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Glyoxal and methylglyoxal in Atlantic seawater and marine aerosol particles: method development and first application during the Polarstern cruise ANT XXVII/4

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Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

Abstract

An analytical method for the determination of the alpha dicarbonyls glyoxal (GLY) and methylglyoxal (MGLY) from seawater and marine aerosol samples is presented. The method is based on derivatisation with *o*-(2,3,4,5,6-Pentafluorobenzyl)-hydroxylamine (PFBHA) reagent, solvent extraction and GC-MS (SIM) analysis. The method showed good precision (RSD < 10 %), sensitivity (detection limits in the low ng L⁻¹ range), and accuracy (good agreement between external calibration and standard addition). The method was applied to determine GLY and MGLY in oceanic water sampled during the POLARSTERN cruise ANT XXVII/4 from Capetown to Bremerhaven in spring 2011. GLY and MGLY were determined in the sea surface microlayer (SML) of the ocean and corresponding bulkwater (BW) with average concentrations of 228 ng L⁻¹ (GLY) and 196 ng L⁻¹ (MGLY). The results show a significant enrichment (factor of 4) of GLY and MGLY in the SML. Furthermore, marine aerosol particles (PM₁) were sampled during the cruise and analyzed for GLY (average concentration 0.19 ng m⁻³) and MGLY (average concentration 0.15 ng m⁻³). On aerosol particles, both carbonyls show a very good correlation with oxalate, supporting the idea of a secondary formation of oxalic acid via GLY and MGLY. Concentrations of GLY and MGLY in seawater and on aerosol particles were correlated to environmental parameters such as global radiation, temperature, distance to the coastline and biological activity. There are slight hints for a photochemical production of GLY and MGLY in the SML (significant enrichment in the SML, higher enrichment at higher temperature). However, a clear connection of GLY and MGLY to global radiation as well as to biological activity cannot be concluded from the data. A slight correlation between GLY and MGLY in the SML and in aerosols could be a hint for interactions of especially GLY between seawater and the atmosphere.

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



1 Introduction

Oceans cover around 2/3 of the earth and most of the manifold exchange processes between oceans and the atmosphere are only rarely understood (Liss and Duce, 1997). Among these interaction processes organic export from the oceans to the atmosphere via gaseous emission or via bubble bursting play an essential role as this is one pathway towards organic compounds on marine aerosols (Liss and Duce, 1997). The two α -dicarbonyls glyoxal (CHOCHO; GLY) and methylglyoxal (CH₃COCHO; MGLY) have attracted increasing attention over the past years because of their potential role in secondary organic aerosol (SOA) formation (Carlton et al., 2007; Fu et al., 2008; Ervens et al., 2011). They can be produced via oxidation of volatile organic compounds (VOCs) and typical precursors are isoprene, toluene, acetylene and acetone (Fu et al., 2008; Warneck, 2003; Shaw et al., 2010). Besides secondary formation, direct emission of both GLY and MGLY to the atmosphere is also reported (Fu et al., 2008; Sinreich et al., 2010). Although the volatility of GLY and MGLY is high and these compounds are found in the gaseous phase, large shifts of GLY and MGLY in the atmosphere towards the particle phase are reported (Ervens and Volkamer, 2010). Aerosol uptake of highly soluble compounds like GLY is most efficient on aqueous aerosols, especially marine aerosols (Liggio et al., 2005), as the solubility of GLY and MGLY in aerosol water is expected to be strongly enhanced due to the present concentrated solutes (Lim et al., 2010). Volkamer et al. (2007) found that glyoxal uptake on aerosols can compete with glyoxal loss in the gas phase due to oxidation processes. Aqueous phase chemistry of GLY and MGLY in aerosol particles leads to further oxidized low volatility products such as oxalic acid (via OH radical reaction) and non radical reaction including the formation of high molecular weight compounds (Tilgner and Herrmann, 2010; Ervens et al., 2011; Lim et al., 2010). Recently Sinreich et al. (2010) attributed the open ocean as an important source for GLY in the atmosphere. They measured concentrations of GLY up to 140 ppt in the atmosphere via differential optical absorption spectroscopy (DOAS) over the remote tropical Pacific Ocean. Due to the short atmospheric lifetime of GLY

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures



Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures



Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

and MGLY of about 2 h these authors assumed a local marine source. They concluded that there must be an airborne formation mechanism of GLY in form of a process at the air-sea interface or via bubble bursting. Shaw et al. (2010) recently described that the oceans also emit GLY and MGLY precursors, especially isoprene. Zhou and Mopper (1997) investigated carbonyls including GLY and MGLY in the open ocean (Sargasso Sea, Atlantic) and found that the surface of the ocean is photochemical active and produces carbonyls. These authors further suggested that besides biological processes also photochemical processes influence carbonyl abundance in seawater (Kieber and Mopper, 1987). In the atmosphere, photochemical activity (e.g. in the oxidation of isoprene) is reported to lead to the formation of GLY and MGLY (Sinreich et al., 2010). Generally, the mechanism of the production regarding GLY and MGLY and the exchange between the surface of the oceans and the atmosphere are not well understood but the ocean is assumed to play an important role (Sinreich et al., 2010).

Analytical methods for GLY/MGLY analysis in environmental samples typically include a derivatisation step in order to reduce reactivity and polarity of the compounds. The most extensively used derivatisation agent for atmospheric samples is 2,4-dinitrophenylhydrazine (DNPH) often in combination with solid phase extraction (SPE) and liquid chromatography-mass spectrometry (LC-MS) or ultraviolet (UV) detection (Liggio and McLaren, 2003; Kampf et al., 2011). Zhou and Mopper (1997) applied DNPH derivatisation followed by SPE-LC-UV for seawater analysis. However, especially for seawater samples problems in SPE due to clogging of the cartridge due to matrix compounds are reported (Mbukwa et al., 2012). Recently a very sensitive HPLC method with fluorescence detection using dansylacetamidooxamine as derivatisation agent was presented for carbonyls in environmental water (Houdier et al., 2011). Kawamura and Ikushima (1993) developed a method for alpha dicarbonyls and atmospheric acids using BF₃/butanol derivatisation and gas chromatography-mass spectrometry (GC-MS). There are several studies using o-(2,3,4,5,6-Pentafluorobenzyl)-hydroxylamine (PFBHA) derivatisation and GC-MS for carbonyls in atmospheric samples (Ortiz et al., 2006; Pang and Lewis, 2013), though less studies exist for these de-

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

vices in high-matrix aqueous samples. One study (Saison et al., 2009) applies PFBHA derivatisation and GC-MS for carbonyls in beer, however for relatively high concentrations ($\mu\text{g L}^{-1}$ range). EPA method 556 applies PFBHA derivatisation followed by hexane extraction and GC-electron capture detection as reference method for carbonyls (including GLY and MGLY) in drinking water.

