

Quantifying water diffusion in high-viscosity and glassy aqueous solutions using a Raman isotope tracer method

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Abstract

Recent research suggests that under certain temperature and relative humidity conditions atmospheric aerosol may be present in the form of a glassy solid. In order to understand the impacts that this may have on aerosol-cloud interactions and atmospheric chemistry, knowledge of water diffusion within such aerosol particles is required. Here, a method is described in which Raman spectroscopy is used to observe D₂O diffusion in high-viscosity aqueous solutions, enabling a quantitative assessment of water diffusion coefficients, D_{water} , as a function of relative humidity. Results for sucrose solutions compare well with literature data at $23.5 \pm 0.3^\circ\text{C}$, and demonstrate that water diffusion is slow ($D_{\text{water}} \sim 5 \times 10^{-17} \text{ m}^2\text{s}^{-1}$), but not arrested, just below the glass transition at a water activity of 0.2. Room temperature water diffusion coefficients are also presented for aqueous levoglucosan and an aqueous mixture of raffinose, dicarboxylic acids and ammonium sulphate: at low humidity, diffusion is retarded but still occurs on millisecond to second timescales in atmospherically relevant-sized particles. The effect of gel formation on diffusion in magnesium sulfate solutions is shown to be markedly different from the gradual decrease in diffusion coefficients of highly viscous liquids. We show that using the Stokes-Einstein equation to determine diffusion timescales from viscosity leads to values which are more than five orders of magnitude too big, which emphasises the need to make measurements of diffusion coefficients. In addition, comparison of bounce fraction data for levoglucosan with measured diffusion data reveals that even when particles bounce the diffusion timescales for water are a fraction of a second for a 100 nm particle. This suggests a high bounce fraction does not necessarily indicate retarded water diffusion.

1. Introduction

Water-soluble compounds are ubiquitous in atmospheric aerosol, which are important for the planet’s climate and hydrological cycle. The ability of aerosol particles to scatter and absorb light impact Earth’s radiation budget directly, and they also influence climate indirectly by mediating cloud properties through their action as cloud condensation nuclei (CCN) and ice nuclei (IN). It is therefore of vital importance to understand aerosol particle phase transitions and interactions with water vapour, and the consequences of these for aerosol optical properties, chemical reactions and health effects (Pöschl, 2005).

In some aqueous solutions, solute crystallization at low relative humidity (RH) or temperature can be inhibited, either partially or completely (Prenni et al., 2001; Parsons et al., 2004; Murray, 2008; Murray and Bertram, 2008; Zardini et al.,

2008; Bodsworth et al., 2010; Murray et al., 2012). This can be due to an increase in viscosity and associated decrease in the rate of diffusion within the solution as it supersaturates or supercools. A semi-solid state may form, the definition of which often includes ultra-viscous liquids, gels and rubbers (Mikhailov et al., 2009). If the viscosity of the solution increases above 10^{12} Pa·s it is said to have formed a glass, or amorphous solid, which may exist out of equilibrium with its surroundings for long timescales (Debenedetti and Stillinger, 2001). It has been proposed that atmospheric aerosol may be present in the form of amorphous solids and semi-solids over a wide range of temperature and relative humidity conditions (Bahreini et al., 2005; Roth et al., 2005; Murray, 2008; Zobrist et al., 2008; Mikhailov et al., 2009; Virtanen et al., 2010; Murray et al., 2012; Perraud et al., 2012; Saukko et al., 2012; Renbaum-Wolff et al., 2013).

The consequences of the existence of atmospheric aerosol in an ultra-viscous or glassy phase are wide-ranging. Highly viscous aerosol particles have been found to act as heterogeneous IN and may play an important role in cirrus cloud formation (Murray et al., 2010b; Wagner et al., 2012; Wilson et al., 2012; Wang et al., 2012; Baustian et al., 2013). Heterogeneous nucleation of ice on glassy aerosol particles well below water saturation may lead to less, but larger, ice crystals which have a greater sedimentation velocity; this may be particularly important in the very cold and dry tropical tropopause region where clouds play a key role in water transport into the stratosphere (Murray et al., 2010b). Reduced diffusion coefficients result in delayed interactions with the gas phase and may impact a particle's optical properties through its ability to take up water (Tong et al., 2011; Bones et al., 2012; Murray et al., 2012). Upon increasing RH, water uptake by amorphous solids occurs at the surface of a particle, proceeding to the interior of the particle only when the outer layer has softened and the rate of diffusion increased (Koop et al., 2011; Zobrist et al., 2011). In this way, the viscosity gradually decreases as the particle undergoes continual deliquescence, and hygroscopic growth occurs at lower relative humidity than for the particle's crystalline counterpart (Mikhailov et al., 2009). When RH is decreased, a core-shell structure may develop as a glassy crust forms around a less viscous interior. In this way water may be trapped inside the particle, even under dry conditions (Koop et al., 2011; Zobrist et al., 2011).

Several studies have suggested that atmospheric aerosol particles are not in equilibrium with surrounding gas phase compounds (Roth et al., 2005; Perraud et al., 2012; Abramson et al., 2013), which may lead to errors in the evaluation of particle-phase and gas-phase concentrations (Liu et al., 2012; Shiraiwa and Seinfeld, 2012; Shiraiwa et al., 2013). The phase of an aerosol particle has been found to affect reactivity (Knopf et al., 2005; Kuwata and Martin, 2012), and the chemical lifetimes of condensed organics can be significantly increased if diffusion is limited (Shiraiwa et al., 2010; Pfrang et al., 2011; Shiraiwa et al., 2011). Highly viscous aerosol may undergo diffusion limited growth, which would result in a different size distribution to that produced by the quasi-equilibrium growth of liquid aerosol (Zhang et al., 2012). In laboratory or field measurements, timescales for equilibration longer than typical thermodenuder residence times may also lead to erroneous volatility results (Saleh et al., 2011), while slow hygroscopic growth may affect hygroscopicity tandem differential mobility analyzer (HTDMA) values (Hersey et al., 2013).

Vaden et al. (2011) and Cappa and Wilson (2011) observed slow evaporation rates of secondary organic aerosol (SOA) and attributed this to condensed phase kinetic limitations to gas-particle partitioning. However, slow evaporation would also be seen in particles containing low volatility compounds, such as aged SOA in which low volatility oligomers may form (Cappa and Wilson, 2011; Saleh et al., 2013). In an experiment directly measuring equilibration timescales at 30°C, Saleh et al. (2013) reported no substantial gas-particle partitioning inhibition due to kinetic limitations within particles. The discrepancies between evaporation- and equilibration-rate measurements highlight the need for more direct measurements of potential kinetic limitations in aerosol particles.

Current techniques used to determine the formation and properties of semi-solid and glassy aerosol often involve some method of establishing the viscosity, quantitatively or qualitatively (Virtanen et al., 2010; Wang et al., 2012; Renbaum-Wolff et al., 2013). The Stokes-Einstein relation is then sometimes used to estimate the diffusion coefficient. However, there is

evidence to suggest that this hydrodynamic description is not generally applicable: the mobility of water and larger molecules deviate near the glass transition (Champion et al., 1997; Rampp et al., 2000; Zhu et al., 2011; Power et al., 2013). In addition, diffusion coefficients do not vary with temperature in the same way as viscosity at $T_g/T > 0.8$ (Parker and Ring, 1995). Since it is diffusion, rather than viscosity, which is key to quantifying the response of aerosol to changing conditions, it is advantageous to directly measure the diffusion coefficients of the species of interest, rather than inferring them from the viscosity.

The diffusion coefficient of water, D_{water} , in atmospheric aerosol is an important parameter because water is often the most mobile component of a glassy aqueous solution (Roberts and Debenedetti, 1999), it is crucial in cloud formation processes, and it can act as a plasticizer (Zobrist et al., 2011; Koop et al., 2011). The uncertainty in the current best estimate of D_{water} in α -pinene SOA spans four orders of magnitude at RHs below 80% (Shiraiwa et al., 2013). Furthermore, the plasticizing effect of water leads to considerable difficulties in measuring the diffusion coefficient of water in glassy aerosol undergoing hygroscopic growth or shrinkage because the diffusion equation becomes non-linear (Tong et al., 2011). Zobrist et al. (2011) used a model to overcome the issue of non-linearity, and were successful in determining diffusion coefficients from water uptake on aqueous sucrose particles with electrodynamic balance and optical techniques.

We have used a similar technique to Zhu et al. (2011) to determine the diffusion coefficient of water in aqueous solutions. Zhu et al. (2011) brought a H_2O -maltose and a D_2O -maltose droplet together until they touched, and used a Raman microscope to quantify diffusion between the droplets. Our approach relies upon the observation of the diffusion of gas phase isotope tracer into a disk of aqueous solution. This involves equilibrating the solution at a chosen RH in H_2O vapour and then switching to D_2O vapour of almost the same RH. The progression of D_2O into the solution disk is then monitored as a function of time and distance which allows a quantitative assessment of the diffusion coefficient. The advantage over techniques which involve changing RH in a stepwise fashion is that the solution to the diffusion equation is greatly simplified since the water content ($\text{D}_2\text{O} + \text{H}_2\text{O}$) of the disk remains constant throughout the experiment – there is no radial solute concentration gradient.

The kinetics of water transport in amorphous inorganic aerosol have previously been investigated by Li et al. (2011), who used Raman spectroscopy to observe the exchange of deuterium with hydrogen, but did not determine diffusion coefficients. Ahlqvist and Taylor (2002) also used H/D exchange to observe diffusion in crystalline and amorphous sugars. The method outlined here allows for the quantitative determination of the diffusion coefficient of water in aqueous solutions relevant to atmospheric aerosol over a wide range of RH and temperature conditions, assuming Fickian diffusion. Here, sucrose is studied to allow direct comparison with literature data for method validation. Measurements are then extended to (i) levoglucosan (a major component of biomass burning aerosol (Decesari et al., 2006)), (ii) raffinose/M5AS (a mixture of raffinose, dicarboxylic acids and ammonium sulfate, (Zobrist et al., 2008)) and (iii) magnesium sulfate (present in seawater aerosol and known to have the interesting property of gel formation at low RH (Zhao et al., 2006)).

2. Experimental methods

2.1 Raman microscopy

A Renishaw InVia Raman spectrometer with a 514 nm laser and motorized XYZ stage was coupled to a temperature and relative humidity controlled chamber (Fig. 1). A dry zero grade N_2 flow was humidified as it passed through a H_2O or D_2O bubbler, whose temperature was regulated by a chiller, and then diluted to the desired dew point with additional N_2 . The dew point of the outflow from the chamber was recorded using a GE Optica hygrometer with model 1311DR sensor. The stage

temperature was maintained at $23.5 \pm 0.3^\circ\text{C}$ using a separate recirculating chiller and measured using a PT100 platinum resistance thermometer. This temperature was chosen for the present study to facilitate comparison with literature data and to reduce experimental duration; experiments at lower temperatures will be the subject of future work. Spectra containing information on the chemical composition of the sample could be continually acquired at predetermined locations on the sample with a laser spot size of $1.3\ \mu\text{m}$. A 600 lines/mm grating was used, enabling Raman intensities to be simultaneously collected across the wavenumber range of interest. All spectral and environmental data were computer logged for subsequent analysis.

2.2 Diffusion in a disk

The two separate locations of the O-H and the O-D bond stretch bands in the Raman spectrum were used to monitor the diffusion of D_2O from the gas phase into a H_2O solution droplet. In order to avoid inhomogeneity in the vertical column of sample which contributed to the Raman signal, a disk of solution was prepared such that its circumference (but neither face) was exposed to the surrounding gases (Fig. 2(a)). In this way, diffusion only occurred horizontally in the radial direction).

Assuming that a disk is homogenous, i.e. the diffusion coefficient does not depend on radial position, Fick's second law is

$$\frac{\partial \varphi(r, t)}{\partial t} = D \nabla^2 \varphi(r, t) \quad (1)$$

where φ is the concentration of the diffusing molecule in $\text{mol}\cdot\text{m}^{-3}$, t is time and D is the diffusion coefficient in m^2s^{-1} . A solution to the above equation for a disk of radius a whose edge is maintained at concentration φ_0 is

$$\varphi(r, t) = \varphi_0 + 2(\varphi_1 - \varphi_0) \sum_{\alpha} \frac{1}{\alpha J_1(\alpha)} e^{\frac{-\alpha^2 D t}{a^2}} J_0\left(\frac{\alpha r}{a}\right) \quad (2)$$

where φ_1 is the concentration at $t = 0$ everywhere in the disk except at the edge ($r = a$) at $t = 0$, J_0 and J_1 are Bessel's functions of order 0 and 1, respectively, and α are the positive roots of the equation $J_0(x) = 0$ (Bowman, 1958).

2.3 D_2O diffusion measurements

Dilute aqueous solutions were prepared with pure water (Milli-Q 18.2 $\text{M}\Omega\cdot\text{cm}$). The solutes used were sucrose (Sigma, >99.5%), levoglucosan (Acros Organics, >99%), magnesium sulfate (Sigma-Aldrich, >99.5%) and raffinose/M5AS (50 wt% D-(+)-raffinose pentahydrate (Alfa Aesar, >99%), 15.4 wt% malonic acid (Alfa Aesar, >99%), 7.1% maleic acid (Fluka, >99%), 7.4% DL-malic acid (Aldrich, >99%), 3.7% methylsuccinic acid (Aldrich, >99%), 8.9% glutaric acid (Alfa Aesar, >99%) and 7.7% ammonium sulfate (Fluka, >99.5%). For each experiment, a $\sim 0.5\ \mu\text{L}$ droplet of solution was placed on a hydrophobic siliconized glass slide (Hampton Research) using a micropipette and allowed to adapt its size as it equilibrated with the surrounding air. Similarly coated glass slides have been used previously and shown not to trigger nucleation (Murray et al., 2012; Murray et al., 2010a; Murray et al., 2011; Broadley et al., 2012; Atkinson et al., 2013). A smaller piece of hydrophobic glass was placed on top of the droplet, supported by a pair of $25\ \mu\text{m}$ -thick Teflon film spacers, in order to create a disk of 200-300 μm radius (Fig. 2(a)). The slide was then placed in a temperature and humidity controlled chamber, initially at 90% RH.

For each experiment, the solution disk was exposed to a flow of nitrogen gas with a controlled partial pressure of H_2O vapour in order to achieve the water activity corresponding to the chosen RH. In these experiments it was essential that the water activity across the disk was constant, so it was important that the disk was exposed to H_2O vapour for a sufficient period of time. This time required for equilibration depended on solute and RH; the lower the RH, the longer the H_2O equilibration time. Hence, it was necessary to estimate the diffusion coefficient and solute concentrations at a particular water activity so that the time required for equilibration could be estimated using the solution to Fick's second law for a cylindrical geometry (Eq. (2)). In this case, φ_0 was the water concentration corresponding to the target water activity, and φ_1 was the water

concentration corresponding to a water activity of 0.9 (i.e. the concentration at the start, defined by the starting RH of 90%). Water concentrations were calculated using relations between water activity and solute mass fraction given in the literature (Ha and Chan, 1999; Zobrist et al., 2011; Lienhard et al., 2012; Wagner et al., 2012), and densities estimated assuming the volume additivity of pure component densities (Tong et al., 2011). The diffusion coefficient used in this estimation was a best initial estimate of the diffusion coefficient of water in the disk at the target water activity, either taken from the literature or based on the trends of previous measurements. By solving Eq. 2 with these conditions, the time after which the water mass fraction in the centre of the disk corresponded to a water activity within 0.01 of the target water activity could be determined – this was used to define the H₂O exposure time. This process sometimes required several iterations: if the final measured diffusion coefficient was found to be lower than that used in the calculation, the experiment was repeated, this time leaving the disk exposed to H₂O for a longer time period. The equilibration time varied between minutes at high humidity to weeks at low humidity. In section 3.1 we show that the measured diffusion coefficients are in good agreement with literature data.

In most experiments, equilibration was done at 23.5°C, but for low water activity experiments which would require impractical H₂O exposure times at 23.5°C, the sample was heated to between 40 and 50°C to speed up diffusion. Once the necessary exposure time at this temperature had elapsed, the sample was cooled back to 23.5°C over a period of several hours, with the RH (and thus water activity) kept constant to within 1% RH. Any uncertainty in water activity associated with this procedure was accounted for in error bars. To confirm that this treatment did not alter the diffusion properties of the sample, two measurements of the water diffusion coefficient in sucrose were made at around 80% RH, one before exposure to higher temperatures and one after a week at 50 °C. They were in good agreement (see section 3.1).

