



SOA formation from  
glyoxal in a 3-D  
model

C. Knote et al.

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# Simulation of semi-explicit mechanisms of SOA formation from glyoxal in a 3-D model

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Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures



Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



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## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures



Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



## Abstract

New pathways to form secondary organic aerosols (SOA) have been postulated recently. Glyoxal, the smallest dicarbonyl, is one of the proposed precursors. It has both anthropogenic and biogenic sources, and readily partitions into the aqueous-phase of cloud droplets and deliquesced aerosols where it undergoes both reversible and irreversible chemistry. In this work we extend the regional scale chemistry transport model WRF-Chem to include a detailed gas-phase chemistry of glyoxal formation as well as a state-of-the-science module describing its partitioning and reactions in the aqueous-phase of aerosols. A comparison of several proposed mechanisms is performed to quantify the relative importance of different formation pathways and their regional variability. The CARES/CalNex campaigns over California in summer 2010 are used as case studies to evaluate the model against observations.

In all simulations the LA basin was found to be the hotspot for SOA formation from glyoxal, which contributes between 1% and 15% of the model SOA depending on the mechanism used. Our results indicate that a mechanism based only on a simple uptake coefficient, as frequently employed in global modeling studies, leads to higher SOA contributions from glyoxal compared to a more detailed description that considers aerosol phase state and chemical composition. In the more detailed simulations, surface uptake is found to be the main contributor to SOA mass compared to a volume process and reversible formation. We find that contribution of the latter is limited by the availability of glyoxal in aerosol water, which is in turn controlled by an increase in the Henry's law constant depending on salt concentrations ("salting-in"). A kinetic limitation in this increase prevents substantial partitioning of glyoxal into aerosol water at high salt concentrations. If this limitation is removed, volume pathways contribute > 20% of glyoxal SOA mass, and the total mass formed (5.8% of total SOA in the LA basin) is about a third of the simple uptake coefficient formulation without consideration of aerosol phase state and composition. All these model formulations are based on very limited and recent field or laboratory data and we conclude that the current

ACPD

13, 26699–26759, 2013

### SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



uncertainty on glyoxal SOA formation spans a factor of 10 in this domain and time period.

## 1 Introduction

Organic matter is a major contributor to atmospheric aerosol load. While a fraction of it is directly emitted as primary particles into the air, a substantial contribution – often the majority – is generated within the atmosphere through chemistry. The progressive oxidation of precursor gases (volatile organic compounds, VOCs) leads to semivolatile products that have lower volatility and higher water solubility than their parent precursors, and can form secondary organic aerosols (SOA). The large number of organic compounds involved proves to be challenging for both measurement and modeling communities (e.g. Goldstein and Galbally, 2007). Recent studies further indicate that SOA formation in the aqueous phase may be important (Lim et al., 2010; Ervens et al., 2011). Glyoxal is the smallest dicarbonyl produced by oxidation of both anthropogenic and biogenic precursors. It is short-lived, so it can serve as an indicator for recent VOC oxidation processes in the atmosphere (Volkamer et al., 2005). Laboratory measurements for glyoxal (Carlton et al., 2007; Noziere et al., 2009; McNeill et al., 2012) confirmed that it partitions into the particle phase under atmospheric conditions due to its high solubility in water and its subsequent aqueous-phase chemistry. In particular, recent work has shown that concentrated salt solutions strongly shift the partitioning of glyoxal towards the liquid-phase (Kampf et al., 2013), where it is a source for SOA with a high oxygen-to-carbon ratio (Waxman et al., 2013). Finally, Holzinger et al. (2013) found that a multi-phase process might be needed to explain the aerosol nitrogen content in the very region and time period we will focus on.

Formation of SOA from glyoxal has been studied with steady-state box models (Volkamer et al., 2007; Lim et al., 2010; Waxman et al., 2013) and with a 1-D transport model employing the Master Chemical Mechanism (Washenfelder et al., 2011) with the difference between measured and modelled glyoxal concentrations attributed

## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures



Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



**SOA formation from glyoxal in a 3-D model**

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



to an aerosol loss process. Volkamer et al. (2007) find that glyoxal could contribute 15% to total SOA in Mexico City for the period investigated. Washenfelder et al. (2011) however derive lower uptake coefficients over California and conclude that glyoxal contribution to total SOA is in the range of 0–4%. Uptake coefficients derived by Liggi-  
5 et al. (2005) were used in global 3-D simulations with simplified chemistry (Fu et al., 2008) where they found that glyoxal contributes additional  $2.6 \text{ TgCa}^{-1}$  to the global SOA source of  $17.5 \text{ TgCa}^{-1}$  from a traditional mechanism ( $\sim 13\%$ ). The partitioning behaviour was hence already studied at urban and global scales. Previous box-model studies were locally constrained by observations but were run in idealized conditions  
10 and lack most of the variability due to emissions, transport and chemistry found at the regional scale. Global-scale 3-D simulations on the other hand lack the degree of detail in their description of the glyoxal atmospheric processing necessary for accurate simulations. Most previous studies did not consider aerosol properties other than surface area, or included reversible partitioning of glyoxal into deliquesced droplets. Waxman et al. (2013) is the only study that incorporated recent findings regarding the “salting-in”  
15 of glyoxal, that is, the strong ( $\sim 3$  orders of magnitude) increase in the effective Henry’s law constant due to glyoxal interactions with salts (Kampf et al., 2013). Waxman et al. (2013) concluded that the catalytic reaction of  $\text{NH}_4^+$  (Noziere et al., 2009) exceeds reactions by OH radicals to form SOA from glyoxal, and pointed out the importance of  
20 particle pH, and phase state for the rate of SOA formation from glyoxal. There is no previous study of glyoxal-SOA formation in a 3-D regional model. A model capable to represent detailed aerosol characteristics, like pH, size resolved chemical composition, and phase state could provide valuable information to assess which parameters determine the regional variability of glyoxal SOA.

25 In this study we investigate the life cycle of glyoxal in the atmosphere at the regional scale over California. We choose summer 2010 as our study period as the Carbonaceous Aerosols and Radiative Effects Study (CARES, Zaveri et al., 2012) and the Research at the Nexus of Air Quality and Climate Change (CalNex, Ryerson et al., 2013) campaigns took place in California during that time, and provide measurements that al-

**SOA formation from glyoxal in a 3-D model**

C. Knote et al.

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[⏪](#)[⏩](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

low constraining glyoxal sources, concentrations, and sinks. A number of studies using this dataset investigated SOA formation (Duong et al., 2011; Ahlm et al., 2012; Gentner et al., 2012; Liu et al., 2012a, b; O'Brien et al., 2012; Rollins et al., 2012; Zhang et al., 2012; Hayes et al., 2013; Hersey et al., 2013; Setyan et al., 2012; Shilling et al., 2013; Zhao et al., 2013), the oxidative environment (Pusede and Cohen, 2012; Warneke et al., 2013) and emissions of compounds related to SOA formation like VOCs (Borbon et al., 2013), NO<sub>x</sub> (McDonald et al., 2012; Brioude et al., 2013), or ammonia (Nowak et al., 2012). We employ the mesoscale chemistry transport model WRF-chem (Grell et al., 2005) which we extended to include a more detailed description of gas-phase chemistry producing glyoxal as well as a new module to describe its partitioning into the aerosol phase. Substantial effort has been put in our study into improving existing emission inventories for this region and period, and in evaluating the model against measurements. Several pathways to form SOA from glyoxal were added, including reversible partitioning. We present results from a number of simulations in which we considered (combinations of) several approaches to this partitioning, and investigate relative contributions and their regional variability. In Sect. 2 we describe the modeling system, including updates made to emissions and gas-phase chemistry, and the modeling of glyoxal partitioning into the particle phase. Section 3 shows how our simulations compare against measurements made during the CARES/CalNex campaigns, and emphasizes the improvements due to the updated emissions inventory. In Sect. 4 we investigate the partitioning behaviour of glyoxal based on the different pathways and its regional variability.

## 2 Modeling

### 2.1 Model setup

Simulations and developments are based on the Weather Research and Forecasting Model coupled with Chemistry (WRF-chem) (Grell et al., 2005; Fast et al., 2006)

**SOA formation from  
glyoxal in a 3-D  
model**

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

in version 3.4.1. Table 1 lists the options chosen for physical parameterizations. The chemical mechanism of the Model for Ozone and Related Chemical Tracers, version 4 (MOZART-4, Emmons et al., 2010) is employed to calculate gas-phase chemical reactions, with the fast Tropospheric Ultraviolet-Visible (fTUV) module (Tie et al., 2003) used for calculating photolysis rates. Aerosol dynamics and associated physical and chemical processes are represented by the Model for Simulating Interactions and Chemistry (MOSAIC, Zaveri et al., 2008) using 4 size bins (39–156 nm, 156–625 nm, 625–2.5  $\mu\text{m}$ , 2.5–10  $\mu\text{m}$ ). Inorganic aerosol thermodynamics within MOSAIC are solved using the multicomponent equilibrium solver for aerosols (MESA, Zaveri et al., 2005). Secondary organic aerosol (SOA) formation from anthropogenic VOCs and semi/intermediate volatility compounds (S/IVOCs) is parameterized with the method of Hodzic and Jimenez (2011), where SOA is formed based on a tracer co-emitted with carbon monoxide. By reaction with OH this tracer is then converted into SOA. Treatment of biogenic SOA follows Shrivastava et al. (2011).

The model domain (Fig. 1) covers the state of California including adjacent states and parts of the Pacific Ocean, and is represented as a  $280 \times 280 \times 40$  cell grid on a Lambert conformal conic projection with a horizontal resolution of 4 km, extending vertically up to 10 hPa. Our simulations are conducted over this region for the period of 29 May until 15 June 2010.

All simulations are initialized and forced at the lateral boundaries by 6 hourly analysis data from the NCEP Global Forecast System (GFS) at  $1.0^\circ$  resolution for meteorology, and 6 hourly MOZART-GEOS5 simulations (based on Emmons et al., 2010) at  $1.9 \times 2.5^\circ$  resolution for chemistry and aerosols. Simulations are nudged to GFS analyses above the boundary layer throughout the simulation. Sea surface temperature (SST) analyses updated every 6 hours from the US Navy Fleet Numerical Meteorology and Oceanography Center ([http://www.usgodae.org/ftp/outgoing/fnmoc/models/ghrsst/docs/ghrsst\\_doc.txt](http://www.usgodae.org/ftp/outgoing/fnmoc/models/ghrsst/docs/ghrsst_doc.txt)) are used instead of default climatologies.

## 2.2 Emissions

Anthropogenic emission estimates were created as a combination of the National Emission Inventory dataset for 2005 (NEI 2005), the California Air Resources Board emission estimates for 2008 (CARB 2008), the inversion-based CO and NO<sub>x</sub> emissions for the LA basin as presented by Brioude et al. (2013), and VOC/CO emission ratios of Borbon et al. (2013). The following steps are applied to create the final anthropogenic emission dataset (see also Fig. 1). The NEI 2005 emissions dataset was used as basis. Over California, it is first replaced by the CARB 2008 inventory. VOCs in both the NEI and CARB inventory were originally speciated for the SAPRC99 mechanism (Carter et al., 2000), and were translated to species available in MOZART. Emissions are then reduced by 35 % for most species to account for the changes between 2005/2008 and 2010 (Table 2). CO and NO<sub>x</sub> emission amounts for grid points in the LA basin are replaced with inversion estimates from Brioude et al. (2013).

