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# Experimental determination of the temperature dependence of water activities for a selection of aqueous organic solutions

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## Abstract

This work presents experimental data of the temperature dependence of water activity in aqueous organic solutions relevant for tropospheric conditions (200–273 K). Water activity ( $a_w$ ) at low temperatures (T) is a crucial parameter for predicting homogeneous

- <sup>5</sup> ice nucleation. We investigated temperature dependent water activities, ice freezing and melting temperatures of solutions, and vapour pressures of a selection of atmospherically relevant aqueous organic systems. To measure  $a_w$  over a wide composition range and with a focus on low temperatures, we use various  $a_w$  measurement techniques and instruments: a dew point water activity meter, an electrodynamic balance
- <sup>10</sup> (EDB), differential scanning calorimetry (DSC), and a setup to measure the total gas phase pressure at equilibrium over aqueous solutions. Water activity measurements were performed for aqueous multicomponent and multifunctional organic mixtures containing the functional groups typically found in atmospheric organic aerosols, such as hydroxyl, carboxyl, ketone, ether, ester, and aromatic groups. The aqueous organic
- <sup>15</sup> systems studied at several fixed compositions over a considerable temperature range differ significantly in their temperature dependence. Aqueous organic systems of 1,4butanediol and methoxyacetic acid show a moderate decrease in  $a_w$  with decreasing temperature. The aqueous M5 system (a multicomponent system containing five different dicarboxylic acids) and aqueous 2-(2-ethoxyethoxy)ethanol solutions both show
- <sup>20</sup> a strong increase of water activity with decreasing temperature at high solute concentrations for *T* < 270 K and *T* < 260 K, respectively. These measurements show that the temperature trend of  $a_w$  can be reversed at low temperatures and that linear extrapolations of high temperature data may lead to erroneous predictions. To avoid this, experimentally determined  $a_w$  at low temperature are needed to improve thermodynamic
- <sup>25</sup> models towards lower temperatures and for improved predictions of the ice nucleation ability of organic–water systems.



## 1 Introduction

Organic compounds account for a large fraction of airborne particulate matter. They constitute around 50% of the total mass of the fine aerosol fraction in the continental mid-latitudes (Saxena and Hildemann, 1996; Novakov et al., 1997; Murphy et al., 2006; Jimenez et al., 2009) while in the tropics they may contribute up to 90% (Yamasoe et al., 2000; Roberts et al., 2002). In the upper troposphere a high fraction of organic aerosols are internally mixed with sulphate aerosols (Murphy et al., 2006, 2007). The organic aerosol fraction is expected to remain in a liquid or amorphous (viscous) state since the large number of organic compounds depresses the temperature at which crystallisation takes place (Marcolli et al., 2004). Studies show that the presence of an organic aerosol fraction may inhibit ice nucleation and growth (DeMott et al., 2003; Cziczo et al., 2004; Peter et al., 2006; Knopf and Lopez, 2009). The organic aerosol fraction, and trace gases (Fuzzi et al., 2006). In turn,

- these interactions influence the physical and chemical properties of aerosol particles such as physical state, hygroscopicity, size, and shape (e.g., Ming and Russell, 2002; Kanakidou et al., 2005; Zobrist et al., 2008; Ciobanu et al., 2009; Mikhailov et al., 2009; Reid et al., 2011; Song et al., 2012). The light scattering intensity of aerosols depends on growth and evaporation of the particles due to uptake and release of water vapour
- <sup>20</sup> driven by changes in ambient relative humidity (RH) (Carrico et al., 2003; Baynard et al., 2006; Zieger et al., 2013). For an accurate description of this process, the hygroscopicity of the involved aerosol compositions have to be known. The hygroscopicities of most organic mixtures depend on temperature. Particle phase water activity  $a_w$  is equal to RH, provided that the aqueous aerosol particles are in equilibrium with the
- <sup>25</sup> surrounding gas phase and that they are sufficiently large so that the Kelvin effect is negligible. However,  $a_w$  data of aqueous organic compounds at low temperatures are scarce. Therefore, most estimates of the RH dependence of the direct aerosol effect rely on data at or close to room temperature. At low tropospheric temperatures, particle

![](_page_2_Picture_5.jpeg)

hygroscopicity may be different, because of the temperature dependence of  $a_w$  and/or water uptake and release may be retarded due to slow diffusion of water into highly viscous and amorphous phases (glasses) (Zobrist et al., 2008, 2011; Bones et al., 2012).

- <sup>5</sup> Homogeneous ice nucleation in the upper troposphere occurs in aqueous aerosol particles that subsequently grow into large ice crystals by deposition of water vapour, in part supplied from evaporating supersaturated cloud droplets. Unless solution droplets become glassy, homogeneous ice nucleation in supercooled aqueous solutions does not depend on the specific nature of the solute; rather knowing the thermodynamic properties of the solution in terms of water activity ( $a_w$ ) is sufficient, which implicitly ac-
- <sup>10</sup> properties of the solution in terms of water activity  $(a_w)$  is sufficient, which implicitly accounts for specific properties of the solutes (Koop et al., 2000; Koop and Zobrist, 2009). Thus,  $a_w$  of solutions is a crucial parameter for homogeneous ice nucleation. Similarly, for heterogeneous freezing in immersion mode,  $a_w$  based models are applicable (Knopf and Alpert, 2013).
- <sup>15</sup> For a correct description of the ice nucleation process, deviations from ideal mixing in aqueous solutions droplets have to be taken into account. This is achieved by using activity coefficients to describe solution non-ideality. For water, with a thermodynamic activity defined on a mole fraction basis, its mole fraction-based activity coefficient is defined as  $\gamma_w = a_w/x_w$ , where  $x_w$  is the mole fraction of water. Activity coefficients may ex-
- hibit considerable temperature dependence, which has to be parametrised explicitly in thermodynamic models in order to give accurate predictions for the level of non-ideality over a large range of temperatures. However, for aqueous organic solutions, thermodynamic models based on the UNIQUAC (UNIversal QUAsi Chemical) model (Abrams and Prausnitz, 1975) or its group contribution version UNIFAC (UNIquac Functional
- <sup>25</sup> group Activity Coefficients) (Fredenslund et al., 1975) may not provide reliable predictions of activity coefficients when they are used outside the temperature range for which they have been parametrised, i.e., at T < 275 K or T > 400 K. Improvement of these models for atmospheric conditions is strongly limited by the limited availability of reliable activity coefficient data for T < 290 K (Saxena and Hildemann, 1997; Lohmann

![](_page_3_Picture_6.jpeg)

et al., 2001; Marcolli and Peter, 2005). Performing new  $a_w$  measurements by combining different measurement techniques (e.g., EDB, DSC, total gas phase pressure measurements) can provide the experimental data required for improved parametrisations of these thermodynamic models, especially for low temperatures. Studies of the

<sup>5</sup> temperature dependence of activity coefficients have been carried out for a few atmospherically relevant inorganic acids and salts such as  $H_2SO_4$ ,  $(NH_4)_2SO_4$ , and  $NH_4NO_3$ (Knopf et al., 2003; Tang and Munkelwitz, 1993; Rodebush, 1918; Clegg et al., 1998), revealing quite distinct tendencies:  $a_w$  of dilute  $H_2SO_4$  solutions are nearly independent of temperature, while in the case of  $NH_4NO_3$ ,  $a_w$  increases with decreasing temperature (Koop, 2004).

For aqueous organic solutions, Zobrist et al. (2003) compared the  $a_w$  data of various poly(ethylene glycol) (PEG) solutions in the stable and supercooled range and noticed that the  $a_w$  of fixed-composition PEG solutions decreases with decreasing temperature. The influence of temperature on  $a_w$  becomes more pronounced with decreasing

- <sup>15</sup> temperature as well as with increasing solute concentration. Zobrist et al. (2008) determined the temperature dependence of water activity coefficients in polyols and sugars at atmospheric pressure in the temperature range from the ice melting curve to 313 K and used these data to convert ice freezing temperatures from the mass fraction composition to the  $a_w$  scale. They found that if the temperature dependence is neglected,
- errors on the order of 10–15% may result for  $a_w$  at the homogeneous ice freezing temperature. These examples show that in the case of aqueous organic solutions, the temperature dependence of  $a_w$  can be atmospherically important.

Low temperature  $a_w$  data from the peer-reviewed literature are mostly restricted to solid–liquid equilibria (SLE). SLE measurements provide data on the melting curves of

<sup>25</sup> ice and/or the organic component, i.e., for the specific solution compositions referring to the solid–liquid phase boundary over a certain temperature range. To measure  $a_w$  at low temperatures over a wide composition and temperature range, we therefore combine different measurement techniques including a dew point water activity meter for  $a_w$  measurements at temperatures higher than 273 K, differential scanning calorimetry

![](_page_4_Picture_7.jpeg)

(DSC) to determine SLE, and hygroscopicity measurements of single levitated particles in an electrodynamic balance (EDB). To complement these measurements, we developed a laboratory setup to measure total gas phase pressure over solutions at low temperatures. A detailed description of the measurement techniques is given in

- Sect. 2. Measurements were performed for binary aqueous organic mixtures covering functional groups and multifunctional organic compounds that are frequently observed in tropospheric aerosols. The organic functional groups considered include hydroxyl (OH), carboxyl (COOH), ketone (CH<sub>n</sub>CO), ester (CCOO), ether (CH<sub>n</sub>O), aromatic carbon (ACH<sub>n</sub>), aromatic carbon-alcohol ("phenol group", ACOH), where "*n*" denotes the number of H atoms in the functional group. These new measurements provide useful data for the improvement of thermodynamic models and for ice applications in nucleing the second second
- data for the improvement of thermodynamic models and for ice applications in nucleation studies.

## 2 Measurement techniques

To perform measurements over a wide concentration and temperature range for at-<sup>15</sup> mospherically relevant organic compounds we use different measurement techniques, each of which covers a certain temperature range.