To determine disk size and location on the motorized XYZ stage, the positions of three points on the disk's circumference were found. This was done by taking three series of spectra along radial tracks traversing the edge of the particle, with 1 µm spacing between each spectrum and a laser spot size of 1.3 µm. For each of these three spectral series, the points at which peaks originating from the sample first appear in the spectra were used to calculate an upper limit on the disk's radius. Similarly, the three points at which the sample peaks reach their maximum intensity were used to determine a lower limit on the disk's radius. The difference between these upper and lower limits was typically around 20 µm. The best estimate for the disk radius was taken to be the mid-point between the upper and lower limits.

Once the water activity of the disk was estimated to be within 0.01 of RH/100, the H₂O vapour was replaced with a flow of D₂O vapour of the same dew-point. Due to the lower saturation vapour pressure of D₂O, this led to a decrease in RH of up to 1.25% (Matsunaga and Nagashima, 1987). We estimate that the complete exchange of gases in the pipework and chamber around the solution disk takes 12±2 s, although in a worst-case scenario where the D₂O and H₂O vapors mix only via gas-phase diffusion in the RH-controlled chamber this exchange may take up to 95 s.

The spectrometer was configured to collect Raman spectra after chosen time intervals at locations across a grid covering a portion of the sample disk (see Fig. 2). The spatial and temporal separations of the spectra were chosen such that, as far as possible, a broad range of D₂O concentrations could be observed over the entire disk radius during the experiment. Each spectrum was measured at a new XY coordinate (and hence radial position), chosen according to the rate of diffusion in each experiment. The D₂O exposure time required to collect enough information to determine a diffusion coefficient was about 10 minutes for a D_{water} of $10^{-10} \text{ m}^2\text{s}^{-1}$, about 2 hours for a D_{water} of $10^{-12} \text{ m}^2\text{s}^{-1}$, about 20 hours for a D_{water} of $10^{-14} \text{ m}^2\text{s}^{-1}$, and about 8 days for a D_{water} of $10^{-16} \text{ m}^2\text{s}^{-1}$.

The Raman software could be used to define a rectangular grid across which spectra could be collected in a raster fashion. At high diffusion coefficients, it was important to collect data across the entire radius of the disk in a short space of time. For this reason, a grid such as that shown in Fig. 2(b) was configured, with a 25 µm spacing in the x direction and a small (up to 5 µm) spacing in the y direction. Limitations existed at low RH because diffusion was so slow as to make it impractical to wait for high D₂O concentrations at the disk centre; it was nevertheless possible to determine diffusion coefficients using the

concentration measurements near the edge of the disk (see Fig. 2(c)). At these slow diffusion coefficients, where many spectra could be collected without the concentration gradient changing significantly, it was less important to cover the entire radius of the disk in a short space of time. This allowed us to collect clusters of more spectra at higher spatial resolution (typically with X and Y spacing of 5 μm), with time breaks between each cluster (Fig. 2(c)). As diffusion progressed, and the O-D band was visible across greater proportions of the radius, the size and number of spectra in each cluster was increased to cover more of the radius.

The time taken to collect each spectrum was typically 1 s, with a further 1 s period required between spectra for the motorized XYZ stage to move to the next location. This therefore defined the upper limit to the diffusion coefficients which could be measured where diffusion occurred on a minute timescale. Some spectra were longer in duration (up to 40 s per spectrum) – these longer acquisitions were used where the extra duration did not compromise the time resolution but did improve the quality of the spectral data. For very slow experiments (e.g. sucrose 20% RH, which involved a 3 week D_2O exposure), there were breaks between several series of spectral acquisitions in order to avoid wear on the laser.

The number of points used in each surface fit varied according to the duration of each experiment, which was directly influenced by the rate of diffusion at each relative humidity. Experiments where diffusion coefficients were large typically involved the collection of several hundred spectra (and thus data points in the surface fit); experiments with the smallest diffusion coefficients usually generated several thousand spectra.

After the exchange had taken place, the process of determining the disk radius by finding the positions of three points on the disk edge was repeated. Decreases in radius of up to 3% were noted; this can be accounted for by the combined effects of a change in volume due to isotope exchange (Bartell and Roskos, 1966), and a change in water activity due to the lower saturation vapour pressure of D_2O (Matsunaga and Nagashima, 1987). The shrinkage was included in the calculation of the error bars associated with each data point.

Raffinose/M5AS was seen to fluoresce at the lowest water activity investigated (0.22), which meant the Raman scattering bands were partially obscured (no significant fluorescence was observed at other humidities or in the other samples). To minimise these effects, the Raman laser was used to photobleach the sample for 500 s prior to each spectral acquisition. Since any photobleaching-induced effects would have been concentrated in a small area due to the small laser spot size, and given that each spectrum was acquired in a new position, we do not expect any effects on the determined diffusion properties from the bleaching technique.

2.4 Analysis of Raman data to determine diffusion coefficients

The Raman spectra of aqueous solutions of organics such as sucrose and levoglucosan are characterized at high wavenumbers by C-H stretch bands at ~ 2800 to 3100 cm^{-1} and O-H stretch bands at ~ 3100 to 3500 cm^{-1} . The C-H stretch band originates entirely from the organic molecules, whilst the O-H band originates from hydroxyl groups both in the organic and in water. The magnesium sulfate spectrum lacks the C-H stretch, whilst the raffinose/M5AS spectrum also features a band arising from the NH_4^+ species which is situated around 3100 cm^{-1} . This band was small (due to the relatively small amount of ammonium in the mixture) and overlapped with the O-H stretch. The O-D stretch band is located at $\sim 2500\text{ cm}^{-1}$, which is $\frac{1}{\sqrt{2}}$ times the O-H stretch frequency, and has a lower integrated intensity than the O-H stretch band by a factor of $\frac{1}{\sqrt{2}}$ (Kohen and Limbach, 2010). Similarly, the ND_4^+ band is located at $\sim 2200\text{ cm}^{-1}$. After switching from H_2O vapour to D_2O flow, the O-H stretch band decreased in area, whilst the O-D band appeared and grew (Fig. 3). In raffinose/M5AS, the NH_4^+ band was also seen to decrease whilst the ND_4^+ band grew. The area of the C-H stretch band did not decrease: hydrogen atoms on oxygen or

nitrogen may exchange with deuterium due to the polarisation of the electron density within the bond, whilst C-H bonds do not exchange as they are not sufficiently polarised.

Using the Levenberg-Marquardt technique (Markwardt, 2008), a set of Gaussian peaks were fitted to the spectral bands (Fig. 4). The background was accounted for using a constant plus a broad, high frequency Gaussian which smoothly matched the low and high frequency ends of the spectra. It should be emphasized that the spectral fits produced here are not intended as accurate band assignments. The sole purpose is to achieve a reasonable, background-corrected quantification of the relative integrated band intensities of the broad O-H and O-D bands.

For sucrose, levoglucosan and raffinose/M5AS, the parameters describing these Gaussian functions (i.e. peak height, width and wavenumber) were constrained so that variable intensity peaks were fitted to the O-H, NH_4^+ , O-D and ND_4^+ bands, and fixed intensity peaks were fitted to the C-H band. A satisfactory fit with a reduced χ^2 value close to 1 was achieved by fitting up to three Gaussians to the O-H band (or the combined O-H and NH_4^+ bands in the case of raffinose/M5AS), up to three Gaussians to the O-D band (or the combined O-D and ND_4^+ bands in the case of raffinose/M5AS), and up to five Gaussians to the C-H band. Constraining the parameters describing these peaks was particularly important in the overlap region between the O-D and C-H bands. So that no C-H intensity was attributed to O-D vibrations, the C-H stretch bands were initially fitted before any D_2O exposure. As the C-H band would not vary during the experiment, the parameters found in this initial fit to the corresponding Gaussian peaks were used to constrain the C-H band fit when D_2O was present. For magnesium sulfate (Fig. 4(d)), up to four Gaussians were fitted to the O-H stretch and up to four to the O-D stretch, as well as a weak Gaussian to account for a slightly raised background at $\sim 2450 \text{ cm}^{-1}$.