Due to the easy-to-use approach of PFBHA derivatisation combined with solvent extraction and the sensitivity and robustness of GC-MS analysis the aim of this work was to validate and apply this analytical method for measuring GLY and MGLY in seawater and in aerosol samples collected during a transatlantic POLARSTERN cruise in spring 2011. In seawater the focus was on the SML, as it is the direct interface between ocean and atmosphere. Connections of GLY/MGLY in the SML and on aerosol particles to environmental parameters such as global radiation, temperature, distance to the coastline and biological activity should help to find hints regarding the production of these compounds and a connection between these carbonyls in the ocean and in the atmosphere.

2 Experimental

2.1 Cruise

Figure 1 presents the POLARSTERN ANT XXVII/4 cruise plot from Capetown (South Africa) to Bremerhaven (Germany) between 20 April 2011 and 20 May 2011. Each day of the cruise is marked with a black square including 5-days backwards trajectories (blue lines), determined in a height of 30 m (NOAA HYSPLIT model, <http://www.arl.noaa.gov>, NOAA Air Resources Laboratory, Silver Spring, MD, USA). Aerosol particle sampling was performed continuously along the cruise. However due to sampling problems only 11 suitable aerosol filters could be loaded (reported in detail in Sect. 3.3.2). Water sampling was performed on 16 days, marked with a cross in Fig. 1.

The green crosses present the sampling points where oceanic water sampling and clean marine aerosol sampling was achieved simultaneously.

2.2 Sampling details

Oceanic water sampling was performed with a rubber boat that was launched from the POLARSTERN ship. For SML sampling, a glass plate (sampling area 2000 cm²) was vertically put in the water and slowly drawn upwards. The film adheres to the surface of the glass and is taken off by framed Teflon wipers (Stolle et al., 2010; van Pinxteren et al., 2012). This sampling technique is easy to use, requires only simple equipment and provides sampling of uniform SML thicknesses of ca. 100 μm (thickness = sampled volume/sampling area). Bulk water (BW) from 1 m depth was collected using a self-made device consisting of a glass bottle mounted on a telescopic rod that regulates sampling depth. After each sampling all material was extensively cleaned with reagent water. All water samples were stored in glass bottles and kept at -20 °C until analysis. Details of the sampling points for seawater sampling together with environmental parameters are listed in Table 1.

A high volume Digital sampler DHA-80 (Walter Riemer Messtechnik, Germany) was installed on top of the observation deck at a height of ca. 30 m. Aerosol particles (PM₁) were collected on preheated 150 mm quartz fiber filters (Minktel, MK 360) at a flow rate of 500 L min⁻¹. Sampling time was 24 h from midnight to midnight (UTC). After sampling, the filters were stored in aluminum boxes and kept at -20 °C in a freezer. After the cruise they were transported in dry ice to the TROPOS laboratories in Leipzig.

Air temperature was achieved from the Scalable Automatic Weather Station SCAWS. Broadband global radiation was measured with a Kipp&Zonen CM21 pyranometer. Data are sampled with a 1 Hz resolution despite a 95 % relaxation time of 5 s. The viewing angle is 180° and correlates with cosine properties. Under a glass dome is a black solar radiation absorbing thermopile and 64 thermocouples. Incoming short-wave radiation heats the top and the resulting temperature gradient of thermocouples generates an electric potential which is proportional to radiation flux density.

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



2.3 Analysis

GLY (40 % w/w) and MGLY (40 %) were purchased from Sigma Chemical Co. (St Louis, MO) Stock solutions with a concentration of 1 mgL^{-1} were prepared and used for 5 days (kept in the fridge at -20°C). From the stock solutions, five calibration standards were prepared in the range of $0.25\text{--}2 \mu\text{gL}^{-1}$ in volumes between 20 mL (for particle analysis) and 100 mL (for seawater analysis). Standards for seawater samples were always set to the same salinity as real samples (35 PSU) by adding 3.5 g sea salt mixture (Sigma Chemical Co.) to 100 mL milliQ water. Internal standards 2-Trifluoromethylbenzaldehyd and 2,2,6,6- Cyclohexanone (Sigma Chemical Co.) were added to the sample in a concentration of $1 \mu\text{gL}^{-1}$ each. For standard addition experiments, the samples were divided in 3–5 aliquots and spiked with different amounts of standard solution. For derivatisation, freshly prepared acidic PFBHA was added to the sample in a 10 000 fold excess corresponding to a concentration of $1 \mu\text{gL}^{-1}$ GLY. The free form of GLY and MGLY is in equilibrium with various forms of hydrated oligomers varying with concentration and solution pH, however, at concentrations below 1 molL^{-1} the monomeric form is dominant (Whipple, 1970). Due to acidic derivatisation conditions, equilibrium is shifted towards the free form of GLY and MGLY (Liggio et al., 2005). Therefore the measurements here represent the sum of the monomer carbonyls, its hydrated form and reversibly formed high molecular weight compounds (Kampf et al., 2012). To ensure complete derivatisation, the reaction mixture was allowed to stand overnight at room temperature. For extraction, 0.5–1 mL of hexane was added as extraction solvent. Formation of emulsion can be a problem using LLE for complex samples, but not observed for our seawater and particle samples. After stirring for 1 h the hexane layer was removed, dried over sodium sulfate and transferred to a GC autosampler vial. $2 \mu\text{L}$ were injected in splitless mode into the GC-MS. A HP-5 column (Agilent technologies, Waldbronn, Germany) was applied (30 m length, 0.25 mm i.D. $0.25 \mu\text{m}$ film thickness) and inlet temperature was set at 250°C and the oven program was as follows: 50°C (2 min) with 3°C min^{-1} to 130°C , with $10^\circ\text{C min}^{-1}$

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures



Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[⏪](#)[⏩](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

to 280 °C. The transfer line was set at 280 °C. For GLY m/z of 181 (quantifier ion) and 448 (qualifier ion), for MGLY m/z of 181 (quantifier ion) and 462 (qualifier ion) were detected. Internal calibration was performed using five concentration levels and dividing the peak areas of GLY and MGLY through the peak areas of the internal standard 2-Trifluormethylbenzaldehyd. For standard addition 2–4 concentration levels were added to a real sample and compared to external calibration. Oxalic acid was measured from aqueous particle extracts (25 % of the filter in 20 mL) as oxalate with ion chromatography, for details see Müller et al. (2010). Dissolved organic carbon was determined from the aqueous particle extracts using a TOC-VCPH analyzer (Shimadzu, Japan) as described in van Pinxteren et al. (2009).

