Polycyclic aromatic hydrocarbons, polychlorinated biphenyls, and chlorinated pesticides in 1 2 background air in central Europe - investigating parameters affecting wet scavenging of polycyclic aromatic hydrocarbons 3 4 Pourya Shahpoury, ¹ Gerhard Lammel, ^{1,2} Adéla Holubová Šmejkalová, ^{3,4} Jana Klánová, ² Petra 5 Přibylová, Milan Váňa^{3,4} 6 7 ¹ Max-Planck Institute for Chemistry, Mainz, Germany 8 ² Research Centre for Toxic Compounds in the Environment, Masaryk University, Brno, Czech 9 Republic 10 ³ Czech Hydrometeorological Institute, Košetice, Czech Republic 11 ⁴ Global Change Research Centre, Academy of Sciences of the Czech Republic, Prague, Czech 12 Republic 13 14 **Abstract** 15 Concentrations of polycyclic aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs), 16 and chlorinated pesticides (CPs) were measured in air and precipitation at a background site in 17 central Europe. Σ PAH concentrations in air and rainwater ranged from 0.7 to 327.9 ng m⁻³ and 18 below analytical method detection limit (<MDL) to 2.1×10^3 ng L⁻¹. The concentrations of PCBs 19 and CPs in rainwater were <MDL. Σ PCB and Σ CP concentrations in air ranged from <MDL to 20 44.6 and <MDL to 351.7 pg m⁻³, respectively. The potential relationships between PAH wet 21 scavenging and particulate matter and rainwater properties were investigated. The concentrations 22 of ionic species in particulate matter and rainwater were significantly correlated, highlighting the 23 importance of particle scavenging process. Overall, higher scavenging efficiencies were found 24 for relatively less volatile PAHs, underlining the effect of analyte gas-particle partitioning on 25 scavenging process. The particulate matter removal by rain, and consequently PAH wet 26 scavenging, was more effective when the concentrations of ionic species were high. In addition, 27

the elemental and organic carbon contents of the particulate matter were found to influence the PAH scavenging.

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1 Introduction

Semi-volatile organic compounds (SOCs), such as polycyclic aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs), and chlorinated pesticides (CPs), can reach remote and background regions through atmospheric transport and deposition processes (Blais, 2005; Hageman et al., 2006; Grimalt et al., 2001). Wet scavenging is an important mechanism through which atmospheric SOCs enter terrestrial and aquatic ecosystems (van Ry et al., 2002). Nonreactive gaseous SOCs are removed from the atmosphere by gas scavenging process, which is controlled by the SOC equilibrium partitioning with falling raindrops, as described by Henry's law. In contrast, particle-associated species are removed from the atmosphere mainly through particle scavenging (Ligocki et al., 1985), a process controlled by physical parameters including particulate matter (PM) characteristics, cloud microphysics, and meteorological conditions (Poster and Baker, 1996). Therefore, factors that affect SOC gas-particle partitioning, such as ambient temperature, relative humidity, PM chemical composition and surface characteristics, and SOC vapor pressure (Lohmann and Lammel, 2004), may play an important role in determining the relative contributions of gas and particle scavenging processes. It was suggested that SOC scavenging efficiencies would be between one to three orders of magnitude higher in case of in-cloud scavenging compared to that of below-cloud scavenging (Ligocki et al., 1985). Potential variability in scavenging efficiencies has also been suggested between rain events originated from warm and cold clouds (Bidleman, 1988). A number of studies observed

variations in scavenging efficiencies of SOCs between different precipitation events (Offenberg and Baker, 2002; Agrell et al., 2002; Liu et al., 2013; Kaupp and McLachlan, 2000; McLachlan and Sellström, 2009; Atlas and Giam, 1988). To this date, studies have mainly focused on determining the relative importance of SOC gas and particle scavenging processes. However, to the best of our knowledge, no comprehensive study has been conducted to understand the causes behind the variability in scavenging efficiencies. Therefore, in this study we aimed at filling this gap by investigating the relationships between a set of aerosol and rainwater properties and SOC scavenging. The present study was conducted at a background site in central Europe, as a follow up with a previously published research (Škrdlíková et al., 2011) in which PAH scavenging efficiencies were determined at that site. The specific objectives of the present study were to (1) measure the concentrations of PAHs, PCBs, and CPs in air and rainwater samples, (2) estimate wet depositional fluxes of target compounds in the study area, (3) determine wet scavenging efficiencies and scavenged mass fractions of the analytes, and (4) investigate the potential relationships between analyte scavenging and meteorological parameters as well as aerosol and rain characteristics. PAHs, PCBs, and CPs were selected because they are widespread atmospheric contaminants and have been found in air at background locations across Europe (Dvorská et al., 2009; Gioia et al., 2007; Roots et al., 2010; Halse et al., 2011). In addition, these chemicals can persist in the environment due to their resistance to photolytic, chemical, and biological degradation (Yolsal et al., 2014).

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2 Experimental

2.1 Sampling site

The sampling was conducted at Košetice observatory located in an open land about 70 km southeast of Prague in the Czech Republic (Fig. S1 in the Supporting Information (SI)). The observatory is classified as a European background site (Dvorská et al., 2008). It is managed by the Czech Hydrometeorological Institute (CHMI) and is part of the European Monitoring and Evaluation Program. The area is covered with snow two to three months per year with the mean annual temperature and precipitation of 7.1 °C and 625 mm. The potential source of pollution in the region is the nearby highway (~ 7 km west of the observatory) connecting Prague to Brno. In addition, due to having dominant westerly winds, atmospherically transported contaminants from the industrial and urban areas of the Czech Republic, notably Prague, may contribute to the pollution in the area.

2.2 Sampling procedure

Rainwater and air samples were collected during the period of December 2011 to January 2014 (Table S1 in the SI). A total of 231 rain samples were collected using an automatic precipitation sampler (Baghira, Czech Republic) equipped with a 1-m² stainless steel collection funnel, a lid, and a 5-L glass collection bottle. The opening and closing of the lid is triggered by a precipitation sensor mounted on the sampler. During sampling, rain drains from the funnel and accumulates in the collection bottle. The collected sample volumes were between 0.5 and 1 L, depending on the amount of rainfall for each event. Larger or smaller volumes were discarded - the latter was not deemed to provide enough analyte mass for trace analysis.

Air samples were collected on quartz fiber filters (OFF) (Grade OM-A, 10.1 cm ID, Whatman, UK) and in polyurethane foam (PUF) plugs (55 cm ID, 50 cm height, Organika, Poland) using a Graseby Andersen PS-1 high-volume sampler (GA, USA) with a flow rate of 17 m³ h⁻¹. For each sampling period, two consecutive PUF plugs and one filter paper were deployed. A total of 162 gaseous and particulate air samples were collected during the period of study. Nevertheless, only samples from 54 events were considered for determining scavenging ratios because only these were followed by rain events. The collection of these samples started between 6 to 35hrs prior to the onset of rain (predicted by CHMI) and stopped when rainfall started (Table S2 in the SI). This sampling approach was chosen in order to sample rainwater from the same air mass as that of PM. In addition, PM has been suggested to be removed from a travelling air parcel by up to 8 and 23% in the first 24 and 72hrs after the onset of rain, respectively (Wiman et al., 1990; Škrdlíková et al., 2011). This means that prolonging the air sampling beyond the onset of rain would potentially lead to underestimating contaminant concentrations in air and, therefore, overestimating scavenging ratios and washed-out mass fractions. In addition, relative humidity substantially increases during rainfall and this would potentially affect gas-particle partitioning of SOCs in the air leading to underestimation or overestimation of gaseous and particulate fraction of contaminants. Field blanks for air samples were prepared at the site following the standard protocol for

mounting QFF and PUF plugs onto the sampler without turning on the sampler. No field blanks

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were generated for rain samples.

2.3 Meteorological and other supporting data

Cloud top heights were estimated from radiosonde measurements over Prague (Station ID. 11520) (IGRA, 2014) (Table S2 in the SI). Meteorological parameters - i.e. near-ground temperature, precipitation type and intensity, cloud base heights (determined through on-site ceilometer measurements), concentrations of PM_{2.5} and PM₁₀, and their ionic species (i.e. SO₄²⁻, NO₃⁻, NH₄⁺), elemental carbon (EC), and organic carbon (OC) contents - were obtained from CHMI (Table S3 in the SI). Aerosol number size distribution data, used for calculating the aerosol surface area, was obtained from the Academy of Sciences of the Czech Republic. Analyte physico-chemical properties were obtained from Estimation Programs Interface Suite 4.11 (USEPA, 2012). The weather charts used to determine the occurrence of frontal passage over the study site were obtained from *Berliner Wetterkarte* (BWK, 2013).

2.4 Chemical analysis and quality control

QFF and PUF samples were extracted with dichloromethane using an automatic extraction system (Büchi B-811, Switzerland). PUF plugs (n = 2) related to each sampling period were extracted together. Field blanks were extracted along with each set of 10 samples. The extracts were concentrated under a gentle stream of nitrogen in ambient temperature and fractionated using a silica column for PAHs and a sulfuric acid modified silica column for PCBs and CPs. The method performance was tested prior to sample analysis and no degradation of target analytes due to acid treatment was detected. Analytes in rainwater were extracted using solid-phase extraction (C18 Speedisks, Bakerbond, the Netherlands). The analytes were later eluted using 40 ml of (1:1) dichloromethane: n-hexane, concentrated under a gentle stream of nitrogen,

138 and passed through a glass column packed with anhydrous sodium sulfate (1cm ID, 3 cm height) 139 to remove residual water. Prior to extraction, all air and rainwater samples and blanks were spiked with a solution containing d₈-naphthalene, d₁₀-phenanthrene, d₁₂-perylene, PCB 30, and 140 141 PCB 185, which were used as recovery standards. 142 Samples were analyzed for 26 parent PAHs (i.e. naphthalene (NAP), acenaphthylene (ACY), acenaphthene (ACE), fluorene (FLN), phenanthrene (PHE), anthracene (ANT), fluoranthene 143 (FLT), pyrene (PYR), benzo(a)anthracene (BAA), chrysene (CHR), benzo(b)fluoranthene 144 (BBF), benzo(k)fluoranthene (BKF), benzo(a)pyrene (BAP), indeno(123-cd)pyrene (IPY), 145 146 dibenz(a,h)anthracene (DHA), benzo(g,h,i)perylene (BPE), benzo(b)fluorene (BFN), 147 benzo(g,h,i)fluoranthene (BGF), cyclopenta(c,d)pyrene (CPP), triphenylene (TPH), 148 benzo(j)fluoranthene (BJF), benzo(e)pyrene (BEP), perylene (PER), dibenz(a,c)anthracene (DCA), anthanthrene (ATT), coronene (COR)), one heterocyclic PAH (i.e. benzo-naphtho-149 150 thiophene (BNT)), and one alkylated PAH (i.e. retene (RET)) using a Hewlett-Packard gas chromatograph (GC 6890) interfaced to a Hewlett-Packard mass selective detector (MS 5973). 151 Seven PCBs (i.e. PCB 28, PCB 52, PCB 101, PCB 118, PCB 153, PCB 138, PCB 180) and 13 152 CPs, namely α -hexachlorocyclohexane (HCH), β -HCH, γ -HCH, δ -HCH, ε -HCH, o, p'-153 dichlorodiphenyldichloroethylene (DDE), p,p '-DDE, o,p '-dichlorodiphenyldichloroethane 154 (DDD), p,p'-DDD, o,p'-dichlorodiphenyltrichloroethane (DDT), p,p'-DDT, pentachlorobenzene 155 (PeCB), hexachlorobenzene (HCB), were analyzed using an Agilent GC (7890) coupled with an 156 Agilent Triple Quadrupole MS/MS (7000B). All analytes were separated on a J&W Scientific 157 capillary column (HP-5ms, 0.25 mm ID, 0.25 µm film thickness). D₁₄-p-terphenyl and PCB 121 158 159 were used as internal standards for PAH and PCB/CP analysis, respectively. The analytical

160 method recoveries for PAHs and PCBs/CPs ranged from 72 to 102% and 88 to 100%, respectively. The measured analyte concentrations were not recovery corrected. 161 Limits of quantification (LOO) for analytes were calculated based on instrument detection limits, 162 163 which in turn are determined using three times the chromatogram baseline noise level. The median LOQs for individual PAHs and PCBs/CPs in air samples were 0.006 and 0.0005 ng m⁻³, 164 respectively. The median LOOs in rainwater samples were 0.05 and 0.45 ng L⁻¹, respectively. 165 LOQ values were used in cases where analyte concentrations in field blanks were <LOQ. The 166 mean concentrations of analytes in five field blanks were subtracted from those in the 167 168 corresponding samples. The concentrations that were lower than mean + 3 standard deviations of 169 those in field blanks were considered below method detection limit (<MDL) and were 170 substituted with LOQ/2 for calculation of averages. Analysis of ionic species in rainwater was done using a Hewlett-Packard capillary 171 electrophoresis system (HP 3D CE). Measurements were performed using an uncoated silica 172 173 capillary column (75 µm ID, 70 cm length). The applied separation voltage was 10kV with hydrodynamic injection of 50 mbar/20 sec. Analytes were detected indirectly at 254 nm 174 wavelength. The electrolyte was composed of 5 mM sodium chromate and boric acid with pH 175 176 adjusted to 8.3 by adding 0.5 mM cetyltrimethylammonium bromide. Quantification was performed using individual calibration curves. Each quantification was performed three times 177 and mean values were used. 178 179

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2.5 Calculations and data analysis

Analyte particulate mass fractions, θ (unit-less), were calculated using equation 1,

$$\theta = C_{ip}/(C_{ig} + C_{ip})$$
 Eq. (1)

- where, C_{ip} is analyte (i) air concentration (ng m⁻³) in particulate phase and C_{ig} is that in gas phase.
- Daily wet deposition fluxes, F_{wet} (ng m⁻²), were determined using equation 2,

$$F_{\text{wet}} = C_{\text{ir}} \times P$$
 Eq. (2)

- where, C_{ir} is analyte concentration in rainwater (ng L⁻¹) and P is the rain volume received per
- collection area (L m⁻²) in each sampling day. Seasonal fluxes were calculated by summing the
- daily fluxes in each season. Total scavenging ratios, W_t (unit-less), were calculated using
- 189 equation 3,

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$$W_{\rm t} = \frac{C_{\rm ir} \times 10^3}{(C_{\rm ig} + C_{\rm ip})}$$
 Eq. (3)

- where, multiplication by 10^3 accounts for conversion from ng L⁻¹ to ng m⁻³. The mass fraction of
- the total analyte burden in air washed out by below- and in-cloud scavenging, c_t (unit-less), is
- calculated according to Škrdlíková et al. (2011) using equation 4,

$$\varepsilon_{\rm t} = \frac{b_{\rm r}}{b_{\rm a}} = \frac{C_{\rm ir} \times P}{(C_{\rm ig} + C_{\rm ip}) \times h}$$
 Eq. (4)

- where, $b_{\rm r}$ and $b_{\rm a}$ are analyte burdens (ng m⁻²) in rainwater and air, respectively, and h (m) is the
- height of the air column subject to precipitation, which corresponds to the cloud height from
- which precipitation originates.

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3 Results and Discussion

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3.1 Concentrations and distribution of analytes

3.1.1. Concentrations in air

201 NAP was targeted for analysis but it did not pass our quality control criteria and, therefore, is not 202 discussed hereafter. In addition, only analytes that were detected in >10% of the samples are discussed herein (Table 1 and 2). ∑ PAH (i.e. the sum of 27 PAHs) concentrations in gas phase 203 ranged from 0.6 to 138.6 (mean \pm standard deviation (SD): 11.3 \pm 15.7) ng m⁻³ (Table 1). PAHs 204 205 with molecular mass <228 Da, namely ACY, ACE, FLN, PHE, ANT, FLT, and PYR, were predominant in gas phase and, on average, accounted for 93% of the \sum PAH concentrations. 206 PHE, FLN, and FLT showed the highest mean contributions accounting for 42, 24, and 12% of 207 the Σ PAH gaseous concentrations, respectively. BKF, BAP, IPY, DHA, BPE, BNT, CPP, BJF, 208 209 BEP, PER, DCA, ATT, and COR were detected in <10% of the gaseous samples (n=162) (Table 1). The \sum PAH concentrations in the particulate phase ranged from 0.1 to 189.3 (mean \pm 210 SD: 9.3±21.2) ng m⁻³ (Table 1). In sum, PAHs with molecular mass > 228 Da were predominant 211 212 in particulate phase and, on average, contributed to 67% of the Σ PAH particulate concentrations. However, FLT and PYR showed the highest individual contributions and, on 213 average, accounted for 13 and 11% of the Σ PAH concentrations, respectively. The sum of the 214 concentrations of gas and particulate phase PAHs ranged from 0.7 to 327.9 ng m⁻³, with the 215 mean \pm SD being 20.6 \pm 35.9 ng m⁻³. The PAH concentration ranges in gas and particulate phase 216 are noticeably higher than those measured by Škrdlíková et al. (2011) (i.e. 0.9 to 34.6 and 0.1 to 217 16.8 ng m⁻³, respectively) for samples collected from the same site during 2007-2008 period, but 218

219 are slightly lower than those reported by Holoubek et al. (2007) for the period between 1996 and 2005 (i.e. 0.4 to 208 and 0.1 to 359 ng m⁻³, respectively). 220 Except for p,p'-DDE, the targeted chlorinated compounds were mainly detected in gas phase (i.e. 221 <10% in particulate phase) (Table 2). The concentrations of p,p'-DDE in particulate phase 222 ranged from <MDL to 6.8 (mean \pm SD: 0.4 \pm 0.7) pg m⁻³. In gas phase, β -HCH, δ -HCH, ε -HCH, 223 and o,p'-DDD were detected in <10% of all samples (n = 162). The concentrations in gas phase 224 ranged from <MDL to 43.5 (mean \pm SD: 8.1 \pm 7.2) pg m⁻³ for \sum PCBs (i.e. the sum of the 225 concentrations of PCB 28, 52, 101, 118, 138, 153, 180), <MDL to 73.8 (11.0±11.6) pg m⁻³ for \sum 226 HCHs (i.e. the sum of the concentrations of a- and y-HCH), <MDL to 104.3 (23.2 \pm 19.8) pg m⁻³ 227 for Σ DDTs (i.e. the sum of the concentrations of o,p'-DDE, p,p'-DDE, p,p'-DDD, o,p'-DDT, and 228 p,p'-DDT) and <MDL to 332.3 (96.1±42.0) pg m⁻³ for Σ CBs (i.e. the sum of the concentrations 229 of PeCB and HCB) (Table 2). On average, PCB 28 (40%), γ-HCH (56%), p,p'-DDE (83%), and 230 HCB (90%) predominated Σ PCB, Σ HCH, Σ DDT, and Σ CB concentrations, respectively. The 231 232 sum of the concentrations of gas and particulate phase PCBs and CPs ranged from <MDL to 44.6 and <MDL to 351.7 pg m⁻³, respectively. The measured concentrations in the present study are 233 234 considerably lower than those reported by Holoubek et al. (2007) for the samples collected from the same location between 1996 and 2005 (i.e. <MDL to 390 pg m⁻³ for Σ PCBs, <MDL to 771 235 pg m⁻³ for Σ HCHs, 1 to 207 pg m⁻³ for Σ DDTs, and <MDL to 831 pg m⁻³ for HCB). As 236 production and use of these compounds are banned in Europe, the relatively low concentrations 237 in the present study could indicate emission due to volatilization from contaminated soil around 238 239 Košetice (Fig. S1). This argument is supported by significant correlations found between the 240 ambient temperature at the sampling site and the measured concentrations of PCB 28 (r = 0.70, P

<0.05, n = 162), γ -HCH (r = 0.74, P <0.05), and p,p '-DDE (r = 0.71, P <0.05). Despite having the

highest mean concentration among the other chlorinated analytes, relatively small correlation was found between the concentrations of HCB and ambient temperature changes (r = 0.25). This could be due to the fact that this compound has different source pathways compared to all other chlorinated chemicals investigated in this study. Although banned, HCB could potentially be released to the environment as an unintended byproduct of organic solvent and aluminum manufacturing and waste burning (EPER, 2014). In addition, this compound is present in some pesticide formulations, such as chlorothalonil, which is currently registered for use in Europe.

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3.1.2. Concentrations in rain

- \sum PAH concentrations in rainwater ranged from <MDL to 2.1×10^3 (mean \pm SD: 173.3 \pm 256.1) ng
- L⁻¹ (Table 3), predominated (mean: 69%) by congeners with <228 Da molecular mass. FLT,
- 253 PHE, and PYR showed the highest individual contributions and, on average, accounted for 35,
- 14, and 10% of the Σ PAH concentrations. The Σ PAH concentration range in the present study
- was higher than the one reported by Škrdlíková et al. (2011) for Košetice (i.e. 7.1 to 485.9 ng L
- but noticeably lower than that measured by Holoubek et al. (2007) (i.e. 2.4 to 6310 ng L⁻¹).
- 257 The concentrations of PCBs and CPs, targeted for analysis in rainwater samples, were below
- LOQs and, therefore, are not discussed hereafter.

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3.2 Wet deposition fluxes

- 261 \sum PAH daily wet deposition fluxes ranged from <MDL to 5.5×10³ (mean ± SD: 632.9±900.1) ng
- 262 m^{-2} (Table 4). \sum PAH seasonal fluxes were noticeably higher during winter and spring compared

to summer and autumn (Table 5), which is in agreement with previous observations in Europe (Kiss et al., 2001; Škrdlíková et al., 2011). Seasonal changes in SOC fluxes could be due to different factors, such as variations in atmospheric concentration, precipitation amount (providing that atmospheric concentrations do not change drastically between seasons), and SOC gas-particle partitioning behavior. The increase in PAH fluxes during winter and spring cannot be explained by changes in precipitation amounts, as indicated in Table 5. However, atmospheric concentrations of PAHs were noticeably higher in winter than they were in other seasons (Fig. 1), in agreement with a previously published study in Europe (Lammel et al., 2011). This may be due to higher emission rate, lower boundary layer mixing height (Birgül et al., 2011), and lower concentrations of OH radicals in winter (Halsall et al., 2001). This may partly explain the higher fluxes seen during winter periods. Finally, lower temperatures in winter could potentially shift partitioning of the more temperature sensitive SOCs towards particulate phase, and particle scavenging, as opposed to gas scavenging, was suggested to be the dominant wet scavenging mechanism for the removal of PAHs from the atmosphere (Bidleman, 1988; Poster and Baker, 1996; Offenberg and Baker, 2002). In the present study, significant negative regressions were found between near-ground temperature and Θ for a number of PAHs including PYR ($r^2 = 0.72$, P < 0.05, n = 150), FLT ($r^2 = 0.69, n = 155$), RET ($r^2 = 0.75, n = 156$), and TPH ($r^2 = 0.73, n = 156$) 162) (Table S4 in the SI), which could provide further explanation for higher fluxes during cold seasons.

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3.3 Scavenging ratios

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Scavenging ratios were only determined for those events with air and rainwater sampled from the same air mass (n = 54). The rejection criterion was that frontal passage did not occur over the site prior (± 1 hour uncertainty) to the onset of rain. Out of 54 pair samples, only 32 met the abovementioned criterion (Table S2 in the SI) and, hence, W_t ratios were determined for 32 rain events. The relative importance of gas and particle scavenging can be calculated theoretically using W_t and gas scavenging ratio obtained through dimensionless Henry's law constant (Offenberg and Baker, 2002; Poster and Baker, 1996). However, this approach was avoided in the present study due to the fact that the equilibrium partitioning of SOCs between gas phase and raindrops, and consequently gas scavenging ratios, is greatly affected by temperature, which may change drastically from cloud to ground level. This process is non-linear and a representative temperature is unknown. This means that Henry's law constants, which are corrected using ground temperatures, may not necessarily represent the true gas scavenging ratios related to falling raindrops. One should also note that theoretical gas scavenging ratios for SOCs in gas phase, being the inverse of their Henry's law constants, may not entirely reflect the underlying mechanism of the compound removal from the atmosphere. For instance, PAHs with higher vapor pressure normally show higher Henry's law constants, as can be seen from experimental data (Bamford et al., 1999). This would result in relatively lower gas scavenging ratios for gaseous PAHs with higher vapor pressure and solubility, which contradicts the concept of SOC gas scavenging.

In the present study, the concentrations of ionic species (i.e. the sum of SO_4^{2-} , NO_3^{-}) in PM and those in adjacent rainwater samples were significantly correlated (r = 0.71, P < 0.05, n = 23). This indicates the efficient removal of PM by rainfall and highlights the importance of particle scavenging in the present study. In general, W_t ratios for individual PAHs ranged from 0 (in cases where concentrations in rainwater were <LOQ) to 3.5×10^5 , with the exception of FLT for which W_t ranged between 3.7×10^3 and 1.3×10^6 (median: 1.4×10^4) (Table 6). Overall, W_t values for relatively more volatile PAHs (log K_{0a} between 6.27 and 7.57), namely ACY, ACE, FLN, ANT, and PHE, were up to 10^4 , whereas the values for the rest of PAHs (log $K_{oa} > 8.70$) were up to 10⁶, but mainly dominated by the values close to 10⁵ (Table 6). Taking into account that SOCs are more efficiently removed from the atmosphere by particle scavenging, the current results may imply that increase in affinity of individual PAHs towards organic phase would overall lead to higher scavenging efficiencies. This, together with other parameters that affect analyte gasparticle partitioning, namely temperature, aerosol surface area and chemical composition (e.g. EC and OC contents) (Junge, 1977; Pankow, 1987; Bidleman, 1988), and factors that affect particle removal rate including rain intensity, aerosol and raindrop size and collision efficiency (Mircea et al., 2000; Poster and Baker, 1996; Slinn et al., 1978), may contribute to the observed variability in measured scavenging ratios. Since chlorinated compounds in the present study were mainly detected in gas phase in the air (Table 2), it is likely that their removal from the atmosphere was determined by gas scavenging. The fact that concentrations of these analytes in rainwater were below detection limits may provide further evidence for this argument, as the magnitude of gas scavenging is negligible. The upper- and lower-bound scavenged mass fractions of target analytes corresponding to cloud base and top heights were determined for 32 events, except in one case for which cloud top

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height was not available (Table S2). \sum PAH upper-bound scavenged mass fractions ranged from 1.2×10^{-3} to 0.8 (median: 5.3×10^{-2}), whereas lower-bound mass fractions were between 5.2×10^{-4} and 0.1 (median: 9.7×10^{-3}) (Table 7). The median ratios for the analyte lower-bound mass fractions were nearly an order of magnitude different from those reported by Škrdlíková et al. (2011) for the same site, except for FLN and CHR, which were found to be in agreement (Table 7). This could potentially be related to the relatively small number of events considered in that study (n = 10). It should be noted that scavenged mass fractions were calculated assuming that analyte concentrations near ground represented those within the air column up to cloud top - i.e. assuming perfect mixing throughout the column. The true scavenged mass fractions could potentially be higher due to a negative vertical concentration gradient (Škrdlíková et al., 2011).

3.4 Factors affecting the scavenging ratios

As can be seen from Tables 6 and S3, W_t ratios for \sum PAHs in the present study ranged from 2.4×10^3 to 2.3×10^5 , with noticeable variability between the events (mean \pm SD: $1.7 \times 10^4 \pm 3.8 \times 10^4$). As discussed earlier, different parameters could contribute to the variability seen in scavenging ratios. In this section, we investigate potential relationships between PAH scavenging efficiencies and precipitation intensity, near-ground temperature, the concentrations of PM_{2.5} and PM₁₀, PM ionic species (i.e. the sum of the concentrations of SO₄²⁻, NO₃⁻, and NH₄⁺), EC and OC contents, PM surface area, and rainwater ionic species (i.e. the sum of SO₄²⁻, NO₃⁻, and Cl⁻). As can be seen from Table S3, the highest \sum PAH W_t was found for the samples collected on 14 April 2012. The ratio was nearly two orders of magnitude higher compared to that obtained for another event with similar precipitation type and ambient temperature (i.e. 11 –

the rest of the parameters were somewhat similar for these two events (Table S3). The concentrations of PM₁₀ as well as those for ionic species in PM and rainwater for the former event were 1.4, 1.8, and 11 times the values found for the latter event. This observation suggests that PM removal by rain, and consequently wet scavenging of PAHs that are sorbed to PM, is enhanced in conditions where PM contains high quantities of soluble species. This argument is supported by considerably higher scavenging ratios found for particulate PAHs (a factor of 50 on average) in the former event. This difference was much smaller for gaseous species (a factor of five on average). In order to explore the relative importance of the abovementioned parameters in relation to scavenging ratios, correlation analysis was performed between these variables. Among all parameters, aerosol surface area was only available for 2012. In addition, aerosol EC and OC contents could only be obtained for 15 rain events out of 32, as they were recorded once every six days (Table S3). Owing to the fact that EC and OC have high importance in SOC gas-particle partitioning (Lohmann and Lammel, 2004), and in order to enhance the comparability of results, for all other parameters, correlation analysis was only performed for the corresponding rain events. The results showed that the concentrations of rainwater ionic species were significantly correlated with Σ PAH scavenging ratios (r = 0.70, P < 0.05, n = 15), whereas weak correlations were found for all other parameters, except the ratio of PM ionic species/PM₁₀, for which a moderate correlation was found (r = 0.34, P > 0.05). However, when the data related to 14 April 2012 (with exceptionally high W_t ratio) was excluded from the analysis, the suggested importance of rainwater and PM ionic species decreased (r = 0.14 and 0.07, respectively, P

>0.05, n = 14). This indicates that the initial analysis may have been overpowered by

12 October 2012). Apart from the concentrations of PM₁₀ and ionic species in PM and rainwater,

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significantly high values of ionic species related to a single event. Without the data point related to 14 April 2012, moderate correlations were obtained between \sum PAH W_t ratios and EC/PM₁₀ (r = 0.34, P > 0.05) and OC/PM₁₀ (r = 0.39, P > 0.05). Although not statistically significant, the current results suggest that the aerosol EC and OC contents are important parameters in determining PAH scavenging efficiencies. This may reflect the affinity of PAHs towards EC and OC, representing adsorption and absorption processes (Lohmann and Lammel, 2004). However, the magnitude of these effects could be different when considering individual PAHs. For instance, the coefficients of correlation between W_t and EC/PM₁₀ ratios for PAHs with moderate to high particulate mass fractions, namely PYR (mean $\theta = 0.40$), CHR (mean $\theta = 0.74$), and BBF (mean $\theta = 0.97$), were 0.38, 0.20, and 0.33 (n = 10), whereas the coefficients for correlation with OC/PM₁₀ ratios were 0.51, 0.53, and 0.03, respectively. Despite not having statistical significance, the results may suggest that the aerosol OC content was more effective than EC for wet scavenging of PYR and CHR. It should be noted that precipitation type (i.e. snow vs. rain) was only available for eight events out of 15. These were mainly identified as rainfall (Table S3), which ruled out the potential impact of precipitation type on scavenging efficiencies. The lack of significant linear relationships between scavenging ratios and aerosol and rainwater properties indicates that the process is not controlled by a single factor but rather by a combination of parameters. The findings of the present study highlight the need for incorporating other PM components, such as EC and OC, in air pollution models for more accurate estimation of particulate PAH scavenging.

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4 Conclusions

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The results suggest that volatilization from contaminated soil was the main source of chlorinated compounds in the air over Košetice. Higher \sum PAH fluxes in winter were possibly related to higher PAH atmospheric concentrations and particulate mass fractions in that season. Particle scavenging was found to be significant in the present study, as indicated by the strong correlation between rainwater and PM ionic species. In sum, greater scavenging ratios were found for less volatile PAHs, highlighting the importance of SOC gas-particle partitioning in determining their scavenging efficiencies. The results suggested that PM removal by rain, and consequently wet scavenging of PAHs sorbed to PM, could be more efficient in cases where the concentrations of ionic species in PM are high. In addition, the high affinity of PAHs towards EC and OC were found to be reflected in the scavenging efficiency. In most model applications which study PAHs, particulates are scavenged either as in conventional air pollution modeling - i.e. accounting for PM ionic composition, but no other PM components (parameterization based on Köhler theory or empirical approaches based on cloud droplet number) (Abdul-Razzak and Ghan, 2000; Gong et al., 2003) - or with insoluble aerosols or aerosol modes (Sehili and Lammel, 2007; Friedman and Selin, 2012). Future studies would need to focus on in-depth analysis of PM chemical composition with the aim to include OC and EC in the parameterizations of particulate PAH wet deposition.

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Table 1. PAH concentrations and detection frequencies in air.

Gaseous Cons. (ng m⁻³) Particulate Cons. (ng m⁻³) Mean ± SD Min. Max. Mean ± SD DF % Min. Max. DF % $7.8 \times 10^{-2} \pm 2.4 \times 10^{-1}$ ACY <MDL 34.5 1.0 ± 3.0 87 <MDL 1.9 53 $1.1 \times 10^{-2} \pm 3.1 \times 10^{-2}$ **ACE** <MDL 3.2 0.3 ± 0.5 90 <MDL 0.2 22 $1.1 \times 10^{-1} \pm 3.4 \times 10^{-1}$ 49 FLN <MDL 26.3 2.7 ± 4.0 98 <MDL 2.5 0.8 ± 2.2 PHE <MDL 45.0 4.7 ± 5.9 98 14.6 72 <MDL $1.3 \times 10^{-1} \pm 4.0 \times 10^{-1}$ $6.3\times10^{-2}\pm1.9\times10^{-1}$ **ANT** <MDL 4.7 81 <MDL 1.5 53 7.0×10^{-2} 3.0×10^{-3} 1.3 ± 1.6 1.3 ± 3.3 99 FLT 13.8 100 26.1 3.9×10^{-3} 3.5×10^{-3} 99 PYR 7.7 0.6 ± 0.8 93 20.7 1.1 ± 2.5 $1.4 \times 10^{-2} \pm 3.7 \times 10^{-2}$ **BAA** <MDL 0.3 38 <MDL 11.1 0.5 ± 1.2 87 $6.0\times10^{-2}\pm6.8\times10^{-2}$ **CHR** <MDL 0.4 95 <MDL 15.8 0.7 ± 1.7 97 $1.3 \times 10^{-2} \pm 8.2 \times 10^{-2}$ **BBF** <MDL 0.9 14 <MDL 13.6 0.7 ± 1.5 96 $6.2\times10^{-3}\pm2.6\times10^{-2}$ BKF <MDL 0.3 4 <MDL 5.0 0.3 ± 0.6 88 $8.7 \times 10^{-3} \pm 4.2 \times 10^{-2}$ 0.5 3 9.1 83 BAP <MDL <MDL 0.4 ± 0.9 $7.2\times10^{-3}\pm4.6\times10^{-2}$ 3 0.5 ± 1.0 81 IPY <MDL 0.5 10.4 <MDL 2 $4.1\times10^{-2}\pm1.1\times10^{-1}$ DHA <MDL 1.2 56 $7.4 \times 10^{-3} \pm 4.2 \times 10^{-2}$ BPE 0.5 3 0.4 ± 0.7 90 <MDL <MDL 6.6 6.5×10^{-3} 5.0 0.2 ± 0.4 100 73 RET <MDL 4.0 0.2 ± 0.5 $5.2\times10^{-2}\pm8.2\times10^{-2}$ **BFN** <MDL 4.7 0.2 ± 0.5 72 0.8 80 <MDL 2.1×10^{-2} $2.4\times10^{-3}\pm2.2\times10^{-3}$ $2.3\times10^{-2}\pm6.2\times10^{-2}$ **BNT** <MDL 6 <MDL 0.5 46 $5.1\times10^{-2}\pm5.4\times10^{-2}$ **BGF** <MDL 0.4 93 <MDL 7.7 0.3 ± 0.8 92 $5.0\times10^{-3}\pm2.2\times10^{-2}$ CPP <MDL 0.2 3 <MDL 10.3 0.3 ± 0.9 73 $1.5 \times 10^{-2} \pm 1.5 \times 10^{-2}$ $1.4 \times 10^{-1} \pm 3.2 \times 10^{-1}$ TPH 87 86 <MDL 0.1 <MDL 2.8 $8.4 \times 10^{-3} \pm 3.9 \times 10^{-2}$ 7.5 91 BJF <MDL 0.4 6 <MDL 0.4 ± 0.9 $1.0 \times 10^{-2} \pm 4.6 \times 10^{-2}$ 5 87 **BEP** <MDL 0.5 <MDL 7.2 0.4 ± 0.8 $6.9 \times 10^{-2} \pm 1.5 \times 10^{-1}$ PER 2 <MDL 1.5 61 2 $2.9 \times 10^{-2} \pm 6.7 \times 10^{-2}$ **DCA** 55 <MDL 0.6 2 $4.7 \times 10^{-2} \pm 1.5 \times 10^{-1}$ ATT <MDL 1.7 48 $1.3\times10^{-1}\pm3.2\times10^{-1}$ 2 **COR** <MDL 3.3 64 $\sum PAHs$ 0.6 138.6 11.3±15.7 0.1 189.3 9.3 ± 21.2

SD: standard deviation; DF: detection frequency (n = 162); <MDL: below method detection limit; \sum PAHs: the sum of the concentrations of individual PAHs with DF >10% in either gas or particulate phase. Concentration ranges and means are only reported for analytes with DF >2%.

Table 2. PCB and CP concentrations and detection frequencies in air.

	Gaseous Cons. (pg m ⁻³)				Particulate Cons. (pg m ⁻³)			
	Min.	Max.	Mean ± SD	DF %	Min.	Max.	Mean ± SD	DF %
PCB 28	<mdl< td=""><td>14.3</td><td>3.2 ± 2.4</td><td>88</td><td></td><td></td><td></td><td>1</td></mdl<>	14.3	3.2 ± 2.4	88				1
PCB 52	<mdl< td=""><td>8.4</td><td>1.5 ± 1.4</td><td>68</td><td></td><td></td><td></td><td>1</td></mdl<>	8.4	1.5 ± 1.4	68				1
PCB 101	<mdl< td=""><td>13.9</td><td>1.2 ± 1.8</td><td>60</td><td></td><td></td><td></td><td>1</td></mdl<>	13.9	1.2 ± 1.8	60				1
PCB 118	<mdl< td=""><td>4.0</td><td>0.3 ± 0.5</td><td>10</td><td></td><td></td><td></td><td><mdl< td=""></mdl<></td></mdl<>	4.0	0.3 ± 0.5	10				<mdl< td=""></mdl<>
PCB 138	<mdl< td=""><td>3.9</td><td>0.5 ± 0.5</td><td>24</td><td></td><td></td><td></td><td>2</td></mdl<>	3.9	0.5 ± 0.5	24				2
PCB 153	<mdl< td=""><td>8.0</td><td>1.0 ± 1.2</td><td>62</td><td><mdl< td=""><td>3.1</td><td>0.3 ± 0.3</td><td>4</td></mdl<></td></mdl<>	8.0	1.0 ± 1.2	62	<mdl< td=""><td>3.1</td><td>0.3 ± 0.3</td><td>4</td></mdl<>	3.1	0.3 ± 0.3	4
PCB 180	<mdl< td=""><td>2.1</td><td>0.3 ± 0.2</td><td>14</td><td><mdl< td=""><td>3.3</td><td>0.2 ± 0.3</td><td>6</td></mdl<></td></mdl<>	2.1	0.3 ± 0.2	14	<mdl< td=""><td>3.3</td><td>0.2 ± 0.3</td><td>6</td></mdl<>	3.3	0.2 ± 0.3	6
α -HCH	<mdl< td=""><td>44.5</td><td>4.4 ± 6.0</td><td>57</td><td></td><td></td><td></td><td>1</td></mdl<>	44.5	4.4 ± 6.0	57				1
β -HCH				2				<mdl< td=""></mdl<>
γ-НСН	<mdl< td=""><td>31.4</td><td>6.6 ± 6.5</td><td>75</td><td></td><td></td><td></td><td>1</td></mdl<>	31.4	6.6 ± 6.5	75				1
δ -HCH	<mdl< td=""><td>2.0</td><td>0.4 ± 0.3</td><td>5</td><td></td><td></td><td></td><td>1</td></mdl<>	2.0	0.4 ± 0.3	5				1
ε -HCH				1				<mdl< td=""></mdl<>
o,p'-DDE	<mdl< td=""><td>1.9</td><td>0.4 ± 0.3</td><td>27</td><td></td><td></td><td></td><td><mdl< td=""></mdl<></td></mdl<>	1.9	0.4 ± 0.3	27				<mdl< td=""></mdl<>
p,p'-DDE	<mdl< td=""><td>87.3</td><td>19.5±15.9</td><td>98</td><td><mdl< td=""><td>6.8</td><td>0.4 ± 0.7</td><td>31</td></mdl<></td></mdl<>	87.3	19.5±15.9	98	<mdl< td=""><td>6.8</td><td>0.4 ± 0.7</td><td>31</td></mdl<>	6.8	0.4 ± 0.7	31
o,p'-DDD	<mdl< td=""><td>2.2</td><td>0.3 ± 0.2</td><td>6</td><td></td><td></td><td></td><td><mdl< td=""></mdl<></td></mdl<>	2.2	0.3 ± 0.2	6				<mdl< td=""></mdl<>
p,p'-DDD	<mdl< td=""><td>2.4</td><td>0.4 ± 0.4</td><td>17</td><td></td><td></td><td></td><td>1</td></mdl<>	2.4	0.4 ± 0.4	17				1
o,p'-DDT	<mdl< td=""><td>8.9</td><td>1.5 ± 1.9</td><td>45</td><td></td><td></td><td></td><td><mdl< td=""></mdl<></td></mdl<>	8.9	1.5 ± 1.9	45				<mdl< td=""></mdl<>
p,p'-DDT	<mdl< td=""><td>11.2</td><td>1.6 ± 2.3</td><td>36</td><td></td><td></td><td></td><td>1</td></mdl<>	11.2	1.6 ± 2.3	36				1
PeCB	<mdl< td=""><td>52.8</td><td>9.6 ± 7.3</td><td>99</td><td></td><td></td><td></td><td>2</td></mdl<>	52.8	9.6 ± 7.3	99				2
HCB	<mdl< td=""><td>279.5</td><td>86.5 ± 40.4</td><td>99</td><td><mdl< td=""><td>62.7</td><td>0.9 ± 6.2</td><td>7</td></mdl<></td></mdl<>	279.5	86.5 ± 40.4	99	<mdl< td=""><td>62.7</td><td>0.9 ± 6.2</td><td>7</td></mdl<>	62.7	0.9 ± 6.2	7
$\sum PCBs$	<mdl< td=""><td>43.5</td><td>8.1±7.2</td><td></td><td></td><td></td><td></td><td></td></mdl<>	43.5	8.1±7.2					
\sum HCHs	<mdl< td=""><td>73.8</td><td>11.0±11.6</td><td></td><td></td><td></td><td></td><td></td></mdl<>	73.8	11.0±11.6					
$\overline{\sum}$ DDTs	<mdl< td=""><td>104.3</td><td>23.2 ± 19.8</td><td></td><td></td><td></td><td></td><td></td></mdl<>	104.3	23.2 ± 19.8					
\sum CBs	<mdl< td=""><td>332.3</td><td>96.1±42.0</td><td></td><td></td><td></td><td></td><td></td></mdl<>	332.3	96.1±42.0					

SD: standard deviation; DF: detection frequency (n = 162); <MDL: below method detection limit; \sum PCBs: the sum of the concentrations of PCB 28, 52, 101, 118, 138, 153, and 180; \sum HCHs: the sum of the concentrations of α - and γ -HCH; \sum DDTs: the sum of the concentrations of o,p'-DDE, p,p'-DDD, o,p'-DDT, and p,p'-DDT; \sum CBs: the sum of the concentrations of PeCB and HCB. Note that only analytes with DF>10% in either gas or particulate phase were considered for calculating the sums. Concentration ranges and means are only reported for analytes with DF>2%.

Table 3. PAH concentrations and detection frequencies in rainwater.

	Concentrations (ng L ⁻¹)						
	Min.	Max.	Mean ± SD	DF %			
ACY	<mdl< td=""><td>23.7</td><td>2.1±2.8</td><td>67</td></mdl<>	23.7	2.1±2.8	67			
ACE	<mdl< td=""><td>7.0</td><td>1.0 ± 1.2</td><td>54</td></mdl<>	7.0	1.0 ± 1.2	54			
FLN	<mdl< td=""><td>41.6</td><td>6.0 ± 5.8</td><td>99</td></mdl<>	41.6	6.0 ± 5.8	99			
PHE	<mdl< td=""><td>268.0</td><td>23.5 ± 29.2</td><td>99</td></mdl<>	268.0	23.5 ± 29.2	99			
ANT	<mdl< td=""><td>41.2</td><td>1.1 ± 3.2</td><td>56</td></mdl<>	41.2	1.1 ± 3.2	56			
FLT	<mdl< td=""><td>1.1×10^{3}</td><td>61.4±144.6</td><td>99</td></mdl<>	1.1×10^{3}	61.4±144.6	99			
PYR	<mdl< td=""><td>282.5</td><td>17.7 ± 27.0</td><td>91</td></mdl<>	282.5	17.7 ± 27.0	91			
BAA	<mdl< td=""><td>76.9</td><td>3.1 ± 6.7</td><td>63</td></mdl<>	76.9	3.1 ± 6.7	63			
CHR	<mdl< td=""><td>141.8</td><td>9.1 ± 14.9</td><td>75</td></mdl<>	141.8	9.1 ± 14.9	75			
BBF	<mdl< td=""><td>85.2</td><td>6.5 ± 13.0</td><td>69</td></mdl<>	85.2	6.5 ± 13.0	69			
BKF	<mdl< td=""><td>42.3</td><td>2.8 ± 4.8</td><td>63</td></mdl<>	42.3	2.8 ± 4.8	63			
BAP	<mdl< td=""><td>50.0</td><td>2.3 ± 4.9</td><td>55</td></mdl<>	50.0	2.3 ± 4.9	55			
IPY	<mdl< td=""><td>58.2</td><td>3.6 ± 7.1</td><td>60</td></mdl<>	58.2	3.6 ± 7.1	60			
DHA	<mdl< td=""><td>3.9</td><td>0.3 ± 0.4</td><td>25</td></mdl<>	3.9	0.3 ± 0.4	25			
BPE	<mdl< td=""><td>50.9</td><td>3.6 ± 6.5</td><td>59</td></mdl<>	50.9	3.6 ± 6.5	59			
RET	<mdl< td=""><td>44.0</td><td>3.1 ± 4.6</td><td>73</td></mdl<>	44.0	3.1 ± 4.6	73			
BFN	<mdl< td=""><td>67.2</td><td>3.1 ± 6.0</td><td>62</td></mdl<>	67.2	3.1 ± 6.0	62			
BNT	<mdl< td=""><td>6.7</td><td>0.5 ± 0.7</td><td>43</td></mdl<>	6.7	0.5 ± 0.7	43			
BGF	<mdl< td=""><td>70.8</td><td>4.9 ± 7.7</td><td>70</td></mdl<>	70.8	4.9 ± 7.7	70			
CPP	<mdl< td=""><td>17.4</td><td>1.1 ± 2.2</td><td>48</td></mdl<>	17.4	1.1 ± 2.2	48			
TPH	<mdl< td=""><td>44.2</td><td>3.5 ± 5.4</td><td>67</td></mdl<>	44.2	3.5 ± 5.4	67			
BJF	<mdl< td=""><td>63.0</td><td>5.6 ± 9.7</td><td>64</td></mdl<>	63.0	5.6 ± 9.7	64			
BEP	<mdl< td=""><td>60.0</td><td>4.8 ± 7.6</td><td>73</td></mdl<>	60.0	4.8 ± 7.6	73			
PER	<mdl< td=""><td>7.8</td><td>0.5 ± 0.9</td><td>39</td></mdl<>	7.8	0.5 ± 0.9	39			
DCA	<mdl< td=""><td>7.7</td><td>0.4 ± 0.7</td><td>27</td></mdl<>	7.7	0.4 ± 0.7	27			
ATT	<mdl< td=""><td>8.0</td><td>0.3 ± 0.7</td><td>18</td></mdl<>	8.0	0.3 ± 0.7	18			
COR	<mdl< td=""><td>22.1</td><td>1.5 ± 2.9</td><td>52</td></mdl<>	22.1	1.5 ± 2.9	52			
∑ PAHs	<mdl< td=""><td>2.1×10^3</td><td>173.3±256.1</td><td></td></mdl<>	2.1×10^3	173.3±256.1				

SD: standard deviation; DF: detection frequency (n = 231); <MDL: below method detection limit; \sum PAHs: the sum of the concentrations of 27 individual PAHs listed on the table.

Table 4. PAH daily wet deposition fluxes (F_{wet}).

 $F_{\text{wet}} (\text{ng m}^{-2})$ Min. Max. Mean ± SD ACY <MDL 94.8 8.4 ± 14.0 76.1 **ACE** <MDL 4.3 ± 8.8 **FLN** <MDL 305.3 29.8 ± 43.7 **PHE** <MDL 816.8 102.1±133.1 **ANT** <MDL 332.4 4.4 ± 22.6 4.0×10^{3} **FLT** 204.1±413.0 <MDL **PYR** <MDL 753.7 66.6±106.0 BAA <MDL 182.7 10.6 ± 23.1 CHR <MDL 407.5 34.0±60.6 **BBF** 569.9 <MDL 21.5 ± 51.4 **BKF** <MDL 165.7 9.8 ± 19.0 **BAP** <MDL 180.5 7.1±17.9 **IPY** <MDL 178.3 12.2 ± 25.9 **DHA** 13.5 <MDL 0.6 ± 1.8 **BPE** <MDL 161.2 12.1 ± 24.5 **RET** 12.7 ± 24.3 <MDL 234.3 **BFN** <MDL 163.5 11.6 ± 23.0 **BNT** 18.1 1.3 ± 2.7 <MDL **BGF** <MDL 240.6 18.4±31.7 **CPP** <MDL 94.5 3.4 ± 9.8 **TPH** <MDL 104.1 12.4 ± 20.0 BJF <MDL 374.2 20.6±43.4 **BEP** <MDL 249.1 16.8 ± 29.8 **PER** <MDL 34.7 1.4 ± 3.6 **DCA** <MDL 16.7 0.8 ± 2.3 ATT <MDL 21.3 0.7 ± 2.5

SD: standard deviation; <MDL: below method detection limit.

66.5

 5.5×10^{3}

 5.0 ± 10.5

632.9±900.1

<MDL

<MDL

COR

 $\sum PAHs$

Table 5. \sum PAH seasonal wet deposition fluxes (F_{wet}) and precipitation amounts.

2012					2013				
Winter	Spring	Summer	Autumn	Winter	Spring	Summer	Autumn		
	$\sum PAH F_{wet} (ng m^{-2})$								
3.0×10^4	2.3×10^4	6.3×10^3	7.3×10^3	4.0×10^4	1.6×10^4	9.4×10^{3}	1.1×10^4		
Precipitation (mm)									
147.8	76.9	272.4	107.2	169.8	131.6	315.0	82.3		

Table 6. PAH scavenging ratios (W_t) .							
Wt							
	Min.	Max.	Median	Log	Log		
				K_{oa}	K_{aw}		
ACY	NA	6.9×10^4	4.7×10^{3}	6.27	-2.33		
ACE	NA	3.0×10^4	4.5×10^{3}	6.31	-2.12		
FLN	863.9	5.0×10^4	4.8×10^{3}	6.79	-2.41		
ANT	NA	8.4×10^4	2.4×10^{3}	7.55	-2.64		
PHE	2.2×10^{3}	2.5×10^4	5.4×10^{3}	7.57	-2.76		
RET	NA	3.2×10^4	6.2×10^3	8.70	-2.35		
PYR	NA	1.1×10^{5}	1.2×10^4	8.80	-3.31		
FLT	3.7×10^{3}	1.3×10^6	1.4×10^4	8.88	-3.44		
BAA	NA	1.5×10^5	4.7×10^{3}	9.07	-3.31		
CHR	NA	1.6×10^{5}	1.3×10^4	9.48	-3.67		
BFN	NA	1.6×10^{5}	8.6×10^{3}	9.57	-3.80		
BGF	NA	1.4×10^{5}	1.4×10^4	9.78	-4.26		
PER	NA	1.5×10^{5}	1.8×10^{3}	10.08	-3.83		
CPP	NA	1.8×10^{5}	563	10.15	-4.45		
BBF	NA	1.4×10^{5}	7.5×10^3	10.35	-4.57		
BJF	NA	1.4×10^{5}	9.2×10^{3}	10.59	-4.48		
TPH	NA	2.0×10^{5}	1.9×10^4	10.69	-5.20		
BKF	NA	1.4×10^{5}	5.6×10^{3}	10.73	-4.62		
BAP	NA	9.1×10^4	1.6×10^{3}	10.86	-4.73		
DCA	NA	1.4×10^{5}	NA	11.11	-4.70		
BEP	NA	1.4×10^{5}	1.3×10^4	11.35	-4.91		
IPY	NA	1.5×10^{5}	7.4×10^{3}	11.55	-4.85		
DHA	NA	1.6×10^{5}	NA	11.78	-5.24		
BPE	NA	1.4×10^{5}	6.5×10^{3}	11.97	-5.27		
COR	NA	1.8×10^{5}	4.3×10^{3}	13.70	-6.06		
BNT	NA	3.5×10^{5}	6.8×10^{3}	NA	-4.36		
ATT	NA	2.3×10^{5}	NA	NA	NA		

 \sum PAHs 2.4×10³ 2.3×10⁵ 8.8×10³ K_{oa} : analyte octanol-air partitioning coefficient; K_{aw} : analyte airwater partitioning coefficient. For ease of interpretation, analytes are sorted based on their log K_{oa} values. Log K_{oa} and log K_{ow} values were obtained from Estimation Programs Interface Suite 4.11(USEPA, 2012); NA: indicates an event in which analyte concentration in rainwater was <LOQ.

Table 7. PAH scavenged mass fractions (ε).

^a Upper-bound ε ^b Lower-bound ε Max. ^c Median Min. Max. Median Min. Median **ACY** NA 2.2 1.4×10^{-2} NA 1.7 2.3×10^{-3} 5.3×10⁻⁴ 8.9×10^{-3} 8.1×10^{-2} 1.8×10^{-3} 5.2×10^{-4} ACE 0.3 NA NA 5.4×10^{-4} 1.5 1.5×10^{-2} 4.3×10⁻⁴ 0.3 3.6×10^{-3} 3.7×10^{-3} FLN 9.4×10^{-3} 1.3×10^{-2} 2.4×10^{-3} **ANT** NA 0.4 NA 0.1 1.3×10^{-3} 2.6×10^{-2} 5.4×10^{-4} 5.1×10^{-2} 1.5×10^{-2} 5.5×10^{-3} PHE 0.3 1.2×10^{-2} 2.8×10^{-3} NA **RET** NA 0.4 NA 0.1 7.5×10^{-3} **PYR** NA 1.1 5.2×10^{-2} NA 0.2 1.0×10^{-2} 2.8×10^{-3} 1.1×10^{-3} 8.1×10^{-2} 1.3×10^{-2} 9.9×10^{-3} 0.2 **FLT** 0.9 1.2×10^{-2} 5.7×10^{-2} 1.9×10^{-3} 4.8×10^{-2} BAANA 0.9 NA 1.2×10^{-2} 1.0×10^{-2} **CHR** NA 1.8 5.2×10^{-2} NA 0.1 3.0×10^{-2} 4.1×10^{-3} **BFN** NA 1.2 NA 0.2 NA 6.1×10^{-2} 9.1×10^{-3} 2.4 NA **BGF** NA 0.1 NA **PER** NA 1.9 NA NA 2.7×10^{-2} NA NA 3.6×10^{-3} 4.6×10⁻⁴ 6.8×10^{-2} CPP 0.5 NA NA NA 2.4×10^{-2} 3.0 5.0×10^{-2} 3.6×10^{-3} **BBF** NA NA NA BJF NA 3.6 2.7×10^{-2} NA 0.2 1.6×10^{-3} NA NA 3.2 7.4×10^{-2} NA 0.3 1.4×10^{-2} NA TPH 6.8×10^{-2} 2.9×10^{-2} 1.9×10^{-3} NA 2.0 NA NA **BKF BAP** NA 1.5 4.1×10^{-3} NA 3.9×10^{-2} 5.2×10^{-4} NA **DCA** NA 2.9 NA NA 0.1 NA NA 7.5×10^{-2} 4.0×10^{-2} 6.5×10^{-3} 2.9 **BEP** NA NA NA 1.7×10^{-2} 0.3 1.8×10^{-3} **IPY** NA 3.5 NA NA NA 3.8×10^{-2} DHA NA 3.1 NA NA NA 2.3×10⁻² 2.5×10^{-3} 3.6 0.2 **BPE** NA NA NA 4.8 2.2×10^{-3} 0.2 7.2×10^{-4} **COR** NA NA NA **BNT** NA 2.8 NA NA 0.2 NA NA 4.7×10^{-2} **ATT** NA 3.8 NA NA NA NA 1.2×10^{-3} 5.3×10^{-2} 5.2×10^{-4} 9.7×10^{-3} \sum PAHs 27 0.8 0.1 NA

^a Upper- and ^b lower-bound ε correspond to cloud base and top height, respectively; ^c median ε values reported by Skrdlíková et al., (2011) for could top heights (n = 10); NA: indicates an event in which analyte concentration in rainwater was <LOQ. Analytes are sorted based on their log K_{oa} values.

Fig. 1. \sum PAH median air concentrations in winter (D.J.F.), spring (M.A.M.), summer (J.J.A.), and autumn (S.O.N.) at Košetice.