VOC/CO emission ratios reported by Borbon et al. (2013) were used to create VOC emission fields for selected VOCs, using the spatial distribution and temporal behaviour of the CO emissions created as described above. These relationships were derived from measurements in the Los Angeles basin during 2010, and are more recent and considered more accurate than estimates from the CARB and NEI inventories. We replaced the VOC emission estimates from the inventories with our inversion based inventory for the species available in Borbon et al. (2013). Figure 2 shows that there are considerable differences in the estimates of speciated VOC emissions between the CARB inventory and the amounts estimated from the CO inversion and the measured VOC/CO emission ratios in the LA basin. An investigation of these differences would be beneficial for the modeling community, but is out of the scope of this work.

The final emissions used in our simulations are a result of an iterative approach to reconcile simulated concentrations with observations. Note that for acetone, benzene, and xylenes the inversion-based emissions are much higher than reported in CARB. Simulations made with the inversion-based amounts showed strongly overestimated

### SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures



Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion





concentrations compared to surface measurements. Hence we reduced these emissions so that simulated concentrations agreed with measurements. Interestingly, the resulting emission amounts for these species were very similar to the ones reported in the inventory. A comparison of a simulation using the original emissions as given from the combination of NEI/CARB inventories and our updated inventory against measurement data is discussed in Sect. 3.

Biogenic emissions from vegetation are calculated as a function of ambient conditions with the Model of Emissions of Gases and Aerosols from Nature (MEGAN) version 2.04 (Guenther et al., 2006). During evaluation we found a large underestimation of isoprene compared to measurements in urban areas like the LA basin (LA ground site), while no such underestimation was found in rural environments (Bakersfield, T1 sites). We attribute this to an insufficient description of vegetation in urban areas in the MEGAN land use database (A. Guenther, NCAR, pers. comm., 2012) and have hence increased emissions of all BVOCs as determined in MEGAN by a factor of 2.5 over grid points with an “urban” land use type in WRF-chem.

### 2.3 The modelled glyoxal lifecycle

The MOZART mechanism describes the formation of glyoxal in the gas phase. Direct anthropogenic emissions of glyoxal are included according the CARB emissions inventory. Glyoxal is removed from the atmosphere by photolysis, OH oxidation, and dry deposition. A constant dry deposition velocity of  $2 \text{ mm s}^{-1}$  is used as derived in Washenfelder et al. (2011) based on nighttime decreases of glyoxal at the LA ground site. Wet deposition is not considered.

#### 2.3.1 Updates to the MOZART-4 gas-phase mechanism

Table 3 lists new and updated species, Table 4 the new and altered reactions, and Table 5 the added photolysis reactions. All changes presented are relative to the mechanism described in Emmons et al. (2010).

## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



Formation of glyoxal in the gas-phase is described in the MOZART mechanism, which already treats glyoxal as an explicit species (Emmons et al., 2010). Three main formation pathways are considered: oxidation of isoprene, of aromatic compounds, and of ethyne. We updated the description of isoprene oxidation in MOZART by adding a 2% molar yield of each glyoxal, methylglyoxal, glycolaldehyde, and hydroxyacetone from the reaction of the isoprene peroxy radical with NO and with NO<sub>3</sub>, the former having recently been observed in laboratory experiments (Volkamer et al., 2005; Gal-  
loway et al., 2011, Thalman et al., 2013). We further included a more explicit treatment of C5 unsaturated hydroxycarbonyl chemistry, a potential additional secondary glyoxal source. Note, however, that while this formation of glyoxal is expected theoretically, it has not been observed in recent laboratory experiments (Thalman et al., 2013). We have further speciated the aromatics (previously lumped as one surrogate) into benzene, toluene and lumped isomers of xylenes with corresponding glyoxal yields for each precursor (Volkamer et al., 2001; Calvert et al., 2002). The treatment of these aromatic species is taken from the Leeds Master Chemical Mechanism (Bloss et al., 2005, <http://mcm.leeds.ac.uk/MCM/>), although the chemistry of some later-generation products is ignored as these multi-functional species are likely prone to heterogeneous removal, are oxidized far from the source region, and/or are not significant sources of glyoxal. Ethyne has also been added as an explicit species and its oxidation chemistry is included; the rate coefficient for its reaction with OH is from the JPL evaluation (Sander et al., 2011) and products are from Hatakeyama et al. (1986). Also, HONO is considered as an additional source of OH radicals through photolysis. Glyoxal production from 2-methyl-3-buten-2-ol (MBO) is not included.

### 2.3.2 Pathways of interaction between gas-phase glyoxal and aerosols

There is no clear scientific consensus on how glyoxal might form SOA. Irreversible SOA formation via both a surface uptake and volume reactions have been proposed (Ervens and Volkamer, 2010, and references therein). Reversible partitioning and oligomerization reactions are observed in laboratory studies (Kampf et al., 2013), including a sub-

stantial increase in effective Henry's law constant due to glyoxal-salt interactions. The interactions between reversible and irreversible formation pathways are still unknown. We have included all these proposed pathways into WRF-chem, and investigate their relative importance with simulations where only selected processes are active. Below we describe how each process is implemented.

Glyoxal can partition into the aerosol phase reversibly and irreversibly (Fig. 3):

(i) Reversible formation of monomer species (glyoxal, glyoxal monohydrate, and glyoxal dihydrate, pool 1 species) and oligomers (pool 2 species) are included according to Kampf et al. (2013), who observed an increase in the Henry's law constant by up to three orders of magnitude (compared to the value over pure water) in particles containing ammonium sulfate. This increase of the effective Henry's law constant,  $K_{h,eq}$ , was attributed to salt-glyoxal interactions ("salting-in"). Our implementation of  $K_{h,eq}$  follows Eq. (3) in Kampf et al. (2013):

$$K_{h,eq} = K_{h,water} / 10^{(-0.24 \min(12.0, (c_{as} + c_{an})))} \quad (1)$$

where  $K_{h,water} = 4.19 \times 10^5 \text{ Matm}^{-1}$  is the Henry's law constant of glyoxal over water,  $-0.24$  the "salting-in" constant of ammonium sulfate, which we assume to be the same for ammonium nitrate, with  $c_{as}$  and  $c_{an}$  the concentrations ( $\text{mol kg}^{-1}$ ) of ammonium sulfate and nitrate respectively. The sum  $c_{as} + c_{an}$  in the calculation above is limited to  $12 \text{ mol kg}^{-1}$ , per the results of Kampf et al. (2013), who found no further increase in  $K_{h,eq}$  at higher concentrations within the timescales of their experiments.

The oligomerization constant  $K_{olig}$  (as used by Ervens and Volkamer, 2010) defines the equilibrium partitioning between monomers (pool 1) and oligomers (pool 2) as

$$\frac{\text{Gly}_{p2,eq}}{\text{Gly}_{p1,eq}} = K_{olig}, \quad (2)$$

with  $\text{Gly}_{p1,eq}$  and  $\text{Gly}_{p2,eq}$  the concentrations in each pool at equilibrium. Kampf et al. (2013) determined that  $K_{olig}$  is approximately unity, for  $(c_{as} + c_{an}) < 12 \text{ mol kg}^{-1}$  and  $0.5$

## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



for  $(c_{as} + c_{an}) > 12 \text{ mol kg}^{-1}$ . They also showed a significant time-dependence of the rate at which the Henry's law equilibrium is established. In particular, the characteristic time scales for gas-particle partitioning of monomer species (glyoxal, monohydrate, and dihydrate) is on the order of minutes at  $(c_{as} + c_{an}) < 12 \text{ mol kg}^{-1}$ , and several hours at  $(c_{as} + c_{an}) > 12 \text{ mol kg}^{-1}$ . Both of these timescales are larger than a typical model timestep in WRF-chem (30 s). To capture this time dependence in our model we calculate explicitly the glyoxal flux from the gas-phase to the monomer pool or vice versa as

$$\frac{\partial \text{Gly}_{p1}}{\partial t} = \frac{1}{\tau_1} \cdot (\text{Gly}_{p1,eq} - \text{Gly}_{p1}) \quad (3)$$

with  $\text{Gly}_{p1}$  the current glyoxal monomer concentration in the aerosol liquid phase and  $\tau_1$  the characteristic time scale for the monomer pool 1. In this work  $\tau_1 = 2.5 \times 10^2 \text{ s}$  for  $(c_{as} + c_{an}) < 12 \text{ M}$ , and  $4.4 \times 10^4 \text{ s}$  for  $(c_{as} + c_{an}) > 12 \text{ M}$  (Kampf et al., 2013). Phase-separation of organic material with low O/C (e.g. Bertram et al., 2011) was not considered in the calculation of the liquid-phase concentration.

The oligomer pool is in equilibrium with the monomer pool, with the flux defined similarly as

$$\frac{\partial \text{Gly}_{p2}}{\partial t} = \frac{1}{\tau_2} \cdot (\text{Gly}_{p2,eq} - \text{Gly}_{p2}). \quad (4)$$

with  $\tau_2$  the characteristic time scale for the oligomer pool, and  $\text{Gly}_{p2}$  the current glyoxal concentration in the oligomer pool. In this work  $\tau_2 = 5.5 \times 10^3 \text{ s}$  for  $(c_{as} + c_{an}) < 12 \text{ mol kg}^{-1}$ , and  $4.7 \times 10^4 \text{ s}$  for  $(c_{as} + c_{an}) > 12 \text{ mol kg}^{-1}$  (Kampf et al., 2013).

(ii) Three pathways for irreversible formation are considered: an ammonium-catalysed volume pathway as presented in Noziere et al. (2009), an OH reaction volume pathway, and a surface uptake (both presented in Ervens and Volkamer, 2010).

The ammonium-catalyzed pathway is parameterized as a bulk (volume) reaction with a rate constant that depends exponentially on both particle acidity (pH) and the activity

SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



of the ammonium ion ( $a_{\text{NH}_4}$ ). Noziere et al. (2009) suggest a second-order rate constant from a two step reaction, but could not identify the second-order rate limiting step. We use this rate constant expression (Noziere et al., 2009, Eq. 12) as

$$k_{\text{II}} = 2 \times 10^{-10} \cdot \exp(1.5 \cdot a_{\text{NH}_4}) \cdot \exp(2.5 \cdot \text{pH}), \quad (5)$$

(in  $\text{M}^{-1} \text{s}^{-1}$ ) and convert into an apparent first-order rate constant ( $k_1$ ) by multiplying with the particulate glyoxal concentration. We approximate  $a_{\text{NH}_4}$  as its concentration ( $a_{\text{NH}_4} = c_{\text{NH}_4}$ ) because the thermodynamic module used (MESA Zaveri et al., 2005) does not provide activity coefficients for individual ions. The relationship between reversible and irreversible pathways is not well understood. Here we assume that the total concentration in the monomer pool, which includes hydrated forms, is the only particulate glyoxal concentration available to the ammonium-catalyzed reaction.