The results of the spectral fitting procedure were used with Eq. (2) is used to find the diffusion coefficient of D_2O in the sample. Unlike in section 2.3, φ is now the D_2O concentration. Because the total amount of water (D_2O plus H_2O) and solute remains constant throughout the experiment, φ can simply be written as the fractional concentration of O-D bonds relative to the total O-D and O-H bonds at each spectral location and time. Thus,

$$\varphi = \frac{A_{OD}}{A_{OD} + \frac{1}{\sqrt{2}} A_{OH}} \quad (3)$$

where A_{OD} is the integrated intensity of the O-D stretch (i.e. the summed areas of the Gaussian curves representing the broad band between 2300 cm^{-1} and 2700 cm^{-1}) and A_{OH} is the integrated intensity of the O-H stretch (i.e. the summed areas of the Gaussian curves representing the broad band between 3100 cm^{-1} and 3500 cm^{-1}). The progression of φ with time at fixed radial positions is shown in Fig. 5(a). φ_0 is here set to 1 (the fractional O-D concentration maintained at the edge of the disk) and φ_1 is here set to 0 (the O-D concentration everywhere except the edge at $t = 0$).

To determine D_{water} at a given RH, a surface fit of Eq. (2) with the diffusion coefficient as the sole variable parameter is performed to the measured values of $\varphi(r, t)$, again using the Levenberg-Marquardt technique. Figure 5(b) shows an example of this surface fit to a set of data, where each measured data point is connected to the fitted surface by a black line for comparison. Due to the finite size of the laser spot (estimated by the manufacturers as $1.3 \mu\text{m}$), measured values of φ are least reliable where the radial concentration gradient is changing rapidly.

It is evident from the complete disappearance of the O-H stretch band in Fig. 3 that H/D exchange occurs not only in the water molecules of the sample, but also in the hydroxyl groups of the organics. A large difference between the rates of these two exchanges would complicate the determination of the diffusion coefficient. However, at 90% RH, complete H/D exchange at the centre of a disk was observed on the timescale of minutes. Given that measurements at RHs lower than $\sim 70\%$ took significantly longer than this, and that the calculated diffusion coefficients were several orders of magnitude lower, we conclude that the limiting step here is diffusion, not H/D exchange rate. We note, however, that for measurements between 70

and 90% RH the procedure may introduce some uncertainty. The determined diffusion coefficient may thus be slightly lower than the true value.

3. Results and discussion

The determined diffusion coefficients of D₂O in sucrose, levoglucosan, magnesium sulfate and raffinose/M5AS as a function of water activity, at 23.5°C, are shown in Fig. 6. The upper error bars shown for D_{water} represent the value of D_{water} calculated using the highest value measured for the upper limit on the disk's radius (usually that calculated from the three edge measurements before exposure to D₂O vapour), with a time delay of 95 s, and include the random error in the surface fit due to errors in φ . The lower error bars shown for D_{water} represent the value of D_{water} calculated using the lowest value measured for lower limit on the disk's radius (usually that calculated from the three edge measurements after exposure to D₂O vapour), with a time delay of 10 s, and also include the random error in the surface fit due to errors in φ . Lines are fitted to the data as follows:

$$\log_{10} D_{\text{water}} = a + b a_w + c a_w^2 + d a_w^3 \quad (4)$$

where a , b , c and d are empirically fitted parameters detailed in Table 1. The fits for all substances converge to the diffusion coefficient for water in water at a water activity of 1.0 (Holz et al., 2000).

The diffusion of D₂O is expected to be slightly slower than that of H₂O. As a guide, the ratio of the self-diffusion coefficients of H₂O and D₂O have been found to lie in the range 1.10 to 1.25 (Mills, 1973; Svishchev and Kusalik, 1994; Liu and Macedo, 1995). However, given that the measured values cover six orders of magnitude, the diffusion coefficient of D₂O can be taken as a good approximation for that of H₂O.

3.1 Sucrose – comparison with literature data

The diffusion coefficients of water in aqueous sucrose solutions are compared to literature data in Fig. 7 and are in good agreement for water activities between 0.5 and 0.7. Below a water activity of 0.5, where the He et al. (2006) and Zobrist et al. (2011) parameterisations deviate from each other, the fit to our data lies between the available literature values, but seems to follow the trend of the free-volume model (He et al., 2006). This is consistent with the observation by Tong et al. (2011) that the diffusion coefficient reported by Zobrist et al. (2011) for a 95% sucrose solution (a water activity of 0.28) is too large to explain their observed droplet size changes.

The slight deviations from literature values at high water activities may be due to a number of factors. When diffusion is fast, the radial progression of the concentration gradient may be limited by something other than water diffusion, possibly the slower H/D exchange in hydroxyl groups (as discussed in section 2.4). The fractional concentration of O-D bonds (relative to the total O-D plus O-H bonds) in the gas phase at the edge of the disk cannot instantaneously become 1 when the switch is made from H₂O to D₂O. There will be an unavoidable period of time in which a mixture of H₂O and D₂O vapours is present; the nearer the duration of this switchover is to the timescale of the measurement, the more likely it is to lead to errors.

3.2 Timescales for water diffusion

The characteristic half-time for diffusion into a spherical particle of radius r at constant water activity is given by (Seinfeld and Pandis, 2006)

$$\tau_{1/2} = \frac{r^2}{\pi^2 D \cdot \ln(2)} \quad (5)$$

Note that this is the diffusion timescale for water diffusing into a droplet at constant water activity and therefore D does not change. It may be of interest to know the timescales over which a droplet would respond to changes in RH under certain atmospheric circumstances. This is not a simple calculation due to the plasticising effect of water in these aqueous solutions – the diffusion coefficient is a function of concentration and would vary radially as a droplet increased or decreased in size. In order to model the response of a droplet to a change in humidity a model of the time response of a droplet undergoing water uptake and loss would be required. This is the subject of ongoing work and beyond the scope of this paper. In this study, we use $\tau_{1/2}$ to give basic information about the rate at which water molecules diffuse within a droplet at a given relative humidity.

Fig. 8(a) shows these timescales for water diffusing into sucrose, levoglucosan and raffinose/M5AS aqueous spherical droplets of radii between 100 nm and 1 mm at 20% RH. For levoglucosan and raffinose/M5AS, water diffusion timescales are faster than 1 s in particles smaller than 250 nm in radius at room temperature. For sucrose these timescales are on the order of minutes to hours. Under these conditions the calorimetry data presented by Zobrist et al. (2008) suggest that aqueous sucrose exists in a glassy state. As Zobrist et al. (2011) point out, even though particles may exist in what is by definition a glassy state, water diffusion is not necessarily arrested. Nevertheless, the kinetic limitation could be important for growth measurements in aerosol instruments which operate on short timescales, such as the HTDMA (Hersey et al., 2013). In addition, diffusion coefficients decrease with temperature and thus under common atmospheric conditions these timescales are expected to be far greater.

Water diffusion coefficients measured in this study have been used to calculate water diffusion timescales as a function of RH in droplet of 100 nm radius, according to Eq. (5); these are plotted as red dots in Fig. 8(b). These timescales have also been calculated using the water diffusion coefficients given in Zobrist et al. (2011) for comparison. To illustrate the magnitude of error that would be encountered when estimating these water diffusion timescales from a viscosity measurement, the Stokes-Einstein relation was used with the viscosity measurements of Power et al. (2013) to estimate D in Eq. (5). The resulting timescales are shown as black crosses. There is a discrepancy of more than 5 orders of magnitude between the two datasets at ~30% RH and ~2 orders at 62% RH. Again, this demonstrates the importance of quantifying diffusion in atmospheric aerosol.

3.3 Levoglucosan – relationship to bounce factor data

Diffusion coefficients of water in levoglucosan were found to be significantly higher than those in sucrose for the same RH and correspond to diffusion in a high-viscosity liquid. This is consistent with the glass transition curve given by Lienhard et al. (2012) which shows that the glass transition temperatures of aqueous levoglucosan solutions of water activities above 0.2 are below 236 K.

