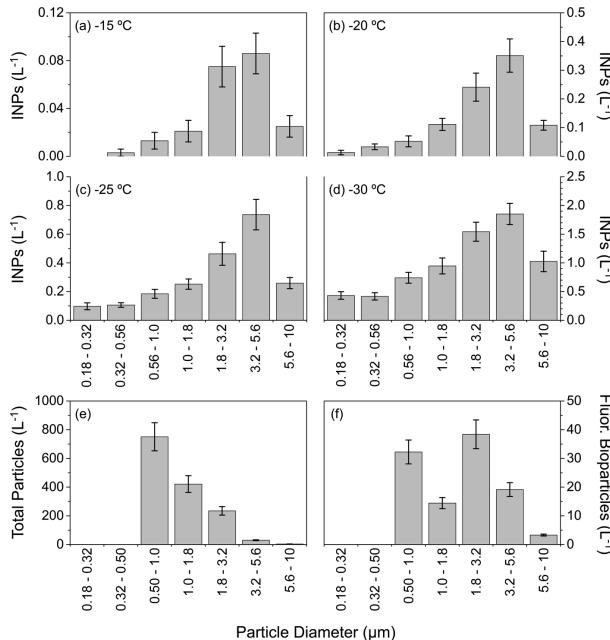
R. H. Mason et al.



**Figure 5.** Number concentrations of INPs active at  $-25^{\circ}\text{C}$  plotted against concentrations of (a) fluorescent bioparticles 0.5–10  $\mu\text{m}$ , (b) total particles 0.5–10  $\mu\text{m}$ , (c) eBC, (d) sodium, (e) MSA, and (f) (wind speed) $^{3.41}$  based on the power law function of Monahan and Muircheartaigh (1980) where wind speed was in units of  $\text{m s}^{-1}$ . Linear fits are shown with corresponding correlation coefficients ( $R$ ) and probability values ( $P$ ).

## Ice nucleating particles at a coastal marine boundary layer site

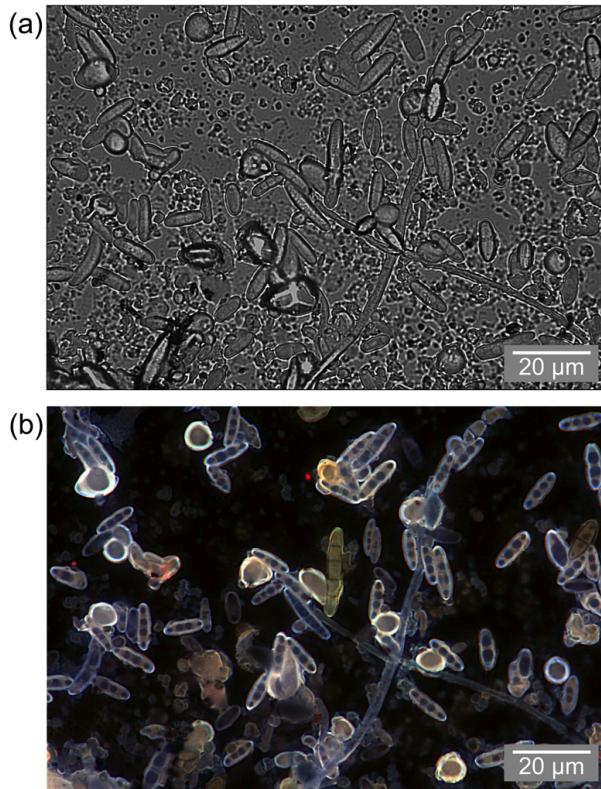
R. H. Mason et al.



**Figure 6.** Mean number concentrations as a function of size for INPs active at (a) –15, (b) –20, (c) –25, and (d) –30 °C, total particles (e) and fluorescent bioparticles (f) using only samples where both the MOUDI-DFT and WIBS-4A were operating. Uncertainties are given as the standard error of the mean. As INP number concentrations can only be determined at temperatures less than the temperature where all droplets are frozen and Eq. (1) becomes undefined, fewer samples are represented at –30 °C. Number concentrations below 0.5 μm were not measured by the WIBS-4A for panels (e) and (f) but plot axes are consistent for easier comparison of the size distributions.

**Ice nucleating  
particles at a coastal  
marine boundary  
layer site**

R. H. Mason et al.

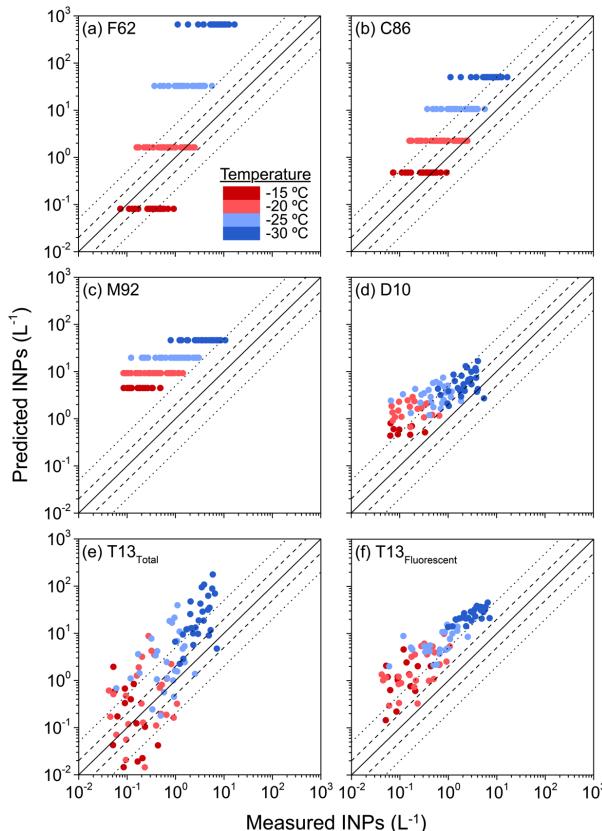


**Figure 7.** Fluorescence microscopy images of an aerosol sample collected on 11 August 2013: (a) bright-field image; (b) an overlay of red, green, and blue fluorescence channels. A blue coloration is characteristic of biological material (Pöhlker et al., 2012).

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

## Ice nucleating particles at a coastal marine boundary layer site

R. H. Mason et al.



**Figure 8.** Predicted vs. measured INP number concentrations based on the parameterizations of (a) Fletcher (1962); (b) Cooper (1986); (c) Meyers et al. (1992); (d) DeMott et al. (2010); and (e and f) Tobo et al. (2013). Details on these parameterizations are given in the Supplement. Data color represents ice nucleation temperatures. This figure uses the format of Fig. 9 in Tobo et al. (2013).

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion