

Acetaldehyde  
exchange above  
a managed temperate  
mountain grassland

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# Acetaldehyde exchange above a managed temperate mountain grassland

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## Abstract

An overview of acetaldehyde exchange above a managed temperate mountain grassland in Austria over four growing seasons is presented. The meadow acted as a net source of acetaldehyde in all four years, emitting between 7 and 28 mgC m<sup>-2</sup> over the whole growing period. The cutting of the meadow resulted in huge acetaldehyde emission bursts on the day of harvesting or one day later. During undisturbed conditions, both uptake and emission fluxes were recorded. The bidirectional nature of acetaldehyde fluxes was also reflected by clear diurnal cycles during certain time periods, indicating strong deposition processes before the 1st cut and emission towards the end of the growing season.

The analysis of acetaldehyde compensation points revealed a complex relationship between ambient acetaldehyde mixing ratios and respective fluxes, significantly influenced by multiple environmental parameters and variable throughout the year. As a major finding of this study, we identified both a positive and negative correlation between concentration and flux on a daily scale, where soil temperature and soil water content were the most significant factors in determining the direction of the slope. In turn, this bidirectional relationship on a daily scale resulted in compensation points between 0.40 ppbv and 0.54 ppbv, which could be well explained by collected ancillary data. We conclude that in order to model acetaldehyde fluxes at the site in Neustift on a daily scale over longer time periods, it is crucial to know the type of relationship, i.e. the direction of the slope, between mixing ratios and fluxes on a given day.

## 1 Introduction

In recent years, technological progress and new measurement techniques facilitated the in situ measurement of volatile organic compounds (VOC) over longer time periods. The quantification of VOC exchange over different ecosystems and at different temporal and spatial scales contributed towards an improved understanding of the complex

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interactions between VOCs and the atmosphere (Goldstein and Galbally, 2007). The emission or uptake of biogenic VOCs (BVOCs) by plants has many underlying causes, most of which are yet not fully understood.

In the past, most studies have focused on the highly reactive isoprenoids over forest due to their significant role in atmospheric chemistry (Sharkey et al., 2008). In contrast, only few field campaigns targeted the group of biogenic short-chained oxygenated volatile organic compounds (BOVOCs) which are abundant throughout the troposphere (e.g. Singh et al., 2004), e.g. formaldehyde, acetaldehyde, acetone, methanol and ethanol (Seco et al., 2007). Especially over grassland, little is known about the interannual and seasonal variability of BOVOC fluxes.

Acetaldehyde ( $\text{CH}_3\text{CHO}$ ) is a reactive compound with an atmospheric lifetime of a few hours during summer (Atkinson, 2000; Atkinson et al., 2006; Possanzini et al., 2002). Its photo-oxidation contributes to the radical budget in the troposphere (Seinfeld and Pandis, 2006) and pollutants like PAN (Roberts, 1990) and  $\text{O}_3$  are formed in the process. Increased  $\text{O}_3$  levels in the troposphere affect plant growth and human health (Kotzias et al., 1997).

Recent studies have reported bidirectional exchange of acetaldehyde with the atmosphere (Fall, 2003; Graus et al., 2013; Karl et al., 2010), but our current knowledge about acetaldehyde production and consumption in plants is still limited (Jardine et al., 2008).

Secondary photochemical production was described as the major source of atmospheric acetaldehyde on a global scale and as much larger than biogenic emissions, which in turn are the dominant direct terrestrial source of atmospheric acetaldehyde (Millet et al., 2010). Acetaldehyde can also be emitted from decaying plant matter (Greenberg et al., 2012) and as a consequence of photo-degradation of colored dissolved organic matter (Kieber et al., 1990; Zhou and Mopper, 1997). Other terrestrial sources include animals (Rumsey et al., 2012), biomass burning and anthropogenic emissions (Langford et al., 2009; Millet et al., 2010).

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Previous studies described a clear dependence of acetaldehyde emission rates on light and temperature (Cojocariu et al., 2004; Filella et al., 2007; Hayward et al., 2004; Kondo et al., 1998; Ku et al., 2000; Schade and Goldstein, 2002). In a fumigation experiment with trees, Kondo et al. (1998) found a linear relationship between acetaldehyde absorption and transpiration under varying light conditions and a clear increase of acetaldehyde uptake with rising total ambient C<sub>2</sub>-C<sub>5</sub> aldehydes volume mixing ratios (VMR) up to 3000 ppbv. In order to explain this continuous sink, the study suggests a biological removal process mediated by stomata and that trees could act as an important sink for C<sub>2</sub>-C<sub>5</sub> aldehydes. Jardine et al. (2008) pointed out that a chemical reaction of acetaldehyde with the leaf surface is unlikely, since it mainly consists of inert alkanes (waxes). Acetaldehyde deposition to the leaf cuticle has been described for leaves of Amazonian floodplain tree species (Rottenberger et al., 2008). Kreuzwieser et al. (2001) described acetaldehyde as relatively insensitive to stomatal conductance for leaves of trees but described an indirect effect by controlling transpiration rates and therefore the amount of ethanol transported to the leaves from the roots. A lack of stomatal influence and physiological parameters on observed acetaldehyde fluxes was also reported by other investigations (Kesselmeier and Staudt, 1999; Kesselmeier, 2001; Martin et al., 1999). In addition, acetaldehyde has been shown to be released from young seedlings during the first days of germination (Stotzky et al., 1976).

Ethanol, an important precursor for acetaldehyde, is produced in plant roots during anoxic conditions, e.g. caused by soil flooding, through alcoholic fermentation and subsequently transported to the leaves via the transpiration stream. In the leaves and mediated by alcohol dehydrogenase (ADH), ethanol is then oxidized, generating acetaldehyde as an intermediate (Kreuzwieser et al., 1999, 2004). As acetaldehyde itself is subsequently oxidized by the action of aldehyde dehydrogenase, Kreuzwieser et al. (2001) considered the emission of acetaldehyde as a leak between its production and metabolism and affiliated diurnal changes of acetaldehyde emission with the amount of ethanol that was produced and transported to the leaves since the enzymatic activity of ADH remained constant. This pathway may be a significant source

of acetaldehyde on a global scale, for example during the wet season in the amazon rain forest when the roots are flooded (Rottenberger et al., 2008). However, Jardine et al. (2008) recently described that this fermentation process may also be active in leaves under aerobic conditions.

5 The dominant sink of atmospheric acetaldehyde is assumed to be its reaction with OH (Atkinson et al., 2006), and to a lesser degree photolysis (Sander et al., 2006). Studies have shown that plants can also act as a sink for acetaldehyde (e.g. Karl et al., 2005; Rottenberger et al., 2004), for example when it is enzymatically oxidized to acetate and subsequently metabolically consumed (Fall, 2003). Recently, both wet  
10 and dry deposition have been described (Custer and Schade, 2007; Karl et al., 2004; Warneke et al., 2002).

An acetaldehyde compensation point ( $C^*$ ) for leaves has been described previously (Kesselmeier, 2001) and is defined as the ambient concentration at which the net exchange between the ecosystem and the atmosphere is zero. With ambient concentrations below  $C^*$ , acetaldehyde is emitted from the leaves, while the compound is taken up when ambient concentrations are above  $C^*$ . When the production within the leaves dominates,  $C^*$  is high, whereas it is low if consumption processes play a more important role (Jardine et al., 2008).  $C^*$  measurements of acetaldehyde are still rare and have only been conducted in a small number of studies, e.g. for Norway spruce  
15 (Cojocariu et al., 2004) and Amazonian tree species (Rottenberger et al., 2004). However, it seems that physiological factors can influence observed compensation points, like species composition and leaf age (Rottenberger et al., 2005). A previous study showed that environmental variables like rising temperatures may increase the  $C^*$ , implying that production is more sensitive to temperature than the consumption in plants  
20 (Karl et al., 2005).

Recent studies described acetaldehyde exchange patterns varying with height in the plant canopy. Flux measurements within the canopy showed net emission in the upper canopy and net uptake deeper within the canopy over a Norway spruce forest (Müller

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et al., 2006) and in the Amazon (Rottenberger et al., 2004). Cojocariu et al. (2004) reported branch enclosure emission rates by spruce trees increasing with canopy height.

Karl et al. (2005) reported emission at the top of a loblolly pine plantation and uptake in the canopy, where the strongest uptake occurred in regions with the highest LAI. The same study observed highest acetaldehyde concentrations at the top of the canopy, where also highest emission fluxes were measured. In turn, lowest concentrations were reported from the lower part of the canopy, where the highest uptake rates of acetaldehyde were quantified. Assuming a fixed  $C^*$ , this exchange pattern cannot be explained (Jardine et al., 2008). The distinct diurnal cycle of acetaldehyde exchange reported by several studies supports the idea that diurnal variation in temperature and radiation influences  $C^*$  and therefore the flux rate (Cojocariu et al., 2005; Jardine et al., 2008).

The present study focuses on acetaldehyde, one of the three major BOVOCs besides methanol and acetone found during undisturbed conditions above a grassland near Neustift, Austria (Bamberger et al., 2010). To this end we investigated four years of diurnal, seasonal and interannual acetaldehyde exchange rates and elucidated observed flux patterns in relation to biotic and abiotic drivers under in situ conditions at ecosystem scale. The objective of this paper is to compare our findings to previous results at leaf or ecosystem level, to elaborate possible causes for observed differences to other studies and to refine our understanding of long term acetaldehyde fluxes. Based on previous findings we hypothesized that during undisturbed conditions (1) emission rates of acetaldehyde are influenced by light and temperature. As shown in an earlier study for methanol (Hörtnagl et al., 2011), we further hypothesized that (2) the cutting of the meadow results in high acetaldehyde emissions that constitute a major influence on resulting acetaldehyde budgets. In addition, as several studies have described a compensation point for acetaldehyde (Cojocariu et al., 2004; Kesselmeier, 2001; Rottenberger et al., 2004), we assumed that (3) a compensation point can be found over our grassland at ecosystem scale and that (4)  $C^*$  varies over the course of a year due to environmental variables and ecosystem structure and function.

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the field site. Ambient air was analyzed for a number of compounds besides acetaldehyde. In 2008, 13 different  $m/z$  values were recorded repeatedly, resulting in a cycle time of 2.82 s until 10 July, after which it was changed to 3.00 s (15 different  $m/z$  values) until 6 November 2008 and after that to 1.80 s (8  $m/z$  values) until 20 November. In 2009 the cycle time was 2.25 s until 6 April (12  $m/z$  values) and then 2.35 s until 11 December (13  $m/z$  values). In both 2011 and 2012, the measurement cycle lasted 1.61 s, targeting 9  $m/z$  values. The dwell time for acetaldehyde ( $m/z$  45) was 0.2 s in all years. During the last five minutes of every half-hour period the instrumental background was recorded by flushing 500 mL of ambient air through a home-built catalytic converter (heated to 350 °C). The PTR-MS was calibrated once a week in 2008 and every 50 h in 2009/2011/2012 by diluting a multi-component gas standard containing VOCs in ppm volume mixing ratios in N<sub>2</sub> (Apel Riemer Inc., USA) in VOC-free ambient air. Typical calibration factors for acetaldehyde were 15/20/13/13 ncps (normalized counts per second) per ppbv in 2008/2009/2011/2012, respectively. The PTR-MS was operated at a drift tube pressure of 2.15 mbar/2.3 mbar and a drift voltage of 550 V/600 V during 2008/2009–2012, respectively. Data were stored in 30 min files and processed to yield VOC volume mixing ratios in ppbv using a homemade program based on MATLAB 7.4.0 (R2007a, The MathWorks, Inc., USA). More information regarding setup, calibration and operation of the PTR-MS instrument at the site can be found in earlier studies (Bamberger et al., 2010, 2011; Hörtnagl et al., 2011). In 2009 a PTR-TOF and a PTR-MS were simultaneously recording VOC fluxes at the field site in Neustift. During this two month period no eddy covariance flux was found at the exact mass of CO<sub>2</sub>H<sup>+</sup> ( $m/z$  44.998), a potential interfering compound of protonated acetaldehyde (CH<sub>3</sub>HCOH<sup>+</sup>  $m/z$  45.0340) in the PTR-MS. Eddy covariance flux values obtained with PTR-TOF at  $m/z$  45.0340 were in excellent agreement with flux values from the PTR-MS at the nominal mass  $m/z$  45. We can therefore rule out erroneous contributions of CO<sub>2</sub>H<sup>+</sup> to acetaldehyde fluxes measured with PTR-MS at  $m/z$  45.