# 2.1 Differential Scanning Calorimetry (DSC)

Solid–liquid equilibria data were obtained by measuring the melting temperatures ( $T_m$ ) of aqueous organic solutions for emulsified samples with a DSC instrument (Q10 from

- <sup>20</sup> TA Instruments) following the procedure described in Zobrist et al. (2008). These measurements also provide homogeneous freezing temperatures ( $T_{hom}$ ) as a function of aqueous solution composition. Water-in-oil emulsions with droplet diameters in the range of 0.5 to 5 µm were prepared by adding four parts by volume of a 5 wt% lanolin/mineral oil solution (Fluka/Aldrich) to one part by volume of an aqueous solution and stirring with a rotor-stator homogenizer (Polytron PT 1300D with a PT-DA 1307/2EC
- **Discussion** Paper **ACPD** 14, 12673–12726, 2014 **Temperature** dependency in aqueous organic **Discussion** Paper solutions G. Ganbavale et al. **Title Page** Introduction Abstract Discussion Paper Conclusions References **Figures** Tables Back Close Discussion Full Screen / Esc **Printer-friendly Version** Paper Interactive Discussion

dispersing aggregate) for 40 s at 7000 RPM. Samples (8 to 10 mg) were pipetted into the DSC pans, which were immediately sealed to prevent any evaporation. All aqueous solutions were made with distilled and deionized water (resistivity > 18.2 MΩ cm, total organic impurities < 5 ppb). Each experiment comprised three subsequent cooling cy-</li>
 <sup>5</sup> cles starting from 293 K. The first and the last cycle were run with a cooling rate of 10 K min<sup>-1</sup> (used as control for the emulsion stability) and the second cycle with a cooling rate of 1 K min<sup>-1</sup> (used for the melting and freezing point evaluation). Ice melting temperatures were evaluated at the melting peak maxima of the heating cycle run at a rate of 1 K min<sup>-1</sup>. A detailed calibration of the DSC resulted in a maximum uncertainty for ice melting temperatures of ±0.8 K (Zobrist et al., 2008).

Aqueous solutions of the following organic compounds were investigated: 1,3propanediol (Aldrich, 98%), 1,5-pentanediol (Fluka,  $\geq$  97%), 1,2-hexanediol (Aldrich, 97%), glycolic acid (Aldrich, 99%), pyruvic acid (Aldrich, 99%), methoxyacetic acid (Aldrich, 99%), 2-ethoxyethyl acetate (Aldrich, 99%), D-sorbitol (Aldrich,  $\geq$  98%), sutrose (sigma,  $\geq$  99%), resorcinol (Aldrich, 99%), 2-(2-ethoxyethoxy)ethanol (Aldrich,  $\geq$  99%), and M5, a multicomponent dicarboxylic acid mixture composed of DL-malic acid (Aldrich, 99%), maleic acid (Aldrich, 99%), malonic acid (Aldrich, 99%), glutaric acid (Aldrich, 99%), and methylsuccinnic acid (Aldrich, 99%). Table 1 lists the compounds used together with a selection of their physical properties. The ice melting  $(T_m(x_{org}))$  and homogeneous freezing temperatures  $T_{hom}(x_{org})$ , obtained by DSC mea-

 $_{20}$  ( $I_m(x_{org})$ ) and homogeneous freezing temperatures  $I_{hom}(x_{org})$ , obtained by DSC measurements for the aqueous organic mixtures at different liquid solution compositions given as mole fraction of the organic component,  $x_{org}$ , are provided in Table 2.

We use the parametrisation by Koop and Zobrist (2009) to calculate  $a_w$  at the melting temperature of ice for a known solution composition. The vapour pressures for solid phase (superscript S) pure hexagonal ice,  $p_{ice}^{\circ,S}$  (=  $p_w^{\circ,S}$ ), and water in equilibrium with the liquid (superscript L) solution,  $p_w^L$ , on the melting curve (SLE) are the same:

$$\rho_{\rm w}^{\rm L}(T=T_{\rm m}(x_{\rm org}))=\rho_{\rm ice}^{\circ,\rm S}(T=T_{\rm m}(x_{\rm org})),$$

![](_page_6_Figure_5.jpeg)

(1)

where vapour–liquid, vapour–solid, and solid–liquid equilibria exist. At SLE, the water activity of the aqueous organic solution,  $a_w^{SLE}(T, p)$ , in equilibrium with ice can therefore be expressed by:

$$a_{\rm w}^{\rm SLE}(T = T_{\rm m}(x_{\rm org}), \rho) = \frac{\rho_{\rm ice}^{\circ, \rm S}(T = T_{\rm m})}{\rho_{\rm w}^{\circ, \rm L}(T = T_{\rm m})},$$

where  $p_w^{\circ,L}(T = T_m)$  is the vapour pressure of pure liquid (supercooled) water at  $T_m$ . We use the Koop and Zobrist (2009) temperature dependent parametrisation which is based on Murphy and Koop (2006) to calculate  $a_w$  at SLE, valid at low (ambient) pressure in the temperature range 173 K < T < 273.15 K:

$$a_{w}^{\text{SLE}}(T,p) = [\exp(9.550426 - 5723.265/T + 3.53068 \cdot \ln(T) - 0.00728332 \cdot T)] \\ \cdot [\exp(54.842763 - 6763.22/T - 4.210 \cdot \ln(T) + 0.000367 \cdot T) \\ + \tanh(0.0415 \cdot (T - 218.8)) \cdot (53.878 - 1331.22/T) \\ - 9.44523 \cdot \ln(T) + 0.014025 \cdot T))]^{-1}.$$

### **2.2** Water activity measurements

5

Water activity measurements were performed using an AquaLab water activity meter (Model 3B, Decagon Devices, USA). The instrument employs the chilled mirror method to determine the dew point temperature of the gas phase in equilibrium with the sample. Infrared thermometry in addition indicates the sample temperature. This <sup>20</sup> instrument allows  $a_w$  measurements in the temperature range from 288 to 313 K for bulk samples. For most measurements the volatile sample block available as an accessory with the instrument was used since several of the organic compounds used have rather low, but potentially significant vapour pressures at the probed temperatures. Experimental errors for the volatile sample block are specified as  $\pm 0.015a_w$  by the manufacturer. The sample block was frequently calibrated and corrected for drifts

![](_page_7_Picture_6.jpeg)

(2)

(3)

and offsets using saturated salt solutions and distilled water samples covering the relevant  $a_w$  range. For solutions of low-volatility organic compounds and water, the standard sample block was used with a specified accuracy of  $\pm 0.003a_w$ . Table 3 provides measured bulk solution  $a_w$  data for sorbitol, resorcinol, glycolic acid, pyruvic acid, and <sup>5</sup> sucrose at 298.15 K. For some selected systems additional bulk  $a_w$  measurements at 279 K were carried out by turning off the temperature control within the AquaLab instrument and performing the measurements in a cold room at a constant temperature of 279 K (with an uncertainty in room temperature of  $\pm 0.5$  K). Water activity measurements were performed for aqueous solutions of 1,4-butanediol, M5, methoxyacetic

acid, and 2-(2-ethoxyethoxy)ethanol, for the temperature range 279 to 313 K.

#### 2.3 Electrodynamic balance (EDB) measurements

The basic experimental setup has been described previously (Krieger et al., 2000; Zobrist et al., 2011). An electrically charged particle (typically 2-20 µm in radius) is levitated in an electrodynamic balance. The balance is hosted within a three wall glass chamber with a cooling agent flowing between the inner walls and an insulation vac-15 uum between the outer walls. A constant flow of a  $N_2/H_2O$  mixture with a controlled H<sub>2</sub>O partial pressure is pumped continuously through the chamber at a constant total pressure, adjustable between 20 and 100 kPa. A charged, liquid particle is injected in the trap using a single particle generator (Hewlett-Packard 51633A ink jet cartridge) and is levitated in the balance, while keeping the temperature constant and increasing or decreasing the relative humidity within the chamber continuously. This is achieved by changing the  $N_2/H_2O$  ratio in the gas phase, using automatic mass flow controllers. The humidity sensor placed close to the particle registers the RH with an accuracy of ±1.5 % RH (U.P.S.I. France, Model G-TUS.13) between 10 and 90 % RH. The sensor was calibrated directly in the trap using the deliguescence relative humidity of different 25

salts at different temperatures in the range of interest. The concentration of the particle can be calculated from the DC voltage compensating the gravitational force when the dry particle mass is known (measured at RH < 10%). An alternative, independent

![](_page_8_Figure_5.jpeg)

measure of concentration is based on Mie-resonance spectroscopy. We use a ball lens type point source LED as a "white light" source to focus the light on the levitated particle and a pierced mirror to collect Mie resonance spectra in a backscattering geometry. Radius information is retrieved from the Mie-resonance spectra as described

- <sup>5</sup> by Zobrist et al. (2011). To convert radius to mass and concentration, we assume ideal volume mixing (linear additivity of molar volumes) to calculate the density of aqueous solutions. For the M5 mixture we use a molar volume of  $86.62 \text{ cm}^3 \text{ mol}^{-1}$  and a molarity based linear parametrisation to calculate the refractive index,  $n_D$ , at 589 nm for aqueous M5 solutions of different concentrations ( $n_D = 1.3334 + 0.01297 \times \text{molarity}$ ).
- <sup>10</sup> Since the optically retrieved concentration data are less noisy and more stable with respect to drifts over longer measurement periods, we use those for the water activity vs. concentration data presented in Tables 10 and 11.

