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Chemical characterization and stable carbon isotopic composition of particulate polycyclic aromatic hydrocarbons issued from combustion of 10 Mediterranean woods

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Abstract

The objectives of this study were to characterize polycyclic aromatic hydrocarbons from particulate matter emitted during wood combustion and to determine, for the first time, the isotopic signature of PAHs from nine wood species and Moroccan coal from the

- ⁵ Mediterranean Basin. In order to differentiate sources of particulate-PAHs, molecular and isotopic measurements of PAHs were performed on the set of wood samples for a large panel of compounds. Molecular profiles and diagnostic ratios were measured by gas chromatography coupled with a mass spectrometer (GC/MS) and molecular isotopic compositions (δ^{13} C) of particulate-PAHs were determined by gas chromatogra-
- phy/combustion/isotope ratio mass spectrometry (GC/C/IRMS). Wood species present similar molecular profiles with benz(a)anthracene and chrysene as dominant PAHs, whereas levels of concentrations range from 1.8 to 11.4 mg g⁻¹ OC (sum of PAHs). Diagnostic ratios are consistent with reference ratios from literature but are not sufficient to differentiate the different species of woods. Concerning isotopic methodology, PAH
- ¹⁵ molecular isotopic compositions are specific for each species and contrary to molecular fingerprints, significant variations of δ^{13} C are observed for the panel of PAHs. This work allows differentiating wood combustion from others origins of particulate matter (vehicular exhaust) using isotopic measurements (with $\delta^{13}C_{PAH} = -28.7$ to -26.6%) but also confirms the necessity to investigate source characterisation at the emission
- ²⁰ in order to help and complete source assessment models. These first results on woodburnings will be useful for the isotopic approach of source tracking.

1 Introduction

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Polycyclic Aromatic Hydrocarbons (PAHs) are ubiquitous compounds in the environment, detected in various compartments such as water (Latimer and Zheng, 2003; Amoako et al., 2011), sediments (Latimer and Zheng, 2003; Srogi, 2007; Van Drooge et al., 2011), soils (Jensen et al., 2007; García-Alonso et al., 2008; Desaules et al.,



2008), organisms (Meador, 2003; Perugini et al., 2007; Srogi, 2007) or air (Ravindra et al., 2008; Van Drooge and Ballesta, 2009; Yan et al., 2009; Sheesley et al., 2011). Most of PAHs come from the incomplete combustion of organic materials and are potentially carcinogenic and mutagenic for human beings (Atkinson and Arey, 1994; Kang et al.,

- ⁵ 2011; IARC, 2011). In the atmosphere, they are distributed between gaseous phase and particles (Eiguren-Fernandez et al., 2004; Ré-Poppi and Santiago-Silva, 2005). Main origins of particulate-PAHs are anthropogenic such as emissions from domestic (residential heating) and industrial activities, agriculture and/or vehicular exhausts. Wood combustion represents a non-negligible origin of particulate-PAHs because of the variety of human activities involving hiemass huming such as building heating.
- the variety of human activities involving biomass burning such as building heating, cooking (Lee et al., 2005) or swidden, slash-and-burn farming, but also because of natural wood combustions (forest fires).

The identification and the apportionment of PAH sources are related to the description of source characteristics, for example molecular and/or isotopic features. Several

- ¹⁵ methodologies of source tracking were developed on the measurements of particulate-PAH concentrations and on the use of molecular profiles and/or diagnostic ratios (Guo et al., 2003; Dallarosa et al., 2005; Ravindra et al., 2008). Some works reported the concentrations of a large range of particulate-PAHs emitted for instance during the combustion of different wood species in the United States of America and contributed
- to the characterization of molecular profiles at the emission (Fine et al., 2001, 2002, 2004). For example, their data are considered in several works as wood combustion fingerprints for source apportionment using Chemical Mass Balance model (Yang and Chen, 2004; Sheesley et al., 2008). Nevertheless, due to the large variety of wood species and the high reactivity of PAH compounds in the atmosphere, involving vari-
- ²⁵ ations of concentrations and diagnostic ratios, it is sometimes risky to apportion PAH sources by the only application of such approaches.

More recently, methodologies based on the molecular stable-carbon isotopic compositions allow source studies by identifying characteristic isotopic fingerprints. Unfortunately, in spite of the conservation of isotopic compositions during the transport of



particulate matter in the atmosphere (O'Malley et al., 1994; Guillon et al., 2012a), only few studies describe the isotopic compositions of PAHs for wood combustion (Ballentine et al., 1996; O'Malley et al., 1997). In order to contribute to the development of source tracking for particulate-PAHs, eleven Mediterranean wood species were burned

- in a combustion chamber to identify molecular and isotopic fingerprints of emitted particulate-PAHs. Concentrations of a large panel of PAHs were measured by GC/MS analysis and the determination of molecular stable-carbon isotopic compositions was performed by GC/C/IRMS analysis. The aim of these measurements was to characterize combustion emissions of several wood species, potentially involved in the pollution
- of Mediterranean area during biomass burning and/or forest fire episodes and largely, to provide molecular isotopic signatures of wood burnings for source assessment in Europe.