3 Results and discussion

3.1 Method development: extraction and derivatisation

For seawater analysis, 100 mL of oceanic water samples (SML and BW samples) and for aerosol analysis up to 20 mL aqueous extract of 25 % of the filter sample were derivatized with PFBHA as described in the experimental part. Repetitive extraction of one and the same filter piece showed complete extraction in the first step. Blank problems due to the derivatisation agent (especially DNPH) for carbonyls are often reported and can be overcome by re-crystallization and further cleaning of the derivatisation reagent with a SPE column (Liggio and McLaren, 2003; Kampf et al., 2011). Using freshly prepared PFBHA solution blank values in the chromatograms were low, still a purification step by passing PFBHA over a SPE column (C18) led to a further reduction of the blank values. Figure 2 shows typical chromatograms for standards and samples using pre-cleaned PFBHA. While derivatised MGLY gave one peak, for derivatised GLY two isomers were found in the chromatograms. The two GLY isomers were assigned based on prior literature, the first eluting peak is referred to *trans* and the second peak to *cis* isomer (Standard Method Number 6252B), (EPA method 556). As

the ratio between *trans* and *cis* GLY was constant (1 : 3) only *cis* GLY was interpreted throughout this study.

3.2 Method validation

The limit of detection (LOD) was determined as the concentration corresponding to the mean blank ($n = 6$) plus three times the standard deviation (Kaiser and Specker, 1956). The values in Table 2 correspond to a sample volume of 100 mL oceanic water and a 20 mL aqueous extract of 25 % of the filter sample (area: 177 cm²) for aerosol particles. LODs can be further lowered by increasing the sample volume or reducing the extraction solvent volume (e.g. by evaporation). The LODs for seawater samples reported here are similar to other applications using GC-MS, for example by Kawamura (1993a) with LODs for carbonyls of 50 ng L⁻¹ (rain and snow samples). Zhou and Mopper (1997) found lower LODs for GLY and MGLY of 3–4 ng L⁻¹ after enrichment via SPE and DNPH-LC-UV analysis probably due to the very high injection volumes of 2000 µL into the LC system. Using highly fluorescence derivatisation agents and LC-FLD analysis, LODs of 16 ng L⁻¹ (GLY) and 12 ng L⁻¹ (MGLY) can be achieved without preconcentration (Houdier et al., 2011). However due to the high matrix content of real samples mostly sample preparation steps for a combination of clean up and analyte enrichment are applied. The LODs achieved for aerosol particle analysis (25 % of filter, aqueous extract of 20 mL, and extraction solvent of 1 mL) are 353 ng L⁻¹ for GLY and 259 ng L⁻¹ for MGLY. The higher LODs for an aqueous extract of 20 mL are caused by higher blank values from the filter material. The LODs for the aerosol particle analysis reported here are slightly lower compared to Kampf et al. (2011) applying DNPH-LC-MS analysis (510 ng L⁻¹ for GLY, 620 ng L⁻¹ for MGLY) and similar to Kawamura (1993a). Solvent extraction yields in this work were estimated by three successive extraction steps with hexane regarding the sum of them as 100 % value. The extraction yields in the first step were 80 % for GLY (76 % for 100 mL; 82 % for 20 mL) and 70 % for MGLY (65 % for 100 mL; 75 % for 20 mL). Repeatability, determined by 6 fold measurements of aqueous standards with a concentration of 1 µg L⁻¹, was very good with

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



relative standard deviations (RSD) below 10%. To test for matrix interferences, internal calibration was compared with standard addition and the graphs are shown in Fig. 3. No interferences with the matrix were found as the slopes of the calibration curves are very close (same for MGLY). Therefore, external calibration could be applied to the oceanic water samples as well as to the aqueous aerosol extracts.

3.3 Field measurements

3.3.1 GLY and MGLY in Atlantic seawater

The results for GLY and MGLY concentrations in SML and bulkwater are listed in Table 3. Average concentration of GLY in the SML was 330 ng L^{-1} ($123\text{--}616 \text{ ng L}^{-1}$). SML concentration for MGLY were on average slightly lower (261 ng L^{-1}) with a higher fluctuation ($77\text{--}760 \text{ ng L}^{-1}$). In BW, average concentrations were 121 ng L^{-1} (GLY) and 94 ng L^{-1} (MGLY). MGLY also showed higher fluctuation and in six BW samples MGLY concentrations were below LOD. There was a weak correlation between GLY and MGLY concentrations in the SML ($R^2 = 0.28$), while no correlation was found for these compounds in BW. The enrichment factor ($EF = c_{\text{SML}}/c_{\text{BW}}$) for GLY and MGLY in the SML varied strongly but was mostly > 1 . The average EF for both compounds was ca. 4 and EF_{GLY} and EF_{MGLY} are slightly correlated ($R^2 = 0.30$). The actual EF is in fact even higher considering that for several BW samples GLY and especially MGLY concentrations were below LOD whereas SML concentrations were detectable. Presently, there are few measured GLY and MGYL concentrations in real seawater samples available from literature. In one application from Zhou and Mopper (1997) GLY and MGLY were measured (besides other aldehydes) in the Sargasso Sea (Atlantic Ocean, near Bahamas). The concentrations are in the same range (lower nM range) compared to the data presented here. The authors also observed enrichment of GLY and MGYL in the SML with higher average enrichment factors than found in the present study but also with higher variability (EF: 1–21 for GLY; EF: 2–13 for MGLY). Regarding the formation and enrichment of carbonyls in seawater, photochemical production has been

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

reported to be a major source of carbonyls in surface water (Kieber et al., 1990). Zhou and Mopper (1997) found higher photoproduction rates of carbonyls in SML compared to BW. Moreover, microbial activity is discussed to be an important factor influencing carbonyl abundance in seawater (Zhou and Mopper, 1997; Kieber and Mopper, 1987).
5 Microorganisms may either take up carbonyl compounds or release them. Kieber and Mopper (1987) suggested microbial activity to be rather a sink than a source in well oxygenated conditions such as the SML. The enrichment of GLY and MGLY found in this work might be an indicator for photochemical production of the carbonyls in the SML, as the SML is more exposed to sunlight. However, only weak correlations were
10 found between global radiation and concentration of GLY and MGLY in the SML as well as enrichment of GLY/MGLY ($R^2 \approx 0.2$, Figs. S1–S4). Global radiation is slightly correlated to air temperature within the SML sampling time ($R^2 = 0.4$, Fig. S5). A correlation to SML concentrations of GLY and MGLY to air temperature was not found (Figs. S7–S8).

15 However, enrichment of the two carbonyls seems to be connected to air temperature, indicated by a positive correlation between EF and temperature ($R^2 = 0.4$, Figs. S9–S10). A connection of GLY and MGLY to biological activity was evaluated by correlating GLY and MGLY concentrations to Chl *a* as indicator for biological activity. Chl *a* satellite data provided by the Ocean Color Web from NASA (<http://oceancolor.gsfc.nasa.gov/>,
20 radius of 78 km, $n = 10$) showed no uniform trend regarding Chl *a* and GLY/MGLY concentration or enrichment in the water samples (Fig. S11–S14). However it must be mentioned that Chl *a* is a very broad indicator for biological activity and does not reflect carbonyl specific microorganisms. Moreover, satellite measurements are sensitive to cloud cover. Therefore, an important issue in further sampling campaigns is the development of analytical analysis of biologic pigments such as Chl *a* and others. Zhou
25 and Mopper (1997) reported that concentrations of carbonyls are higher in coastal areas compared to the open ocean but enrichment in the SML showed the opposite trend (higher enrichment in open ocean, lower enrichment in coastal waters). This is in agreement to Carlson (1983), who reported the same trend for DOC in general. Re-

garding GLY and MGLY in the present work the trend for decreasing concentrations in the SML towards the open ocean is weak ($R^2 = 0.14$ for GLY, $R^2 = 0.05$ for MGLY, Figs. S15–S16). Higher enrichment towards the ocean was very slightly observed for MGLY ($R^2 = 0.2$, Fig. S18).

In summary the enrichment factor > 1 implies photochemical production of the carbonyls that is also supported by the trend of a higher enrichment at higher temperatures. However, a direct correlation to global radiation was less pronounced. No indications of biological carbonyl production or depletion could be concluded from our data. Furthermore, only small dependencies regarding carbonyls in seawater and the distance to coastline were found.

3.3.2 GLY/MGLY on aerosol particles

Sampling problems and artifacts

As reported in Sect. 2.1, PM_1 aerosol particles were collected along the entire cruise. For several sampling days and at certain wind directions, the exhaust plume affected the filter sampling, resulting in a contamination of the filters. Therefore, all filters with elemental carbon values above 60 ng m^{-3} were excluded. Furthermore, some filters needed to be excluded due to water entrance into the Digital sampler inlet at high wind speeds resulting in wet filters. Overall we achieved 11 filter samples (24 h sampling each) which were suggested to be of marine origin due to their low values of elemental carbon and air masses from the open ocean.

At the moment it is not possible to assess positive and negative artifacts during aerosol sampling and the GLY and MGLY concentrations reported here may be affected by uncertainties such as adsorption of organic vapors, volatilization or further reactions of the analytes. However, care was taken that directly after sampling and during transport to the laboratories, filters were cooled throughout at -20°C until analysis. Quartz filters are employed in similar applications for the sampling of GLY and MGLY (Kawamura et al., 2010; Pavuluri et al., 2010; Li and Yu, 2005). Miyazaki et al. (2010)

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



reported that levels of field blanks were generally less than 17% of mass concentrations for diacids and related compounds (such as GLY and MGLY) for quartz filters.

GLY/MGLY concentrations on aerosol particles

The concentrations of GLY and MGLY in the PM₁ samples (Fig. 4) ranged between 0.11 and 0.41 ng m⁻³ (GLY) and 0.06 and 0.29 ng m⁻³ (MGLY). Due to strong acidic conditions during derivatisation, GLY and MGLY values represent the sum of the monomer carbonyls, its hydrated form and reversibly formed high molecular weight compounds (Liggio et al., 2003). The very good correlation between GLY and MGLY ($R^2 = 0.82$) suggest a similar transport and/or formation mechanism for these two compounds in the atmosphere. GLY concentrations are always slightly higher than MGLY (factor 1.3) which might be due to the increased solubility of GLY ($K_H = 3.6 \times 10^5 \text{ M atm}^{-1}$; MGLY: $K_H = 3.7 \times 10^3 \text{ M atm}^{-1}$ data from Fu et al., 2008) which might lead to a stronger uptake to the aerosol phase. As GLY and MGLY are short living compounds in the atmosphere with an atmospheric lifetime of ca. 2 h in an marine environment (Sinreich et al., 2010), GLY and MGLY are not expected to originate from continental sources but rather from the open sea. The concentrations for GLY and MGLY reported here are in the same order of magnitude as reported for marine aerosol particles by other authors in different places. No measured GLY and MGLY data for aerosol particles over the Atlantic Ocean were found in the literature. Kawamura et al. (2010) observed GLY (average: 0.55 ng m⁻³) and MGLY (average 0.31 ng m⁻³) on arctic aerosols showing a seasonal variation with a peak in April being produced during polar sunrise events. Over the western North Pacific the concentration of MGLY concentrations were in average 0.5 ng m⁻³ while GLY concentrations were much lower (0.06–0.12 ng m⁻³) (Miyazaki et al., 2010). In tropical Indian aerosols GLY and MGLY were found in concentrations between 4 and 23 ng m⁻³ (Pavuluri et al., 2010). An explanation for the higher concentrations could be that the air was mainly coming from the Arabian Sea, the Indian Ocean but also partly continentally influenced. Generally higher concentrations of GLY

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

and MGLY on aerosol particles are reported in anthropogenic areas compared to remote areas. In megacities GLY and MGLY can be up to hundreds of ng m^{-3} (for example up to 218 ng m^{-3} in Hong Kong (Li and Yu, 2005; Ho et al., 2007; Kawamura and Yasui, 2005). Liggio and McLaren (2003) reported the alpha dicarbonyls in the lower ng m^{-3} range at urban and rural sites in Canada (GLY: $0.43\text{--}3.3 \text{ ng m}^{-3}$; MGLY: $0.03\text{--}1.2 \text{ ng m}^{-3}$). At a background measurement site in the Rhine–Main area in Germany, concentration were in average 1 ng m^{-3} for GLY and 1.4 ng m^{-3} for MGLY (Kampf et al., 2011). In a correlation of GLY and MGLY to NO_x the authors showed an anthropogenic influence on the SOA formation potential of GLY and MGLY. In another study the authors found 0.81 ng m^{-3} for GLY and 0.31 ng m^{-3} for MGLY in a Boreal forest in Finland (Kampf et al., 2012). In a very recent study, Kawamura (2013) presented GLY and MGLY aerosol concentrations with a maximum of 70 ng m^{-3} and 120 ng m^{-3} at Mt. Tai (North China). The corresponding gas phase concentrations were by a factor of 10–20 higher. They found a strong correlation to levoglucosan, suggesting biomass burning to be an important source for alpha dicarbonyls.

For the data presented here, a very good correlation between the two carbonyls and oxalate was found (Fig. 5), regarding the absolute concentrations as well as their DOC fraction on the aerosol particles. The strong correlation is partly caused by the last sampling point close to Europe (highest GLY, MGLY and oxalate concentrations) but still pronounced without this data point ($R^2 = 0.4$ for GLY and MGLY). The correlation between GLY and MGLY to oxalate might show a linkage towards a secondary formation of oxalic acid via these alpha dicarbonyls, that is a detailed described pathway in the literature (Tilgner and Herrmann, 2010; Ervens et al., 2011). Tilgner and Herrmann (2010) showed that oxalic acid is formed preferably in the atmospheric aqueous phase due to aldehyde precursors such as GLY and MGLY. Rinaldi et al. (2011) suggested that cloud-mediated oxidation of gaseous GLY might be an important source for oxalate on aerosols. Oxalate concentrations in this work are by a factor of 29 higher than the sum of GLY and MGLY concentrations. This might be due to multiple potential precursors for oxalic acid or higher accumulation of oxalic acid in the aerosol particles due

to its much lower volatility compared to GLY and MGLY. Positive correlations between alpha dicarbonyls and oxalic acid were also reported by Kawamura (2013) regarding aerosol particles from Mt. Tai in China as well as for Arctic aerosols where oxalate concentrations were on average by a factor of 19 higher than GLY and MGLY together (Kawamura et al., 2010).

Regarding the data presented here, the last sampling point (17 May 2011) was very close to European mainland (Fig. 1) measurements. Although air masses were marine and EC concentrations below 60 ngm^{-3} , the higher concentration of GLY and MGLY (and also oxalate) might be caused by stronger anthropogenic influences such as enhanced ship traffic in this region. Different authors (Liggio and McLaren, 2003; Kampf et al., 2011) report stronger GLY and MGLY abundance in higher anthropogenic influenced areas probably due to an increased concentration of aromatic precursors. Also for oxalic acid, higher concentrations are reported in areas where a continental influence becomes noticeable (Warneck, 2003; Kawamura and Usukura, 1993). Probably other than ocean driven mechanisms and different precursors are important in this area close to Europe. Disregarding the last sampling point, a positive correlation between GLY and MGLY concentration to temperature ($R^2 = 0.42$, $R^2 = 0.30$, respectively, $n = 10$, Figs. S19–S20) was found. Global radiation correlated to MGLY ($R^2 = 0.42$; $n = 10$, Fig. S21) but weakly to GLY ($R^2 = 0.10$, $n = 10$, Fig. S22). Photochemical production from precursors such as isoprene is reported to be the dominant source for GLY and MGLY, but photolysis and oxidation with OH also represent sinks for these carbonyls in the atmosphere (Fu et al., 2008). Besides photochemical activity, Ervens and Volkamer (2010) revealed further important parameters that strongly affect GLY on aerosols such as particle constitution (liquid water content, pH, chemical composition). Therefore, a combination of environmental factors might affect GLY and MGLY abundance on aerosols. For oxalate concentrations determined in this work, correlation to global radiation was quite strongly pronounced ($R^2 = 0.60$, $n = 10$, Fig. S23) indicating photochemical production to be a dominant source. This might indicate that GLY and

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[⏪](#)[⏩](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

MGLY production could result from the photooxidation of a VOC precursor in the gas phase.

In contrast to seawater results, a negative correlation of GLY and MGLY to Chl *a* was observed for GLY ($R^2 = 0.54$, Fig. S24) and less pronounced for MGLY ($R^2 = 0.28$, $n = 6$, Fig. S25). Miyazaki et al. (2010) also observed increased concentrations for GLY on marine aerosol particles in times of lower biological activity, while MGLY concentrations were hardly affected. Less GLY on aerosols at higher biological activity might support the hypothesis that microbial activity represents rather a sink than a source for these compounds. However, in seawater no such correlation between Chl *a* and carbonyl concentrations were found for our data. As mentioned earlier, for more detailed investigation of dependencies between carbonyls and microbiological activity, the development of analytical methods of Chl *a* and carbonyl specific microorganisms is needed.

3.3.3 GLY/MGLY in the SML and aerosol

Six sampling points were obtained, where seawater and clean marine aerosol was sampled in the same time range (Fig. 1; green crosses). It needs to be considered however that SML are punctual sampling spots whereas aerosol sampling is a mean of 24 h. A positive correlation between the carbonyls in the SML and in aerosol particles was found for GLY ($R^2 = 0.33$; less pronounced for MGLY: $R^2 = 0.12$) presented in Fig. 6. There are multiple ways of exchange processes between oceans and atmosphere for the carbonyls that could be reasons for a positive correlation. Turekian et al. (2003) summarized three main processes for sea to air transfer of organics: (1) gaseous transfer from the ocean into the atmosphere and gas-to-particle conversion, (2) incorporation of material from the organic rich SML during wind driven aerosol production and (3) accumulation of organics to the surface of bubbles and its subsequent injection to the atmosphere (bubble bursting). Although the last process is mostly expected for surface active material, also non-surface active organics can be associated to the surface active agent (Duce, 1976). Generally, all these processes are possible

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

for GLY and MGLY – as well as for their precursors – including subsequent deposition back to the ocean. Due to the high solubility of GLY and MGLY their net flow is expected from air to sea (Zhou and Mopper, 1990). Sinreich et al. (2010) observed high concentrations of gaseous GLY over the Pacific Ocean and suggested an airborne source of GLY in form of a process at the air sea interface or by a bubble bursting process.

In sum the slight correlation of the alpha dicarbonyls in the SML and in the aerosol particles reported here could be a hint for interactions of these compounds – especially GLY – in seawater and in the atmosphere although specific mechanisms cannot be concluded from this work.

4 Conclusions

This work presents a suitable and easy to use method for the analysis of GLY and MGLY in seawater and aerosol particles. To our knowledge, these data present the first measured GLY and MGLY concentrations on Atlantic aerosol particles and the corresponding seawater samples. In seawater the enrichment in the SML implies a photochemical production of GLY and MGLY though a clear connection to global radiation was not observed. GLY and MGLY on marine aerosols are strongly connected to each other, suggesting similar formation mechanisms. Moreover they are both correlated to oxalate. A connection to biological activity regarding GLY and MGLY on aerosols was found however the reason for this connection is unclear as in seawater this correlation was not observed. The results of GLY and MGLY in marine aerosol and in the ocean water give a first insight towards interaction processes between ocean and atmosphere, though detailed mechanisms and net fluxes cannot be concluded from our data. Further investigations of important precursors of GLY and MGLY (especially isoprene), gas phase sampling of the carbonyls and a larger dataset are necessary for a better understanding of GLY and MGLY interactions between air and sea.

Supplementary material related to this article is available online at:
[http://www.atmos-chem-phys-discuss.net/13/15301/2013/
acpd-13-15301-2013-supplement.pdf](http://www.atmos-chem-phys-discuss.net/13/15301/2013/acpd-13-15301-2013-supplement.pdf).

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ACPD

13, 15301–15331, 2013

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

Glyoxal and methylglyoxal in Atlantic seawaterM. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

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Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

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Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

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ACPD

13, 15301–15331, 2013

Glyoxal and methylglyoxal in Atlantic seawaterM. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

Table 1. Sampling points and environmental parameters.

Sampling date	Local sampling time	Coordinates latitude	longitude	Distance to coast (km) ^a	Air temperature ^b (°C)	Global radiation ^b (W m ⁻²)
23 Apr 2011	10:05 (UTC+1)	24°24.47′ S	9°18.50′ E	560	19.3	402
24 Apr 2011	12:04 (UTC)	20°48.90′ S	6°4.26′ E	780	21.6	326
25 Apr 2011	12:03 (UTC)	17°22.89′ S	3°2.86′ E	920	22.9	413
29 Apr 2011	12:05 (UTC)	8°42.44′ S	4°22.05′ W	1496	26.4	538
30 Apr 2011	13:02 (UTC)	5°30.22′ S	7°3.21′ W	1056	27.8	478
1 May 2011	12:02 (UTC−1)	2°29.84′ S	9°33.70′ W	760	28.2	551
2 May 2011	08:05 (UTC−1)	0°0.75′ S	11°37.64′ W	600	27.4	– ^c
3 May 2011	12:01 (UTC−1)	3°32.22′ N	14°33.95′ W	484	27.3	541
4 May 2011	12:04 (UTC−1)	6°45.68′ N	17°14.39′ W	460	27.6	575
5 May 2011	12:01 (UTC−1)	9°52.69′ N	19°50.63′ W	430	25.2	563
8 May 2011	12:04 (UTC−1)	13°32.49′ N	20°31.61′ W	380	22.9	552
9 May 2011	12:03 (UTC−1)	23°39.87′ N	20°15.97′ W	410	20.4	374
12 May 2011	11:00 (UTC−1)	25°39.95′ N	18°3.28′ W	328	20.2	508
13 May 2011	12:19 (UTC)	34°57.29′ N	12°55.29′ W	498	18.1	489
16 May 2011	07:08 (UTC−1)	37°34.94′ N	12°5.52′ W	340	17.0	404

^a shortest distance to coastline estimated via google earth,

^b at the sampling time,

^c not available.

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Table 2. Validation data for GLY and MGLY.

	GLY	MGLY
RSD, (%) $n = 6$	8.9	5.4
extraction yield (%)	80	70
LOD _{seawater} (ng L ⁻¹) ^a	54	50
LOD _{aerosol extract} (ng L ⁻¹) ^b	353	259
LOD _{aerosol particle} (ng m ⁻³) ^c	0.05	0.04

^a for 100 mL saltwater,

^b for 20 mL aqueous extract,

^c for 545 m³ air volume.

[Title Page](#)
[Abstract](#)
[Introduction](#)
[Conclusions](#)
[References](#)
[Tables](#)
[Figures](#)
[Back](#)
[Close](#)
[Full Screen / Esc](#)
[Printer-friendly Version](#)
[Interactive Discussion](#)


Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



Table 3. SML thickness, GLY and MGLY concentrations in SML and BW and corresponding EF.

Sampling date	SML thickness (μm)	GLY			MGLY		
		SML	BW	EF	SML	BW	EF
23 Apr 2011	132	257	< LOD	> 5.8	234	< LOD	> 5.9
24 Apr 2011	122	239	71.1	3.4	111	< LOD	> 3.4
25 Apr 2011	132	123	83.8	1.5	130	< LOD	> 1.5
29 Apr 2011	–	312	134	2.3	172	50.9	3.4
30 Apr 2011	142	419	59.4	7.1	325	< LOD	> 8.1
1 May 2011	146	284	75.2	3.8	145	51.4	2.8
2 May 2011	128	453	75.1	6.0	760	246	3.1
3 May 2011	127	276	69.7	4.0	305	68.1	4.5
4 May 2011	93	492	93.7	5.3	650	< LOD	> 16
5 May 2011	93	389	101	3.9	179	213	0.8
8 May 2011	108	360	93.9	3.8	253	150	1.7
9 May 2011	134	208	114	1.8	225	143	1.6
12 May 2011	111	204	216	0.9	77.2	< LOD	> 1.9
13 May 2011	–	616	257	2.4	199	63.8	3.1
16 May 2011	113	313	322	1.0	143	186	0.8
Average*		330	121	3.5	261	94.2	4.0

* For average values concentrations < LOD were set at LOD (GLY: 54 ngL^{-1} ; MGLY: 50 ngL^{-1}).

Glyoxal and methylglyoxal in Atlantic seawater

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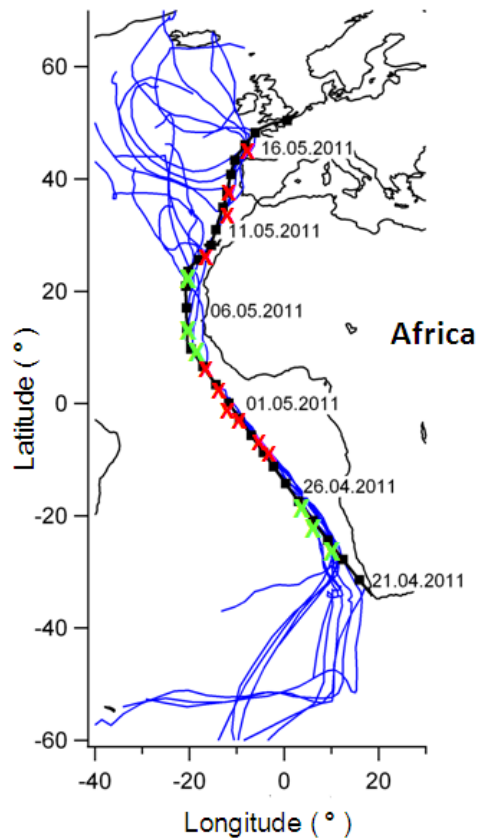
[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[◀](#)[▶](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

Fig. 1. Cruise plot including seawater sampling (red crosses), simultaneous seawater and aerosol particle sampling (green crosses) and backwards trajectories (blue lines).

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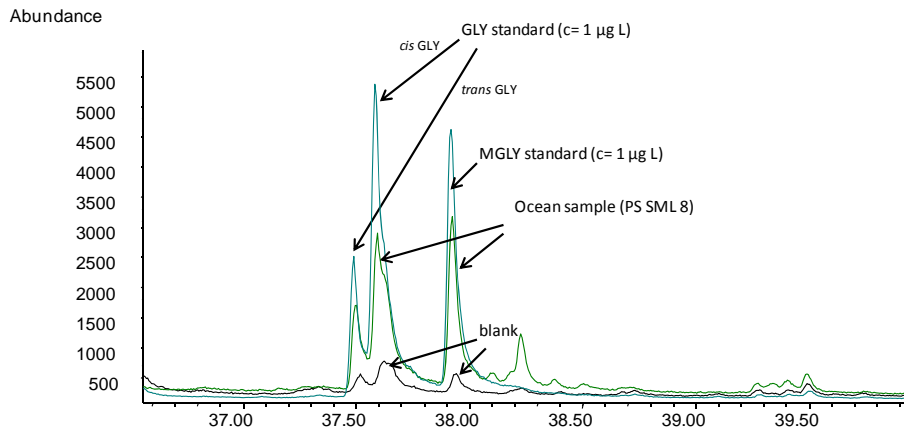
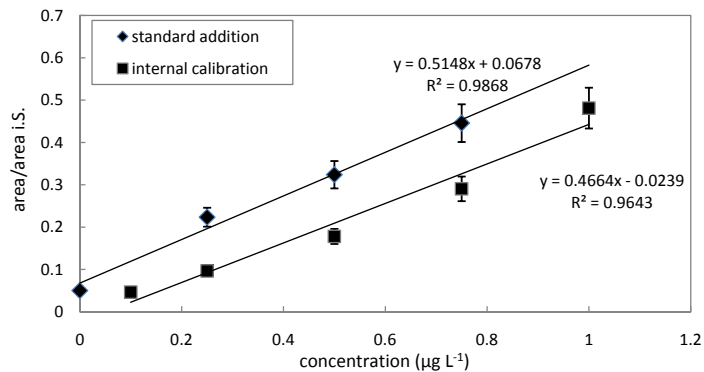


Fig. 2. SIM chromatograms of GLY and MGLY standard, ocean sample and blank (milliQ water + PFBHA), 100 mL sample volume each, PFBHA was pre-cleaned over a SPE C18 cartridge.

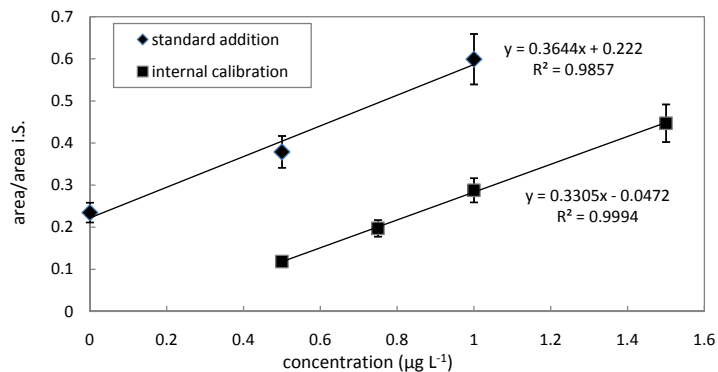
[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[⏪](#)[⏩](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and H. Herrmann



(a)



(b)

Fig. 3. calibration curve for internal calibration and standard addition for GLY (a) 100 mL seawater sample, (b) 20 mL aqueous extract of aerosol particle sample.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and H. Herrmann