Saukko et al. (2012) determined the bounced fraction of aqueous levoglucosan aerosol by impacting the aerosol onto a substrate and counting the number of particles which rebounded – a high bounce fraction is considered to indicate high viscosity, whilst a low bounce fraction implies low viscosity. As noted by Saukko et al. (2012), high-viscosity liquids and semi-solids have similar bounce behaviour to glasses and crystalline solids: bounce fractions of around 0.8 were measured for both high-viscosity liquid levoglucosan and crystalline ammonium sulfate. Fig. 9 shows diffusion half-lives of 100 nm radius levoglucosan droplets as a function of water activity, as calculated using Eq. (5), compared with bounce fraction data. Even at low RH, this characteristic timescale for diffusion in levoglucosan is less than 0.1 s. Zobrist et al. (2011) measured water diffusion coefficients in glassy particles (which would presumably have similarly high bounce fractions to low RH levoglucosan) as low as $10^{-24} \text{ m}^2 \text{ s}^{-1}$, which, according to Eq. (5), would have diffusion half-lives of over 100 years. Thus, these

results reiterate what is already accepted about the bounce technique: although bounce fraction measurements are useful in determining particle phase in the relevant size range, they alone cannot yet distinguish between particles in which diffusion of a small molecule such as water occurs on timescales of milliseconds and those in which it takes years (Saukko et al., 2012). This underlines the importance of quantitative diffusion measurements for the understanding of high-viscosity atmospheric aerosol.

3.4 MgSO_4 gel formation

Fig. 10 shows the diffusion surface fits for magnesium sulfate between 82 and 51% RH. There is a distinct shift from Fickian diffusion between 62 (Fig. 10(c)) and 51% RH (Fig. 10(d)), where it is no longer possible to find a fit to the data using Eq. (2). There is also a corresponding structural change in the sample. At 62% and higher RH, the sample appeared homogeneous when inspected with an optical microscope, whereas at 51% RH there was some structure within the disk (Fig. 11(a)). This structure had a strong effect on D_2O diffusion. Fig. 11(b) shows a map of ϕ for the area highlighted in Fig. 11(a) after one day's exposure to D_2O vapour. Within the approximately triangular structure D_2O replaced the H_2O , whereas very little or no exchange occurred in the other regions.

Magnesium sulfate is known to form a gel due to the formation of a complex ion-induced network; this network structure has been shown to inhibit water mass transfer within the gel (Chan et al., 1998; Zhang and Chan, 2000). The transition to a gel has been variously reported to occur between 55 and 30% RH (Wang et al., 2005; Zhao et al., 2006; Li et al., 2011; Davies et al., 2012), in agreement with our findings. The marked change in the water diffusion process at the gel transition is completely different to the gradual decrease in the diffusion coefficient seen in the aqueous organic liquids. For this reason, it is important to note the clear distinction between semi-solids such as gels or rubbers and highly viscous liquids.

4. Summary

The existence of aerosol as highly viscous liquids and amorphous glassy solids may have a profound effect on their interactions with water vapour and other chemical species in the atmosphere. It is therefore important to quantify diffusion in ultra-viscous and amorphous solid states. In this paper we describe a new experimental system for quantifying the diffusion coefficient of water molecules in aqueous solutions, with diffusion coefficients measured between 10^{-16} and $10^{-10} \text{ m}^2\text{s}^{-1}$. This range corresponds to solutions from the glassy to liquid states. We show that this methodology, which makes use of D_2O as an isotopic tracer, produces water diffusion coefficients for sucrose solutions in good agreement with those in the literature. New data for water diffusion coefficients in aqueous solutions of MgSO_4 , levoglucosan and a mixture of raffinose, carboxylic acids and ammonium sulfate are also presented for the first time.

Using these measured water diffusion coefficients we predict diffusion timescales for aerosol-sized particles (~ 100 nm) at room temperature under constant RH conditions, and show that just below the glass transition it takes minutes before equilibrium with water vapour is achieved. This is a vastly shorter time scale than one would expect based on the Stokes-Einstein equation, which has been used to relate diffusion to viscosity in the past. Our results reinforce previous studies showing that the Stokes-Einstein relationship is inappropriate in the regime of the glass transition (Champion et al., 1997; Rampp et al., 2000; Power et al., 2013). Even though the diffusion timescale is much shorter than predicted by the Stokes-Einstein equation, it is still long in comparison to the timescale needed when using thermodenuders and HTDMAs (seconds). Hence, some caution must be applied to the interpretation of results from measurements. In addition, we show for levoglucosan that a high bounce fraction does not necessarily mean long water diffusion timescales.

It is important to note that we have only measured diffusion coefficients for water, whereas diffusion of important gaseous species (e.g. NO₂, SO₂, H₂O₂ and O₃) into aerosol is important for chemistry, and diffusion of organic molecules is important for aerosol growth and evaporation. It is unclear how well (or poorly) the Stokes-Einstein equation predicts diffusion of these other atmospherically important species and dedicated diffusion measurements are therefore highly desirable.

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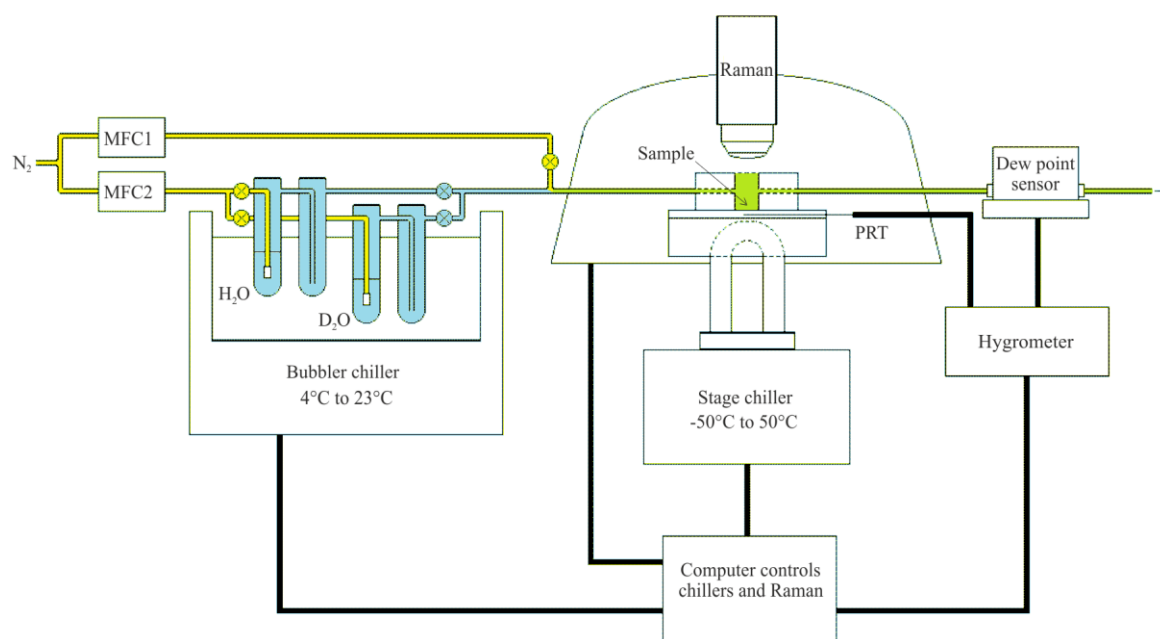


Figure 1. Humidity and temperature controlled Raman spectroscopy setup. Nitrogen gas flows were regulated using two mass flow controllers (MFC1 and MFC2) and either remained dry or were humidified in a bubbler. The two flows were mixed at a known ratio to produce the desired relative humidity at the stage, which was also temperature controlled. A hygrometer logged temperature and humidity data.