2 Experimental procedure

2.1 Description of wood species

¹⁵ A large variety of wood species from the Mediterranean Basin was selected for the study of particulate matter emitted during wood combustion. Collection of woods was performed in the natural park of "Calanques de Marseille" (France). Table 1 summarizes the main characteristics of the selected woods: nine plant species and eucalyptus Moroccan coal which is commonly used in North African countries for cooking preparation

²⁰ ration.

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Note that some woods were studied several times in order to compare different conditions of combustion and the way of wood cutting prior to experiments. Organic and elemental carbon (OC and EC) measurements were not performed on the entire panel of wood species. After collection, all wood samples were dried up at 50 °C during two days.



2.2 Setup for wood combustion

Wood combustion experiments were carried out at Lannemezan in the combustion facility of the Laboratory d'Aérologie (UMR5560, Toulouse, France) made up of three rooms: the fireplace, the mixing chamber (120 m^3) and the laboratory for measure-

- 5 ment monitoring. The fireplace was equipped with a chimney allowing the circulation of smoke to the mixing chamber by a trap door. The internal surface of this room was rendering. Approximately 100 grams of each wood were burned in the fireplace and once combustion was stabilized, smoke was injected in the mixing chamber by opening the trap door. After 5 min, the door was closed and two fans were activated to homogenize
- the repartition of products of combustion (particles and gases) in all the volume of the mixing chamber prior sampling. Samples were collected during 20 min (Table 1). At the end of each combustion experiment, the mixing chamber was vented in order to renew the full atmosphere between two experiments.

2.3 Sample collection

- ¹⁵ Wood combustion aerosol particles with aerodynamic diameters less than 2.5 μ m were collected on quartz filters (150 mm diameter, Whatman, Q-MA) using a high-volume sampler (Mégatec Digitel DA-80, 30 m³ h⁻¹). Collection was performed during around 20 min corresponding to a sampling volume of (10 ± 1.3) m³ for the different species, except for eucalyptus Moroccan coal (sampling time 77 min corresponding to a volume
- of 36.6 m³). Field blanks (consisting in exposed filter without any air filtration) were collected before each combustion experiment. Prior to sampling, all the quartz fibre filters were heated at 500 °C during 4 h. After sampling, filters were wrapped in aluminium foil, sealed in polyethylene bags and stored at -20 °C. To perform different kind of analysis on the same panel of wood combustions, filters were punched: 0.64 cm² of each filter
 were used for the quantification of particulate-PAHs, 1 cm² for the analysis of organic
- and elemental carbon (OC/EC) and 16.62 cm² of filters were used to determine the isotopic compositions of PAHs by GC/C/IRMS.



EC and OC measurements were performed on 1.5 cm^2 of each filter using a Thermo-Optical Transmission (TOT) method on a Sunset Lab analyser (Birch and Cary, 1996) implemented with the EUSAAR_2 method (Cavalli et al, 2010).

2.4 Analytical procedure

concentrations by NIST.

5 2.4.1 Validation on standard reference materials

Three standard reference materials were used for the validation of the full analytical procedure from extraction to quantification steps. Urban Dust (SRM 1649a) and Diesel exhaust (SRM 2975 and SRM 1650b) were purchased from NIST (Gaithersburg, MD, USA). As ambient particle samples, these materials are very complex containing a large panel of organic and inorganic compounds. SRM 1649a was collecting during 12 months in the Washington DC area in 1976–1977 and represents a time-integrated sample of urban area (Wise and Watters, 2007). Four-cycle diesel engines operating in a large variety of conditions during 200 h allowed the collection of SRM 1650b which was representative of heavy-duty diesel engine emissions (Wise and Watters, 2006). SRM 2975 was collected from a filtering system designed for an industrial diesel-powered forklift (Wise and Watters, 2009). The certified value for total carbon (organic and elemental) is (0.1768 ± 0.0019) g g⁻¹ for SRM 1649a particle. This value was not available for Diesel exhausts. These sets of particles are certified for PAH

2.4.2 Extraction by pressurised fluid extraction

Pressurised fluid extractions (PFE) were performed using accelerated solvent extractor ASE 350 (Dionex SA). A large range of parameters were reported in the literature for the extraction of particulate-PAHs. This technique was validated and largely employed for the certification of NIST materials such as SRM 1649a or SRM 2975 (Schantz et al., 1997). In our study, extractions were performed in dichloromethane ("For residue



and pesticides analysis" type, Acros Organics) under following parameters: $T = 100 \,^{\circ}$ C, $P = 100 \,^{\circ}$ Bars, heat time of 6 min with 2 static cycles of 8 min. Different sizes of ASE stainless steel cells were employed for the different extraction required in this study in order to optimize the volume of solvent against the volume occupied by the filter or the punch in the cell. Therefore, standard reference materials deposed on glass fibre filters, punches of filters (0.64 cm²) cut off for the quantification of PAHs and punches (16.62 cm²) of blank filters were extracted in 10 ml cells whereas remains of filters used for the determination of isotopic compositions in 34 ml cells. Prior to the extraction, a cellulose filter (19.8 mm diameter, Dionex SA) was placed at the bottom of the cell and one third of the cell was filled up with 2.0-mm-diameter glass beads (A556, Roth) previously cleaned and heated at 450 °C during 4 h. Filters folded in four were placed in the cell and the volume was completed with glass beads in order to minimize the dead volume. For quantitative measurements by GC-MS, internal standards (20 µl per samples) were added gravimetrically in the cell and then, the cell was completely filled with clear based.