### 2.4 Total gas phase pressure measurements

We use total gas phase pressure measurements of binary aqueous organic solutions to determine *a*<sub>w</sub> at low temperatures over a wide concentration range. The organic components of the binary systems were selected such that their vapour pressure contribution to the total pressure is irrelevant, i.e., substantially lower than the vapour pressure of water in the considered temperature range. Given this prerequisite, measured total pressures can be evaluated as being the vapour pressures of water in vapour-liquid equilibrium with the binary solutions at measured temperatures and known composi-

- 20 equilibrium with the binary solutions at measured temperatures and known compositions. Since we attribute measured total pressures entirely to water vapour, we are restricted to binary mixtures with low organic vapour pressures that lie within or below the measurement uncertainty of the total pressure over the whole investigated composition range. The experimental setup used for the total gas phase pressure measurements
- is shown in Fig. 1. The setup consists of a round bottom flask (500 mL) in which an aqueous organic mixture of a particular composition is filled. For the total gas phase pressure measurements, the flask is immersed in a thermostated ethanol bath (initially set to 223 K) whose temperature is slowly ramped up to 290 K. The flask can be

![](_page_9_Figure_7.jpeg)

evacuated to ~  $10^{-6}$  Pa using a vacuum pump and cooled in liquid N<sub>2</sub> for the purpose of degassing residual air naturally present in the samples. Glass beads (~ 50 g, ~ 3 mm diameter) are added to the flask to prevent undesirable foaming of the solution during degassing cycles, which may occur when a frozen solution thaws relatively quickly at

- room temperature after cooling to liquid nitrogen temperature. Foaming may often lead to splashing of solution droplets up to the neck of the flask, where the droplets remain without contact to the rest of the sample. If such droplets were present above the level of the ethanol bath into which the flask is submerged to maintain a temperaturecontrolled environment (Julabo, FP50 thermostat), they would be at a different temper-
- <sup>10</sup> ature than the bulk solution and present a cause of systematic errors. The pressure increase during the temperature ramp is registered by two pressure heads operating in the pressure ranges  $P_1$  (Pfeiffer Vacuum, CMR-262, range: 1 to 1 × 10<sup>4</sup> Pa) and  $P_2$ (Pfeiffer Vacuum, PKR-251, range: 1 × 10<sup>-2</sup> to 110 Pa).
- A typical experiment involves the following operational procedure: a binary aqueous solution mixture (volume of 3 to 5 mL) is added to the flask. To remove the residual gases (e.g., N<sub>2</sub>, O<sub>2</sub>, Ar, and CO<sub>2</sub>) from the solution, a thorough degassing procedure is carried out for which the flask is placed in a liquid N<sub>2</sub> bath (at T  $\approx$  77 K) and the valve to the vacuum pump is opened to remove the gas phase above the solution until a low pressure of  $\sim 10^{-6}$  Pa is reached. Thereafter, the valve is closed again and the solution is slowly brought back to room temperature. These warming and cooling
- cycles are carried out about 2–3 times so that all the residual gases are removed. These degassing cycles are assumed to lead only to a very small loss of water via the gas phase, so that the mixture composition remains practically unaffected. Before the actual measurement starts, the sample flask is once more cooled in liquid  $N_2$  and
- then transferred to the ethanol bath, which is held at 223 K, and further evacuated until a constant pressure is reached to allow the removal of any remaining dissolved gases from the solution and ensuring that the total pressure measured by the pressure sensors corresponds, within uncertainty, to the one of water vapour alone. The valves to the pressure heads are then opened and the total gas phase pressure measurement is

![](_page_10_Figure_6.jpeg)

carried out in the temperature range from 228 to 290 K by increasing the temperature of the ethanol bath at a constant rate of  $10.6 \,\mathrm{K} \,\mathrm{h}^{-1}$ , over a time period of 350 min. The measured constant leak rate of the instrument, which we measured for the same time interval as the actual experiment, is subtracted from the total pressure data and from

- <sup>5</sup> the resulting values, water vapour pressure data for the particular system compositions and temperatures are obtained. Pure water-ice was measured as a reference sample. From this we estimate the error in the measured water activity to be  $\pm 0.015$ . For the investigated systems,  $a_w$  values are derived by dividing the measured water vapour pressure by the calculated liquid-state vapour pressure of pure water (liquid-state saturation vapour pressure) at the prescribed temperature of the ethanol bath using the
- uration vapour pressure) at the prescribed to parametrisation by Murphy and Koop (2006).

## 3 Results

The total gas phase pressure measurements were performed for four aqueous organic mixtures namely for aqueous solutions of 1,4-butanediol, M5, methoxyacetic acid and <sup>15</sup> methoxyacetic acid. These measurements were complemented by bulk solution  $a_w$ measurements using the water activity meter in the temperature range from 313 to 279 K, and SLE ice melting and freezing data measured with the DSC in the temperature range from 273 to ~ 200 K, to obtain  $a_w$  data coverage over a wide range of temperatures and concentrations. In the case of aqueous M5 mixtures, additional  $a_w$  measurements to temperatures as low as 233 K were obtained from single-particle measurements using the EDB.

#### 3.1 1,4-Butanediol

Figure 2 shows  $a_w$  measurements for aqueous 1,4-butanediol solutions ob tained from the different measurement techniques. The ice melting curve as a function of  $a_w$  (dashed blue curve) is calculated using the parametrisation by Koop and Zobrist

![](_page_11_Figure_7.jpeg)

(2009). The dash-dotted lines represent the parametrisation by Zobrist et al. (2008) for the different compositions given in terms of mass percent (wt%). Zobrist et al. (2008) used experimental ice melting temperatures and bulk  $a_w$  data and applied a composition and temperature dependent parametrisation for water activities of specific mixtures

- <sup>5</sup> with 7 coefficients fitted to their data sets. The solid lines (both coloured and grey) show the derived  $a_w$  values from the total gas phase pressure measurements for concentrations from 20 to 90 wt% over a temperature range from 240 to 290 K. The measured data is considered reliable within the uncertainty of the method (±0.03 of  $a_w$ ). The grey part of these lines do not reflect the  $a_w$  of the solutions; rather, they are substantially
- influenced by the presence of solid phases and/or reflect undesired partial pressure contributions from residual gases (N<sub>2</sub>, O<sub>2</sub>), which may be present to a small extent in the solutions even after several cycles of degassing. Such artefacts caused by residual, highly volatile (supercritical) gases become much more important towards low temperatures relative to the much lower partial pressure signal from water, essentially enlarging
- <sup>15</sup> the uncertainties of the derived water activities beyond a practical limit for a meaningful data evaluation. The grey portions of the curves merge within uncertainty on the SLE phase boundary of the aqueous solution and ice for  $a_w > 0.76$  and analogously on the 1,4-butanediol SLE line for  $a_w < 0.70$ . Eutectic melting is observed at ~ 243 to 245 K as a horizontal peak to low  $a_w$  because the sample temperature remains constant during
- <sup>20</sup> the melting process while the prescribed temperature in the surrounding ethanol bath, which is used to derive the  $a_w$  data shown in Fig. 2, keeps increasing. The area of the phase diagram below the melting curve of one of the components, i.e., the green and blue shaded areas, is not accessible for  $a_w$  measurements by bulk techniques, because the presence of solid phases alters the initial (known) composition of the re-
- <sup>25</sup> maining liquid solution. The dips visible on the curves around  $T \approx 275$  K of the total gas phase pressure measurements of the 20 and 40 wt% mixtures, are artefacts, which are probably due to the melting of tiny droplets that may have splashed to the neck of the flask while degassing the solution prior to the actual measurements.

![](_page_12_Figure_6.jpeg)

All measurements taken together provide a consistent picture of the temperature dependence of  $a_w$  of aqueous 1,4-butanediol, namely a more or less constant decrease of  $a_w$  with decreasing temperature, which is well described by the parametrisation of Zobrist et al. (2008) (for bulk solutions at least until the intersection with the melting <sup>5</sup> curves). Table 4 provides  $a_w$  data obtained from the Aqualab water activity meter. Table 12 lists the  $a_w$  data derived from the total gas phase pressure measurements.

## 3.2 Methoxyacetic acid

Figure 3 shows  $a_w$  measurements of aqueous methoxyacetic acid covering the concentration range from 10 to 90 wt% and temperatures from 240 to 298 K using different experimental techniques. Water activity of the aqueous solution in equilibrium with ice, i.e., SLE for concentrations from 10 to 60 wt% covering a temperature range of 251 to 272 K are obtained using DSC measurements (pentagons). The grey solid lines at lower concentrations represent the  $a_w$  data derived from total gas phase pressure measurements which merge on the ice melting curve ( $a_w > 0.8$ ), while the grey

- <sup>15</sup> solid lines at higher concentrations ( $a_w < 0.7$ ) follow the melting curve of methoxyacetic acid. Eutectic melting is observed at  $T \approx 253$  K. The colour shaded region below the ice-melting and the methoxyacetic acid melting curves are not accessible for total gas phase pressure measurements, because the presence of solid phases influences the solution concentration (above the eutectic melting line), due to which the measured  $a_w$
- will not correspond to the initial composition of the solution being measured. The broad horizontal peaks to high  $a_w$  on the 50, 60, and 70 wt% lines of the total gas phase pressure measurements indicate an increased total pressure whose origin is unclear, but points to artefacts (perhaps due to the melting of small solution droplets, splashed to the neck of the glass flask, that are of lower organic concentrations and/or at higher temperature).

The measured  $a_w$  using the water activity meter and the  $a_w$  data derived from total pressure measurements indicate a small temperature dependence and are consistent within 2 to 5% at higher concentration wt% of methoxyacetic acid. A reason for

![](_page_13_Picture_6.jpeg)

this deviation might be that full equilibration of the solution was not reached after the melting of solid methoxyacetic acid over the timescale of the experiment. In the more dilute composition range with respect to the organic component, deviations  $< 2 \% a_w$  are found, indicating the principle validity of the total pressure data in the temperature range considered reliable. Measurements and the parametrisation for the temperature dependence of water activity by Zobrist et al. (2008) (dashed-dotted lines in Fig. 3) indicate a small  $a_w$  decrease with the decrease in the temperature. The bulk  $a_w$  data from the water activity meter measurements is provided in Table 5. The calculated  $a_w$  data obtained from the total pressure experiments for various solution compositions

<sup>10</sup> are given in Table 13.

## 3.3 2-(2-Ethoxyethoxy)ethanol

Water activity measurements for aqueous solutions of 2-(2-ethoxyethoxy)ethanol (also known as carbitol) in the concentration range from 10 to 90 wt % of organic and the temperature range from 230 to 298 K are presented in Fig. 4. DSC measurements were performed for a concentration range of 10 to 40 wt% covering the temperature range from 248 to 272 K. Bulk  $a_w$  measurements using the water activity meter in the

- temperature range from 279 to 298 K are listed in Table 6 and shown as coloured circles in Fig. 4. Eutectic melting was not observed since the melting point of pure 2-(2-ethoxyethoxy)ethanol ( $T_m = 197$  K) is lower than the considered temperature range.
- <sup>20</sup> The  $a_w$  data measured using the water activity meter and DSC data are in good agreement with the parametrised temperature dependence by Zobrist et al. (2008), the latter is displayed by the coloured dash-dotted lines. Both the  $a_w$  measurements and the parametrisation indicate a decrease in  $a_w$  with decrease in temperature. The  $a_w$  derived from the total gas phase pressure measurements are in good agreement with the  $a_w$
- <sup>25</sup> data obtained from the water activity meter and the parametrisation for T > 265 K and for mass fractions of the organic component,  $w_{org}$ , of 60 wt %  $\leq w_{org} \leq$  90 wt %. At lower temperatures,  $a_w$  strongly increases with decreasing temperature for  $w_{org} \geq$  80 wt %. At T = 225 K, all the solid lines converge. Such a temperature dependence of water

![](_page_14_Figure_7.jpeg)

activity is expected for solutions that exhibit a phase transition (as seen when the curves bend and follow the melting curves in the previous systems, where two solid phases exist at lower T). Another possibility for such a convergence would be that a phase transition in terms of a liquid-liquid phase separation occurs at low temper-

- atures. It should be noted that measurement artefacts would not be expected to lead to a convergence of the curves onto the ice melting curve at a common temperature. Complementary measurements using the EDB could not be performed for this system due to the comparably high vapour pressure of 2-(2-ethoxyethoxy)ethanol at room temperature, which leads to fast evaporation during injection of the particle into the EDB.
- At lower  $w_{\text{org}}$  (10 to 50 wt%) the solid coloured lines show good agreement with the parametrisation by Zobrist et al. (2008). The water activity data derived from the total gas phase pressure measurements for various concentrations are provided in Table 14.