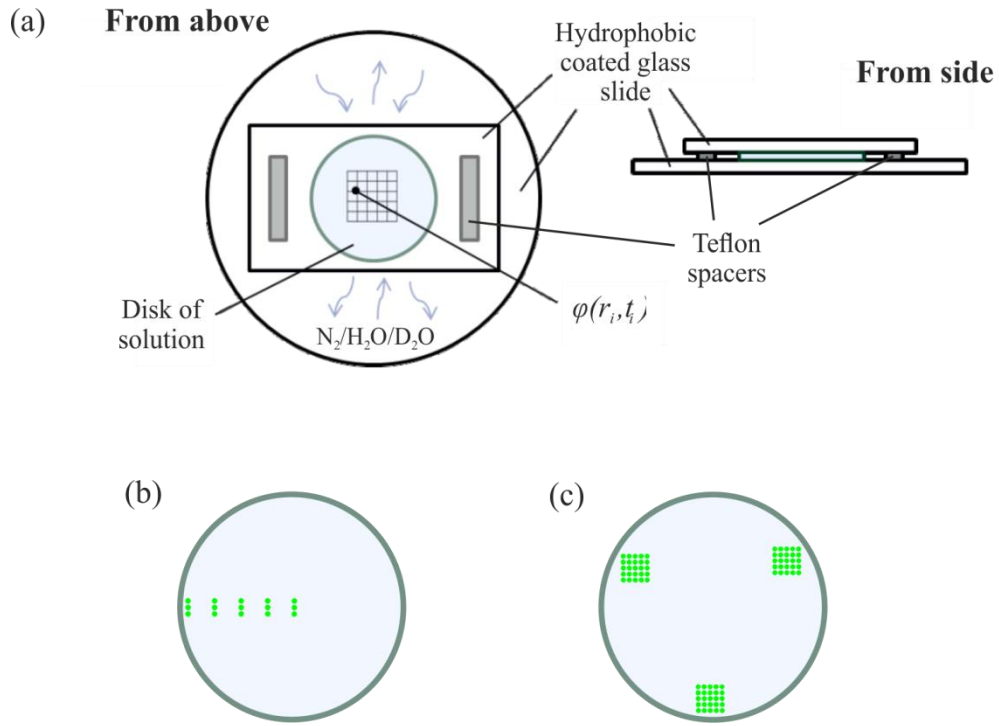


Figure 2. (a) Arrangement of a disk on the slide in the D₂O diffusion experiment. The slide was placed in the RH and temperature controlled chamber. A typical disk of solution had a size of around 200-300 μm , and the laser spot had a size of 1.3 μm . The 25 μm -thick spacers and top slide were held in place using vacuum grease. Each Raman spectrum was obtained at a predetermined location on a grid, whose size and spacing was chosen to suit the rate of diffusion in each experiment. Other spacer thicknesses were tested to ensure that results were the same and there were no edge effects. (b) Spectral collection configuration for diffusion measurements at high RH. (c) Spectral collection configuration for diffusion measurements at low RH.

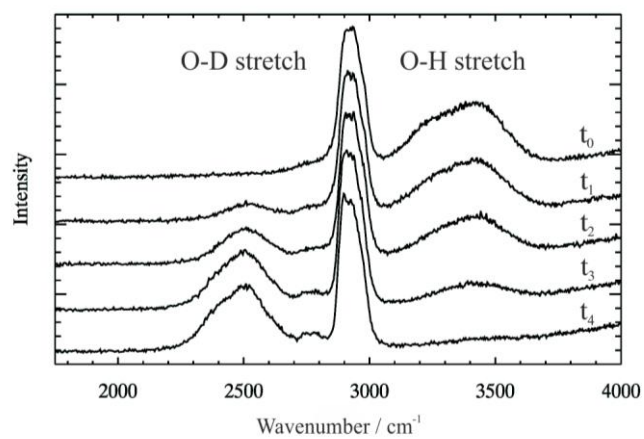


Figure 3. Raw Raman data showing the gradual decrease in the O-H stretch (at 3100 to 3500 cm^{-1}) band and increase in the O-D stretch (at 2300 to 2700 cm^{-1}) band after switching from H_2O to D_2O vapour. The central band (at 2800 to 3100 cm^{-1}) is the C-H stretch.

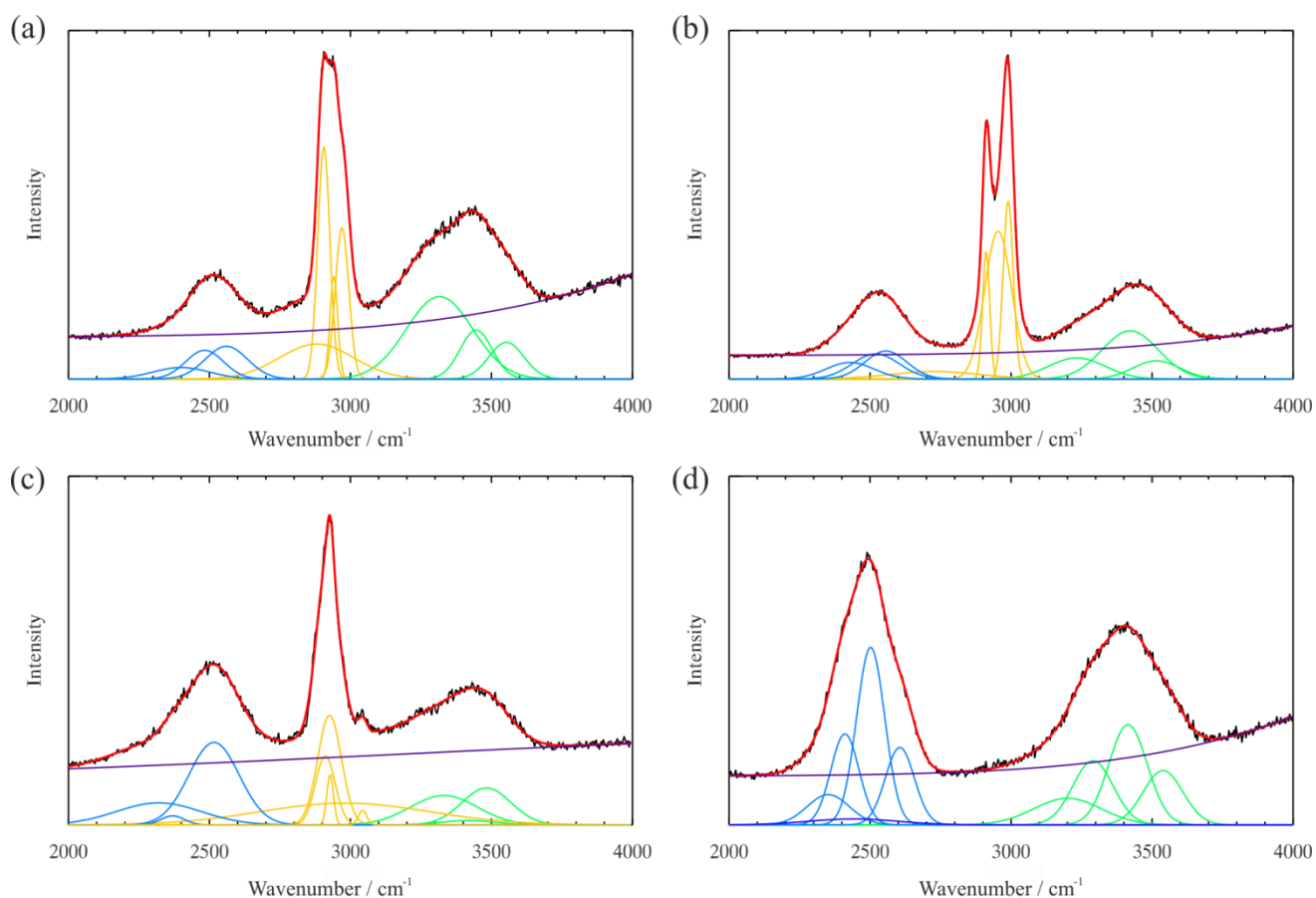


Figure 4. Example fits to the Raman spectra of (a) sucrose, (b) levoglucosan, (c) MgSO_4 and (d) raffinose/M5AS. Raw data is shown in black, green peaks are fitted to the O-H (and NH_4^+ in raffinose/M5AS) stretch, blue peaks are fitted to the O-D (and ND_4^+ in raffinose/M5AS) stretch, orange peaks are fitted to the C-H stretch, and the background fit is shown in purple. The sum of all fitted peaks is shown in red. Green and blue peaks were allowed to vary as H/D exchange took place; orange peaks were more tightly constrained.