## 2.4 Flux calculations

Half-hourly fluxes were calculated using ambient air measurements of the first 25 min of each 30 min period due to the zero calibration during the last five minutes. Because of the disjunct nature of the VMRs, the vDEC method (Karl et al., 2002) was applied to calculate acetaldehyde fluxes as the maximum covariance between the turbulent departures of the 20 Hz vertical wind speed and the lower resolved acetaldehyde VMRs. First, a homemade program was used for the time lag search between the two time series and subsequently the post-processing software *EdiRe* (University of Edinburgh) for final flux calculations by using a subsample of the horizontal wind data as given by the sampling rate of the PTR-MS. Means and turbulent departures were calculated by Reynolds (block) averaging. Hörtnagl et al. (2010) showed that the vDEC method yields unbiased flux estimates but is characterized by a larger random uncertainty compared to the true EC.

Among all targeted VOCs the determination of the tubing induced delay time by optimizing the correlation coefficient of the VOC signal with the vertical wind velocity (McMillen, 1988) worked best for methanol, of which the frequency distribution of found lag times showed a peak around 1.5 s in all four years. Lag times for acetaldehyde were searched in a window of  $\pm 3$  s around this peak. If the time delay was outside of the pre-defined window, the acetaldehyde lag time was set to the peak of the methanol distribution (i.e. 1.5 s).

Raw acetaldehyde fluxes were then corrected for high pass (block averaging) and low-pass (lateral sensor separation, dynamic frequency response, scalar and vector path averaging, frequency response mismatch and the attenuation of concentration fluctuations down the sampling tube) filtering according to Moore (1986), Massman (2000) and Aubinet et al. (2000). Frequency-response corrections were based on a site-specific model cospectrum described by Wohlfahrt et al. (2005). Instrumentation, data treatment and quality control of CO<sub>2</sub>, sensible and latent heat fluxes have been described by Wohlfahrt et al. (2008) and Hammerle et al. (2008).

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## 2.5 Quality control

Half-hourly acetaldehyde fluxes were excluded from further analysis if (i) the third rotation angle exceeded  $10^\circ$  (McMillen, 1988), (ii) the stationarity test for acetaldehyde fluxes exceeded 60% (Foken and Wichura, 1996), (iii) the deviation of the integral similarity characteristics was larger than 60% (Foken and Wichura, 1996), (iv) the maximum of the footprint function (Hsieh et al., 2000) was outside the boundaries of the meadow, (v) the measured background signal of acetaldehyde was higher than its ambient concentration (averaged over half an hour) and (vi) the background drift was greater than the sum of the standard deviations of the two adjacent background measurements before and after the flux averaging period. Acetaldehyde VMR data points were flagged as an outlier if the difference between a specific data point and the averaged signal of the respective half-hour was higher than 20 times the theoretical standard deviation (noise). On days influenced by management no outliers were removed, as large fluctuations in acetaldehyde concentrations were found to be physically realistic. Half-hours with more than five outliers were rejected and not used in further analyses.

In total over all four years, 38 558 half-hourly fluxes of acetaldehyde were recorded, of which 27 648 (72%) passed all quality tests and were used in the present study.

## 2.6 Ancillary data

Meteorological measurements included total (PAR) and the fractions of diffuse ( $PAR_{\text{dif}}/PAR$ ) and reflected ( $PAR_{\text{refl}}/PAR$ ) photosynthetically active radiation (BF3H, Delta-T, Cambridge, UK), net radiation ( $R_{\text{net}}$ ; measured by CNR1, Kipp & Zonen, Delft, Netherlands), air temperature ( $T_{\text{air}}$ ) and humidity at 2 m height measured by the means of a ventilated temperature/humidity sensor (RFT-2, UMS, Munich, Germany), soil heat flux (SHF) quantified by means of heat flux plates (3 replicates at 0.05 m depth, corrected for the change in heat storage above that depth; HFP01, Hukseflux, Delft, Netherlands), soil temperature ( $T_{\text{soil}}$ ) at 0.05 m depth (TCAV thermocouple, Campbell

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Scientific, Logan, UT, USA), volumetric soil water content (SWC) (ML2x, Delta-T De-  
vices, Cambridge, UK) and precipitation (52202, R. M. Young, Traverse City, MI, USA).  
All data were collected continuously by a data logger (CR10X, Campbell Scientific,  
Logan, UT, USA). The green plant area index (GAI) was assessed (i) in a destruct-  
5 tive fashion by harvesting the plant matter of square plots (0.09 m<sup>2</sup>, 3–5 replicates)  
and subsequent plant area determination (Li-3100, Li Cor, Lincoln, NE, USA) and (ii)  
from measurements of canopy height which was related to destructively measured GAI  
(Wohlfahrt et al., 2008). Continuous time series of the GAI were derived by fitting appro-  
10 priate empirical functions to measured data separately for each growing phase before  
and after cutting events. A more detailed list of all auxiliary parameters measured at  
this site is given by Wohlfahrt et al. (2008) and Hammerle et al. (2008).

### 2.7 Statistical analyses

Statistical analyses were done using *Statistica 9* (StatSoft, Inc.), *SigmaPlot 11* (Systat  
Software, Inc.) and *Excel 2010* (Microsoft, Inc.). The *partial correlation* in multiple linear  
15 regression analyses gives the correlation between two variables after controlling for the  
effect of all other variables in the equation. The squared *semi-partial correlation* was  
used to express the unique variance accounted for by a specific predictor, relative to  
the total variance of a dependent variable. *Tolerance* is a measure of redundancy and  
defined as 1 minus the squared multiple correlation of a variable with all other indepen-  
20 dent variables in the regression equation. The higher the tolerance value of a predictor,  
the less redundant is its contribution to the regression. To determine significant differ-  
ences between group means in an analysis of variance (ANOVA) setting, the Unequal  
N HSD post hoc test, a modification of the Tukey's HSD test, was used. For statistical  
analyses, only days or half-hours where all parameters were available were included.

## 3 Results

### 3.1 Overview

Figure 1 shows 844 daily average values of acetaldehyde fluxes over four measurement years, with 140 (2008), 249 (2009), 248 (2011) and 207 values (2012) for each single year. Both emission and deposition fluxes could be observed in all four years (Fig. 1). Between 2008 and 2012, the influence of management events, characterized by elevated emission fluxes, could be observed on a total of 37 days, all of which resulted in a daily net emission of acetaldehyde. During undisturbed conditions emission and deposition were recorded on 333 (41 %) and 474 days (59 %), respectively, and 80 days (10 %) showed strong daily average deposition fluxes below  $-0.1 \text{ nmol m}^{-2} \text{ s}^{-1}$ , while 79 days resulted in net emission fluxes that exceeded  $0.1 \text{ nmol m}^{-2} \text{ s}^{-1}$  (Fig. 1). The average flux over all 4 yr during undisturbed conditions was exactly zero, and only slightly higher with management events included ( $0.04 \text{ nmol m}^{-2} \text{ s}^{-1}$ ).

Highest daily average acetaldehyde fluxes in each year were observed on the day of the 2nd cutting of the meadow, reaching emission values between  $1.89\text{--}2.69 \text{ nmol m}^{-2} \text{ s}^{-1}$ . During undisturbed conditions, maximum emissions of  $0.75 \text{ nmol m}^{-2} \text{ s}^{-1}$  were observed in June and August 2008. Highest deposition fluxes of more than  $-0.3 \text{ nmol m}^{-2} \text{ s}^{-1}$  were found in June 2012 ( $-0.37 \text{ nmol m}^{-2} \text{ s}^{-1}$ ) and August 2011 ( $-0.31 \text{ nmol m}^{-2} \text{ s}^{-1}$ ), deposition fluxes of  $-0.45 \text{ nmol m}^{-2} \text{ s}^{-1}$  on 6 December 2009, one week after the start of continuous snow cover, were mainly caused by erratic nighttime fluxes but nevertheless passed all quality control criteria (Fig. 1).

In contrast to emission rates, which reached their maximum on days when the meadow was cut, peak VMRs in each year were generally found 1–2 weeks after the 1st and 2nd cut, but also in November. The highest VMR was recorded in June 2012 (2.3 ppbv). Between 2008 and 2012 and including management, the daily average VMR of acetaldehyde was above 0.5 ppbv on 379 days (45 % of all recorded days).

Highest PAR values were similar in all four years, ranging from 663.7 in 2011 to 688.7  $\mu\text{mol m}^{-2} \text{ s}^{-1}$  in 2008. Yearly average air temperatures of between 6.7 (2012)

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and 7.1 °C (2008, 2011) were close to the 2001–2007 average of 6.7 °C at the same site. The highest daily average temperature was recorded at the end of June 2012 (23.5 °C), the lowest at the beginning of February 2012 (−17.3 °C) (Fig. 1). Half-hourly temperatures peaked at 32.5 °C on 30 June 2012, the lowest value was recorded on 6 February 2012 (−22.4 °C). On days where VOC measurements were performed, the average daily air temperature was between −3.5–23.5 °C. Soil temperature at a depth of 5 cm was naturally dampened compared to air temperature, peaking at 22.1 °C in June 2008 (Fig. 1). During winter months, soil temperatures were typically around zero, with the exception of January 2008 when temperatures fell as low as −2.8 °C on a daily average timescale.

Soil water content exhibited a similar pattern in all four years. Generally, peak values of up to 0.46 m<sup>3</sup> m<sup>−3</sup> were found around snow melt, after which SWC continually decreased to values as low as 0.08 m<sup>3</sup> m<sup>−3</sup> by mid-May (2008, 2011) or start of June (2009, 2012), a decline only temporarily interrupted by rain events. During the second half of the year, water content increased steadily with each precipitation event. Precipitation ranged between 499 (2012) and 648 (2008) mm yr<sup>−1</sup>, well below the 2001–2007 average of 765 mm yr<sup>−1</sup>. During the measurement campaigns, rain was recorded on 77 (2008), 124 (2009), 97 (2011) and 81 (2012) days (Fig. 1).

On average, relative humidity was around 80 %, with the lowest value of 43 % at the end of April 2012. Naturally, vapor pressure deficit was highest during warmer months between April and August with average values around 0.5 kPa and reached peak values of 1.5 kPa on 2 days at the end of May 2008. GAI was close to zero after snowmelt, reached 7.1–7.8 m<sup>2</sup> m<sup>−2</sup> right before the 1st cut and was then reduced to 1.3–2.0 m<sup>2</sup> m<sup>−2</sup> due to harvesting. Maximum values before the 2nd and 3rd cut decreased compared to the 1st cut and were in the range of 6.7–7.1 m<sup>2</sup> m<sup>−2</sup> and 5.1–6.3 m<sup>2</sup> m<sup>−2</sup>, respectively. After the 3rd cut GAI first increased and later decreased.