### 3.4 M5 (multicomponent dicarboxylic acid) mixture

Figure 5 shows  $a_w$  measurements for aqueous solutions of M5 over a concentration range from 10 to 84 wt% M5 (i.e., up to the solution saturation limit at 298 K) in the temperature range from 225 to 313 K. Bulk  $a_w$  measurements performed in the temperature range from 279 to 313 K are represented by the coloured circles and are listed in Tables 7 and 8. Humidity cycles of single levitated particles in the EDB were performed in the temperature range from 233 to 289 K and are represented by filled coloured diamonds. M5 mixture compositions and  $a_w$  data for the aqueous solutions are listed in Tables 9–11. The total gas phase pressure measurements are represented by solid and dashed lines (dashed for M5 mass fractions from 75 to 84 wt%) in Fig. 5. For mass fractions  $w_{M5} > 50$  wt% both EDB measurements and total gas phase pressure measurements show an increase in  $a_w$  with decreasing temperature, with a stronger effect of bights appearents the presented by an and total gas phase pressure measurements show an increase in  $a_w$  with decreasing temperature, with a stronger effect

<sup>25</sup> at higher concentrations. No simple (pseudo-binary) eutectic melting was observed because in case of the total gas phase pressure measurements individual components of the M5 mixture crystallise at different temperatures and mass fractions of M5. We assume that the total pressure data for  $w_{M5} > 70$  wt% (shown by dashed coloured lines

![](_page_15_Figure_6.jpeg)

in Fig. 5) are influenced by the crystallisation of certain components of the M5 mixture. These data are not considered to reflect the water activity of the initial mixtures and are therefore not tabulated. The same might be true for the bulk  $a_w$  data points at 279 K and concentrations of 80 and 84 wt%, which might explain their high water activity. Ta-

<sup>5</sup> ble 15 provides the composition of the M5 + water mixtures used for the total gas phase pressure measurements; corresponding water activities are listed in Table 16. Thus it should be noted, both M5 and 2-(2-ethoxyethoxy)ethanol solutions show a similar effect of increase in  $a_w$  with decrease in temperature at higher concentrations.

## 4 Discussion

# 10 4.1 Measurement techniques: scope and limitations

Water activity as a function of solute concentration can be measured either for bulk solutions or by single particle measurement techniques. To measure *a*<sub>w</sub> of bulk solutions, commercial dew point water activity meters are probably the best choice because they have high accuracy and cover a wide concentration range, thus, providing *a*<sub>w</sub> data from saturated to dilute solutions. However, their accuracy decreases for water activities close to 1. Therefore they are less suited for very dilute solutions. Moreover, the commercially available water activity meters typically provide data for temperatures > 273 K and experiments are time consuming when point-to-point measurements have to be performed over a wide temperature range.

- <sup>20</sup> The DSC technique provides accurate measurements of solid–liquid equilibrium curves. DSC measurements are performed at fixed composition and can provide  $T_m$ ,  $T_{hom}$  and glass transition temperatures ( $T_g$ ) of aqueous solutions. From the melting point depression of ice in aqueous solutions, accurate values of  $a_w$  on the SLE phase boundary of the aqueous solution and ice can be obtained, because the vapour pres-
- <sup>25</sup> sures of ice and water at atmospheric temperatures are well known (see Sect. 2.1). When oil-in-water emulsions instead of bulk samples are measured, homogeneous

![](_page_16_Picture_7.jpeg)

freezing temperatures  $T_{\text{hom}}$  can also be obtained. This is especially useful because homogeneous ice nucleation temperatures of aqueous solutions can be parametrised in terms of water activity following Koop et al. (2000); Koop and Zobrist (2009). The drawback of the DSC technique is that only  $a_w$  data on the ice melting curve, thus, over a limited range of compositison, are obtained.

The electrodynamic balance and optical tweezers in addition offer a possibility to measure the water activity via the mass or size change of hygroscopic, micrometresized particles Krieger et al. (2012). In both techniques, the particles are levitated without contact to any surface. Hence the supersatured concentration range can be investigated when the particles are exposed to humidity cycles. In the absence of liquid/gas phase diffusion limitations and given the relatively large diameters of solution

uid/gas phase diffusion limitations and given the relatively large diameters of solution droplets (Kelvin effect can be ignored), RH of the gas phase corresponds with  $a_w$  of the condensed phase. Thus, the water content of the levitated particle in terms of a mass or molar fraction can be calculated from the mass data when the dry particle mass

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- <sup>15</sup> is known. The more accurate radius data obtained from the Mie scattering pattern of an illuminating light beam can be used to derive mass or mole fractions of solute but need refractive index data and density data for the measurement evaluation. The available temperature range depends on the cooling and heating possibilities of the specific setup. Since the particles are airborne without contact to any surface (walls),  $a_w$  of
- highly supersaturated solutions can be obtained, covering thus a concentration range that is not accessible by standard bulk methods.

Total gas phase pressure measurements over aqueous solutions can provide water activities when the measured total pressure can be completely ascribed to water vapour. Since this technique needs bulk volumes,  $a_w$  data can be obtained from dilute <sup>25</sup> up to saturated solutions. When the sample is kept in an externally regulated thermostat, temperature profiles can be run that provide water activity data of solutions at fixed concentration covering a wide temperature range. Apart from deriving  $a_w$  from the measured data, the total gas phase pressure measurements can also provide information about eutectic melting points and solid–liquid equilibria. However, accurate

![](_page_17_Picture_6.jpeg)

measurements of water activities rely on efficient removal of the residual gases from the solution to make sure that the measured total pressure corresponds to the water vapour pressure over the solution. Great care needs to be taken during the removal of residual gases since there is always the danger of overpumping which may change

- the concentration of the solution (removal of significant amounts of water). The total gas phase pressure measurements are also prone to artefacts. The freezing/thawing process can lead to composition heterogeneity in the sample. Some solutions show a strong foaming during melting leading to tiny droplets settling on the walls of the solution flask. These drops can influence the gas phase pressure when they are at
- a higher temperature than the bulk solution or when they are of a different composition due to arising heterogeneity during the freezing and thawing processes. To cover a large concentration and temperature range, organic substances with a low melting point and a high water solubility are required. These conditions strongly reduce the number of compounds for which total gas phase pressure measurements may provide
- <sup>15</sup> a large extension to composition/temperature ranges that are not covered by other methods. These drawbacks together with the rather difficult handling of the instrumental setup render total gas phase pressure measurements to be less attractive than the alternative techniques such as DSC and EDB measurements. A good temperature and concentration coverage can be achieved when  $a_w$  bulk measurements are combined <sup>20</sup> with EDB experiments.

## 4.2 Hydrogen bonding effects on water activity in aqueous solutions

Hydrogen bonds are strong electrostatic dipole-dipole interactions that occur between covalently bound hydrogen atoms and the free electron pair of a highly electronegative atom, such as nitrogen (N), oxygen (O) or fluorine (F). They have some features of covalent bonding since they are directional and can lead to interatomic distances shorter than the sum of the van der Waals radii of the involved atoms. Hydrogen bonds have a strong influence on the activity of the constituents in a solution. In aqueous solutions of alcohols and organic acids, organic–organic, organic–water, and water–water

![](_page_18_Figure_6.jpeg)

hydrogen bonds can form. In general, organic–water hydrogen bonds decrease  $a_w$  while association of organic molecules among each other leads to an increase in water activity. The strength and average number of hydrogen bonds per molecule depends on concentration and temperature. A strong increase of  $a_w$  with decreasing tempera-

ture can be rationalized in terms of association of solute molecules among each other. Therefore, the analysis of the hydrogen bonds present in aqueous solutions can help to understand the temperature dependence of water activity.

The carboxyl group of organic acids can build hydrogen bonds with water or other dicarboxylic acids. The hydrogen atoms of the carboxyl group form hydrogen bonds <sup>10</sup> with the free electron pair of the oxygen atoms of a carboxyl group of another acid molecule. Similarly, for dicarboxylic acids when the hydrogen bonds connect to another dicarboxylic acid molecule this leads to an association of dicarboxylic acids and formation of a dimer, which reduces the effective number of dissolved species and leads to a relative increase in water activity. Hydrogen bonds between water and dicarboxylic acids lead to a relative decrease of water activity.

Aqueous solutions of poly(ethylene glycol) (PEG) have attracted much attention because of their extraordinary mixing behavior with water and their importance in pharmaceutical and biomedical appliances (Dormidontova, 2002). 2-(2-ethoxyethoxy)ethanol shares with PEG the  $-CH_2-CH_2-O-$  repetition unit as the main structural feature. Different from PEG with two terminal hydroxyl groups, 2-(2-ethoxyethoxy)ethanol carries

one terminal methyl and one terminal hydroxyl group, which makes it less hydrophilic. The  $-CH_2-CH_2-O$  repetition unit lends 2-(2-ethoxyethoxy)ethanol and PEG a higher solubility than typical for ethers. This behavior can be rationalized by the increased structuring of water around PEG molecules. This structuring is a consequence of hy-

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drogen bonding and the ability of the PEG structure to fill out the natural cavities in the hydrogen bonded network of water. The increased structuring of the water is reflected in the large negative excess entropy of the solution, while its large positive excess heat capacity is due to the temperature sensitivity of the structure (Kjellander and Florin, 1981). This interplay of entropic and enthalpy contributions to the Gibbs energy leads

![](_page_19_Picture_6.jpeg)

to a closed loop miscibility gap at elevated temperature for aqueous PEG solutions with PEG molecular weights of 2200 g mol<sup>-1</sup> and higher (e.g., Dormidontova, 2004; Kjellander and Florin, 1981; Zobrist et al., 2003).