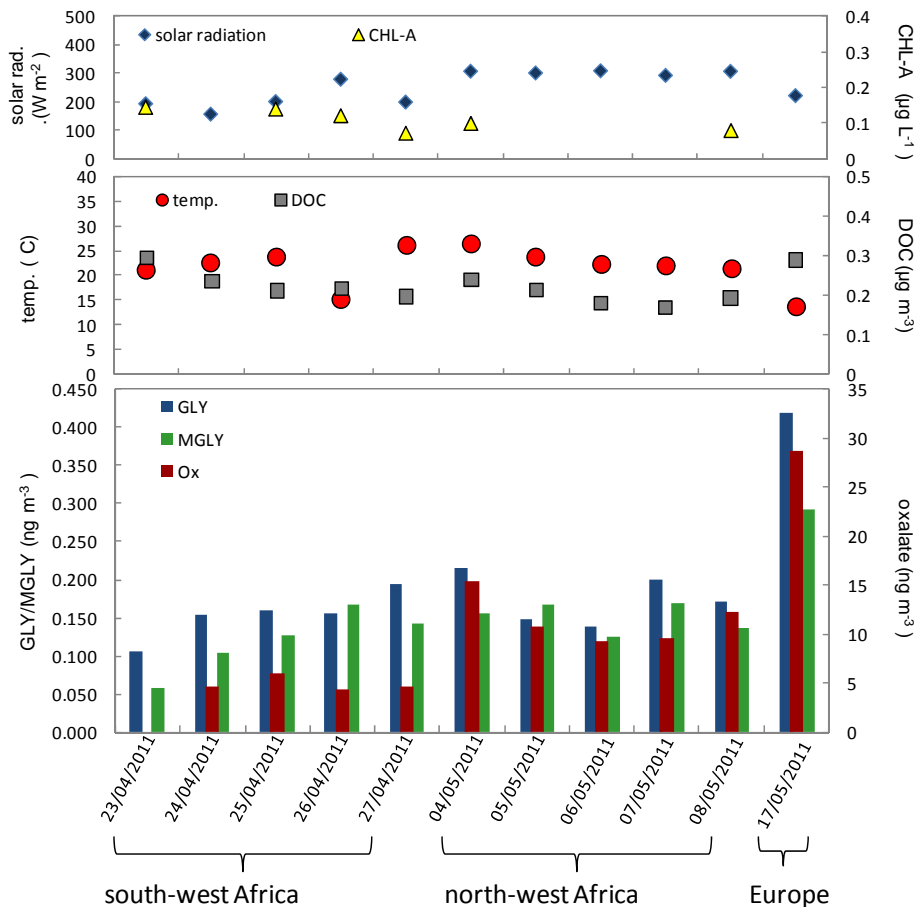


Fig. 4. Concentration of GLY, MGLY and oxalate together with DOC and environmental parameters.

Title Page

Abstract Introduction

Conclusions References

Tables Figures

◀ ▶

◀ ▶

Back Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



Glyoxal and methylglyoxal in Atlantic seawater

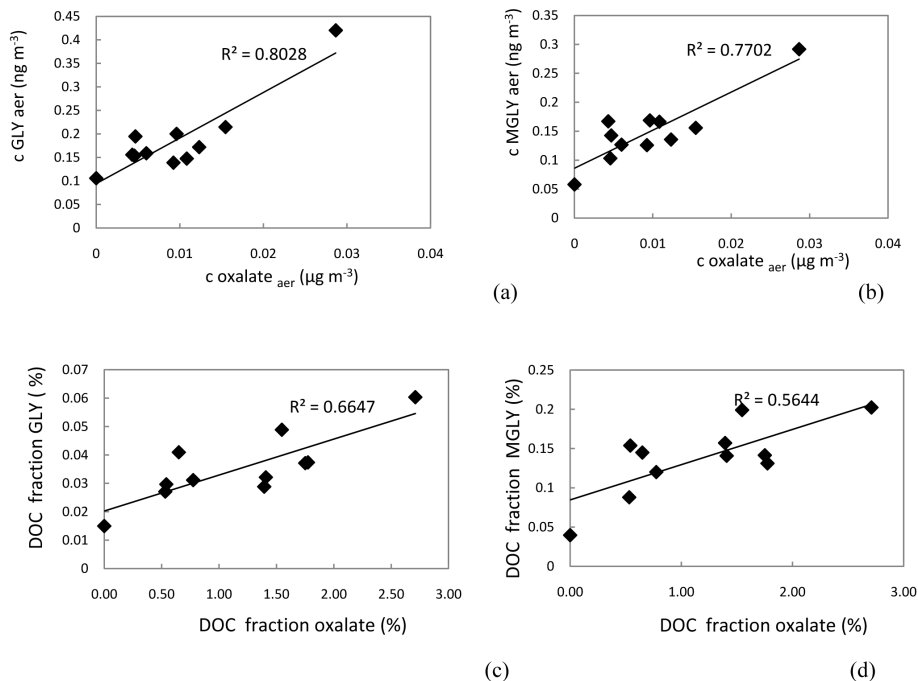
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Fig. 5. Correlation of GLY/MGLY and oxalate: (a, b) concentration, (c, d) DOC fraction. (DOC fraction was calculated from GLY, MGLY and oxalate concentrations and their contribution to DOC concentrations on a carbon basis).

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[⏪](#)[⏩](#)[⏴](#)[⏵](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

Glyoxal and methylglyoxal in Atlantic seawater

M. van Pinxteren and
H. Herrmann

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

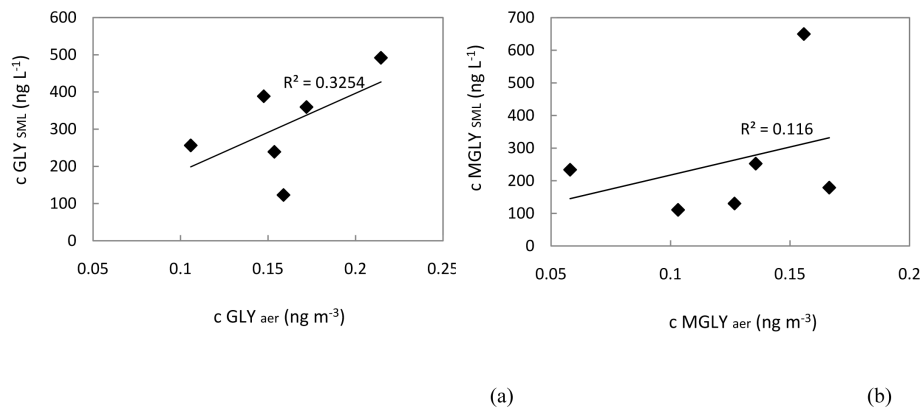


Fig. 6. Correlation of **(a)** GLY and **(b)** MGLY in SML and in aerosol particle.