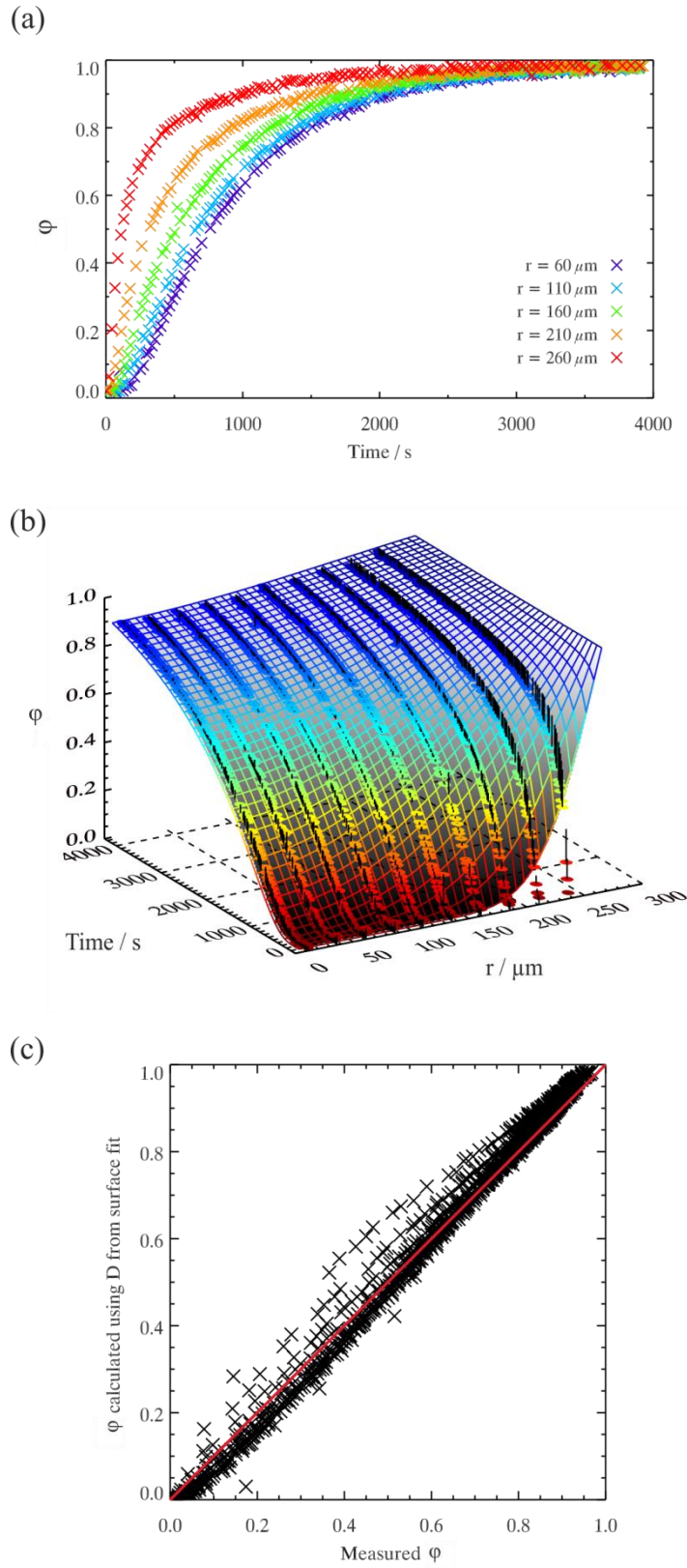


Figure 5. (a) Plot of fractional concentration of O-D bonds, ϕ , as a function of time after switch from H_2O to D_2O vapour in a $278\text{ }\mu\text{m}$ (radius) levoglucosan disk at 60% RH. Colours indicate radial distance (where $r = 0\text{ }\mu\text{m}$ is the centre of the disk): red data points are near to the edge, blue are near the centre. This was a one-off experiment in which data was repeatedly collected at five specific radial positions, for the purpose of visualising how concentration changed at a fixed location over time. (b) Plot of the fractional concentration of O-D bonds, ϕ , as a function of radial distance and time after switch from H_2O to D_2O vapour

in the levoglucosan disk at 50% RH. The grid shows the surface fit to the data, the collected data points are joined to the grid by black lines. The colour corresponds to the value of φ . (c) A one-to-one plot showing the measured φ vs φ according to the fit, also for levoglucosan at 50% RH.

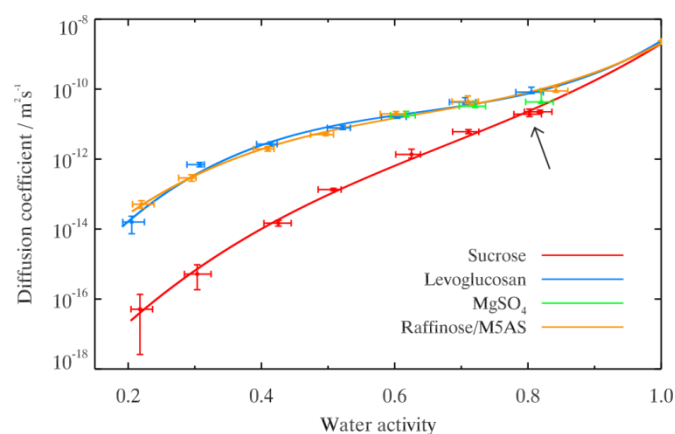


Figure 6. Measured water diffusion coefficients of sucrose, levoglucosan, magnesium sulfate and raffinose/M5AS. Lines are fitted to the data with an empirical polynomial fit (Eq. (4)). The arrow indicates a measurement performed on a sucrose disk which had previously been held at 50°C for a week. We infer from the fact that this data point agrees well with its neighbour that equilibrating the samples at high temperatures does not alter their diffusion properties.

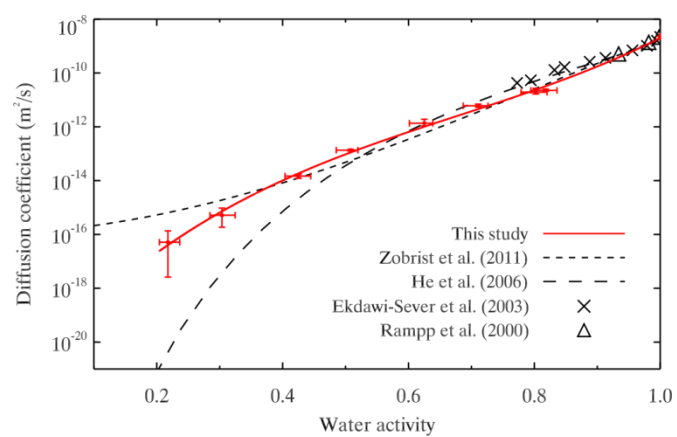


Figure 7. Measured diffusion coefficients of water in aqueous sucrose solutions, with comparison to the parameterisations for sucrose solutions given in Zobrist et al. (2011) and He et al. (2006), and the NMR measurements of Ekdawi-Sever et al. (2003) and Rampp et al. (2000).

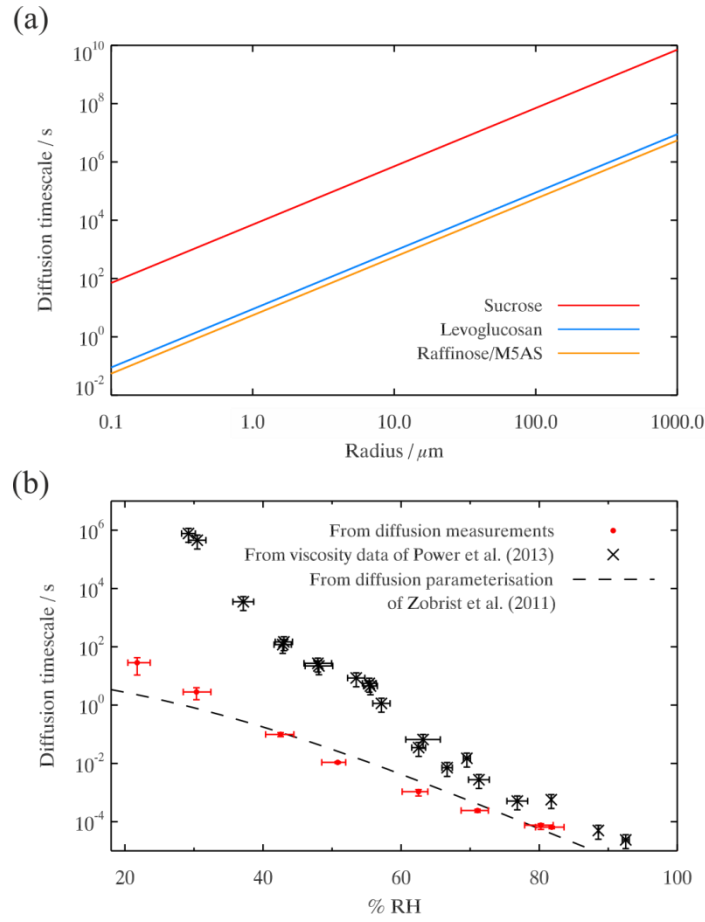


Figure 8. (a) Water diffusion timescales as a function of radius for spherical droplets at 20% RH, calculated according to Eq. (5). Sucrose is shown in red, levoglucosan in blue and raffinose/M5AS in orange. (b) Water diffusion timescales (Eq. (5)) as a function of relative humidity for 100 nm particles. Red dots show timescales calculated using water diffusion coefficients measured in this study, the dashed black line shows these timescales using the water diffusion coefficient parameterization given in Zobrist et al. (2011). Black crosses are calculated using the Stokes-Einstein relation with viscosity data from Power et al. (2013) (error bars correspond to a water molecular diameter uncertainty of 0.2 ± 0.1 nm).