## 3.2 Inter-annual variability

Figure 2 shows half-hourly fluxes during the four years of measurements. For each year highest fluxes of acetaldehyde were recorded on the day of the 2nd cut or one day later with peak values of 11.0–16.5 nmol m<sup>-2</sup> s<sup>-1</sup>. Including days influenced by management, deposition from the atmosphere to the meadow was observed for 54 % of all half-hourly acetaldehyde fluxes, strong uptake with fluxes below -0.3 nmol m<sup>2</sup> s<sup>-1</sup> for 8 %. During undisturbed conditions, 98 % of all fluxes were between -1 and 1 nmol m<sup>-2</sup> s<sup>-1</sup>.

The meadow acted as a net source of acetaldehyde in all four years (Fig. 3). The largest efflux was recorded in 2008 with an emission of 27.8 mg C m<sup>-2</sup> in 183 days, followed by 2012 (17.5 mg C m<sup>-2</sup> over 239 days). Lower emissions were observed for 2009 (7.4/269) and 2011 (9.9/270). Cumulative carbon emissions on days influenced by management events were substantial (Fig. 3). In 2008, 11.0 mg C m<sup>-2</sup> were emitted over a time period of four days that were influenced by two cutting events. 2009 and 2011 were similar: in both years all 3 cuts and the spreading of the manure were captured by the acetaldehyde measurements, resulting in 12 days influenced by anthropogenic actions that resulted in a total emission of 17.7 (2009) and 17.9 (2011) mg C m<sup>-2</sup>. Highest emissions as a direct consequence of management actions were found in 2012 (20.9 mg C m<sup>-2</sup> over 9 days), although management dates were only partially (3rd cut) or not at all (manure spreading) covered by our measurements (Fig. 3). On a daily average scale, no deposition was observed on management days. When days with management influence were excluded from the analysis, only 2008 acted as a source of acetaldehyde (16.8 mg C m<sup>-2</sup>), while all other years acted as a sink with cumulative deposition fluxes of -10.4 (2009), -8.0 (2011) and -3.5 (2012) mg C m<sup>-2</sup>.

With the exception of 2008, distinct time periods characterized by consecutive days with net acetaldehyde uptake were observed in each year (Fig. 3). For example, starting on 15 April 2009 the meadow acted as a strong sink for acetaldehyde over the next 49 days, divided into two stages distinguished by uptake rates. During the first

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16.4), decreased sharply the following day (1.9–3.1) and subsequently leveled off to reach pre-cut, close-to-zero fluxes by day 5 after the cut. Emissions caused by the 1st or 3rd cutting of the meadow were lower, with maximum emissions of 5.3–14.7 and 4.3–8.4  $\text{nmol m}^{-2} \text{s}^{-1}$  on the day of cutting, respectively, and dropped off more rapidly after the management event. In most years no cutting influence could be seen by day 3 after the cut. One exception to this pattern was the 3rd cut in September 2011, when the cut grass was left on the field for drying and maximum emissions were observed during the turning of the grass in the afternoon one day after the cut. During this event, acetaldehyde continued to exhibit a clear diurnal emission cycle until 5 days after the harvesting (Fig. 4). Often, peak emissions were observed during the turning of the hay or immediately after all grass was collected from the field. For example, the 1st cut in 2011 started at 9 a.m. and maximum emissions during the harvesting were observed about one hour later (2.5  $\text{nmol m}^{-2} \text{s}^{-1}$ ). The hay was left on the field, turned at 12 p.m. (3.5) and collected at 3 p.m. Maximum emissions were then observed at 4 p.m., immediately after all grass was removed from the field (6.8) (Fig. 4). This pattern was even more pronounced in 2012, when maximum emissions during the 1st cut were relatively low (2.2). However, acetaldehyde emissions increased considerably one day later during the turning of the hay at 11.30 a.m. (14.7), then decreased sharply until 1 p.m. (4.3), increased significantly during the collection of the hay at 2 p.m. (12.4) and then immediately dropped off to zero at 3 p.m. (Fig. 4, see also Sect. 4.1).

The spreading of manure was only covered by measurements in 2009 and 2011, where emissions between 1.6–2.7  $\text{nmol m}^{-2} \text{s}^{-1}$  were observed, about one order of magnitude higher than maximum fluxes one day earlier during undisturbed conditions (Fig. 4). In 2011, the spreading of manure on the meadow was split over two days and started on 18 October. On the first day, fluxes increased rapidly to 2.7  $\text{nmol m}^{-2} \text{s}^{-1}$  when the manure was brought out directly in the footprint at 3 p.m. Acetaldehyde fluxes started to exhibit a clear diurnal cycle for several days thereafter, with peak emissions of 2.7  $\text{nmol m}^{-2} \text{s}^{-1}$  at 7 a.m. on 19 October during the thawing of the nightly frost-

cover on plants and manure (Fig. 4). Unfortunately, in 2011 the days directly before fertilization were not covered by BVOC measurements.

High emission fluxes of acetaldehyde were generally accompanied by high volume mixing ratios. Maximum half-hourly VMRs of 4.2 ppbv were registered on the day of the 2nd cut 2012, the same day when also the highest emission flux values were measured (Fig. 4). After cutting events, maximum concentrations of acetaldehyde continued to remain elevated over several days, gradually decreasing to pre-cut values.

### 3.4 Seasonal variability

Figure 5 shows the diurnal course of acetaldehyde fluxes and VMRs before, in-between and after management events in all four investigated years.

The most distinct uptake was observed in 2009 after snowmelt and before the 1st cut, when acetaldehyde deposition to the meadow started early in the morning during a period of increasing VMRs and lasted until midnight, maximum uptake was  $-0.19 \text{ nmol m}^{-2} \text{ s}^{-1}$  around noon. A clear diurnal uptake pattern characterized by two uptake peaks was observed in 2011, when the daily maximum deposition flux of  $-0.15 \text{ nmol m}^{-2} \text{ s}^{-1}$  was recorded in the morning, shortly after the maximum VMR of 0.71 ppb (Fig. 5). After a period of close-to-zero fluxes and low VMRs around noon, uptake of acetaldehyde started again in the afternoon when acetaldehyde concentrations started to increase. The exchange pattern was similar but less distinct in 2012, when local maxima and minima for both fluxes and VMRs were often observed during the same half-hour or within an hour (Fig. 5). On average, flux and VMR patterns between snow melt and the 1st cut of the year exhibited a clear diurnal cycle, whereby highest uptake rates before noon were recorded shortly after the daily maximum acetaldehyde concentration and in the afternoon during rising VMRs.

In the period after the 1st and before the 2nd cut, low uptake rates were constantly recorded during daytime in 2009 and 2011, while exchange patterns during the other two years showed higher variability (Fig. 5). The average 4 yr diurnal cycle shows a ten-

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significant ( $p < 0.05$ ) in 11 of 14 time periods and highly significant ( $p < 0.001$ ) once, before the 1st cut in 2009 during the period of highest acetaldehyde deposition (Table 1, Fig. 5).

Among the collected parameters only two – the net ecosystem CO<sub>2</sub> exchange (NEE) and acetaldehyde VMR – were highly significant during at least one time period, while  $T_{\text{air}}$  was the only parameter that was not significant in any period (Table 1). When all data were pooled, NEE had the highest positive partial correlation (PC) among all parameters in combination with a high tolerance value. Its influence was similar on days with close-to-zero fluxes (group  $f_0$ ), but diminished clearly on days with acetaldehyde uptake ( $f_-$ ) or emission ( $f_+$ ). Over the course of the vegetation period, PCs and tolerance values of NEE increased towards the end of the year (Table 1). The other two ecosystem fluxes, latent evaporation (LE) and sensible heat flux ( $H$ ), were also significant on a 4 yr scale, but their respective PC with acetaldehyde fluxes was considerably lower when compared to NEE.  $H$  was positively correlated with acetaldehyde emission ( $f_0$ ) and significant during time periods in single years whenever NEE was also significant (Table 1). Due to the bidirectional influence of LE and  $H$ , their PCs often changed in sign between years, often resulting in very low PCs when data were pooled for specific time periods. The proportion of unique variance relative to the total variance of acetaldehyde fluxes accounted for by all ecosystem fluxes combined (NEE, LE and  $H$ ) over all 4 yr was 9%.

In contrast to NEE, acetaldehyde VMR was characterized by a highly significant negative and positive PC on uptake and deposition days, respectively, but its PC was very low when all data were pooled and on days with fluxes around zero. During most of the vegetation period and especially at the beginning, partial correlations were found to be positive, but changed in sign towards the end of the measurement campaign after the 3rd cut (Table 1). However, a highly significant positive PC was found in 2009 between the 2nd and 3rd cut, a period of around-zero fluxes (Fig. 5).

Radiation variables showed significant correlations especially at the start of the vegetation period before the 1st cut, and none of the radiation parameters yielded significant

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results after the 3rd cut. Although PAR,  $PAR_{dif}/PAR$  and  $R_{net}$  were significant on a 4 yr scale, they explained only 3 % of the total acetaldehyde flux variance when combined (Table 1).  $PAR_{refl}/PAR$  had little influence when all data were pooled, but had a high and significant negative PC during the period when acetaldehyde deposition fluxes were highest, before the 1st cut in 2009 (Table 1, Fig. 5).

On a 4 yr scale, SWC had a positive PC with acetaldehyde emission and was characterized by high tolerance values, underlining its unique contribution to the regression equation. It also had the highest PC among all parameters in a multiple linear regression, found in a period of low acetaldehyde uptake between the 2nd and 3rd cut in 2012 (Table 1, Fig. 5). The PC between SHF and acetaldehyde VMRs was positive on deposition days ( $f_-$ ) and negative on emission days ( $f_+$ ). Although SHF was not significantly correlated with acetaldehyde exchange over the course of the vegetation period, its PCs were considerably higher between the 2nd and 3rd cut than during other time periods (Table 1).  $T_{soil}$  was the only soil parameter that was not significant on any of the 4 yr scales, but in the time period between snow melt and 1st cut of two consecutive years, when its PC changed from positive to negative. The negative PC found for vapor pressure deficit (VPD) when all data were pooled was very similar to values found throughout the four years. Its strongest effect on acetaldehyde fluxes was observed in 2012 in the period between 2nd and 3rd cut, when uptake strongly increased with VPD.

A simple linear regression analysis resulted in SWC, NEE and  $H$  having highly significant positive correlations, which were very similar in magnitude to their respective PCs (Table 1). No significant correlations were found for all other parameters with the exception of  $PAR_{dif}/PAR$ , which in contrast to its PC was negatively correlated and significant.

### 3.6 Compensation point

As shown before, the correlation between acetaldehyde VMRs and fluxes based on daily average values was not constant, changed over the course of a single year as well as between years and was generally weak or statistically not significant (Table 1).





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ters during different times of the year (Fig. 8). In addition, this approach enabled the calculation of  $C^*$  by inter- or extrapolating the values of the two bins closest to the zero line and looking at the zero-crossing with the  $x$  axis. During different time periods,  $C^*$  was found between 0.05 (in the period snowmelt–1st cut) and 0.19 ppbv (2nd–3rd cut, Fig. 8a). When all data were pooled, the compensation point was 0.13 ppbv (Fig. 8b).