The OH pathway is a simplified version of what has been developed in Ervens and Volkamer (2010). Liquid-phase OH is assumed to be in equilibrium with the gas-phase with a Henry's law constant of  $25 \text{ Matm}^{-1}$ . Glyoxal is converted irreversibly into SOA by reaction with OH with a rate constant  $k_{\text{OH}} = 1.1 \times 10^9 \text{ M}^{-1} \text{ s}^{-1}$  (Buxton et al., 1997). This rate constant has been derived for dilute solutions (cloud droplets), and is suggested to be a lower limit for more concentrated solutions like aerosols (Ervens and Volkamer, 2010). Like the ammonium reaction also the OH reaction uses the total glyoxal concentration in the monomer pool.

The third irreversible pathway describes a surface uptake parameterized by Ervens and Volkamer (2010) as

$$\frac{\partial \text{Gly}_g}{\partial t} = 0.25 \cdot \gamma_{\text{phot}} \cdot A \cdot \omega \cdot \text{Gly}_g \quad (6)$$

where  $\gamma_{\text{phot}} = 1.0 \times 10^{-3}$  is the uptake coefficient,  $A$  the aerosol surface area concentration ( $\text{cm}^2 \text{ cm}^{-3}$ ), and  $\omega$  the mean gas-phase velocity of glyoxal under current conditions ( $\text{cm s}^{-1}$ ).

## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



This latter pathway was used in previous global (Fu et al., 2008; Stavrou et al., 2009) and other model studies (Volkamer et al., 2007; Washenfelder et al., 2011; Waxman et al., 2013) for scenarios without explicit glyoxal gas-to-particle partitioning and particle-phase chemistry, when the uptake onto particles was approximated to depend only on aerosol surface area (and not phase state or chemical composition). There is no experimental evidence so far to determine whether this surface uptake can also take place on aerosols of any phase state, or whether a liquid phase is necessary.

The system of equations is integrated numerically using a 4th-order Runge–Kutta method (Press, 1992) at each call to the chemistry subsystem in WRF-chem for each size bin in MOSAIC. Only size bins which MOSAIC considers mixed-phase or completely deliquesced are considered, except for the simulations in which we mimic previous modeling studies. All 4 glyoxal SOA reservoirs (2 reversible pools, volume pathway and surface uptake) in each size bin are treated as prognostic variables and hence subject to all physical processes acting upon aerosols in WRF-chem.

### 2.4 Comparison of the formation pathways for glyoxal SOA

We conducted a total of 7 simulations listed in Table 6 to evaluate our model and investigate the effects of different pathways and their interactions. In RAW and BASE, glyoxal has no interactions with aerosols. The BASE simulation is used to evaluate model performance against measurement data. In the RAW simulation we use the unmodified emission inventories to show the effects of our updated inventory. The SIMPLE case emulates previous modeling studies like Volkamer et al. (2007); Fu et al. (2008) or Washenfelder et al. (2011), in which SOA formation from glyoxal is parameterized as surface uptake, only depends on available aerosol surface area, and does not consider aerosol chemical composition or phase state. The VOLUME case assumes that SOA is formed through a volume (ammonium-catalyzed) pathway – rather than a surface process. In the HYBRID case, both volume and surface processes are active, and both require a mixed-phase or deliquesced aerosol. In both the VOLUME and HYBRID studies reversible partitioning is considered as well, and the increase in the Henry's

law constant due to “salting-in” (Kampf et al., 2013) potentially increases the glyoxal available in the aqueous-phase. Two additional sensitivity studies were made: FAST assumes that the increase in Henry’s law constant due to “salting-in” does not have a time dependence. Hence in FAST glyoxal in the monomer pool (pool 1) is assumed to be in instantaneous equilibrium with the gas-phase according to the “salted-in” Henry’s law constant. The behaviour of glyoxal oligomers (pool 2) is unchanged. With FAST\_PH we investigate how the kinetic limitation of glyoxal partitioning at high salt concentrations affects contributions from oligomers, and assess the potential contribution of glyoxal to SOA in an environment with reduced aerosol acidity. As described in the previous section the characteristic time scales to fill the monomer and oligomer pools change for salt concentrations  $> 12 \text{ mol kg}^{-1}$ . This translates into a slower partitioning of glyoxal into the particle-phase, and consequently no further increase in glyoxal for the ammonium-catalyzed reaction pathway. FAST\_PH is based on FAST, hence also here we assume instantaneous equilibration for pool 1 (monomers). In FAST\_PH we additionally keep the fast characteristic time scales for pool 2 (oligomers) at high salt concentrations. We further calculate ammonium-catalyzed reaction rates at a pH increased by 2 over the actual value. The exponential dependence of the ammonium-catalyzed reaction in Eq. (5) suggests a much stronger contribution at higher pH, and indeed this dependence is largely responsible to explain findings in Mexico City (Volkamer et al., 2007; Waxman et al., 2013), where  $\sim 15\%$  of SOA is formed from glyoxal. All cases except SIMPLE consider the aerosol phase state, and in those no glyoxal SOA formation occurs once aerosols are considered dry by the aerosol module.

### 3 Model evaluation

In order to investigate the atmospheric fate of glyoxal, the model needs to accurately simulate meteorology (transport patterns, radiation), the gas-phase precursors of glyoxal (isoprene, aromatics, ethyne), the oxidation capacity of the atmosphere (OH), photolysis, and ambient aerosol characteristics. We performed a sim-

## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion











inability of the model to simulate local conditions may stem from strongly overestimated isoprene concentrations at Bakersfield (Fig. 4), which would lead to a suppression of OH, and consequently to an underestimation of glyoxal concentrations.

The main sinks for glyoxal in the atmosphere are reaction with OH, photolysis, and dry and wet deposition. OH concentrations (Fig. 6) are on average underestimated in the model when compared to measurements at the Pasadena ground site. While there is substantial uncertainty in how well models represent the hydroxyl radical, OH measurements have large uncertainties as well (Mao et al., 2012). Further, local conditions at the Pasadena ground site might not reflect the average concentrations over grid cell under consideration ( $4 \times 4$  km). No significant biases have been found when comparing photolysis rates of  $O(^1D)$ ,  $NO_2$ , methylglyoxal and glyoxal against measurements on-board the NOAA WP-3D aircraft (see also Figure S3 in the supplement). There were no major precipitation events during the period of study, hence wet deposition is negligible (and not included in our simulations). We did not consider uptake and possible chemical reactions of glyoxal in cloud droplets.

## 4 Regional variability of glyoxal and its SOA formation

In the following we give a general overview of the regional variability of glyoxal and associated SOA formation over the whole model domain, and focus on two regions with distinct characteristics (Fig. 1): the LA basin region as representative of the densely populated South Coast Air Basin, and the eastern slopes of the Central Valley, as more rural region with high isoprene emissions (Dreyfus et al., 2002).

### 4.1 The photochemical budget of glyoxal

Photochemical source and sink strengths were tracked throughout the model runs and used to derive mean budgets for the two focus regions (Fig. 8): within the LA basin, aromatics (37%), isoprene (28%) and ethyne (35%) contribute with comparable shares to

## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



glyoxal formation. This is in contrast to the findings of Washenfelder et al. (2011), where aromatics and isoprene contribute in total about 87 % with equal shares, with ethyne only 11 %. This difference is likely due to the fact that we account for averaged regional contributions from the whole LA basin, compared to the budget at the Pasadena site itself as reported by Washenfelder et al. (2011). Pasadena is a rather residential neighbourhood with more vegetation (i.e. isoprene) emissions and less strong emitters of aromatics and ethyne than other, more industrial and traffic-intense areas in the LA basin. It is informative to compare this against a source budget over the isoprene-rich eastern slopes of the Central Valley. Here, isoprene gains importance as expected, now being responsible for three quarters of glyoxal formation (74 %).

OH and photolysis sinks both contribute substantially to total photochemical loss of glyoxal, with an equal share over the eastern slopes of the Central Valley, and OH being larger in the LA basin (60 %), due to strong pollutant sources and consequently higher radical concentrations. Washenfelder et al. (2011) find a comparable result in Pasadena, even though the photolysis sink is larger than the reaction with OH in their analysis, consistent with previous studies on local (Volkamer et al., 2007) and on global scales (Myriokefalitakis et al., 2008). Here again we suggest that these differences are likely due to the fact that we are analyzing an average airmass over the LA basin.

When analyzing the vertical variability of sources and sinks, we find a local maximum of both glyoxal production and removal rates between 500 and 1000 m AGL (about the top of the boundary layer) in LA, which is not seen on the eastern slopes of the Central Valley. This is caused by higher concentrations of isoprene in these layers, probably the result of advection from regions with higher vegetation density. The eastern slopes of the Central Valley were explicitly chosen as a region with high (local) isoprene emissions, hence ground level isoprene concentrations are the highest and the maximum aloft is not seen.

## SOA formation from glyoxal in a 3-D model

C. Knote et al.

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[⏪](#)[⏩](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

## 4.2 Environmental parameters affecting SOA formation from glyoxal

The panels in Fig. 9 give an overview of averaged quantities that affect the interactions of glyoxal with aerosols. Regions where the aerosol is usually dry (aerosol water content  $< 0.1 \mu\text{g m}^{-3}$  on average) are masked. As the glyoxal-aerosol interactions most likely require a liquid-phase, no SOA formation occurs there in our simulations, except in the SIMPLE case. The LA basin is the hotspot for both aerosol surface area as well as gas-phase glyoxal. Additionally, the eastern slopes of the Central Valley (our second focus region) show elevated glyoxal concentration due to high availability of its precursor isoprene. Only slightly higher aerosol surface area concentrations are observed there. As expected, highest aerosol water concentrations are found over the oceans, but also several kilometers onshore. Only towards the northern part of the domain does the aerosol have substantial water concentrations further inland. Note the sharp contrast in aerosol water content between the LA basin and its surroundings. The pH of aerosol particles ranges between 1.5 and 5 and is typically around 3.5. Clearly visible is the pH lowering effect of emissions in the sea shipping lanes (e.g. parallel to  $34.5^\circ \text{N}$ ). Aerosol-phase ammonium salt concentrations are in general highest over land due to higher emissions and lower water concentrations. Ammonium sulfate and ammonium nitrate concentrations nonetheless exhibit very different spatial patterns. A stronger background contribution leads to a more dispersed pattern for sulfate concentrations, and we find the highest values over Nevada and the northern end of the Central Valley. Ammonium nitrate concentrations on the other hand are only substantial within the Central Valley and the LA basin. Note that we show salt concentration in the liquid phase of aerosol, which is not equivalent to absolute aerosol mass concentrations. For example, ammonium sulfate concentrations (in air) are comparably low over Nevada, but due to the low aerosol liquid water content in this region we find high values of the concentration in the aerosol water.

This combination of environmental factors makes coastal regions (extending several tens of kilometers from the shoreline) with strong biogenic and anthropogenic

### SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



emissions very susceptible to formation of SOA from glyoxal, as all prerequisites (deliquesced aerosol, high surface area, high inorganic salt concentrations) are fulfilled in these regions. Note that while the surface uptake in HYBRID, FAST and FAST\_PH currently only depends on the availability of a mixed-phase or deliquesced aerosol, both the reversible and ammonium-catalyzed pathway have additional dependencies on the chemical characteristics of the aerosol: (i) The ammonium-catalyzed reaction has an exponential dependence on the activity of the ammonium ion and aerosol pH (Noziere et al., 2009). (ii) The increase in the Henry's law constant for the reversible processes depends exponentially on the total concentration of ammonium sulfate and ammonium nitrate in deliquesced aerosols, with an upper limit at  $12 \text{ mol kg}^{-1}$  (Kampf et al., 2013). A substantial area of the model domain is above this limit on average.

### 4.3 SOA formation from glyoxal

A previous study using a surface uptake parameterization by Washenfelder et al. (2011) concluded that glyoxal would make a small contribution to SOA mass, between  $0\text{--}0.2 \mu\text{g m}^{-3}$  or 0 and 4 %, in California at the time period investigated. It is nonetheless instructive to look at the different total amounts and spatial patterns of SOA formed from glyoxal in the different sensitivity studies (Fig. 10). In the LA basin we find average SOA concentrations between 5 and  $10 \mu\text{g m}^{-3}$ . Depending on the pathways investigated, glyoxal contributes between 1.0 % (VOLUME) and 15 % (SIMPLE) of this SOA concentration. In the Central Valley, SOA mass is typically between 1 and  $4 \mu\text{g m}^{-3}$ , with highest values above the eastern slopes, and contributions from glyoxal between 0.22 % (VOLUME) and 9.4 % (SIMPLE).

#### 4.3.1 Spatial distribution

All simulations show that the LA basin is the hotspot for SOA contribution from glyoxal, due to the high availability of precursors and existing aerosols. Most simulations also show elevated glyoxal SOA concentrations over the eastern slopes of the Central

Valley. In SIMPLE the highest amounts of glyoxal SOA are formed, HYBRID, FAST and FAST\_PH form about half the amounts of SIMPLE, and the VOLUME simulation a factor of 15 less.

In SIMPLE we do not consider aerosol chemical composition, which allows for constant SOA production as long as there is aerosol surface area available. All other simulations require the presence of a liquid-phase in the aerosol, including the surface uptake in HYBRID, FAST and FAST\_PH. The VOLUME simulation shows that all volume process if used as parameterized based on previous studies contribute about 1 % SOA in the LA basin (0.22 % in ESCV). Once we add a surface process (now dependent on a liquid-phase in the aerosol) about 5 % (LA) of SOA is formed from glyoxal. Removing the kinetic limitation of gas-particle partitioning in FAST adds 0.3 % in the LA basin and 0.2 % in ESCV. Shifting the pH in FAST\_PH adds additional 0.1 % of SOA in both regions.

### 4.3.2 Diurnal evolution and vertical distribution

In Fig. 11 we present diurnal cycles and vertical distribution for the relevant parameters affecting glyoxal SOA production efficiency. Figure 12 shows the average diurnal cycle and vertical distribution of different glyoxal SOA reservoirs in all 4 size bins, for all 5 simulations over the LA focus region. Figure 13 shows the same for the eastern slopes of the Central Valley (ESCV) focus region.

In both focus regions we find (Fig. 11) that average particle acidity is decreased at sunrise (increase in average particle pH to 4.0–4.5) which lasts until around noon, when acidity drops back to base levels (pH of 3). We suggest that strong contributions from local, fresh emissions in the morning in both LA and ESCV lead to this observed increase in particle pH. This is accompanied by an increase in salt concentrations and (in LA) decreasing aerosol water content. In ESCV the aerosol water content is very low, and remarkably constant throughout the day. The same holds true for aerosol surface area with average values of 30–50  $\mu\text{m}^2 \text{cm}^{-3}$  in ESCV compared to 100–200  $\mu\text{m}^2 \text{cm}^{-3}$  in LA. There, a diurnal cycle is visible with lower values in the morning and an early

## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



afternoon peak. Vertically, aerosols in the LA basin quickly dry out above 500 m and become almost completely dry above 1000 m. This is in sharp contrast to the findings for ESCV, where aerosol water constant is even increasing with height. Here we also find decreasing pH values and salt concentrations with height.

5 In all simulations we find that (where applicable) the surface uptake and the OH pathway are the main contributors to SOA formed from glyoxal (Figs. 12 and 13). In LA (Fig. 12) the simulations show a distinct noon peak in aerosol mass both from the surface uptake (corresponding to the peak in surface area) and the OH pathway (due to highest OH concentrations at noon, Fig. 6). The picture is different over the eastern slopes of the Central Valley (ESCV, Fig. 13). Here, the peak of glyoxal SOA mass is shifted towards the evening without any of the governing environmental parameters showing a good correlation. This is a strong indication of different origins of glyoxal SOA: locally produced (LA) versus advected (ESCV).

15 Comparison of the different formation pathways reveals that glyoxal SOA formed through the volume pathways alone (VOLUME) is a factor of 10 lower than from a simple surface uptake (SIMPLE), even though the diurnal cycles are similar (Figs. 12 and 13). Within the volume pathways, OH reaction is the dominant source of SOA, with the ammonium pathway contributing about 5 % (10 % in ESCV) and reversible partitioning < 5 % mass.

20 Once we combine a surface process with a volume process (HYBRID), now considering aerosol phase-state in the surface uptake, we form around  $0.12 \mu\text{g m}^{-3} / 0.001 \mu\text{g m}^{-3}$  in LA/ESCV respectively. Around 80 % (90 % in ESCV) of this mass is from the surface uptake. If we assume that gas-particle partitioning of glyoxal has no time dependence (instantaneous equilibration, FAST simulation), then the contribution of volume pathways increases notably, now forming about 40 % of glyoxal in LA and 20 % in ESCV. Still, the OH pathway is dominant. Finally, if we assume that there is no kinetic limitation at high salt concentrations and we pretend to be in an environment with less acidic particles (FAST\_PH) we find that, while the overall contribution of volume pathways to glyoxal SOA stays the same, between 10–20 % of this contribu-

## SOA formation from glyoxal in a 3-D model

C. Knote et al.

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[⏪](#)[⏩](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)



tion is now from the ammonium pathway, and further we can identify 5–10% of mass as oligomers. Simulation chamber data (Kampf et al., 2013) in contrast would have suggested around 40% of SOA existing in reversible forms (monomers, oligomers).

Looking at the vertical distribution we find in the ESCV focus region (Fig. 13) that concentrations almost linearly decrease with altitude, comparable to the vertical evolution of glyoxal turnover rates (Fig. 8). In LA we find that all simulations except HYBRID show a constant or even increasing glyoxal SOA mass between 0 and 750 m. Notably, in FAST and FAST\_PH the relative contribution of volume processes to total glyoxal SOA increases with height in these levels.

The FAST\_PH simulation was a sensitivity study to investigate the partitioning behaviour in regions where the aerosol pH is higher on average (e. g. Mexico City). The expectation that this would strongly increase the contribution of glyoxal to SOA – as the ammonium-catalyzed pathway depends exponentially on pH – was not confirmed by our results. While we see minor increases over FAST, the limiting factor is not the ammonium-catalyzed reaction rate (Eq. 5), but the availability of glyoxal in the particle-phase, governed by reversible partitioning.

In terms of size distribution, the reversible and ammonium-catalyzed pathway put most glyoxal SOA mass into bins 2 (156–625 nm) and 3 (625 nm–2.5  $\mu$ m), with the ammonium-catalyzed pathway favouring bin 3. This is different for the surface uptake, which also adds substantial amount of mass to the smallest size bin (39–156 nm), and adds the majority into bin 2 (156–625 nm).

### 4.3.3 Effects of the aerosol sink on gas-phase glyoxal

To understand the impact of the aerosol sink for glyoxal on its gas-phase concentrations we show averaged absolute concentrations and differences against a simulation without aerosol sink (BASE) in Fig. 14.

In ESCV the effects of all parameterizations tested on gas-phase glyoxal concentrations are negligible except for the SIMPLE case. Here, we find an average decrease of 5 pptv (or 10% of a 50 pptv daily average), with the largest differences during night-

## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion





based on a smog chamber study with model sulfate aerosols and subsequent measurements of gas-phase and aerosol characteristics employing filter based and on-line mass spectrometry techniques. The rate constant of glyoxal + OH has been investigated in dilute solutions e. g. by Buxton et al. (1997), but more complex chemistry has been suggested (e.g. Ervens and Volkamer, 2010). The ammonium-catalyzed pathway rate constant (Noziere et al., 2009) has been derived from a laboratory study in which glyoxal-containing bulk solutions (glass vials) have been analyzed spectrometrically. The surface uptake coefficient finally was derived using an imbalance approach in which ambient measurements of glyoxal are combined with a box-model study (Volkamer et al., 2007). While laboratory studies benefit from a controlled environment and allow for more precise measurements, the transferability of the results to ambient situations can be questioned. The imbalance approach on the other side is based on ambient conditions, but can bear uncertainties that are difficult to constrain. Furthermore, it is not obvious that a discrepancy found between models and data can necessarily be attributed to the loss of glyoxal to aerosols. When we translate these cautionary remarks to the sensitivity studies we conducted, the picture looks as follows: SIMPLE is based solely on imbalance calculations, VOLUME only on laboratory results. HYBRID uses both, as do FAST and FAST\_PH. The latter two simulations are pure sensitivity studies, as there is no laboratory evidence so far to remove the kinetic limitation, and no basis to increase the pH value. Whether a surface uptake is indeed happening or not, and whether it can also work on aerosols without a liquid phase (the SIMPLE case), is to be determined by the measurement community. In case the surface uptake does happen, and it does not require a liquid phase it will be the dominant SOA formation pathway in the period and region investigated here. Only in this case would we agree with previous modeling studies based on simple uptake coefficients, otherwise we conclude that these studies overestimate the amount of SOA formed from glyoxal.

**SOA formation from glyoxal in a 3-D model**

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



## 5 Conclusions

We investigated the formation and regional variability of SOA from glyoxal over the domain of California during the CARES/CalNex measurement campaigns employing the regional chemistry-transport model WRF-chem. We extended several parts of the model to represent in more detailed the glyoxal life cycle, and added several hypothetical mechanisms to form SOA from glyoxal. A very simple parameterization as it has been used in previous (global) modeling studies was compared with a number of more complex mechanisms. We considered surface as well as volume processes, and added reversible formation based on recent laboratory studies.

The LA basin had the highest overall concentrations of SOA from glyoxal of the whole study domain in all simulations. A comparison of the simple surface uptake approach (SIMPLE) and ones considering aerosol phase state, volume and reversible processes (VOLUME, HYBRID) revealed that the simple approach leads to much higher concentrations (on average  $0.5 \mu\text{g m}^{-3}$  or 15 % of total SOA for SIMPLE, compared to  $0.04 \mu\text{g m}^{-3}$  (1 %) for VOLUME and  $0.15 \mu\text{g m}^{-3}$  (5.5 %) for HYBRID). The phase-state constrained surface uptake in the more complex simulation (HYBRID) dominated with minor contributions (< 20 %) from reversible formation and volume pathways. A kinetic limitation caps the increase in the Henry's law constant and reduces the time scales for its increase above a certain salt concentration in the aerosol liquid phase. Once we remove this limitation for the gas-particle partitioning of glyoxal and assume instantaneous equilibration (FAST) the volume pathways contribute around 40 % to glyoxal SOA mass (20 % in ESCV). Further keeping a fast equilibration rate with the oligomer pool at high salt concentration leads to contributions up to 5 % glyoxal SOA mass from oligomers (FAST\_PH). We find a minor further increase in glyoxal SOA formed from the ammonium-catalyzed pathway at the expense of the OH pathway once we artificially increase the aerosol pH handed over to the glyoxal SOA module (FAST\_PH). This increase is not exponential, as was expected from the exponential dependence of the rate constant. This is explained by the fact that, based on our current parameteriza-

### SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures



Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



tion, the volume pathways are always limited by the availability of glyoxal in the liquid-phase, i.e. by the (time-dependent) increase in the Henry's law constant. The effect of the aerosol sink on average gas-phase glyoxal concentrations ranges between 0 and 15 %, with volume reactions (and not surface uptake) being the pathway able to reproduce a noon maximum difference in concentrations as previously found. Unless it can be shown in laboratory studies that there is indeed a surface uptake that forms SOA from glyoxal and that it is independent of phase state and chemical composition we conclude that previous modeling studies based on uptake coefficients overestimated SOA formation from glyoxal.

In this work we investigated the formation of SOA from glyoxal in California during the CARES/CalNex measurement campaigns so that we could constrain glyoxal sources, transformation and sinks with the available measurement data sets. Other studies (e.g. Volkamer et al., 2007; Waxman et al., 2013) indicate that other regions might have substantially larger contributions of SOA from glyoxal. We expect that especially the Eastern US is very susceptible to glyoxal SOA formation due to high relative humidity, high availability of all precursors as well as substantial aerosol concentrations.

**Supplementary material related to this article is available online at <http://www.atmos-chem-phys-discuss.net/13/26699/2013/acpd-13-26699-2013-supplement.pdf>.**

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## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



## SOA formation from glyoxal in a 3-D model

C. Knote et al.

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

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**SOA formation from glyoxal in a 3-D model**

C. Knote et al.

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

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**SOA formation from  
glyoxal in a 3-D  
model**

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



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**SOA formation from glyoxal in a 3-D model**

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



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**SOA formation from glyoxal in a 3-D model**

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



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**SOA formation from glyoxal in a 3-D model**

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

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**SOA formation from  
glyoxal in a 3-D  
model**

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



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SOA formation from  
glyoxal in a 3-D  
model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

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**SOA formation from glyoxal in a 3-D model**

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures



Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



**Table 1.** Chosen parameterization for selected physical processes in WRF.

| Process                  | Method                                |
|--------------------------|---------------------------------------|
| Cloud microphysics       | Morrison double-moment scheme         |
| Radiation                | RRTMG short- and longwave             |
| Land surface             | Noah Land Surface Model               |
| Urban surface            | Urban Canopy Model                    |
| Planetary boundary layer | Mellor-Yamada Nakanishi and Niino 2.5 |
| Cumulus parameterization | Grell 3-D ensemble                    |



**SOA formation from glyoxal in a 3-D model**

C. Knote et al.

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)**Table 2.** Emissions scaling factors ( $E = E_{\text{orig}} \cdot \text{Scaling Factor}$ ), for 2010 relative to 2005/2008.

| Species                     | Scaling Factor |
|-----------------------------|----------------|
| SO <sub>2</sub>             | 0.55           |
| NO, NO <sub>2</sub>         | 0.8            |
| CO                          | 1.2            |
| NH <sub>3</sub>             | 1.0            |
| glyoxal, methylglyoxal      | 1.0            |
| all other gaseous emissions | 0.65           |
| all aerosol emissions       | 1.0            |

**Table 3.** New/updated chemical species in MOZART-4.

| Symbolic name | Atomic composition  | Comments  |
|---------------|---|---|
| C2H2          | C <sub>2</sub> H <sub>2</sub>                                     | ethyne  |
| HCOOH         | HCOOH   | formic acid   |
| HOCH2OO       | HOCH <sub>2</sub> OO  | radical formed in HO <sub>2</sub> addition to formaldehyde              |
| HONO          | HONO  |   |
| TOLUENE       | C <sub>6</sub> H <sub>5</sub> (CH <sub>3</sub> )                  | unlumped, represented sum of benzene + toluene + xylenes in old version |
| BENZENE       | C <sub>6</sub> H <sub>6</sub>                                     |   |
| PHENOL        | C <sub>6</sub> H <sub>5</sub> OH                                  |   |
| BEPOMUC       | C <sub>6</sub> H <sub>6</sub> O <sub>3</sub>                      | Unsaturated epoxide-dialdehyde  |
| BENZO2        | C <sub>6</sub> H <sub>6</sub> (OH)(OO) <sub>2</sub>               | Bicyclic peroxy radical from OH addition to benzene                     |
| PHENO2        | C <sub>6</sub> H <sub>5</sub> (OH) <sub>2</sub> (OO) <sub>2</sub> | Bicyclic peroxy radical from OH addition to phenol                      |
| PHENO         | C <sub>6</sub> H <sub>5</sub> (OH) <sub>2</sub> (OO)O             | Bicyclic oxy radical from OH addition to phenol                         |
| PHENOOH       | C <sub>6</sub> H <sub>5</sub> (OH) <sub>2</sub> (OO)OOH           | Bicyclic hydroperoxide from OH addition to phenol                       |
| C6H5O2        | C <sub>6</sub> H <sub>5</sub> O <sub>2</sub>                      |   |
| C6H5OOH       | C <sub>6</sub> H <sub>5</sub> OOH                                 |   |
| BENZO2OH      | C <sub>6</sub> H <sub>6</sub> (OH)(OO)OOH                         | Bicyclic hydroperoxide from OH addition to benzene                      |
| BIGALD1       | HC(=O)CH=CHCH=O   | Unsaturated dialdehyde  |
| BIGALD2       | CH <sub>3</sub> C(=O)CH=CHCH=O                                    | Unsaturated dicarbonyl  |
| BIGALD3       | HC(=O)C(CH <sub>3</sub> )=CHCH=O                                  | Unsaturated dialdehyde  |
| BIGALD4       | CH <sub>3</sub> C(=O)CH=C(CH <sub>3</sub> )CH=O<br>(and isomers)  | Unsaturated dicarbonyls from xylene oxidation                           |
| MALO2         | HC(=O)CH=CHC(=O)OO  | Acyl radical from "BIGALD1" photolysis                                  |

## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

Table 3. Continued.

| Symbolic name | Atomic composition                          | Comments   |
|---------------|---|--|
| PBZNIT        | $C_6H_5C(O)OONO_2$                          | Peroxybenzoyl nitrate  |
| TEPOMUC       | $C_7H_8O_3$                                 | Unsaturated epoxide-dialdehyde   |
| BZOO          | $C_6H_5CH_2O_2$                             | Peroxy radical formed following OH abstraction from toluene            |
| BZOOH         | $C_6H_5CH_2OOH$                             |  |
| BZALD         | $C_6H_5CHO$                                 | Benzaldehyde   |
| ACBZO2        | $C_6H_5C(O)OO$                              | Acylperoxy radical obtained from benzaldehyde                          |
| DICARBO2      | $CH_3C(=O)CH=C(CH_3)C(=O)OO$<br>and isomers | Acylperoxy radical obtained from photolysis of unsaturated dicarbonyls |
| MDIALO2       | $HC(=O)C(CH_3)=CHC(=O)OO$<br>and isomer     | Acylperoxy radical obtained from photolysis of unsaturated dicarbonyls |
| XYLENES       | $C_6H_4(CH_3)_2$                            | Lumped isomers of xylene   |
| XYLOL         | Isomers of $C_6H_3(CH_3)_2(OH)$             |  |
| XYLOLO2       | Isomers of $C_6H_3(CH_3)_2(OH)_2(OO)_2$     | Bicyclic peroxy radical from OH addition to xylenols                   |
| XYLOLOOH      | Isomers of $C_6H_3(CH_3)_2(OH)_2(OO)OOH$    | Bicyclic hydroperoxide from OH addition to xylenols                    |
| XYLENO2       | Isomers of $C_6H_4(CH_3)_2(OH)(OO)_2$       | Bicyclic peroxy radicals from OH addition to xylenes                   |
| XYLENOOH      | Isomers of $C_6H_3(CH_3)_2(OH)_2(OO)OOH$    | Bicyclic hydroperoxides from OH addition to xylenes                    |



**Table 4.** Continued.

| Reactants      | Products   | Rate Constant(s)   |
|----------------|--|--|
| MALO2 + NO2    | → M  | $k_0 = 8.5E-29 \cdot (300/T)^{6.5}$ ;<br>$k_i = 1.1E-11 \cdot (300/T)$ ; $f = 0.6$ |
| MALO2 + NO     | → .4 GLYOXAL + .4 HO2 + .4 CO  | $7.5E-12 \cdot \exp(290/T)$  |
| MALO2 + HO2    | → .16 GLYOXAL + .16 HO2 + .16 CO   | $4.3E-13 \cdot \exp(1040/T)$   |
| OH + TOLUENE   | → .18 CRESOL + .10 TEPOMUC + .07 BZOO + .65 TOLO2<br>+ .28 HO2                                     | $1.7E-12 \cdot \exp(352/T)$  |
| OH + CRESOL    | → .20 PHENO2 + .73 HO2 + .07 PHENO   | 4.7E-11  |
| BZOO + HO2     | → BZOOH  | $7.5E-13 \cdot \exp(700/T)$  |
| OH + BZOOH     | → BZOO   | $3.8E-12 \cdot \exp(200/T)$  |
| BZOO + NO      | → BZALD + NO2 + HO2  | $2.6E-12 \cdot \exp(365/T)$  |
| OH + BZALD     | → ACBZO2   | $5.9E-12 \cdot \exp(225/T)$  |
| ACBZO2 + NO2   | → PBZNNIT  | $k_0 = 8.5E-29 \cdot (300/T)^{6.5}$ ;<br>$k_i = 1.1E-11 \cdot (300/T)$ ; $f = 0.6$ |
| PBZNNIT        | → ACBZO2 + NO2   | $k(\text{ACBZO2} + \text{NO2})$<br>$\cdot 1.111E28 \cdot \exp(-14\,000/T)$         |
| ACBZO2 + NO    | → C6H5O2 + NO2   | $7.5E-12 \cdot \exp(290/T)$  |
| ACBZO2 + HO2   | → .4 C6H5O2 + .4 OH  | $4.3E-13 \cdot \exp(1040/T)$   |
| TOLO2 + NO     | → NO2 + .6 GLYOXAL + .4 CH3COCHO + HO2 + .2 BIGALD1<br>+ .2 BIGALD2 + .2 BIGALD3                   | $2.6E-12 \cdot \exp(365/T)$  |
| DICARBO2 + HO2 | → .4 OH + .07 HO2 + .07 CH3COCHO + .07 CO + .33 CH3O2  | $4.3E-13 \cdot \exp(1040/T)$   |
| DICARBO2 + NO  | → NO2 + .17 HO2 + .17 CH3COCHO + .17 CO + .83 CH3O2  | $7.5E-12 \cdot \exp(290/T)$  |
| MDIALO2 + HO2  | → .4 OH + .33 HO2 + .07 CH3COCHO + .14 CO + .07 CH3O2 + .07 GLYOXAL                                | $4.3E-13 \cdot \exp(1040/T)$   |
| MDIALO2 + NO   | → NO2 + .83 HO2 + .17 CH3COCHO + .35 CO + .17 CH3O2<br>+ .17 GLYOXAL                               | $7.5E-12 \cdot \exp(290/T)$  |
| DICARBO2 + NO2 | → M  | $k_0 = 8.5E-29 \cdot (300/T)^{6.5}$ ;<br>$k_i = 1.1E-11 \cdot (300/T)$ ; $f = 0.6$ |
| MDIALO2 + NO2  | → M  | $k_0 = 8.5E-29 \cdot (300/T)^{6.5}$ ;<br>$k_i = 1.1E-11 \cdot (300/T)$ ; $f = 0.6$ |
| OH + XYLENES   | → .15 XYLOL + .23 TEPOMUC + .06 BZOO + .56 XYLENO2<br>+ .38 HO2                                    | 1.7E-11  |
| OH + XYLOL     | → .30 XYLOLO2 + .63 HO2 + .07 PHENO  | 8.4E-11  |
| XYLOLO2 + NO   | → HO2 + NO2 + .17 GLYOXAL + .51 CH3COCHO   | $2.6E-12 \cdot \exp(365/T)$  |
| XYLOLO2 + HO2  | → XYLOLOOH   | $7.5E-13 \cdot \exp(700/T)$  |
| OH + XYLOLOOH  | → XYLOLO2  | $3.8E-12 \cdot \exp(200/T)$  |
| XYLENO2 + HO2  | → XYLENOOH   | $7.5E-13 \cdot \exp(700/T)$  |
| OH + XYLENOOH  | → XYLENO2  | $3.8E-12 \cdot \exp(200/T)$  |
| XYLENO2 + NO   | → NO2 + HO2 + .34 GLYOXAL + .54 CH3COCHO<br>+ .06 BIGALD1 + .2 BIGALD2 + .15 BIGALD3 + .21 BIGALD4 | $2.6E-12 \cdot \exp(365/T)$  |

**SOA formation from glyoxal in a 3-D model**

C. Knote et al.

Title Page

Abstract Introduction

Conclusions References

Tables Figures

◀ ▶

◀ ▶

Back Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



## SOA formation from glyoxal in a 3-D model

C. Knote et al.

**Table 5.** New photolysis reactions. ( $J_{\text{HONO}}/J_{\text{NO}_2}$  is the photolysis rate for HONO/ $\text{NO}_2$  calculated by fTUV.)

| Reactant         | Products                            | Rate                          |
|------------------|-------------------------------------|-------------------------------|
| HONO + $h\nu$    | → OH + NO                           | $J_{\text{HONO}}$             |
| BIGALD1 + $h\nu$ | → .6 · MALO2 + HO2                  | $0.140 \cdot J_{\text{NO}_2}$ |
| BEPOMUC + $h\nu$ | → BIGALD1 + 1.5 · HO2 + 1.5 · CO    | $0.100 \cdot J_{\text{NO}_2}$ |
| TEPOMUC + $h\nu$ | → .5 · CH3CO3 + HO2 + 1.5 · CO      | $0.100 \cdot J_{\text{NO}_2}$ |
| BIGALD2 + $h\nu$ | → .6 · HO2 + .6 · DICARBO2          | $0.200 \cdot J_{\text{NO}_2}$ |
| BIGALD3 + $h\nu$ | → .6 · HO2 + .6 · CO + .6 · MDIALO2 | $0.200 \cdot J_{\text{NO}_2}$ |
| BIGALD4 + $h\nu$ | → HO2 + CO + CH3COCHO + CH3CO3      | $0.006 \cdot J_{\text{NO}_2}$ |

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



## SOA formation from glyoxal in a 3-D model

C. Knote et al.

**Table 6.** List of simulations conducted.

|         |  |
|---------|--|
| BASE    | Reference simulation where glyoxal does not interact with aerosols.  |
| RAW     | Like BASE, but using the unmodified emission inventories.  |
| SIMPLE  | Only surface uptake, active independent of aerosol phase state or composition, with $\gamma_{\text{phot}} = 3.3 \times 10^{-3}$ . Comparable to global model simulations (e.g. Fu et al., 2008).                           |
| VOLUME  | Reversible partitioning, OH and ammonium-catalyzed volume reactions.   |
| HYBRID  | Reversible partitioning, OH and ammonium-catalyzed volume reactions, and surface uptake, with $\gamma_{\text{phot}} = 1.0 \times 10^{-3}$ . Also surface uptake considers phase state. Comparable to Waxman et al. (2013). |
| FAST    | Like HYBRID, but glyoxal partitions into the liquid-phase (monomers, pool 1) instantaneously.  |
| FAST_PH | Like HYBRID, but keeping fast time scales for the oligomer pool at high salt concentrations. Further the dependence of SOA formation of the ammonium-catalyzed channel shifted by $\Delta\text{pH} = -2$ .                 |

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

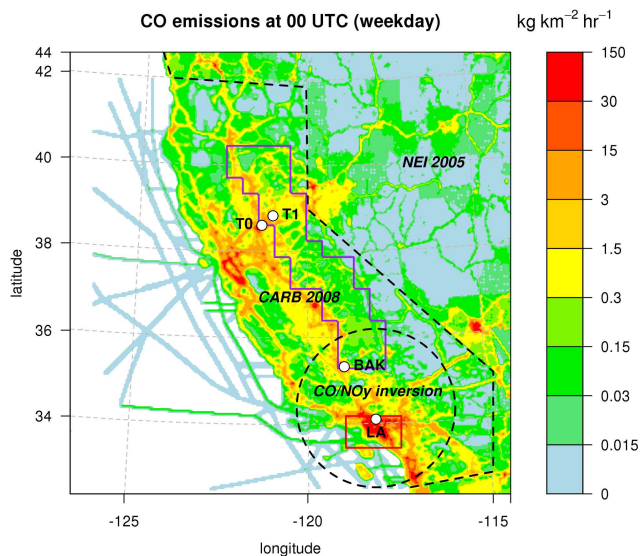
Printer-friendly Version

Interactive Discussion



## SOA formation from glyoxal in a 3-D model

C. Knote et al.

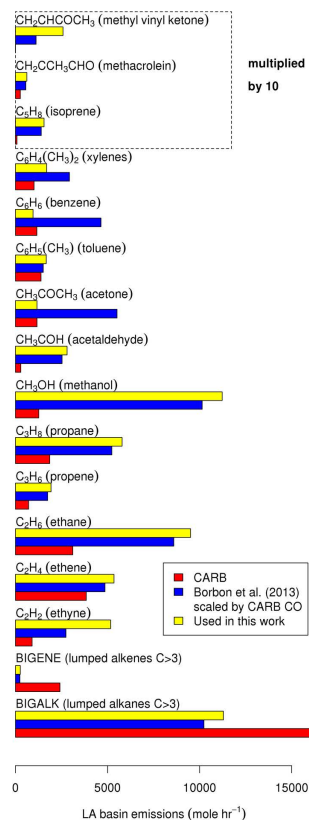


**Fig. 1.** Model domain and schematic illustrating the emissions merging (see text), with CO emissions ( $\text{kg km}^{-2} \text{hr}^{-1}$ ) as visual guideline in the background. Measurement “super-sites” from CARES and CalNex shown: T0 American River College, Sacramento. T1 Northside School, Cool. BAK Cooperative Extension Kern County, Bakersfield. LA California Institute of Technology campus, Pasadena. Two focus regions are marked: the LA basin (red, in the following called “LA”), and the isoprene-rich eastern slopes of the Central Valley (purple, called “ESCV”).



SOA formation from  
glyoxal in a 3-D  
model

C. Knote et al.



**Fig. 2.** VOC emissions in the LA basin (rectangular box from  $33.6^\circ\text{N}$ ,  $119.0^\circ\text{W}$ , to  $34.3^\circ\text{N}$ ,  $117.7^\circ\text{W}$ ) for selected MOZART-4 species as given by the CARB 2008 inventory (red), scaling factors in Borbon et al. (2013) multiplied by CARB 2008 CO emissions (blue), and the final emissions used, a result of the emissions preparation procedure outlined in the text (yellow).

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

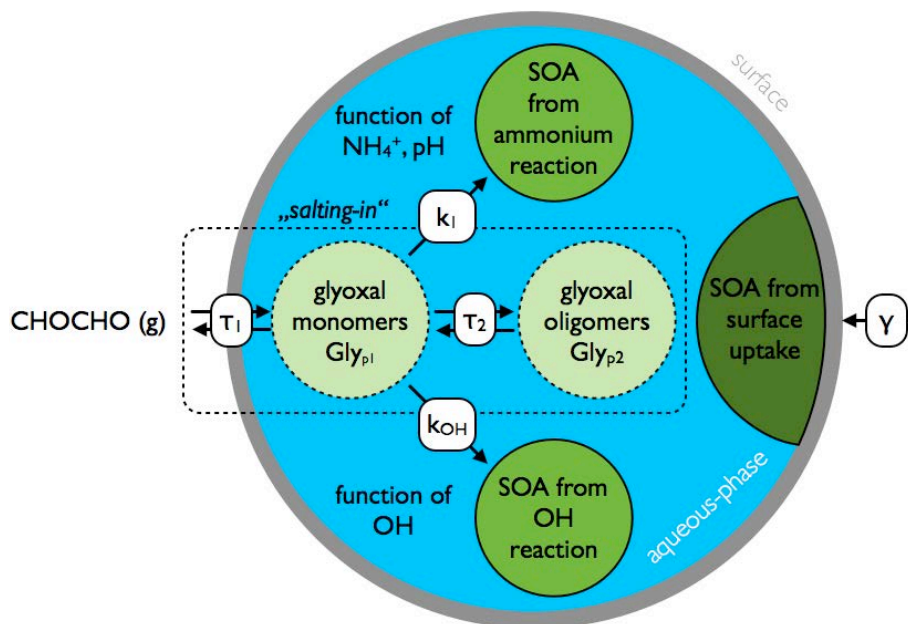
Full Screen / Esc

Printer-friendly Version

Interactive Discussion

## SOA formation from glyoxal in a 3-D model

C. Knote et al.



**Fig. 3.** Schematic of pathways that can lead to SOA formation from glyoxal.  $\gamma_{\text{phot}}$  is the uptake coefficient of the surface reaction.  $k_i$  the rate constant of the ammonium-catalyzed reaction,  $k_{\text{OH}}$  the one for the OH reaction.  $\tau_1$  and  $\tau_2$  are the characteristic time scales to fill the monomer/oligomer pool ( $\text{Gly}_{p1}$  and  $\text{Gly}_{p2}$  respectively, as defined in Kampf et al., 2013). SOA in monomer and oligomer reservoirs is reversible. SOA in surface uptake, and ammonium and OH reaction reservoirs is irreversible.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



SOA formation from  
glyoxal in a 3-D  
model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures



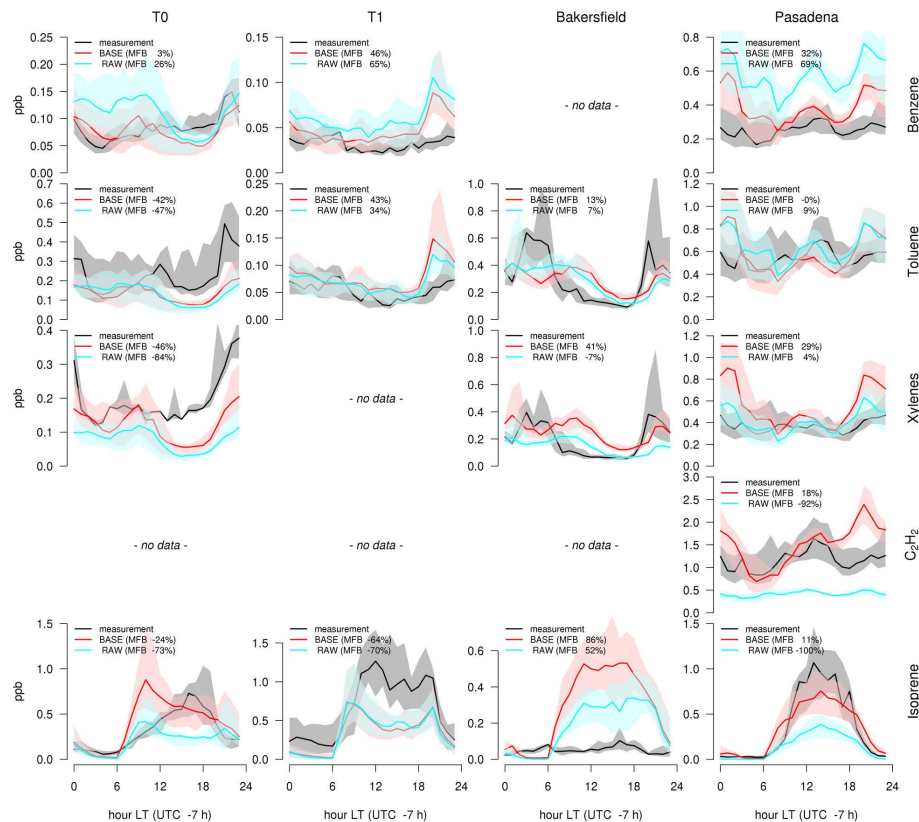
Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



**Fig. 4.** Averaged (29 May 00:00 UTC–16 June 00:00 UTC) diurnal cycles of measurements at the supersites (black) and model results using the original (RAW, blue) and updated emissions inventory (BASE, red) for the main precursors of glyoxal. Shown are the mean as solid line and the 25–75 % range as shaded areas. MFB indicates mean fractional bias against measurements.

## SOA formation from glyoxal in a 3-D model

C. Knote et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures



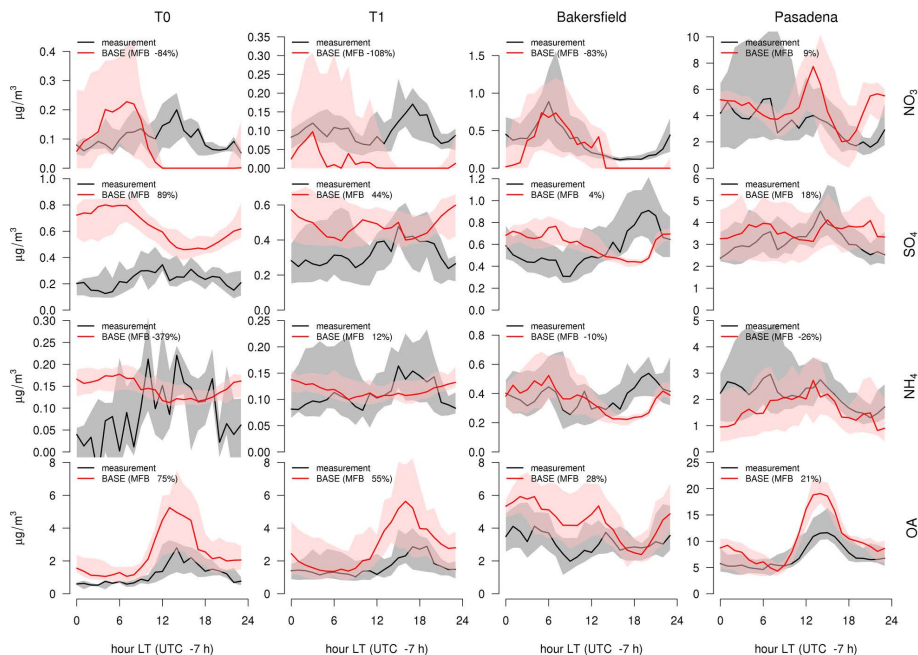
Back

Close

Full Screen / Esc

Printer-friendly Version

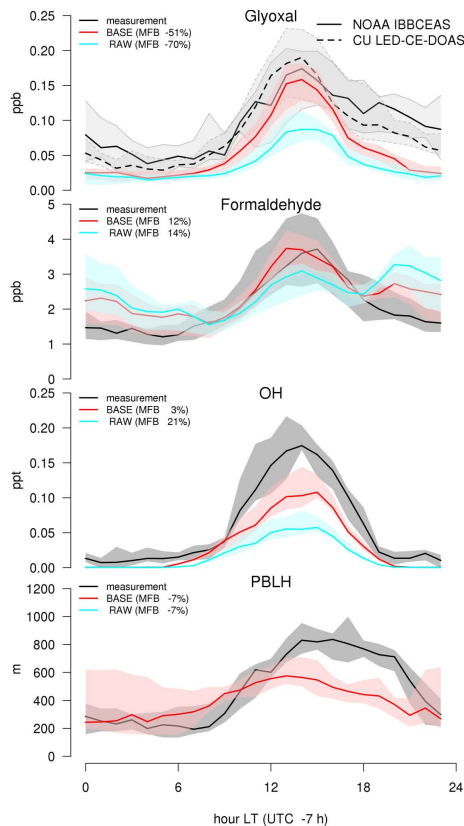
Interactive Discussion



**Fig. 5.** Averaged (29 May 00:00 UTC–16 June 00:00 UTC) diurnal cycles of model results (BASE, red) against aerosol mass spectrometer (AMS) measurements (black) at the four supersites for nitrate, sulfate, ammonium and organic sub-micron non-refractory aerosol mass. MFB indicates mean fractional bias against measurements.

## SOA formation from glyoxal in a 3-D model

C. Knote et al.



**Fig. 6.** Averaged (29 May 00:00 UTC–16 June 00:00 UTC) diurnal cycles of glyoxal, formaldehyde, OH and boundary layer height (PBLH) at the LA ground site with model results using the original (RAW, blue) and updated emissions inventory (BASE, red). Glyoxal has been measured by two instruments at the LA ground site which are shown separately: IBBCEAS (Washenfelder et al., 2008) and CU LED-CE-DOAS (Thalman and Volkamer, 2010).

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

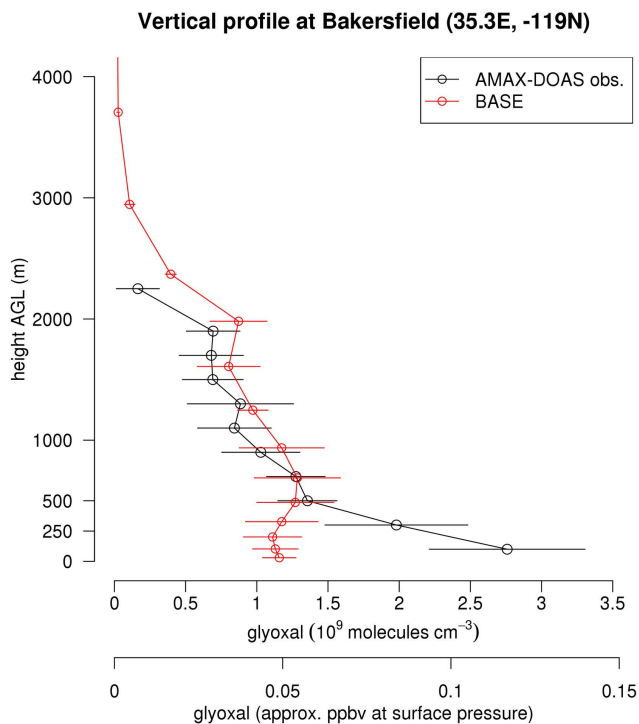
Printer-friendly Version

Interactive Discussion



## SOA formation from glyoxal in a 3-D model

C. Knote et al.

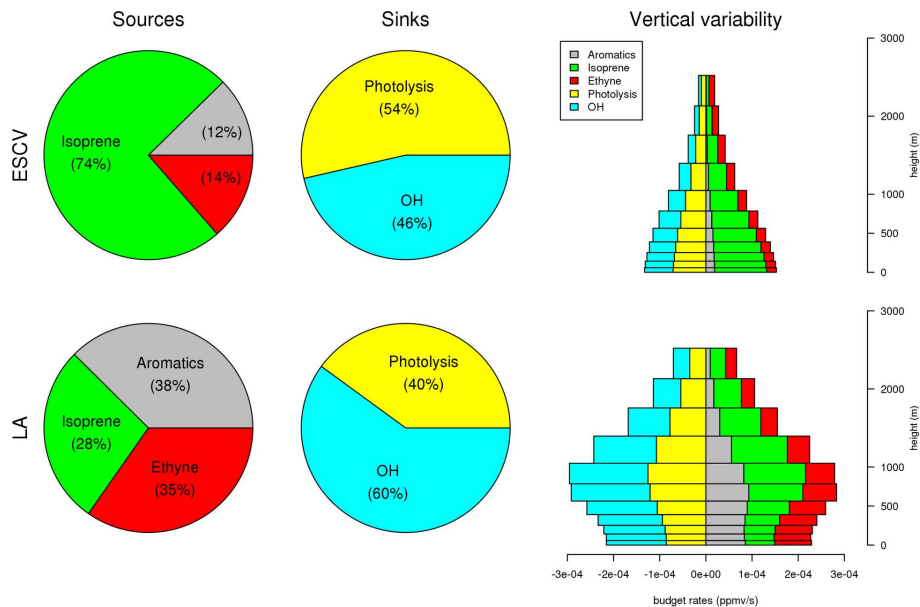


**Fig. 7.** Comparison against vertical profiles of glyoxal as measured by AMAX-DOAS (black) on 15 June (low approach over Bakersfield at about 21:25–21:40 UTC. Ascent and descent averaged  $1\sigma$  level errors). Model (red) error bars are 1 standard deviation of the 3 hourly values (11:00, 12:00, 13:00 LT).

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

SOA formation from glyoxal in a 3-D model

C. Knote et al.



**Fig. 8.** Relative importance of different photochemical sources and sinks of glyoxal in the LA basin and eastern slopes of the Central Valley focus region as time average over the simulation period in the lowest model level (pies), and its vertical variability (barplot).