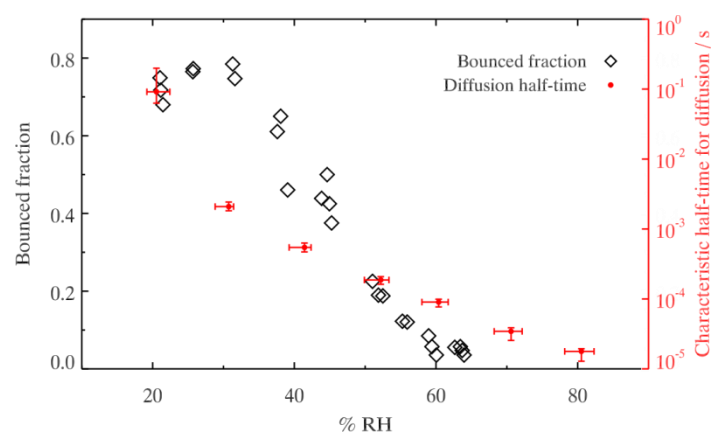


Figure 9. Characteristic timescales for diffusion of D_2O in 100 nm aqueous levoglucosan aerosol particles (red dots), compared with bounce fraction data from Saukko et al. (2012) (black diamonds).

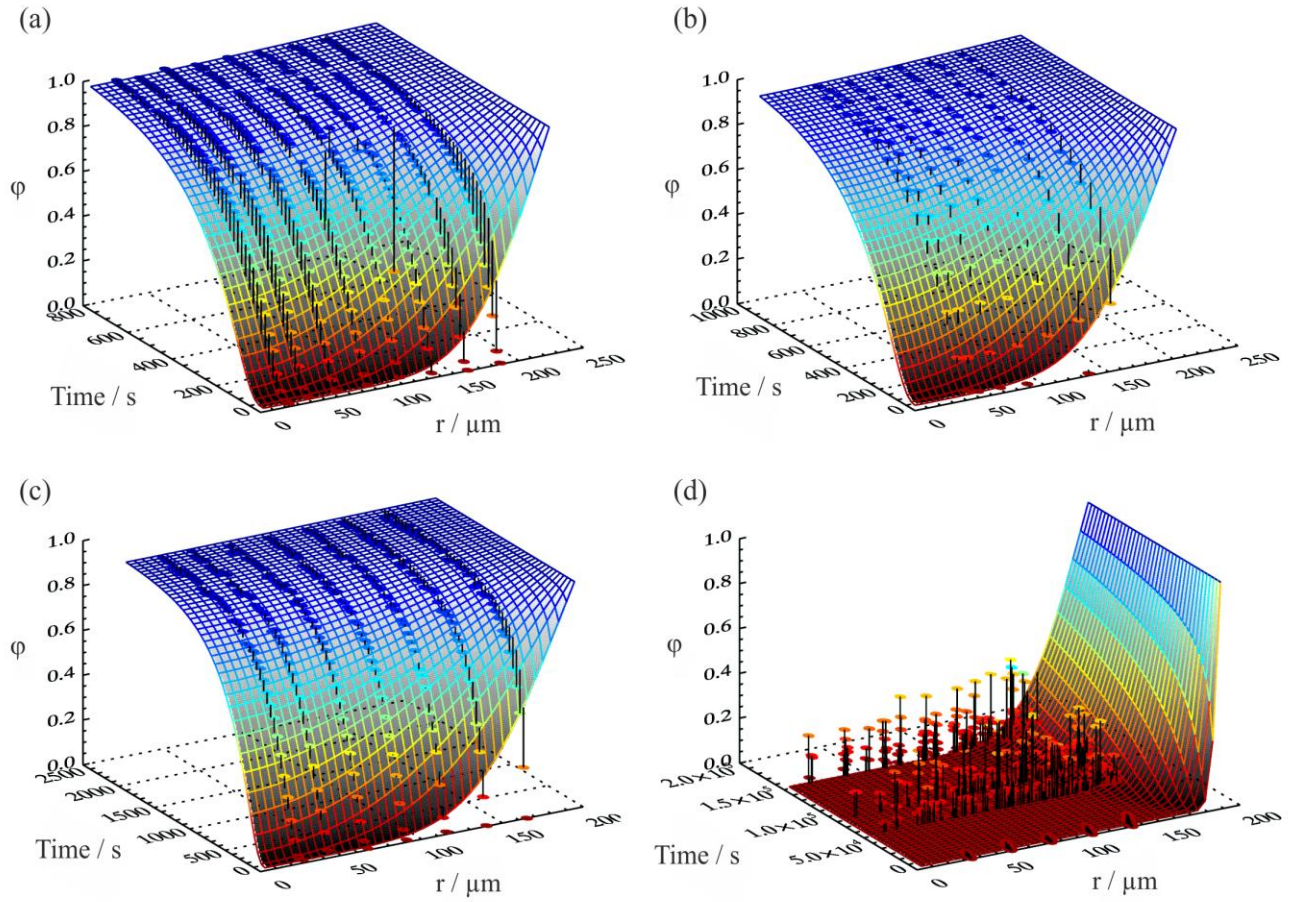


Figure 10. Surface fits to ϕ as a function of radial position and time, using Eq. (2), at an RH of (a) $82 \pm 2\%$, (b) $72 \pm 2\%$, (c) $62 \pm 2\%$ and (d) $51 \pm 2\%$ for magnesium sulfate. The quality of the fit is poor at 51% RH, which reflects the lack of homogeneity in the sample.

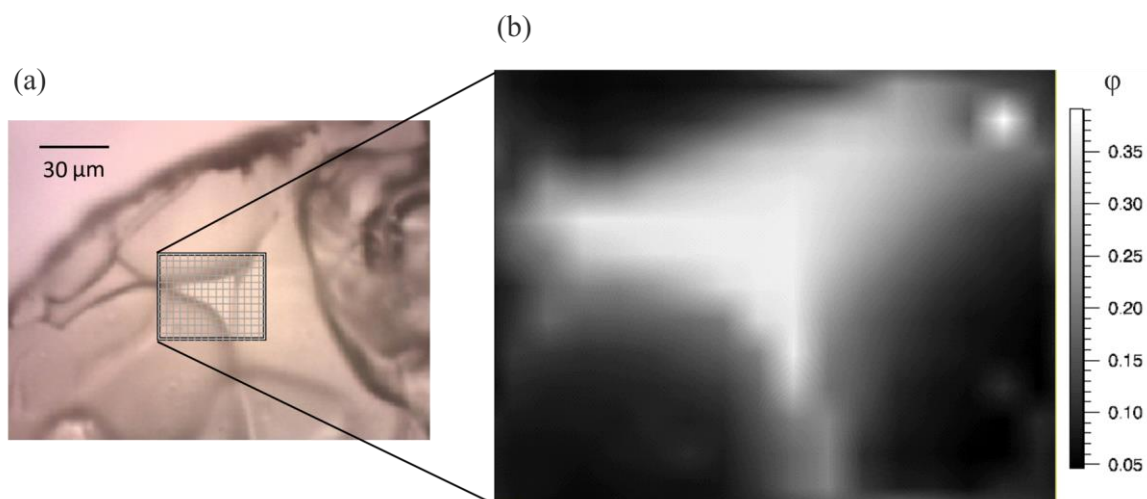


Figure 11. (a) Visible inhomogeneities in the magnesium sulfate disk at 51% RH. This disk was exposed to D_2O vapour for one day before a Raman map was acquired across the grid shown. (b) Fractional concentration of O-D bonds after one day. It appears that D_2O diffused faster into the central triangular shape than the surrounding regions.

	a	b	c	d
Sucrose	-20.89	25.92	-26.97	13.25
Levogluconan	-18.41	31.10	-44.43	23.12
Raffinose/M5AS	-17.21	24.00	-32.50	17.02

Table 1. Fit parameters a to d used in Eq. (4) for $D_{\text{water}}(a_w)$.