In addition, Fig. 8c investigates the relationship between binned VMR and flux in the four groups previously established in Fig. 7. For days where the relationship between half-hourly values of VMRs and fluxes yielded a negative correlation with  $r^2 > 0.2$  (group N1, Fig. 7b) the compensation point was 0.10 ppbv, and 0.09 ppbv when  $r^2$  was between 0.05 and 0.2 (N2, Fig. 8c). On days with a positive slope  $k$  (Fig. 7b) the relationship between bin averaged VMRs and fluxes was less clear. For high  $r^2$  (P1), emission fluxes increased linearly with VMRs above a concentration of 0.54 ppbv, while acetaldehyde exchange fluctuated around zero at VMRs below (Fig. 8c). In group P2, deposition fluxes were observed above 0.59 ppbv (Fig. 8c). On days with no clear relationship ( $r^2 < 0.05$  group F) between VMR and flux on a half-hourly scale,  $C^*$  was 0.06 ppbv.

Figures 7 and 8 show that the relationship between VMRs and fluxes is complex and that the correlation between the two parameters can be both negative and positive. For the purpose of modeling acetaldehyde fluxes at our study site it is important to better understand the relationship between acetaldehyde concentration and flux, in particular whether a significant correlation between these two parameters on a given day exists, and if it is positive or negative. Therefore, an ANOVA was performed to determine significant environmental differences between the groups established in Fig. 7b. The analysis revealed that only groups N1 and P1 were significantly different from each other, with  $T_{\text{soil}}$  and SWC both being higher in P1 (15.7 °C and 0.29 m<sup>3</sup> m<sup>-3</sup>, respectively) than in N1 (12.1 °C and 0.24 m<sup>3</sup> m<sup>-3</sup>). Daily average values of both parameters and in both groups are illustrated in Fig. 9.

Table 2 examines to what extent measured ancillary data can explain  $C^*$  and slope  $k$ , both resulting from the analysis in Fig. 7b, in forward stepwise regression analyses.

Generally, results for N1 and P1 can be regarded as the most reliable, as both groups are characterized by a relatively high correlation between VMR and flux.

Among the different groups between 66–75 % of the  $C^*$  variance could be explained, whereby acetaldehyde VMR was highly significant and positively correlated with  $C^*$  in N1, N2 and P2, but was not used in P1, where instead VPD emerged as the most important regressor with the highest partial correlation of any parameter in the  $C^*$  regression analysis. While SHF was an important predictor in N1 and not in P1, the opposite was the case for PAR. NEE was the only parameter that was used in the regression equation of both N1 and P1, being positive in the former group and negative in the latter, but not significant in either. Most significant variables were found for N2, where  $T_{\text{air}}$  was positively, but  $T_{\text{soil}}$  negatively correlated with  $C^*$ , a significant difference to P2, where the two PCs changed in sign. Similarly, the PCs of both PAR and NEE were positive in N2, but negative in both P1 and P2. N2 was also the only group where the other two ecosystem fluxes, LE and  $H$ , were significant (Table 2). Four parameters were not significant in any of the groups (SWC,  $\text{PAR}_{\text{dif}}/\text{PAR}$ ,  $\text{PAR}_{\text{refl}}/\text{PAR}$ ,  $R_{\text{net}}$ ).

The regression analysis of slope  $k$  resulted in substantial differences between groups N and P. While the analysis resulted in only one variable being significant in P1 and P2, 9 and 5 parameters were found at  $p < 0.05$  for N1 and N2, respectively, explaining 86 and 35 % of the observed total variance of slope  $k$  between acetaldehyde VMRs and fluxes (Table 2). Similar to the  $C^*$  analysis, VMR was an important predictor in all groups except in P1, where VPD was the most significant parameter and, in contrast to N1, positively correlated with slope  $k$ . Results in N1 and N2 were similar, with SWC and VMR being positively, PAR negatively correlated. The two temperature parameters  $T_{\text{air}}$  and  $T_{\text{soil}}$  were only significant in N1 and had a negative and positive PC with slope  $k$ , respectively. Other significant parameters in N1 with PCs similar to  $T_{\text{soil}}$  were LE,  $H$  and  $\text{PAR}_{\text{refl}}/\text{PAR}$ . NEE and  $\text{PAR}_{\text{dif}}/\text{PAR}$  were significant only in N2, while SHF and  $R_{\text{net}}$  were not significant in any of the groups (Table 2).

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## 4 Discussion

### 4.1 Flux variability

The present study shows that air temperature cannot explain ecosystem acetaldehyde fluxes at the present site. The influence of  $T_{\text{air}}$  on observed flux patterns was statistically insignificant during each of the time periods shown in Table 1 and in addition virtually zero in a simple linear regression. Although PAR yielded significant results during certain time periods, no consistent relationship with acetaldehyde exchange was found. These findings are in contrast to our hypothesis and an important difference to other VOCs (e.g. terpenoids), which often exhibit a distinct temperature- or light-driven emission pattern that is widely used in modeling approaches (Guenther et al., 1995, 2006, 2012).

Soil parameters seemingly influenced observed emission patterns to a larger degree than most other ancillary data (Table 1). The significant influence of SWC on emission fluxes can be explained by increasingly anoxic conditions near the plant roots and as a consequence thereof increased production of ethanol, an important precursor of acetaldehyde (Kreuzwieser et al., 2004). However, the significant positive correlation of SWC with acetaldehyde fluxes in both the simple and multiple linear regression when all data are pooled seemed to contradict results by Asensio et al. (2007), who reported increasing soil uptake of acetaldehyde at higher soil moistures (Table 1).

While it is difficult to find a general explanation for acetaldehyde deposition processes at ecosystem scale, a look at specific, shorter time periods reveals possible driving forces and the seemingly important role of SWC for acetaldehyde transport to the meadow. For example, after the meadow became snow-free on 29 March 2009, environmental conditions changed rapidly over the next 2 weeks.  $T_{\text{soil}}$ ,  $T_{\text{air}}$ ,  $H$ , LE and SHF all increased significantly, but acetaldehyde fluxes were still close to zero. This changed on 15 April when the meadow started to incorporate acetaldehyde, the same time when  $\text{CO}_2$  uptake started and many parameters further increased, most notably  $T_{\text{soil}}$ ,  $T_{\text{air}}$  and LE, while SWC and VMR decreased. Then, during the period of the high-

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rent ethanol measurements might be indispensable for deciphering the reasons behind acetaldehyde exchange patterns at ecosystem scale.

The positive correlation of VMRs with acetaldehyde emissions on uptake days (group  $f_+$ , Table 1) might be misleading, as emissions of acetaldehyde from the meadow also cause ambient VMRs to rise. This observation for acetaldehyde is similar to an earlier study at the same site, where a highly positive PC between methanol VMRs and fluxes, pooled over 2 yr, was reported (Hörtnagl et al., 2011). However, no such relationship could be seen for acetaldehyde when all 4 yr of data were pooled, which resulted in a close-to-zero PC between the two parameters as a direct result of their bidirectional relationship on uptake or emission days in combination with the non-existent influence of VMRs on acetaldehyde exchange on days with very low or zero fluxes (Table 1). The negative correlation of acetaldehyde VMRs on days with net uptake (group  $f_-$ , Table 1) is more in line with studies reporting an acetaldehyde compensation point (e.g. Kesselmeier, 2001) and discussed below (see 4.2).

The amount of carbon emitted from the meadow as a result of the net acetaldehyde exchange in 2009, 2011 and 2012 corresponded to 2.5–4.2% of the carbon emitted due to methanol emissions during the same time period, while the respective value was considerably higher in 2008 (11.2%). One reason for the latter could be the late start of the measurement campaign, where the period between snowmelt and the 1st cut – a period of high uptake rates in all other years – was only partially captured (Figs. 3 and 5, see also 3.2).

A comparison of eddy covariance acetaldehyde fluxes during undisturbed conditions with other studies at ecosystem scale is difficult, as earlier publications over agricultural grassland sites mainly focused on the effect of cutting or other compounds like methanol over shorter time scales (Brunner et al., 2007; Davison et al., 2008; Karl et al., 2001a, 2001b, 2001c; Olofsson et al., 2003). Warneke et al. (2002) reported close-to-zero fluxes right before the cutting of an alfalfa field in August, while Graus et al. (2013, see addendum) reported average leaf level fluxes of  $0.07 \text{ nmol m}^{-2} \text{ s}^{-1}$  for switch grass over a 7 day period in September, both of which are somewhat similar to

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average fluxes of  $0.01 \text{ nmol m}^{-2} \text{ s}^{-1}$  during August and September in this study. Also in September, Custer and Schade (2007) found a median flux of  $-0.01 \text{ nmol m}^{-2} \text{ s}^{-1}$  (interquartile range:  $-0.03$  to  $0.02$ ) over 19 days above ryegrass. These numbers closely resemble results from Neustift, where we observed a median flux of  $-0.02 \text{ nmol m}^{-2} \text{ s}^{-1}$  ( $-0.09$  to  $0.06$ ) in September.

Maximum acetaldehyde fluxes in Neustift were considerably higher than exchange rates reported from various forest ecosystems, where previous studies reported high fluxes of  $0.2\text{--}1.2 \text{ mg m}^{-2} \text{ h}^{-1}$  (Karl et al., 2002, 2003; Rinne et al., 2007), while maximum flux rates in Neustift were  $1.5 \text{ mg m}^{-2} \text{ h}^{-1}$  during undisturbed conditions and  $2.6 \text{ mg m}^{-2} \text{ h}^{-1}$  during cutting. However, the maximum average emission from diurnal cycles (Fig. 5) was only  $0.04 \text{ mg m}^{-2} \text{ h}^{-1}$ , which is much lower than the  $0.6$  and  $0.2 \text{ mg m}^{-2} \text{ h}^{-1}$  reported by Schade and Goldstein (2001) and Kaser et al. (2013), respectively, above a ponderosa pine plantation and lower than the  $0.1 \text{ mg m}^{-2} \text{ h}^{-1}$  over a tropical rain forest (Karl et al., 2004).

Results from measurements including harvesting and drying of harvested plant material compare well to other studies. Davison et al. (2008) reported maximum acetaldehyde emissions of  $19.4 \text{ nmol m}^{-2} \text{ s}^{-1}$  on the day of cutting and a daytime (7 a.m.–5 p.m.) average of  $5.1 \text{ nmol m}^{-2} \text{ s}^{-1}$  over a 3 day drying phase in June for a meadow similar to Neustift near Oensingen, Switzerland. While maximum fluxes during the 1st cut were similar ( $5.3\text{--}14.7 \text{ nmol m}^{-2} \text{ s}^{-1}$ ), daytime averages were generally lower ( $0.6\text{--}1.6 \text{ nmol m}^{-2} \text{ s}^{-1}$ ). Similarly, Davison et al. (2008) described average daytime fluxes of  $8.4 \text{ nmol m}^{-2} \text{ s}^{-1}$  on the day of cutting, much higher than in Neustift ( $0.3\text{--}2.5 \text{ nmol m}^{-2} \text{ s}^{-1}$ ). In Oensingen, average daytime emissions decreased by 53 % one day after the cut, and with the exception of the 1st cutting in 2012, when emissions increased by one order of magnitude, we observed the same decrease in Neustift (54 % on average). Average daytime VMRs in Neustift also compared well, decreasing from  $0.81$  ppbv on the day of cutting to  $0.48$  ppbv 3 days later (Oensingen: from  $1.84$  ppbv to  $0.29$  ppbv).

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an laboratory experiment (Karl et al., 2001b), 3–80  $\mu\text{g}\text{gdw}^{-1}$  for cut clover (*Trifolium repens*; Brilli et al., 2012; De Gouw et al., 2000; Kirstine et al., 1998) and between 6–9  $\mu\text{g}\text{gdw}^{-1}$  for the day of harvesting and two drying days (Warneke et al., 2002). Brilli et al. (2012) also give numbers for *Dactylis glomerata* (1  $\mu\text{g}$  acetaldehyde  $\text{gdw}^{-1}$  over 24 h after cutting) and *Ranunculus acris* (3). In comparison, when the cut grass was left on the field in Neustift, we observed acetaldehyde emissions of 1–19  $\mu\text{g}\text{gdw}^{-1}$  on the day of cutting and 6–33  $\mu\text{g}\text{gdw}^{-1}$  over two days of drying, with maximum fluxes in August. Despite the different plant composition these numbers are similar to the findings of Warneke et al. (2002), who reported 6  $\mu\text{g}\text{gdw}^{-1}$  on the cut day and 15  $\mu\text{g}\text{gdw}^{-1}$  cumulative during cutting and drying. The same study described acetaldehyde VMRs of around 2 ppbv directly before and up to 8 ppbv during the cutting, much higher than in Neustift (around 0.5 and 2.5–4.5 ppbv, respectively).

Acetaldehyde fluxes described by Warneke et al. (2002) were generally very similar to the present study: after close-to-zero fluxes right before the cutting, acetaldehyde emissions increased rapidly and reached a maximum on the day after the harvesting, with a stronger increase during the drying phase than during the cutting itself. Similar emission bursts were observed for methanol at the same site by Hörtnagl et al. (2011), and in the case of acetaldehyde the reason for these massive emissions may be similar, when acetaldehyde in the liquid phase inside the damaged plant cells is suddenly exposed to the atmosphere as a consequence of the wounding. The question if these pools are always present or if acetaldehyde is formed directly as a consequence of the harvesting, for example from pyruvate by an unknown mechanism upon wounding (Loreto et al., 2006), merits further study. Loreto et al. (2006) reported acetaldehyde emissions remote from the wounding site for *Phragmites* leaves on leaf level scale, where emissions from ethanol transported to the leaves seemed unlikely, as no ethanol emissions were associated with the acetaldehyde burst. Remotely triggered emissions due to mechanical stress could also play a role in Neustift, for example after the 1st cut in 2011 when maximum emissions were observed directly after all grass had been

removed from the field and all liquid phase acetaldehyde at the wounding site may already have escaped to the atmosphere (Fig. 4, see also 3.3).

Loreto et al. (2006) further described the absence of acetaldehyde emissions from *Phragmites* leaves when the leaves were immediately placed in water instead of leaving them in air, and further reported the possibility of a strong stomatal control on acetaldehyde emissions because of wounding or high light exposure as well as a delayed response to high temperatures. These observations on leaf level could be one explanation for the low acetaldehyde emissions on the day of the 1st cut in 2012, followed by massive emissions of acetaldehyde on the following two days. On the day of cutting, a light rain contributed to the already wet conditions in the canopy by falling on the cut grass that was still on the field, possibly preventing the majority of acetaldehyde that was solved in the liquid phase from escaping from the wounding site to the atmosphere by keeping them adsorbed on or within the cut grass. On the next day when weather conditions were sunny and dry, emissions increased sharply before the majority of adsorbed acetaldehyde was released in two massive burst during the turning and collection of the hay (Fig. 4). This observation could be similar to earlier studies, where increased acetaldehyde emissions from drying alfalfa after rainfall (Warneke et al., 2002) and from wetted plant material in the laboratory (Warneke et al., 1999) were reported. After all grass was removed from the field, fluxes suddenly dropped to zero, indicating that all previous emissions after the cut were due to the harvesting itself, the temperature-driven drying process or the grass turning. One day after all grass was removed emissions began to rise tenfold as soon as sunlight reached the vegetation in the morning while also surface conductance was high (data not shown). Up to this point only little light had reached the short vegetation below the overlying grass cover for nearly 24 h and light conditions for the lower canopy were already relatively dark before the harvesting due to high GAI and low PAR. If the observation of a possible strong stomatal control on acetaldehyde emissions because of wounding or high light exposure described by Loreto et al. (2006) was to be examined at ecosystem scale, the conditions during this morning would be a good comparison to respective labo-

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ratory experiments. This pattern of high emissions after all grass was removed from the field was observed several times (e.g. 1st cut 2011, see also Sect. 3.3). Generally, these emission bursts of acetaldehyde during and after harvesting events confirm our hypothesis and could influence the lower atmospheric chemistry on a local scale.

Regarding anthropogenic management actions the spreading of liquid manure on the field may constitute another, minor source of acetaldehyde. Earlier studies described the strong impact of fertilization on biological and biochemical processes (Ros et al., 2006). For example, Seewald et al. (2010) reported elevated acetaldehyde emissions from soil samples and a change of microbial community composition after the application of mineral fertilizer in combination with sewage sludge compost. Therefore, increased acetaldehyde emissions during and some days after manure spreading in 2009 and to a lesser extent in 2011 could be a direct effect of increased microbial activity in the soil. The emission spike on 19 October 2011 at 7 a.m. followed by high deposition fluxes 30 min later could be a combination of multiple factors: first, acetaldehyde production in the soil increased during the first part of fertilization on 18 October, but emissions remained low and fluctuated around zero, maybe held back or dampened by starting dew formation later in the evening. Then, one day later, built-up acetaldehyde was released instantly in the morning, the same time when the nightly frost-cover on the meadow thawed and adsorbed acetaldehyde was revolatilized (Seco et al., 2007). This resulted in a huge emission spike (Fig. 4) and high ambient VMRs. Now, with acetaldehyde VMRs in the soil being low, a deposition process started according to Fick's first law of diffusion and acetaldehyde was transported to the meadow, resulting in a net uptake of the compound. However, the exact origin of acetaldehyde fluxes cannot be determined with the eddy covariance method, simultaneous soil measurements of acetaldehyde would definitely expand the room for interpretation of observed exchange patterns.

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## 4.2 Compensation point

Over the course of a year the relationship between acetaldehyde VMRs and fluxes changed constantly. This complex interaction was reflected by varying values for  $C^*$  throughout the year, which is in accordance with our hypothesis. From a very general perspective, our results confirm findings by Karl et al. (2005) who found varying compensation points of loblolly pine needles, with  $C^*$  between 3.7 and 8.5 ppb, which is by more than one order of magnitude higher than values found for the grassland at Neustift (Fig. 7, see 3.6) but similar to other tree studies (Cojocariu et al., 2004). Although we did not find  $C^*$  exponentially increasing like described by Karl et al. (2005), we identified  $T_{\text{air}}$  and  $T_{\text{soil}}$  as significant parameters (Table 2).

The main difference of the present study to other  $C^*$  investigations is the existence of both a positive and a negative relationship between mixing ratios and fluxes, reflecting the complex relationship at ecosystem scale (Fig. 7). Table 2 shows that NEE plays a highly significant role in group N2, meaning a low compensation point in times of high  $\text{CO}_2$  uptake, which would facilitate the deposition of acetaldehyde into the plant and fits flux observations in this study. In the same group increasing  $T_{\text{soil}}$  lead to decreasing  $C^*$  values, which could in part be a consequence of microbial processes in the soil that facilitate acetaldehyde uptake. Due to a high goodness of fit between acetaldehyde VMR and flux, groups N1 and P1 yielded the most reliable results and will be discussed in the following.

Some of our observations are in line with findings by Jardine et al. (2008), who described  $C^*$  as a function of light, temperature and ambient acetaldehyde concentrations at leaf level. However, it seems that at ecosystem scale different parameters influence  $C^*$  during different time periods, strongly depending on whether the correlation between VMR and flux is positive or negative (Table 2). The high correlation between VMR and  $C^*$  in N1 seems to reflect Fick's first law of diffusion. During periods when both  $C^*$  and ambient VMRs are low and mixing ratios start to rise, acetaldehyde starts to diffuse into the plants, i.e. the leaf-internal acetaldehyde levels increase. This in turn causes

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the gradient between internal and ambient acetaldehyde VMRs to decrease, slowing or stopping the diffusion process. For the gradient to build up again, it needs either less internally stored acetaldehyde or higher ambient VMRs. As shown before, the meadow in Neustift only emits minor amounts of acetaldehyde during undisturbed conditions (Fig. 3), therefore an increase of ambient acetaldehyde mixing ratios might be a more likely scenario. At this point, with VMRs further increasing, plants have already taken up considerable amounts of acetaldehyde and leaf-internal acetaldehyde levels are relatively high, reflected by a high  $C^*$ . Therefore, high compensation points at ecosystem scale at the study site in Neustift might be the consequence of preceding periods of acetaldehyde uptake in combination with little or no emission fluxes, which would mean that an increase of  $C^*$  at our meadow could be controlled by external factors as opposed to leaf-internal processes (Jardine et al., 2008).

During periods of low ambient VMRs and low  $C^*$  the canopy might further be more susceptible to uptake of acetaldehyde that was transported to the meadow from external sites. It might be of note that during the strong uptake period before the 1st cut 2009 analyzed in 4.1,  $C^*$  remains about the same (Fig. 7), indicating that leaf-internal acetaldehyde might be consumed by an unknown process. Table 2 further shows that low SHF resulted in lower  $C^*$ , meaning that uptake of acetaldehyde is facilitated. This seems to confirm our findings in Table 1, where decreasing SHF resulted in more acetaldehyde uptake on deposition days.

The negative slope between acetaldehyde mixing ratios and fluxes calculated in Fig. 7 is influenced significantly by multiple parameters in N1 (Table 2), most notably VPD and PAR, whereby the former is positively, the latter negatively correlated with slope  $k$ . During periods of high VPD there may be more LE with plants drawing more water through their roots, which could result in increased ethanol oxidation in the leaves and higher leaf-internal acetaldehyde. This in turn would increase  $C^*$ , resulting in a slower deposition of ambient acetaldehyde to the canopy. Similarly, high PAR is often associated with high  $T_{\text{air}}$ , both of which can increase acetaldehyde emission (Loreto et al., 2006). In such case, increased acetaldehyde production in the plant would lead

to an increased  $C^*$  and in less uptake of ambient acetaldehyde. This internal production might also be fueled by high SWC (more ethanol production) and high LE (more transport of ethanol to the leaves), both of which decreased the steepness of slope  $k$  in N1 (Table 2).

Discussing the positive correlation between acetaldehyde mixing ratios and respective fluxes is challenging (Table 1, Figs. 6, 8), as it seems plausible that high ambient VMRs are found during and as a result of periods of high acetaldehyde emission at or near the study site. Also, environmental parameters were not able to explain slope  $k$  in groups P1 and P2 satisfyingly (Table 2), which is an apparent contrast to groups N1 and N2 and indicates that there might be other factors involved than environmental drivers available in this study.

## 5 Conclusions

Our analysis of acetaldehyde fluxes measured over four growing seasons between 2008–2012 show that in contrast to other VOCs, air temperature and photosynthetic active radiation do not explain observed acetaldehyde exchange patterns at the investigated grassland ecosystem.

The cutting of the meadow resulted in massive bursts of acetaldehyde, either released on the day of harvesting, during drying when the grass was left on the field or immediately after all grass was removed from the meadow. Emission values from this study compared well to laboratory or in situ measurements investigating cutting-related acetaldehyde emissions. Due to management events, the meadow acted as a source of acetaldehyde in all four years, while it was a net sink of acetaldehyde in three of four years with management actions excluded.

In comparison to days with high acetaldehyde emission, high deposition days during undisturbed conditions were characterized by higher  $\text{CO}_2$  uptake rates in combination with lower soil water content and lower sensible heat flux, while acetaldehyde VMRs were similar and higher than on zero-flux days.

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The calculation of acetaldehyde compensation points revealed a complex relationship between ambient acetaldehyde mixing ratios and respective fluxes that changed constantly throughout the year. While the correlation between these two parameters was negative during certain time periods as expected, it was positive during others. It was further shown that slope  $k$  associated with negative correlations is influenced by multiple environmental parameters, each of which increase or slow down deposition from the atmosphere to the canopy. In order to model acetaldehyde fluxes at the site in Neustift it is important to know the relationship between mixing ratios and fluxes on a given day.