- PEG with molecular weight below 2000 g mol<sup>-1</sup> do not show liquid-liquid phase separation close to room temperature. However liquid-liquid equilibria are observed for aqueous triethylene glycol solutions (Salabat, 2010) and smaller poly(ethylene glycols) when a salt is added (e.g., Marcolli and Krieger, 2006; Ciobanu et al., 2009). Liquidliquid phase separation below room temperature is not easily accessible because of the competition with crystallisation of either the PEG, the ice phase or both phases depending on solution composition and temperature. Ciobanu et al. (2009) have in-
- vestigated in detail liquid-liquid phase separation of aqueous PEG-400 solutions when ammonium sulfate is added as a salting-out agent. This phase separation seems to persist or even grow with decreasing temperature. At the onset of liquid-liquid phase separation water activity lines of different concentration converge at a high  $a_w$  value.
- <sup>15</sup> Therefore the strong increase of  $a_w$  with decreasing temperature in aqueous M5 and 2-(2-ethoxyethoxy)ethanol solutions may be rationalized in terms of approaching a low temperature miscibility gap that is not experimentally accessible because it falls in the concentration/temperature range where ice forms.

# 4.3 Atmospheric implications

- <sup>20</sup> In the upper troposphere, one pathway for cirrus cloud formation is by means of homogeneous ice nucleation on/in liquid aqueous aerosol particles, which subsequently grow into supermicron-sized ice crystals by condensing water vapour. Water-activitybased ice nucleation theory can be employed to predict homogeneous ice nucleation temperatures and corresponding ice nucleation rate coefficients for aqueous solution
- <sup>25</sup> droplets without explicit knowledge of the nature of the solute (Koop et al., 2000; Koop and Zobrist, 2009). While for known mixture compositions the ice melting temperature and the corresponding equilibrium water activity can be measured and/or parametrised precisely (Koop et al., 2000; Koop, 2004; Knopf and Rigg, 2011), the same level of

![](_page_20_Picture_7.jpeg)

information is not accessible experimentally in the case of supercooled aqueous solution droplets, which exhibit homogeneous freezing of ice at a temperature lower than the corresponding melting point, denoted as *T*<sub>hom</sub>. As is well understood in the case of the ice melting process, with the melting temperature being a function of aqueous solution composition (melting point depression), similar behavior is observed in droplet freezing experiments for the composition and temperature dependence of the homogeneous freezing process. However, the water activity at the freezing temperature is typically not accessible in experiments and/or subject to relatively large uncertainty.

Koop et al. (2000) suggest that  $a_w$  at the homogeneous freezing temperature  $T_{hom}(x_{org})$  can be obtained from the corresponding  $a_w$  determined at the melting temperature  $T_m(x_{org})$  of the solution at the same composition with the assumption that  $a_w$  does not change significantly over the temperature difference from  $T_m(x_{org})$  to  $T_{hom}(x_{org})$ . This approach has been tested and shown to be a good approximation for a variety of inorganic solutions (Koop et al., 2000; Koop, 2004), but may lead to significant errors in predictions of the homogeneous freezing temperatures for aqueous organic solutions. Figure 6 shows  $a_w$  at melting and freezing conditions for aqueous organic solutions investigated in this work. The  $T_{hom}(x_{org})$  and  $T_m(x_{org})$  temperatures

for the investigated in this work. The  $T_{hom}(x_{org})$  and  $T_m(x_{org})$  temperatures for the investigated organic solutions are measured using the different measurement techniques stated in this paper. The  $a_w$  data for the ice melting curve are calculated using the parametrisation by Koop and Zobrist (2009). By applying the assumption

that solutions do not show significant change in  $a_w$  with temperature from  $T_m(x_{org})$  to  $T_{hom}(x_{org})$ , i.e., neglecting the temperature dependence, the  $a_w$  values evaluated this way show considerable scatter around the ice-freezing curve in Fig. 6a. The homogeneous ice freezing curve, shown as dashed black line in Fig. 6, is calculated by shifting

<sup>25</sup> the melting curve by  $\Delta a_w = 0.313$ , as suggested by Koop and Zobrist (2009). The  $a_w$  data derived with this assumption in most cases lie below the ice freezing curve. This result could be interpreted as freezing occurring at lower temperatures than predicted by the water-activity-based ice nucleation theory for the known mixture compositions in Fig. 6a. However, since the water-activity-based freezing curve parametrisation is

![](_page_21_Figure_5.jpeg)

actually very successful in describing the freezing behavior of many inorganic solutions, for which the experimental or predicted temperature dependence in water activity is small, it is more likely the case that assuming constant water activity from  $T_m(x_{org})$  to  $T_{hom}(x_{org})$  is the reason for the apparent discrepancy. Figure 6b shows the same experimental homogeneous freezing data, but in contrast to Fig. 6a, the corresponding water activities at the freezing temperature are estimated by using Eq. (3) at  $T_{hom}(x_{org})$  with a constant  $\Delta a_w = 0.313$  added to the  $a_w$  value determined on the melting curve for the same mixture. With this method of  $a_w$  data evaluation, the experimental data show much less scatter and outline in good approximation a single "experimental" freez-

- <sup>10</sup> ing curve. This freezing curve described by the ensemble of aqueous organic solution data is in relatively good agreement with the estimated homogeneous freezing curve according to Koop and Zobrist (2009). A slight, yet systematic deviation of the experimental data towards higher freezing temperatures/lower water activities is found, especially for the freezing temperatures below 220 K. These deviations may result in
- <sup>15</sup> significantly lower values of homogeneous nucleation rate coefficients ( $J_{hom}$ ) (Knopf and Rigg, 2011). A change of  $a_w$  by 0.025 may result in a change of  $J_{hom}$  by 6 orders of magnitude, which may significantly affect predictions of the onset of ice crystal formation in cloud microphysical models. A difference of 3 orders of magnitude in  $J_{hom}$ could delay or accelerate homogeneous ice nucleation by about an hour in an atmo-<sup>20</sup> spheric cloud formation simulation shown by Knopf and Rigg (2011).

The strong increase of  $a_w$  with decreasing temperature of aqueous M5 and 2-(2ethoxyethoxy)ethanol solutions at low temperatures and high solute concentrations has consequences for the hygroscopic growth of these systems at low temperatures. First, the water uptake assuming thermodynamic equilibrium between the gas and the

<sup>25</sup> condensed phase will be much smaller. Second, the low water content at these low temperatures will promote high viscosities or even glass formation leading in addition to kinetic limitations of water uptake.

![](_page_22_Figure_5.jpeg)

## 5 Conclusions

Water activity measurements for selected atmospherically relevant aqueous organic systems were carried out using different experimental techniques with the aim of covering a broad concentration and temperature range. Hygroscopicity measurements of

- single levitated aerosol particles with an electrodynamic balance cover RH between 10 and 90 % RH and temperature from 200 to 300 K. DSC measurements provide the melting and freezing point data at various solution concentrations and can be used to derive composition and temperature dependent water activities. To complement these measurements, total gas phase pressure measurements were performed for aqueous
   organic mixtures. The measured a<sub>w</sub> data obtained from the different experiments are consistent with each other and reveal that organic solutes exhibit differing water activity
- temperature dependence; both substantially increasing and decreasing  $a_w$  values with decreasing temperatures below 298 K are observed.

More accurate *a*<sub>w</sub> data at low temperatures are needed in the context of applications of homogeneous ice nucleation theory at upper tropospheric temperatures. The experiments presented in this study provide new equilibrium data sets useful for the development and improvement of thermodynamic activity coefficient models, such as UNIFAC (UNIquac Functional group Activity Coefficients) and AIOMFAC (Aerosol Inorganic–Organic Mixtures Functional groups Activity Coefficients). In turn, improved

thermodynamic models can be used for more accurate predictions of the temperature dependence of activity coefficients of water and other solution constituents, as well as equilibrium compositions of multiphase systems for mixtures and environmental conditions, for which experimental data is unavailable.

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![](_page_23_Picture_7.jpeg)

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![](_page_27_Picture_13.jpeg)

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**Table 1.** Selected physical properties of organic components used for the experiments: molar mass (*M*), melting point ( $T_m$ ), boiling point ( $T_b$ ) at standard pressure (101.325 kPa), and structure.

Organic compounds	Chemical formula	M (a mol <sup>-1</sup> )	T (K)	<i>T.</i> (K)	Structure
			, m (14)	7 <sub>B</sub> (10)	
1,3-propanediol	$C_3H_8O_2$	76.094	246.15	487.55	но он
1,4-butanediol	$C_4H_{10}O_2$	90.121	292.15	501.15	
1,5-pentanediol	$C_5H_{12}O_2$	104.148	257.15	512.15	но он
1,2-hexanediol	C <sub>6</sub> H <sub>14</sub> O <sub>2</sub>	118.174	318.15	496.65	ОН
sorbitol	$C_6H_{14}O_2$	182.172	372.15	568.15	но сн
					$\gamma \gamma$
resorcinol	$C_6H_6O_2$	110.111	383.15	549.2	HO
					OH OH
					HO
					HOMMAN
sucrose	C <sub>12</sub> H <sub>22</sub> O <sub>11</sub>	342.116	459.15	decomposes	OH O
glycolic acid	$C_2H_4O_3$	76.051	353.15	385	HOLOH
					С
pyruvic acid	$C_3H_4O_3$	88.062	284.65	438.37	
methoxyacetic acid	$C_3H_6O_3$	90.077	281.15	477.5	OH
2-ethoxyethyl acetate	$C_{6}H_{12}O_{3}$	132.158	212.15	428.71	$\sim$
2-(2-ethoxyethoxy)ethanol (carbitol)	$C_6H_{14}O_3$	134.174	193.15	475.49	HO

![](_page_29_Figure_2.jpeg)

#### Table 1. Continued.

Organic compounds	Chemical formula	$M (\text{g mol}^{-1})$	7 <sub>m</sub> (K)	<i>Т</i> <sub>b</sub> (К)	Structure
					HO
malic acid	$C_4H_6O_5$	134.087	405.15	509.13	Т сн о о
malonic acid	$C_3H_4O_4$	104.061	409.15	339.99	но он
maleic acid	C4H4O4	116.011	403.15	256	
olutaric acid	C₅H₀O₄	132.042	369.65	575.96	но
3	-5 6-4				HO
methylsuccinic acid	$C_5H_8O_4$	132.042	383–388	decomposes	Ĩ ↓ <sup>oh</sup>