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

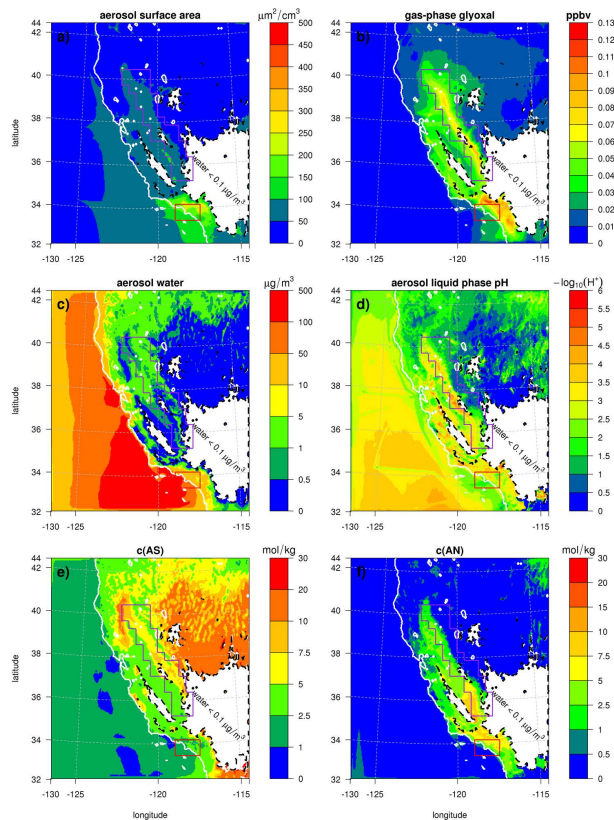
Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



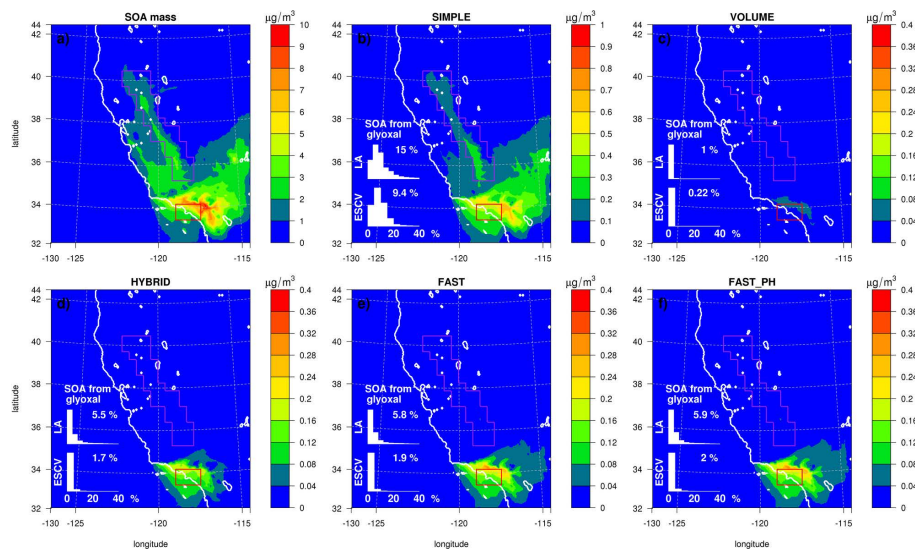


**Fig. 9.** Maps of averaged aerosol related quantities in the lowest model layer (from top-left row-wise): **(a)** aerosol surface area, **(b)** gas-phase glyoxal, **(c)** aerosol water, **(d)** aerosol pH, **(e)** ammonium sulfate and **(f)** ammonium nitrate salt concentrations in aerosol water. White lines delineate land mass. Regions with predominantly dry aerosols (avg. water content  $< 0.1 \mu\text{g m}^{-3}$ ) were masked. Salt concentrations and pH show mean for timesteps where an aqueous aerosol phase existed. Red and purple outline indicate LA and ESCV focus regions respectively.



## SOA formation from glyoxal in a 3-D model

C. Knote et al.



**Fig. 10.** Average total SOA (top left panel) and glyoxal SOA masses (all other panels) in the lowest model vertical layer for the different studies. Note the differing scales. For each study, a histogram of the relative mass contribution of glyoxal to total SOA mass in the LA (red outline) and ESCV (purple) focus regions, as well as the average, is shown in the lower left corner of each panel.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

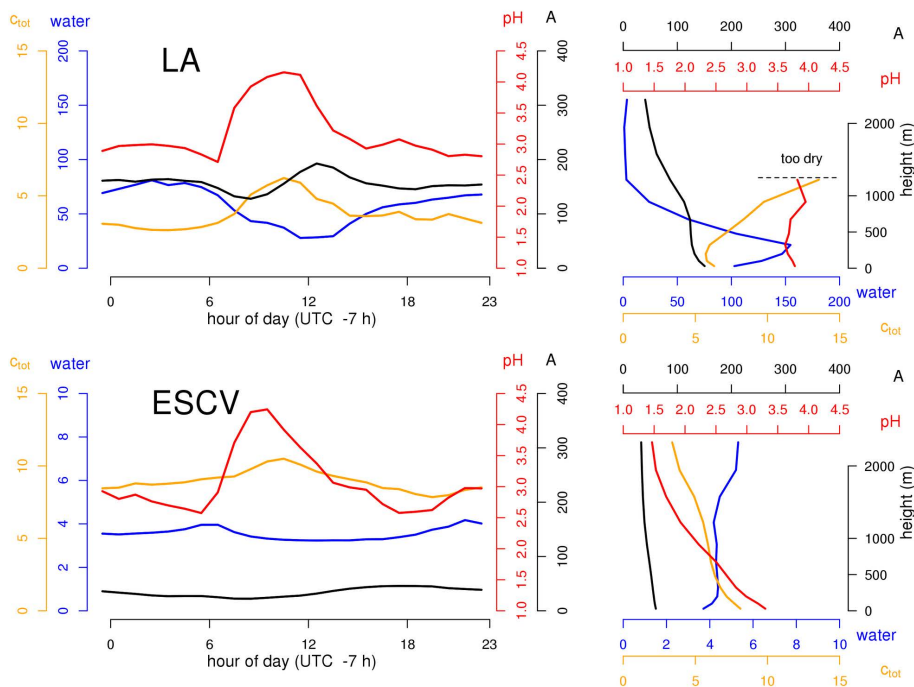
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Interactive Discussion



## SOA formation from glyoxal in a 3-D model

C. Knote et al.



**Fig. 11.** Averaged diurnal cycles (left panels) and vertical evolution (right) of environmental parameters affecting SOA formation from glyoxal in the two different focus regions: salt concentration in deliquesced aerosols  $c_{tot}$  (mol kg<sup>-1</sup>), aerosol water content ( $\mu\text{m}^2 \text{cm}^{-3}$ ), particle acidity (pH), and aerosol surface area  $A$  ( $\mu\text{m}^2 \text{cm}^{-3}$ ). Note different scales for water content between LA and ESCV. Values for pH and  $c_{tot}$  masked above 1200 m in LA due to too few data points for statistically sound averaging.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

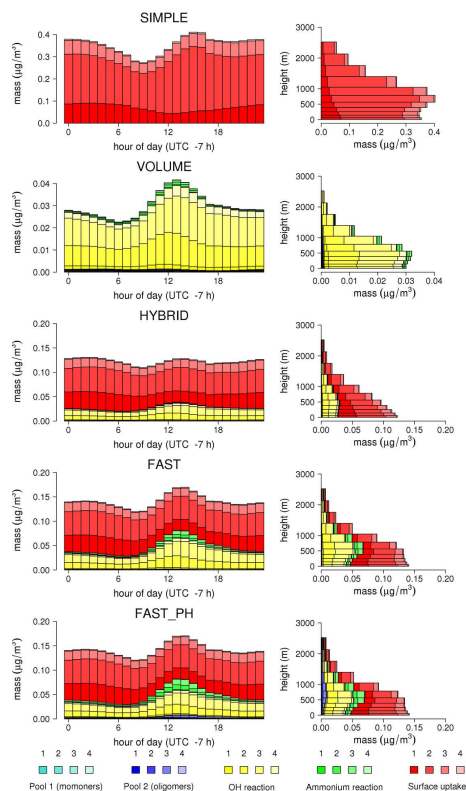
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C. Knote et al.



**Fig. 12.** Contributions of different formation pathways to glyoxal SOA in the LA basin focus region for the different studies given as diurnal cycle in the lowest model layer averaged over the simulation period (left panels), and as average vertical distribution (panels on the right). Size bins start with the smallest (1: 39–156 nm, 2: 156–625 nm, 3: 625 nm–2.5 µm, 4: 2.5–10 µm). Note the different scales for each run.

SOA formation from glyoxal in a 3-D model

C. Knote et al.

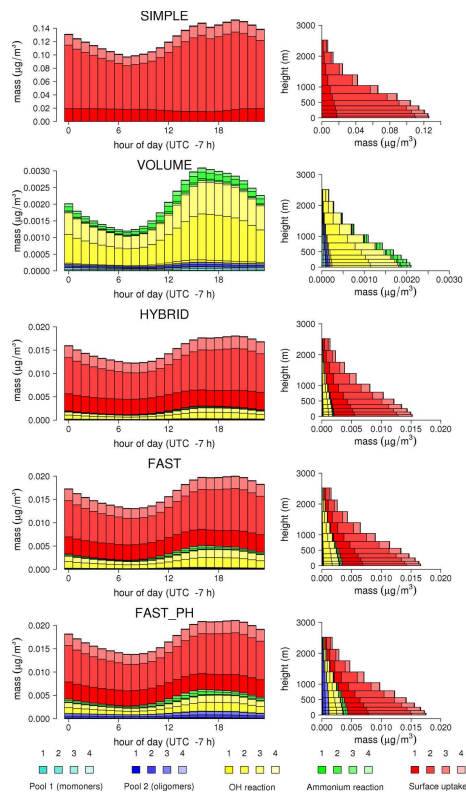


Fig. 13. Like Fig. 12, but for the ESCV focus region. Note the different scales.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

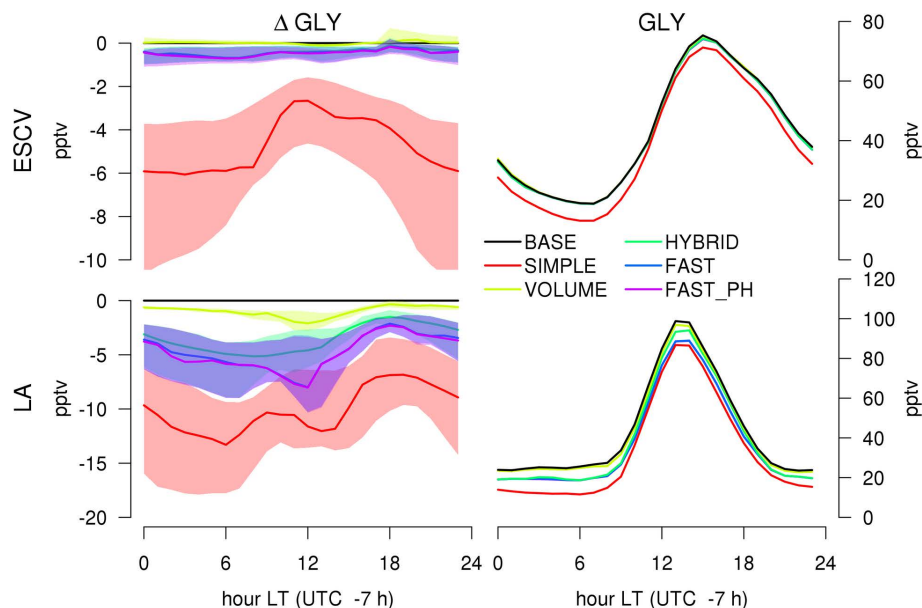
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Interactive Discussion



## SOA formation from glyoxal in a 3-D model

C. Knote et al.



**Fig. 14.** Changes in gas-phase glyoxal due to the different parameterizations of the aerosol sink relative to the BASE simulation. Shown are diurnal cycles for differences relative to the BASE simulation (left plots) and absolute concentrations (right plots) in the two focus regions, averaged over the simulation period. Shaded areas in differences represent the 25–75 % value region.