Eddy covariance measurements of acetaldehyde at ecosystem scale would benefit greatly when performed in combination with simultaneous soil measurements quantifying VOC exchange, measurements within the canopy to learn more about (VOC) gradients and laboratory experiments targeting ethanol production in plants.

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**Table 1.** Partial correlations (PC) and tolerance (tol) of a multiple linear regression analysis and correlation coefficients ( $r$ ) of a simple linear regression analysis using daily average values of air temperature ( $T_{\text{air}}$ ), soil temperature ( $T_{\text{soil}}$ ) and soil water content (SWC) in 5 cm depth, soil heat flux (SHF), net ecosystem  $\text{CO}_2$  exchange (NEE), latent (LE) and sensible ( $H$ ) heat flux, photosynthetically active radiation (PAR), fraction of diffuse ( $\text{PAR}_{\text{dif}}/\text{PAR}$ ) and reflected ( $\text{PAR}_{\text{refl}}/\text{PAR}$ ) PAR, total net radiation ( $R_{\text{net}}$ ), vapor pressure deficit (VPD) and acetaldehyde volume mixing ratios. Management events were excluded from the analysis. Bold numbers highlight  $p < 0.05$ , except bold underlined numbers resulted in  $p < 0.001$ . all... all fluxes,  $f - \dots$  deposition fluxes  $< -0.03 \text{ nmol m}^{-2} \text{ s}^{-1}$ ,  $f + \dots$  emission fluxes  $> 0.03 \text{ nmol m}^{-2} \text{ s}^{-1}$ ,  $f_0 \dots$  fluxes between  $-0.03$  and  $0.03 \text{ nmol m}^{-2} \text{ s}^{-1}$ .

time period		Multiple Linear Regression																					
		2008–2012		$f_-$		$f_+$		$f_0$		2009		snow melt–1st cut		2008–2012		2008		2009		1st cut–2nd cut		2008–2012	
	all																						
$T_{\text{air}}$	PC	0.07	-0.12	0.15	0.11	0.09	-0.19	0.25	0.06	-0.05	0.13	-0.06	-0.19	-0.05									
	tol	0.04	0.04	0.04	0.03	0.02	0.01	0.03	0.04	0.05	0.03	0.03	0.02	0.06									
$T_{\text{soil}}$	PC	-0.06	0.10	-0.08	-0.09	-0.21	<b>0.29</b>	<b>-0.40</b>	-0.08	0.25	-0.31	0.08	0.07	0.07									
	tol	0.06	0.08	0.06	0.04	0.05	0.02	0.05	0.08	0.17	0.12	0.18	0.08	0.17									
SWC	PC	<b>0.11</b>	0.06	<b>0.19</b>	0.11	0.10	0.16	-0.20	0.01	0.22	0.03	0.15	0.28	<b>0.18</b>									
	tol	0.60	0.57	0.57	0.62	0.18	0.11	0.16	0.26	0.29	0.27	0.24	0.26	0.76									
SHF	PC	-0.05	<b>0.16</b>	-0.16	<b>-0.14</b>	-0.07	0.14	-0.12	0.00	0.07	0.00	-0.01	0.10	0.02									
	tol	0.13	0.11	0.12	0.11	0.16	0.06	0.08	0.19	0.03	0.04	0.04	0.04	0.10									
NEE	PC	<b>0.28</b>	-0.02	0.10	<b>0.20</b>	0.19	<b>0.25</b>	-0.07	0.10	0.18	0.08	-0.23	0.24	0.15									
	tol	<b>0.37</b>	0.30	0.36	0.31	0.17	0.16	0.18	0.21	0.10	0.18	0.13	0.13	0.27									
LE	PC	<b>0.12</b>	0.07	-0.02	-0.09	-0.03	0.14	0.04	0.00	0.19	0.16	0.01	0.19	<b>0.17</b>									
	tol	0.13	0.13	0.13	0.11	0.08	0.09	0.07	0.12	0.13	0.03	0.06	0.06	0.11									
$H$	PC	<b>0.08</b>	-0.01	0.02	<b>0.14</b>	0.11	<b>0.29</b>	0.01	0.12	0.00	0.04	-0.06	0.32	-0.02									
	tol	0.24	0.19	0.23	0.23	0.09	0.23	0.12	0.18	0.17	0.10	0.08	0.13	0.20									
PAR	PC	<b>0.13</b>	-0.03	0.11	-0.03	0.10	0.14	0.15	<b>0.20</b>	0.03	0.06	-0.24	-0.07	0.06									
	tol	0.03	0.02	0.03	0.03	0.02	0.01	0.02	0.03	0.01	0.01	0.01	0.01	0.02									
$\text{PAR}_{\text{dif}}/\text{PAR}$	PC	<b>0.11</b>	-0.05	0.12	-0.03	0.09	0.19	0.03	<b>0.18</b>	0.05	0.06	-0.32	0.09	0.02									
	tol	0.13	0.11	0.14	0.12	0.06	0.04	0.07	0.08	0.05	0.04	0.04	0.03	0.05									
$\text{PAR}_{\text{refl}}/\text{PAR}$	PC	0.02	0.05	0.14	<b>0.18</b>	-0.31	0.07	-0.11	-0.02	-0.10	-0.08	<b>0.36</b>	-0.07	<b>0.21</b>									
	tol	0.67	0.85	0.83	0.42	0.39	0.54	0.54	0.79	0.10	0.18	0.12	0.19	0.47									
$R_{\text{net}}$	PC	<b>-0.08</b>	-0.07	0.01	0.13	-0.07	<b>-0.30</b>	-0.06	<b>-0.18</b>	0.10	-0.09	0.29	0.03	-0.03									
	tol	0.06	0.06	0.04	0.06	0.11	0.04	0.05	0.09	0.04	0.03	0.03	0.04	0.07									
VPD	PC	<b>-0.13</b>	-0.11	-0.12	0.11	-0.01	0.07	<b>-0.28</b>	-0.09	-0.07	-0.16	-0.16	0.13	<b>-0.16</b>									
	tol	0.11	0.09	0.12	0.11	0.08	0.05	0.08	0.08	0.04	0.06	0.03	0.05	0.07									
acetaldehyde VMR	PC	0.02	<b>-0.29</b>	<b>0.38</b>	0.00	-0.11	0.12	0.10	<b>0.15</b>	0.29	0.17	0.09	0.29	0.04									
	tol	0.72	0.65	0.74	0.67	0.30	0.36	0.43	0.70	0.41	0.27	0.19	0.17	0.42									
multiple $r^2$		<b>0.15</b>	<b>0.30</b>	<b>0.21</b>	<b>0.14</b>	<b>0.60</b>	<b>0.31</b>	<b>0.47</b>	<b>0.22</b>	<b>0.59</b>	0.29	<b>0.54</b>	<b>0.64</b>	<b>0.26</b>									
	N	647	255	157	235	60	76	63	213	34	49	45	41	169									

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**Table 1.** Continued.

		Multiple Linear Regression								Simple Linear Regression		
		2008	2009	2nd cut–3rd cut		2008–2012	2008	3rd cut–snow cover		2008–2012	<i>r</i>	N
				2011	2012			2009	2012	2008–2012		
$T_{air}$	PC	0.06	-0.16	-0.39	0.52	0.07	0.08	0.10	-0.17	0.04	<i>r</i>	-0.03
	tol	0.01	0.03	0.01	0.01	0.03	0.01	0.02	0.02	0.03	N	781
$T_{soil}$	PC	0.10	0.22	-0.06	-0.54	-0.10	0.03	-0.11	0.40	-0.08	<i>r</i>	-0.01
	tol	0.03	0.06	0.03	0.02	0.08	0.02	0.02	0.02	0.05	N	781
SWC	PC	-0.20	-0.07	-0.21	<b>0.74</b>	-0.04	0.29	-0.14	0.42	-0.09	<i>r</i>	<b>0.18</b>
	tol	0.29	0.25	0.19	0.16	0.62	0.19	0.09	0.23	0.55	N	<u>777</u>
SHF	PC	-0.26	0.15	0.35	-0.36	-0.14	-0.05	-0.03	0.05	-0.01	<i>r</i>	0.03
	tol	0.03	0.04	0.02	0.01	0.05	0.05	0.06	0.06	0.11	N	777
NEE	PC	-0.09	-0.09	0.39	-0.11	<b>0.33</b>	0.42	<b>0.43</b>	0.04	<b>0.27</b>	<i>r</i>	<b>0.21</b>
	tol	0.07	0.18	0.26	0.06	0.31	0.39	0.69	0.43	0.74	N	<u>676</u>
LE	PC	-0.08	-0.07	0.48	-0.22	0.00	-0.06	0.09	0.33	0.05	<i>r</i>	-0.01
	tol	0.03	0.03	0.02	0.02	0.07	0.09	0.04	0.11	0.15	N	691
$H$	PC	0.08	0.31	-0.07	-0.40	-0.04	-0.29	<b>0.44</b>	-0.27	0.04	<i>r</i>	<b>0.14</b>
	tol	0.08	0.12	0.15	0.08	0.21	0.05	0.12	0.03	0.16	N	<u>720</u>
PAR	PC	0.24	-0.27	-0.25	0.03	<b>0.19</b>	-0.17	-0.20	-0.01	-0.04	<i>r</i>	0.03
	tol	0.01	0.01	0.01	0.01	0.02	0.01	0.02	0.03	0.05	N	781
PAR <sub>diff</sub> /PAR	PC	0.38	-0.33	0.11	-0.15	0.13	-0.16	-0.12	-0.03	-0.06	<i>r</i>	<b>-0.07</b>
	tol	0.02	0.03	0.04	0.02	0.06	0.03	0.13	0.04	0.12	N	<u>777</u>
PAR <sub>refl</sub> /PAR	PC	-0.17	0.23	0.10	0.33	-0.12	0.47	0.12	0.24	0.08	<i>r</i>	0.04
	tol	0.19	0.15	0.14	0.04	0.55	0.04	0.61	0.26	0.74	N	777
$R_{net}$	PC	0.20	-0.09	0.08	0.37	0.03	-0.01	-0.12	0.02	0.04	<i>r</i>	-0.01
	tol	0.02	0.06	0.02	0.01	0.06	0.19	0.12	0.07	0.16	N	777
VPD	PC	0.00	-0.02	-0.23	<b>-0.66</b>	-0.16	-0.06	-0.09	-0.15	0.06	<i>r</i>	-0.05
	tol	0.05	0.07	0.04	0.05	0.10	0.05	0.07	0.05	0.15	N	781
acetaldehyde VMR	PC	0.17	<b>0.69</b>	0.38	0.17	0.06	-0.24	-0.29	0.12	<b>-0.32</b>	<i>r</i>	-0.06
	tol	0.40	0.32	0.18	0.21	0.47	0.12	0.39	0.31	0.61	N	771
multiple $r^2$		<b>0.67</b>	<b>0.64</b>	<b>0.76</b>	<b>0.85</b>	<b>0.21</b>	0.70	<b>0.48</b>	0.49	<b>0.23</b>		
N		32	41	28	24	125	25	51	33	124		