![](_page_30_Figure_2.jpeg)

<b>Table 2.</b> Homogeneous ice freezing $T_{hom}(x_{ora})$ , and ice melting $T_m(x_{ora})$ temperatures of the
investigated aqueous organic solutions. Solution compositions are expressed in mole fraction
x(organic) and temperature in K <sup>*</sup> .

x(1,3-propanediol)	$T_{\rm hom}(x_{\rm org})$	$T_{\rm m}(x_{\rm org})$	x(1,5-pentanediol)	$T_{\rm hom}(x_{\rm org})$	$T_{\rm m}(x_{\rm org})$
0.02637	231.23	270.47	0.01924	232.38	270.91
0.05656	223.55	266.78	0.04240	225.04	269.23
0.09225	212.74	262.08	0.06971	214.41	266.51
0.13630	200.20	255.69	0.10300	200.14	263.28
			0.15035		259.59
x(glycolic acid)	$T_{\rm hom}(x_{\rm org})$	$T_{\rm m}(x_{\rm org})$	x(pyruvic acid)	$T_{\rm hom}(x_{\rm org})$	$T_{\rm m}(x_{\rm org})$
0.02549	230.37	270.54	0.02223	231.72	276.76
0.05612	224.03	267.46	0.04879	223.68	267.06
0.09191	217.77	262.99	0.08090	210.84	261.65
0.13601	205.90	258.56	0.12026	209.45	253.58
x(1,2-hexanediol)	$T_{\rm hom}(x_{\rm org})$	$T_{\rm m}(x_{\rm org})$	x(sucrose)	$T_{\rm hom}(x_{\rm org})$	$T_{\rm m}(x_{\rm org})$
0.01664	232.13	271.57	0.01040	232.00	272.00
0.03658	225.01	271.54	0.01624	230.68	271.34
0.06099	223.21	271.32	0.02303	227.51	270.35
0.09197	223.17	271.05	0.03056	225.33	269.08
0.13202	222.20	270.69	0.03937	221.09	267.75
0.18637	221.35	269.71	0.04870	217.10	265.75
0.25834	216.14	268.86	0.06064		263.15
			0.07317		260.95

 $^*$  The accuracies of the freezing and melting point measurements are  $\pm 0.5\,K$  and  $\pm 0.4\,K,$  respectively.

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#### Table 2. Continued.

x(methoxyacetic acid)	$T_{\rm hom}(x_{\rm org})$	$T_{\rm m}(x_{\rm org})$	x(sorbitol)	$T_{\rm hom}(x_{\rm org})$	$T_{\rm m}(x_{\rm org})$
0.02180	231.96	271.48	0.01103	233.35	272.19
0.04757	244.69	268.98	0.02422	230.00	270.63
0.07904	218.43	265.95	0.04077	225.40	268.68
0.11765	208.79	262.64	0.06174	218.67	265.94
0.16709	193.55	257.67	0.08948	208.42	261.84
0.22937		251.31	0.12867		255.75
x(resorcinol)	$T_{\rm hom}(x_{\rm org})$	$T_{\rm m}(x_{\rm org})$	x(2-ethoxyethyl acetate)	$T_{\rm hom}(x_{\rm org})$	$T_{\rm m}(x_{\rm org})$
0.01818	231.55	271.98	0.01516	232.65	272.24
0.03956	228.20	270.67	0.03318	228.62	271.22
0.06567	226.60	268.84	0.05509	207.81	270.75
0.09848	223.28	267.15			
x(2-(2-ethoxyethoxy)ethanol)	$T_{\rm hom}(x_{\rm org})$	$T_{\rm m}(x_{\rm org})$			
0.01463	232.52	272.03			
0.03245	224.30	269.16			
0.05412	210.51	265.18			
0.08215		259.56			

\* The accuracies of the freezing and melting point measurements are  $\pm 0.5$  K and  $\pm 0.4$  K, respectively.

![](_page_32_Figure_3.jpeg)

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**Table 3.** Bulk water activity  $(a_w)$  measurements<sup>\*</sup> at 298.15 K of different aqueous organic solutions. Solution compositions are expressed in mole fraction *x*(organic).

x(sorbitol)	a <sub>w</sub>	x(resorcinol)	a <sub>w</sub>	x(glycolic acid)	a <sub>w</sub>	x(pyruvic acid)	a <sub>w</sub>	x(sucrose)	a <sub>w</sub>
0.0110	0.981	0.01818	0.985	0.02564	0.962	0.02267	0.966	0.01040	0.992
0.0242	0.956	0.03956	0.950	0.05588	0.934	0.04868	0.925	0.01624	0.988
0.0408	0.934	0.06567	0.938	0.09209	0.887	0.08068	0.883	0.02303	0.977
0.0617	0.898	0.09848	0.893	0.13644	0.838	0.12020	0.836	0.03056	0.971
0.0895	0.834	0.13992	0.877	0.19154	0.767	0.16986	0.778	0.03937	0.955
0.1287	0.752	0.19312	0.843	0.25957	0.665	0.23467	0.704	0.04870	0.939
0.1872	0.621	0.27629	0.836	0.35447	0.570	0.32089	0.596	0.06064	0.914
				0.48560	0.451	0.45005	0.434	0.07317	0.888
						0.61773	0.193	0.16527	0.692

\* The accuracy of the water activity measurements is specified as  $\pm 0.015$  (absolute range) in  $a_w$  and  $\leq 0.1$  K in temperature.

**Table 4.** Bulk water activity  $(a_w)$  measurements<sup>\*</sup> of water (1) + 1,4-butanediol (2) solutions at three different temperatures. Solution compositions are given in mole fraction  $(x_2)$  of the organic component (2).

<i>x</i> <sub>2</sub>	$a_{\rm w}(T=289.15{\rm K})$	$a_{\rm w}(T=298.15{\rm K})$	$a_{\rm w}(T = 313.15{\rm K})$
0.02192	0.978	0.985	0.982
0.04731	0.940	0.959	0.965
0.08020	0.918	0.935	0.958
0.11570	0.888	0.899	0.914
0.16604	0.844	0.860	0.871
0.23358	0.787	0.808	0.816
0.31787	0.717	0.733	0.745
0.44309	0.611	0.626	0.631
0.61890	0.424	0.445	0.450

 $^*$  The accuracy of the water activity measurements is specified as ±0.015 (absolute range) in  $a_{\rm w}$  and  $\leq$  0.1 K in temperature.

![](_page_34_Figure_3.jpeg)

**Table 5.** Bulk water activity  $(a_w)$  measurements<sup>\*</sup> of water (1) + methoxyacetic acid (2) solutions at three different temperatures. Solution compositions are given in mole fraction  $(x_2)$  of the organic component (2).

<i>x</i> <sub>2</sub>	$a_{\rm w}(T=279.15{\rm K})$	$a_{\rm w}(T=289.15{\rm K})$	$a_{\rm w}(T=298.15{\rm K})$
0.02185	0.953	0.961	0.969
0.04762	0.947	0.934	0.958
0.07433	0.922	0.920	0.939
0.11760	0.897	0.874	0.912
0.16659	0.866	0.844	0.880
0.22976	0.830	0.789	0.807
0.31637	0.702	0.720	0.715
0.44445	0.555	0.560	0.564
0.63961	0.346	0.346	0.354

\* The accuracy of the water activity measurements is specified as  $\pm 0.015$  (absolute range) in  $a_w$  and  $\leq 0.1$  K in temperature.

![](_page_35_Figure_3.jpeg)

**Table 6.** Bulk water activity  $(a_w)$  measurements<sup>\*</sup> of water (1) + 2-(2-ethoxyethoxy)ethanol (2) solutions at three different temperatures. Solution compositions are given in mole fraction( $x_2$ ) of the organic component (2).

<i>x</i> <sub>2</sub>	$a_{\rm w}(T=279.15{\rm K})$	$a_{\rm w}(T = 289.15 {\rm K})$	$a_{\rm w}(T = 298.15 {\rm K})$
0.01455	0.943	0.995	0.997
0.03275	0.935	0.986	0.982
0.05434	0.916	0.978	0.975
0.08182	0.900	0.929	0.933
0.11827	0.862	0.870	0.904
0.16747	0.822	0.839	0.866
0.23851	0.742	0.772	0.813
0.28670	0.703	0.738	0.780
0.34830	0.645	0.690	0.711
0.43139	0.582	0.613	0.641
0.54853	0.485	0.498	0.540
0.71022	0.328	0.333	0.367

\* The accuracy of the water activity measurements is specified as  $\pm 0.015$  (absolute range) in  $a_w$  and  $\leq 0.1$  K in temperature.

![](_page_36_Figure_3.jpeg)

<b>Table 7.</b> Bulk water activity $(a_w)$ measurements <sup>*</sup> of aqueous M5 solutions at 289.15, 298.15 and
313.15 K. The total organic mass fraction of the solution is given in the first column in wt% and
the composition in terms of individual components is given in mole fractions of the dicarboxylic
acids constituting the M5 mixture.

wt%	x(DL-malic)	x(malonic)	x(maleic)	x(glutaric)	x(methylsuccinic)	a <sub>w</sub> (289.15)	<i>a</i> <sub>w</sub> (298.15)	<i>a</i> <sub>w</sub> (313.15)
10.01	0.00247	0.00537	0.00193	0.00450	0.00200	0.984	0.991	0.989
20.01	0.00545	0.01184	0.00423	0.00991	0.00441	0.966	0.974	0.977
30.01	0.00908	0.01978	0.00711	0.01659	0.00735	0.945	0.951	0.960
39.68	0.01370	0.02852	0.01065	0.02492	0.01106	0.917	0.922	0.930
49.97	0.01962	0.04270	0.01528	0.03577	0.01587	0.873	0.877	0.880
54.99	0.02332	0.05074	0.01819	0.04251	0.01889	0.844	0.853	0.860
60.01	0.02766	0.06021	0.02157	0.05043	0.02241	0.804	0.807	0.807
69.98	0.03904	0.08503	0.03042	0.07120	0.03158	0.694	0.700	0.702
79.87	0.05631	0.12246	0.04385	0.10263	0.04556	0.542	0.543	0.543
84.41	0.06768	0.14725	0.05279	0.12339	0.05478	0.450	0.447	0.458

\* The accuracy of the water activity measurements is specified as  $\pm 0.003$  (absolute range) in  $a_w$  and  $\leq 0.1$  K in temperature.