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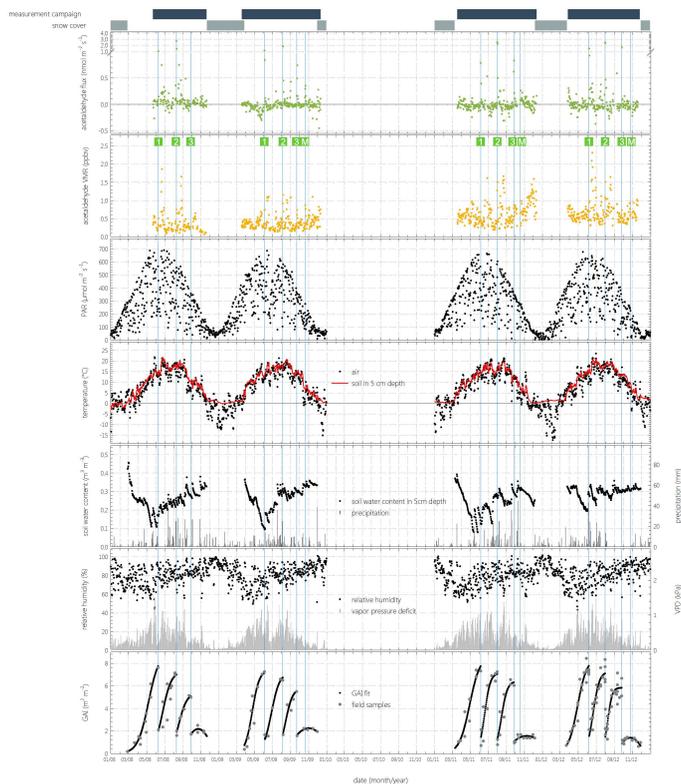
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**Table 2.** Partial correlations of forward stepwise multiple linear regression analyses of the compensation point ( $C^*$ ) and slope  $k$  calculated per day from half-hourly acetaldehyde VMR and flux values, using daily average values of air temperature ( $T_{\text{air}}$ ), soil temperature ( $T_{\text{soil}}$ ) and soil water content (SWC) in 5 cm depth, soil heat flux (SHF), net ecosystem  $\text{CO}_2$  exchange (NEE), latent (LE) and sensible ( $H$ ) heat flux, photosynthetically active radiation (PAR), fraction of diffuse ( $\text{PAR}_{\text{dif}}/\text{PAR}$ ) and reflected ( $\text{PAR}_{\text{refl}}/\text{PAR}$ ) PAR, total net radiation ( $R_{\text{net}}$ ), vapor pressure deficit (VPD) and acetaldehyde volume mixing ratios (VMR). Bold numbers highlight  $p < 0.05$ , except bold underlined numbers resulted in  $p < 0.001$ . Groups represent direction of correlation and goodness of fit between acetaldehyde VMR and flux: N1...negative slope,  $r^2 > 0.2$ , N2...negative slope,  $0.05 < r^2 < 0.2$ , P1...positive slope,  $r^2 > 0.2$ , P2...positive slope,  $0.05 < r^2 < 0.2$ . All management data were excluded from the analysis.

dependent variable group	$C^*$				slope $k$				
	N1	N2	P1	P2	N1	N2	P1	P2	
	negative		positive		negative		positive		
correlation flux vs. VMR	high	low	high	low	high	low	high	low	
$T_{\text{air}}$	-0.26	<b>0.28</b>		-0.26	<b>-0.54</b>				
$T_{\text{soil}}$		<b>-0.30</b>		<b>0.31</b>	<b>0.60</b>		0.29		
SWC				-0.15	<b>0.49</b>	<b>0.31</b>			
SHF	<b>0.42</b>	-0.16		0.06	0.30				
NEE	0.33	<b>0.34</b>	-0.34	<b>-0.37</b>		<b>-0.28</b>			
LE		<b>-0.20</b>			<b>0.49</b>	0.10			
$H$	0.22	<b>0.36</b>			<b>0.56</b>				
PAR		<b>0.25</b>	<b>-0.59</b>	-0.24	<b>-0.78</b>	<b>-0.28</b>		-0.16	
$\text{PAR}_{\text{dif}}/\text{PAR}$		0.15		0.02	<b>-0.39</b>	<b>-0.21</b>			
$\text{PAR}_{\text{refl}}/\text{PAR}$					<b>0.47</b>				
$R_{\text{net}}$		-0.09				0.17		0.21	
VPD			<b>0.84</b>	<b>0.34</b>	<b>0.75</b>		<b>-0.42</b>		
acetaldehyde VMR	<b>0.69</b>	<b>0.73</b>		<b>0.72</b>	<b>0.65</b>	<b>0.38</b>		<b>-0.30</b>	<b>0.66</b>
multiple $r^2$	<b>0.66</b>	<b>0.68</b>	<b>0.75</b>	<b>0.71</b>	<b>0.86</b>	<b>0.35</b>	0.19	<b>0.15</b>	
N	29	127	28	61	29	127	28	61	

## Acetaldehyde exchange above a managed temperate mountain grassland

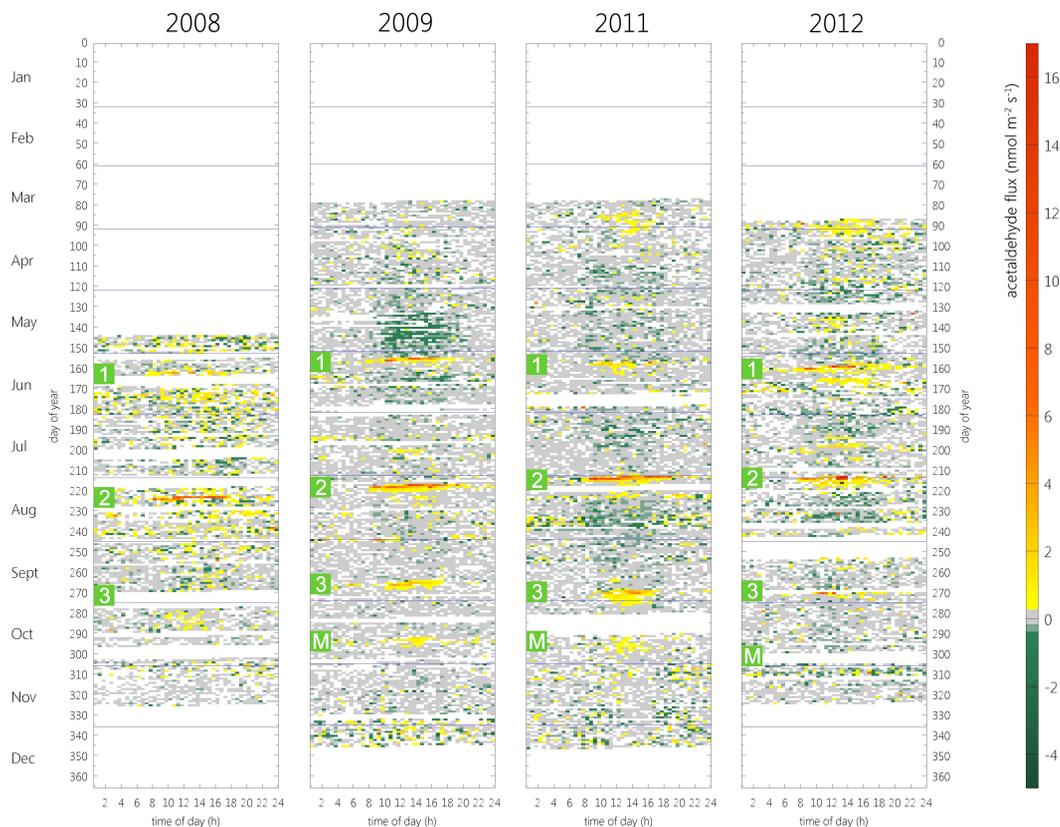
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**Fig. 1.** Daily averages of acetaldehyde flux and volume mixing ratio (VMR), photosynthetically active radiation (PAR), air temperature, soil temperature in 5 cm depth, soil water content in 5 cm depth, relative humidity, vapor pressure deficit, green plant area index (GAI) and daily sums of precipitation over four years of VOC measurements from 2008–2009 and 2011–2012. Vertical lines show management dates, numbers 1, 2 and 3 in green squares mark the 1st, 2nd and 3rd cutting of the meadow, respectively, while M denotes manure spreading.

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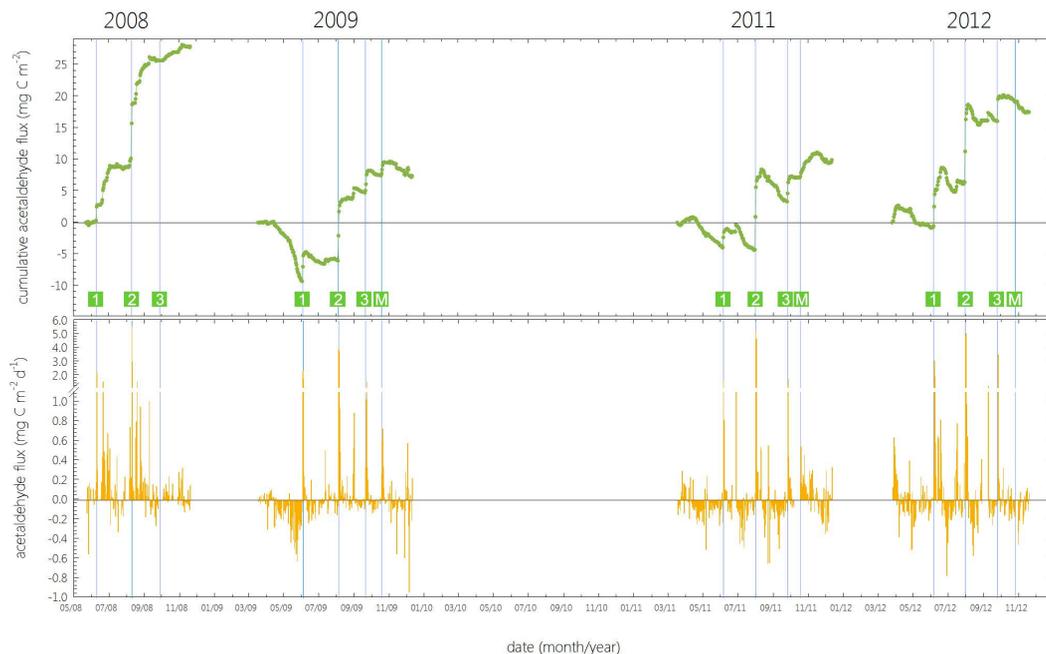
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**Fig. 2.** Half-hourly acetaldehyde fluxes over four years of VOC measurements. Numbers 1, 2 and 3 in green squares mark the 1st, 2nd and 3rd cutting of the meadow, respectively, while M denotes manure spreading. Horizontal blue lines show the start and end of months.

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**Fig. 3.** Cumulative and daily average acetaldehyde fluxes over four years of VOC measurements. Vertical lines show management dates, numbers 1, 2 and 3 in green squares mark the 1st, 2nd and 3rd cutting of the meadow, respectively, while M denotes manure spreading.

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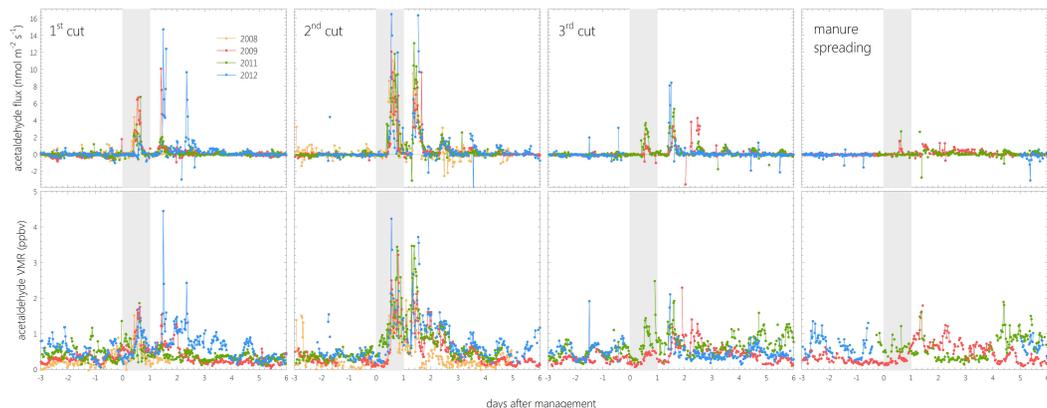
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**Fig. 4.** The influence of management events on acetaldehyde fluxes and volume mixing ratios (VMR) during four years of VOC measurements, the filled rectangle marks the day of the management action.