![](_page_37_Figure_3.jpeg)

**Table 8.** Bulk water activity  $(a_w)$  measurements<sup>\*</sup> of aqueous M5 solutions at 279.15 K. The total organic fraction of the solution is given in the first column in wt% and the composition in terms of individual components is given in mole fractions of the dicarboxylic acids constituting the M5 mixture.

wt%	x(DL-malic)	x(malonic)	x(maleic)	x(glutaric)	x(methylsuccinic)	$a_{\rm w}$
10.01	0.00339	0.00602	0.00335	0.00259	0.00112	0.986
20.01	0.00768	0.00794	0.00921	0.00824	0.00248	0.968
30.04	0.01262	0.01436	0.01509	0.01368	0.00386	0.945
40.10	0.01910	0.03182	0.01385	0.02043	0.00556	0.907
50.07	0.02461	0.04022	0.02371	0.03281	0.00792	0.880
60.09	0.03001	0.04019	0.04088	0.05020	0.01840	0.804
70.21	0.04604	0.07285	0.06374	0.05439	0.02133	0.683
75.06	0.05403	0.08763	0.06039	0.06100	0.04357	0.625
80.07	0.06972	0.08695	0.08148	0.07131	0.05207	0.568

\* The accuracy of the water activity measurements is specified as  $\pm 0.003$  (absolute range) in  $a_w$  and  $\leq 0.1$  K in temperature.

![](_page_38_Figure_3.jpeg)

Table 9. EDB measurements: the total organic fraction of the solution is given in the first column
in wt% and the composition in terms of individual components is given in mole fractions of the
dicarboxylic acids constituting the M5 mixture.

wt%	x(DL-malic)	x(malonic)	x(maleic)	x(glutaric)	x(methylsuccinic)
10.01	0.00247	0.00537	0.00193	0.00450	0.00200
20.01	0.00545	0.01184	0.00423	0.00991	0.00441
30.01	0.00908	0.01978	0.00711	0.01659	0.00735
39.68	0.01370	0.02852	0.01065	0.02492	0.01106
49.97	0.01962	0.04270	0.01528	0.03577	0.01587
54.99	0.02332	0.05074	0.01819	0.04251	0.01889
60.01	0.02766	0.06021	0.02157	0.05043	0.02241
69.98	0.03904	0.08503	0.03042	0.07120	0.03158
79.87	0.05631	0.12246	0.04385	0.10263	0.04556
84.41	0.06768	0.14725	0.05279	0.12339	0.05478

![](_page_39_Figure_2.jpeg)

<b>Table 10.</b> Water activity $(a_w)$ measurements <sup>*</sup> of aqueous M5 solutions from EDB hygroscopic
growth curves evaluated at the indicated weight fractions. The corresponding M5 solution com-
positions in terms of organic components are given in Table 9.

wt%	a <sub>w</sub> (289 K)	a <sub>w</sub> (273 K)	a <sub>w</sub> (263 K)	a <sub>w</sub> (244 K)	a <sub>w</sub> (233 K)
10.01	0.987				
20.01	0.969				
30.01	0.947				
39.68	0.915				
49.97	0.874	0.867			
54.99	0.839	0.839	0.841		
60.01	0.805	0.802	0.805		
69.98	0.694	0.699	0.714		
79.87	0.540	0.562	0.572	0.648	0.667
84.41	0.448	0.470	0.490	0.569	0.626
90.00	0.318	0.329	0.362	0.429	0.544

\* The accuracy of the water activity measurements is specified as  $\pm 0.015$  (absolute range) in  $a_w$  and  $\leq 0.1$  K in temperature.

![](_page_40_Figure_3.jpeg)

Table 11. Water activity	$(a_w)$ measurements <sup>*</sup>	* of aqueous	M5 solutions	from EDB	hygroscopic
growth curves evaluated	at the indicated weight	ght fractions.			
		(0501()	(2221())		

wt%	a <sub>w</sub> (268 K)	a <sub>w</sub> (253 K)	a <sub>w</sub> (236 K)
60.00	0.791		
65.00	0.767		
70.00	0.725		
73.00		0.700	
75.00	0.676		
80.00	0.602	0.611	
85.00	0.504	0.507	
90.00	0.364	0.385	
95.60	0.172	0.216	0.347
97.20			0.264

\* The accuracy of the water activity measurements is specified as  $\pm 0.015$  (absolute range) in  $a_{\rm w}$  and  $\leq 0.1$  K in temperature.

![](_page_41_Figure_3.jpeg)

<i>x</i> <sub>2</sub>	a <sub>w</sub> (270.15 K)	<i>x</i> <sub>2</sub>	а <sub>w</sub> (273.15 К)	<i>x</i> <sub>2</sub>	a <sub>w</sub> (275.15 K)
0.24441	0.762	0.24441	0.766	0.23200	0.812
0.26933	0.751	0.26933	0.754	0.24441	0.768
0.31705	0.688	0.31705	0.694	0.26933	0.755
0.37368	0.654	0.37368	0.658	0.31705	0.697
0.44332	0.604	0.44332	0.606	0.37368	0.660
		0.52556	0.502	0.44332	0.607
				0.52556	0.504
<i>x</i> <sub>2</sub>	<i>a</i> <sub>w</sub> (278.15 K)	<i>x</i> <sub>2</sub>	a <sub>w</sub> (280.15 K)	<i>x</i> <sub>2</sub>	a <sub>w</sub> (283.15 K)
0.24441	0.772	0.04797	0.971	0.04797	0.971
0.26933	0.757	0.11823	0.900	0.11823	0.901
0.31705	0.702	0.16701	0.868	0.16701	0.868
0.37368	0.663	0.23200	0.813	0.23200	0.813
0.44332	0.608	0.24441	0.775	0.24441	0.776
0.52556	0.507	0.26933	0.759	0.26933	0.761
0.64080	0.396	0.31705	0.706	0.31705	0.710
		0.37368	0.665	0.37368	0.667
		0.44332	0.608	0.44332	0.610
		0.52556	0.508	0.52556	0.510

**Table 12.** Measured water activities of water (1) + 1,4-butanediol (2) mixtures derived from total gas phase pressure measurements<sup>\*</sup>, listed for a selection of temperatures in the range 270 K < T < 291 K. Solution compositions are given in mole fractions of the organic ( $x_2$ ).

![](_page_42_Figure_3.jpeg)

#### Table 12. Continued.

<i>x</i> <sub>2</sub>	а <sub>w</sub> (285.15 К)	<i>x</i> <sub>2</sub>	а <sub>w</sub> (288.15 К)	<i>x</i> <sub>2</sub>	a <sub>w</sub> (290.15 K)
0.04797	0.969	0.04797	0.967	0.04797	0.969
0.11823	0.901	0.11823	0.901	0.11823	0.906
0.16701	0.869	0.16701	0.868	0.16701	0.873
0.23200	0.814	0.23200	0.815	0.23200	0.818
0.24441	0.778	0.24441	0.780	0.24441	0.786
0.26933	0.763	0.26933	0.766	0.26933	0.770
0.31705	0.712	0.31705	0.716	0.31705	0.724
0.37368	0.667	0.37368	0.669	0.37368	0.673
0.44332	0.611	0.44332	0.611	0.44332	0.615
0.52556	0.512	0.52556	0.513	0.52556	0.517
0.64080	0.403	0.64080	0.404	0.64080	0.406

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**Table 13.** Measured water activities of water (1) + methoxyacetic acid acid (2) mixtures derived from total gas phase pressure measurements<sup>\*</sup>, listed for a selection of temperatures in the range 268 K < 7 < 291 K. Solution compositions are given in mole fractions of the organic ( $x_2$ ).

X_	a (268.15 K)	Xo	<i>a</i> (270.15 K)	Xo	<i>a</i> (273.15 K)	Xo	<i>a</i> (275.15K)
0.40740	0.007	0.01001	0.750	0.04.004	0.740	0.00007	0.005
0.43/12	0.607	0.31301	0.750	0.31301	0.749	0.22937	0.835
0.53152	0.509	0.43/12	0.608	0.43/12	0.608	0.31301	0.749
0.63995	0.328	0.53152	0.510	0.53152	0.511	0.43/12	0.608
		0.63995	0.331	0.63995	0.336	0.53152	0.511
						0.63995	0.337
<i>x</i> <sub>2</sub>	a <sub>w</sub> (278.15 K)	<i>x</i> <sub>2</sub>	<i>a</i> <sub>w</sub> (280.15 K)	<i>x</i> <sub>2</sub>	<i>a</i> <sub>w</sub> (283.15 K)	<i>x</i> <sub>2</sub>	a <sub>w</sub> (285.15 K)
0.02180	0.983	0.02180	0.983	0.02180	0.983	0.02180	0.983
0.04757	0.974	0.04757	0.974	0.04757	0.975	0.04757	0.975
0.07904	0.960	0.07904	0.958	0.07904	0.954	0.07904	0.952
0.11765	0.924	0.11765	0.921	0.11765	0.918	0.11765	0.916
0.16696	0.893	0.16696	0.888	0.16696	0.885	0.16696	0.884
0.22937	0.832	0.22937	0.831	0.22937	0.830	0.22937	0.829
0.31301	0.748	0.31301	0.748	0.31301	0.747	0.31301	0.747
0.43712	0.607	0.43712	0.605	0.43712	0.604	0.43712	0.603
0.53152	0.511	0.53152	0.510	0.53152	0.509	0.53152	0.509
0.63995	0.340	0.63995	0.341	0.63995	0.343	0.63995	0.344
<i>x</i> <sub>2</sub>	a <sub>w</sub> (288.15 K)	<i>x</i> <sub>2</sub>	a <sub>w</sub> (290.15 K)				
0.02180	0.983	0.02180	0.987				
0.04757	0.976	0.04757	0.980				
0.07904	0.946	0.07904	0.949				
0.11765	0.915	0.11765	0.918				
0.16696	0.882	0.16696	0.884				
0.22937	0.829	0.22937	0.830				
0.31301	0.745	0.31301	0.747				
0.43712	0.601	0.43712	0.602				
0.53152	0.507	0.53152	0.508				
0.63995	0.346	0.63995	0.348				

![](_page_44_Figure_3.jpeg)

**Table 14.** Measured water activities of water (1) + 2-(2-ethoxyethoxy)ethanol (2) mixtures derived from total gas phase pressure measurements<sup>\*</sup>, listed for a selection of temperatures in the range 265 K < T < 291 K. Solution compositions are given in mole fractions of the organic ( $x_2$ ).

	<i>x</i> <sub>2</sub>	$x_2 = a_w(265.15 \text{ K}) = x_2 = a_w(265.15 \text{ K})$		a <sub>w</sub> (268.15 K)	<i>x</i> <sub>2</sub>	a <sub>w</sub> (270.15 K)	<i>x</i> <sub>2</sub>	a <sub>w</sub> (273.15 K)
	0.54409	0.469	0.54409	0.477	0.54409	0.481	0.54409	0.488
	0.42719	0.573	0.42719	0.583	0.42719	0.589	0.42719	0.598
	0.34418	0.644	0.34418	0.654	0.34418	0.660	0.34418	0.669
	0.23774	0.732	0.23774	0.742	0.23774	0.748	0.23774	0.756
	0.16753	0.801	0.16753	0.811	0.16753	0.817	0.16753	0.824
	<i>x</i> <sub>2</sub>	a <sub>w</sub> (275.15 K)	<i>x</i> <sub>2</sub>	a <sub>w</sub> (278.15 K)	<i>x</i> <sub>2</sub>	a <sub>w</sub> (280.15 K)	<i>x</i> <sub>2</sub>	a <sub>w</sub> (283.15 K)
	0.54409	0.492	0.54409	0.497	0.54409	0.501	0.54409	0.506
	0.42719	0.603	0.42719	0.610	0.42719	0.616	0.42719	0.622
	0.34418	0.674	0.34418	0.681	0.34418	0.685	0.34418	0.692
	0.23774	0.762	0.23774	0.769	0.23774	0.774	0.23774	0.782
	0.16753	0.828	0.16753	0.835	0.16753	0.839	0.16753	0.845
	0.11810	0.878	0.11810	0.883	0.11810	0.886	0.11810	0.891
			0.08217	0.932	0.08217	0.932	0.08217	0.935
			0.05412	0.961	0.05412	0.961	0.05412	0.961
			0.03245	0.988	0.03245	0.989	0.03245	0.990
	<i>x</i> <sub>2</sub>	a <sub>w</sub> (285.15 K)	<i>x</i> <sub>2</sub>	a <sub>w</sub> (288.15 K)	<i>x</i> <sub>2</sub>	a <sub>w</sub> (290.15 K)		
	0.54409	0.509	0.54409	0.514	0.54409	0.518		
	0.42719	0.627	0.42719	0.634	0.42719	0.643		
	0.34418	0.696	0.34418	0.704	0.34418	0.711		
	0.23774	0.787	0.23774	0.795	0.23774	0.815		
	0.16753	0.849	0.16753	0.855	0.16753	0.864		
	0.11810	0.895	0.11810	0.900	0.11810	0.907		
	0.08217	0.938	0.08217	0.941	0.08217	0.946		
	0.05412	0.962	0.05412	0.964	0.05412	0.967		
	0.03245	0.992	0.03245	0.994	0.03245	0.998		

![](_page_45_Figure_3.jpeg)

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**Table 15.** Mixture compositions at different overall organic mass fractions of the aqueous M5 system used for total gas phase pressure measurements. Component mole fractions (x) of the dicarboxylic acids constituting the M5 mixture are given, with water accounting for the remaining fraction.

wt%	x(DL-malic acid)	x(malonic acid)	x(maleic acid)	x(glutaric acid)	x(methylsuccinic acid)
20	0.00545	0.01183	0.00428	0.00992	0.00441
40	0.01353	0.02815	0.01064	0.02453	0.01085
50	0.01959	0.04236	0.01526	0.03581	0.01672
60	0.02746	0.06049	0.02135	0.04971	0.02241
70	0.03833	0.08327	0.02997	0.06984	0.03090
75	0.04547	0.09924	0.03549	0.08310	0.03700

 $a_{\rm w}(265.15\,{\rm K})$   $a_{\rm w}(268.15\,{\rm K})$   $a_{\rm w}(270.15\,{\rm K})$   $a_{\rm w}(273.15\,{\rm K})$ wt% 20 40 50 0.872 0.881 0.886 0.890 60 0.822 0.823 0.820 0.819 70 0.724 0.719 0.719 0.709 wt%  $a_{\rm w}(275.15\,{\rm K})$  $a_{\rm w}(278.15\,{\rm K})$  $a_{\rm w}(280.15\,{\rm K})$  $a_{\rm w}(283.15\,{\rm K})$ 20 0.982 0.982 0.981 40 0.943 0.939 0.934 50 0.892 0.890 0.888 0.887 60 0.817 0.817 0.817 0.815 0.706 70 0.701 0.699 0.696 *a*<sub>w</sub>(285.15 K) wt%  $a_{\rm w}(288.15\,{\rm K})$  $a_{\rm w}(290.15\,{\rm K})$ 20 0.980 0.980 0.982 40 0.933 0.932 0.934 0.886 50 0.884 0.934 60 0.814 0.812 0.826 0.695 70 0.695 0.701

**Table 16.** Measured water activities of water (1) + M5 (2) mixtures derived from total gas phase pressure measurements<sup>\*</sup>, listed for a selection of temperatures in the range 265 K < T < 291 K. The total organic mass fraction of the solution is given in the first column.

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**Figure 2.** Measured water activities of aqueous 1,4-butanediol solutions vs. temperature. The different colours indicate the solution compositions in wt% of the organic component. The solid curves show data derived from the total gas phase pressure measurements. The coloured portion of the solid curves represents the temperature range for which the measurements are considered reliable within the uncertainty of the method ( $\pm 0.03$  of  $a_w$ ). Water activities derived from DSC measurements on the ice melting curve are represented by pentagons. Bulk  $a_w$  measurements using the water activity meter are represented by solid circles. The dash-dotted lines show the composition and temperature dependent  $a_w$  parametrisation by Zobrist et al. (2008). The blue dashed line is the ice melting curve (Koop and Zobrist, 2009). In the colour shaded regions one or both components are supersaturated with respect to the solid phase and therefore, above the eutectic temperature (~ 243–245 K), at equilibrium one solid phase coexists with the remaining solution.

![](_page_49_Figure_2.jpeg)

![](_page_50_Figure_0.jpeg)

**Figure 3.** Measured water activities of aqueous methoxyacetic acid solutions vs. temperature. The different colours indicate the solution compositions in wt% of the organic component. The solid curves show data derived from the total gas phase pressure measurements. The coloured portion of the solid curves represents the temperature range for which the measurements are considered reliable within the uncertainty of the method ( $\pm 0.03$  of  $a_w$ ). Water activities derived from DSC measurements on the ice melting curve are represented by pentagons. Bulk  $a_w$  measurements using the water activity meter are represented by solid circles. The dash-dotted lines show the composition and temperature dependent  $a_w$  parametrisation by Zobrist et al. (2008). The blue dashed line is the ice melting curve (Koop and Zobrist, 2009). In the colour shaded regions one or both components are supersaturated with respect to the solid phase and therefore, above the eutectic temperature (~ 251 K), at equilibrium one solid phase coexists with the remaining solution.

![](_page_50_Figure_2.jpeg)

![](_page_51_Figure_0.jpeg)

**Figure 4.** Experimental water activities vs. temperature for aqueous 2-(2-ethoxyethoxy)ethanol solutions. The different colours indicate the solution compositions in wt% of the organic component. The solid curves show data derived from the total gas phase pressure measurements. The coloured portion of the solid curves represents the temperature range for which the measurements are considered reliable. Water activity data obtained from DSC measurements on the water-ice melting curve Zobrist et al. (2008) are represented by solid pentagons. The solid circles show the data from bulk  $a_w$  measurements with the water activity meter. The dash-dotted lines show the composition and temperature dependent  $a_w$  parametrisation by Zobrist et al. (2008). The blue dashed line is the ice melting curve (Koop and Zobrist, 2009).

![](_page_51_Figure_2.jpeg)

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**Figure 5.** Experimental water activities vs. temperature for aqueous M5 solutions. The solid and dashed curves are from the total gas phase pressure measurements. The coloured dashed curves (75–84 wt%) represent measurements for which the solution was not in a homogeneous liquid state but partly crystallised. The coloured portion of the solid curves represents the temperature range for which the measurements are considered reliable (grey elsewhere). Water activity data obtained from DSC measurements on the water-ice melting curve from Zobrist et al. (2008) are shown by solid pentagons. The solid circles show the data form bulk  $a_w$  measurements with the water activity meter. The dash-dotted lines show the composition and temperature dependent  $a_w$  parametrisation by Zobrist et al. (2008). The blue dashed line is the ice melting curve (Koop and Zobrist, 2009). The  $a_w$  data from EDB measurements are represented by coloured diamonds.

![](_page_52_Figure_2.jpeg)

![](_page_53_Figure_0.jpeg)

**Figure 6.** Homogeneous freezing  $T_{hom}(x_{org})$  and melting points  $T_m(x_{org})$  as a function of  $a_w$  for various aqueous organic solutions. The blue dashed line in panels **(a, b)** is the homogeneous ice-melting curve  $(T_m)$  (Koop and Zobrist, 2009). The black dashed line is the homogeneous ice-freezing curve  $(T_{hom})$  calculated by shifting the melting curve by  $\Delta a_w = 0.313$  (Koop and Zobrist, 2009). The grey dotted line and grey dashed-dotted lines indicate a deviation of 2.5 and 5% in  $a_w$ , respectively. In panel **(a)**,  $a_w$  at  $T_{hom}(x_{org})$  is taken to be equal as  $a_w$  determined at  $T_m(x_{org})$  i.e., with the assumption that there is no significant change in  $a_w$  with temperature from  $T_m(x_{org})$  to  $T_{hom}(x_{org})$ . In panel **(b)**,  $a_w$  at  $T_m(x_{org})$  is determined using Eq. (3) and the corresponding  $a_w$  at the freezing temperature are estimated by using Eq. (3) at  $T_{hom}(x_{org})$  with a constant  $\Delta a_w = 0.313$  added to the  $a_w$  value determined on the melting curve.

![](_page_53_Figure_2.jpeg)