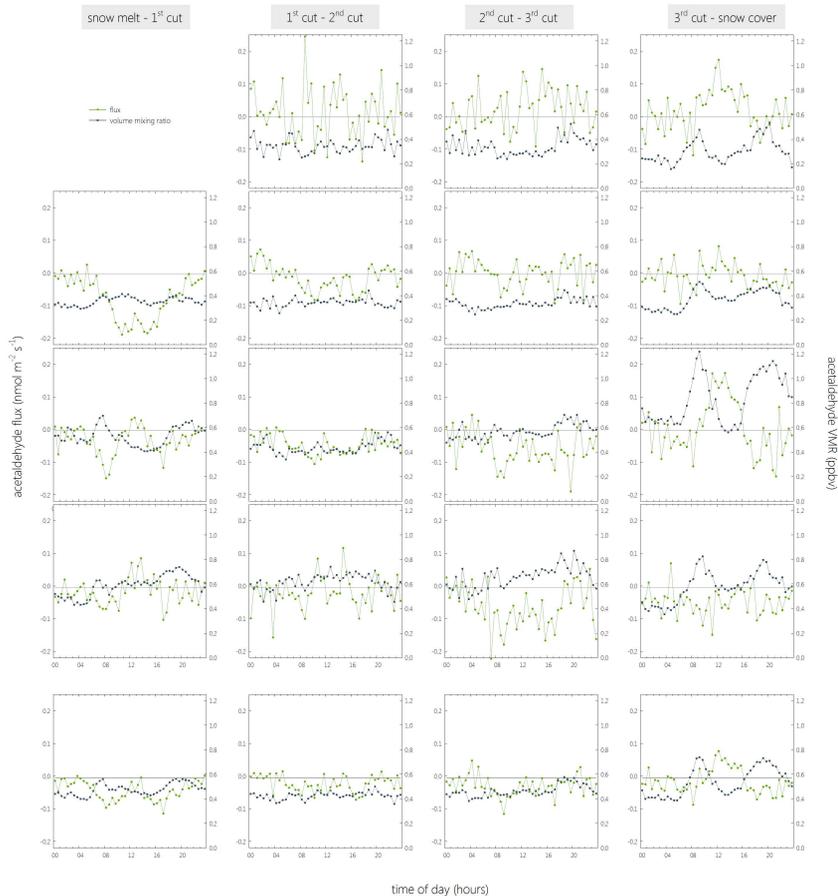
2008

2009

2011

2012

all 4 years



**Fig. 5.** Diurnal cycles of acetaldehyde fluxes and volume mixing ratios (VMR) during different time periods over four years of VOC measurements. Management data were excluded from the analysis.

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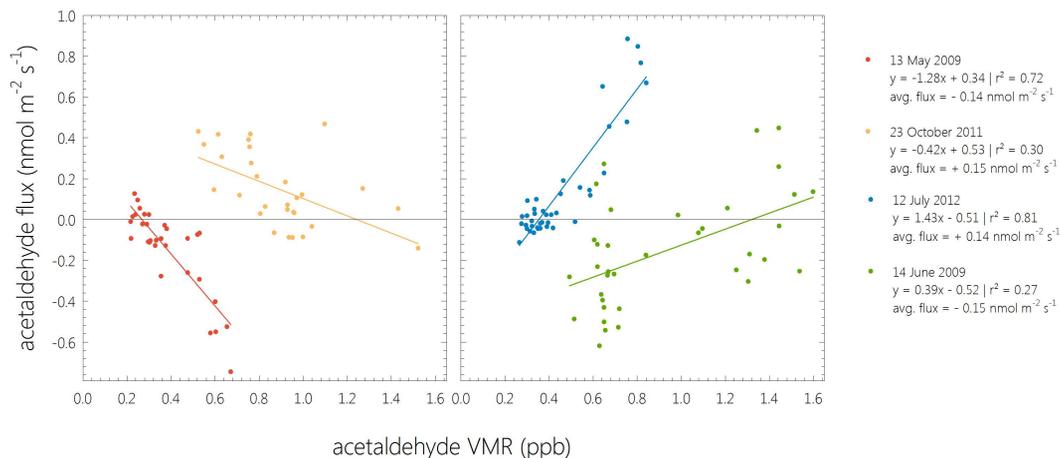
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**Fig. 6.** Four days with different relationships between half-hourly acetaldehyde volume mixing ratio (VMR) and flux including linear regression lines, showing negative and positive correlation between the two parameters on days with net emission and net deposition of acetaldehyde.

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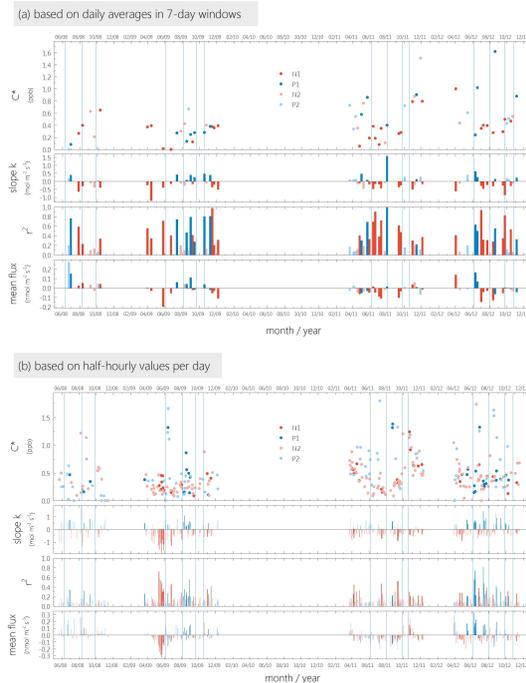
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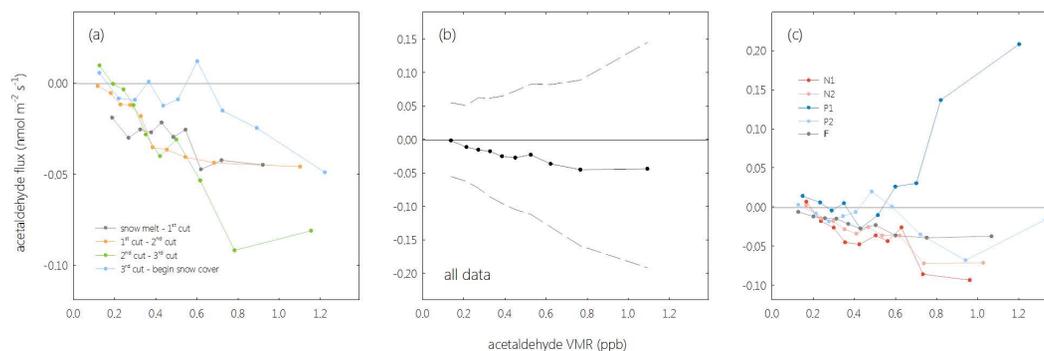
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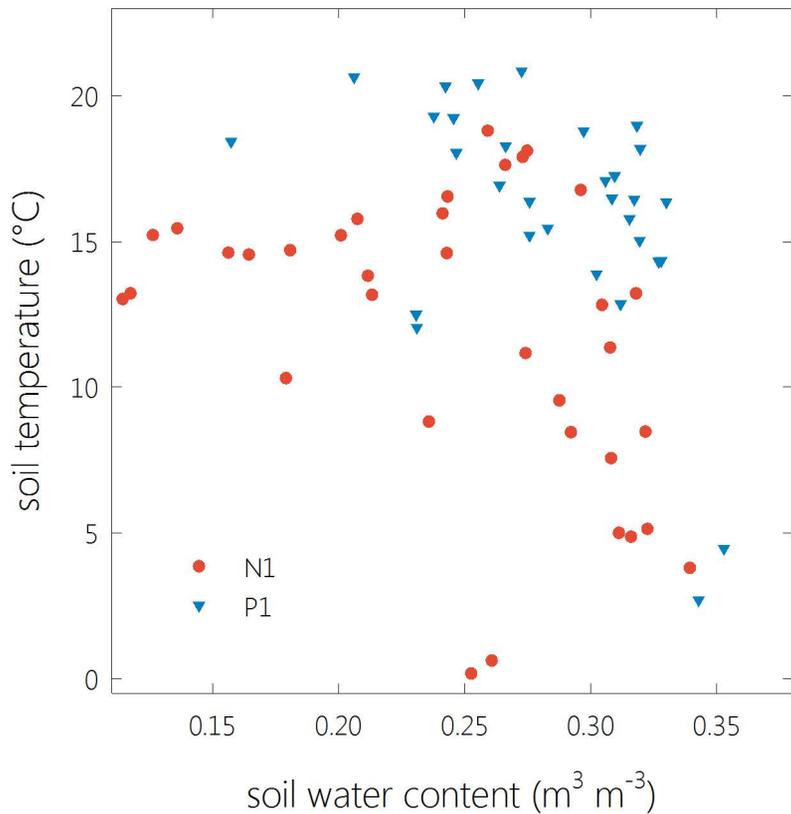
**Fig. 7.** Results of a simple linear regression between acetaldehyde VMR as the independent variable and flux as the dependent parameter, quantifying the compensation point ( $C^*$ ), slope  $k$ ,  $r^2$  and the respective acetaldehyde flux on two different time scales. Panel (a) shows  $C^*$  calculated from daily average values in 7 day windows when a minimum of four days was present in each window. Panel (b) illustrates  $C^*$  and corresponding parameters calculated from half-hourly values per day. Colors represent direction of correlation and goodness of fit between acetaldehyde VMRs and fluxes: N1... negative slope,  $r^2 > 0.2$ , N2... negative slope,  $0.05 < r^2 < 0.2$ , P1... positive slope,  $r^2 > 0.2$ , P2... positive slope,  $0.05 < r^2 < 0.2$ . All management data were excluded from the analysis. Vertical lines show management dates.

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**Fig. 8.** Bin averages of half-hourly acetaldehyde volume mixing ratio (VMR) and flux **(a)** during different time periods and over all four years (551–725 values per bin), **(b)** using all available data from all four years (2542 values per bin) and **(c)** in groups defined by the direction of correlation and goodness of fit between half-hourly acetaldehyde VMRs and fluxes: N1... negative slope,  $r^2 > 0.2$ , N2... negative slope,  $0.05 < r^2 < 0.2$ , P1... positive slope,  $r^2 > 0.2$ , P2... positive slope,  $0.05 < r^2 < 0.2$ , F... zero slope,  $r^2 < 0.05$ . All management data were excluded from the analysis.



**Fig. 9.** Daily average values of soil temperature ( $T_{\text{soil}}$ ) and soil water content (SWC) in two groups defined by the direction of correlation and goodness of fit between half-hourly acetaldehyde VMRs and fluxes, where N1... negative slope,  $r^2 > 0.2$ , P1... positive slope,  $r^2 > 0.2$ . All management data were excluded from the analysis.