¹⁵ with glass beads. After the extraction, filtration was performed in the PFE cell through the cellulose filter.

Moreover, the cells filled with the cellulose filter and glass beads were pre-extracted using the same extraction conditions as for the samples.

2.4.3 Purification procedure on micro-columns

- After extraction and filtration, the extract was concentrated in approximately 500 µl in the extraction solvent using a vacuum evaporation system (RapidVap, Labconco), under the following conditions: vortex motion (70%), vacuum (900 mBars) and heat (51 °C). Different purification steps were then required to remove all undesirable organic compounds that could interfere in the PAH analysis. The extract was first purification and interfere in the PAH analysis.
- fied on a micro-column containing activated copper (copper powder, 40 mesh, Sigma-Aldrich) which allowed sulphur elimination, and alumina (aluminium oxide, 150 Basic Type T, Merck, VWR) in order to remove macromolecules present in these complex matrices such as lipids or pigments, by eluting PAHs with 3 × 5 ml of dichloromethane.



The extract was then concentrated with low heating (T = 40 °C) under nitrogen stream (N₂, 99.995% purity, Linde Gas) and the solvent was changed to iso-octane (2,2,4-trimethyl pentane, HPLC grade, Scharlau). Finally, the sample was fractionated on a micro-column of silica (0.063–0.200 mm diameter, Merck). The first fraction containing alkanes was eluted with 2 ml of pentane (Ultra Resi-analyzed type, JT Baker) and the second fraction containing PAHs with 3 × 5 ml of pentane/dichloromethane (65/35, v/v). Again, this fraction was concentrated under nitrogen stream and low heating and the solvent was changed in isooctane prior to the analysis.

In the case of GC/MS analysis, the final volume was not a critical point due to the use of internal and "syringe" standards. Analyte losses were evaluated by checking the internal standards recovery yields. "Syringe" standards were added to the sample and measured gravimetrically prior to the analysis by GC/MS for the quantification of the internal standards.

In the case of GC/C/IRMS analysis, neither internal standard nor syringe standard were used because molecular isotopic composition of a compound could be measured only if no coelution with other compounds appeared. Under our analytical conditions, native and perdeuterated PAHs were coeluted involving an external calibration of the GC/C/IRMS system.

2.4.4 Quantification of PAHs

- Gas chromatography coupled with a mass spectrometer (GC/MS) analyses were performed for the quantification of PAHs in the samples and for the validation of HPLC fractionation. Quantitative analyses were performed using gas chromatography (HP model Series 6890) coupled to a quadrupole mass spectrometer (HP model 5973). Two different columns were used. First, for PAHs quantification, the capillary column
- was a $30 \text{ m} \times 0.25 \text{ mm}$ ID $\times 0.25 \mu \text{m}$ film thickness Rxi-17 (50% of diphenyl and 50% of dimethyl-polysiloxane, Restek) in order to analyse some isomers classically coelutated in usual chromatographic conditions. One μI was injected into the GC (7683 autosampler injector, Agilent Technologies) in splitless mode (purge delay = 90s, purge



flow = 60 ml min⁻¹, 25 psi pulse during 90 s). The injector temperature was 280 °C and the oven temperature was held at 60 °C during 2 min, then programmed from 60 °C to 320 °C at a rate of 10 °C min⁻¹ and held at 320 °C during 25 min. Helium (He, 99.9996 % purity, Linde Gas) was used as carrier gas at the constant flow of 1.3 ml min⁻¹. The

- ⁵ interface temperature was kept to 290 °C during analysis. Ionization was realized by electron impact mode (70 eV) and mass detection was performed in selected ion monitoring (SIM) mode (dwell time = 30 ms, electron multiplier voltage = (1620 ± 50) V, solvent delay = 5 min). The ion molecular m/z were chosen for native and perdeuterated PAHs.
- ¹⁰ PAH concentrations were measured using internal standard quantification and moreover, internal standards were quantified using "syringe" standards. This double quantification allowed the calculation of internal standard recovery yields and hence, to check that internal standards, and therefore native PAHs, were not lost along the analytical procedure. As described before in Sect. 2.4.2, internal standards were added prior to
- the extraction in the ASE cell and "syringe" standards at the end of sample preparation before the GC/MS analysis. All the standards used in this study are presented in Table 2. This GC/MS method was validated by the application of our analytical procedure on SRM 1649a and SRM 1650b. Percentages of recovery yields for internal standards vary from 50 % to 97 % depending on the compounds (see Sect. S1 in the Supplement).

Then, for the validation of HPLC fractionation, a classical method was applied on all the collected fractions. Analysis were performed by GC/MS using an HP model Series 5890 Gas Chromatograph coupled to an HP model 5972 mass-selective detector (quadrupole) (Agilent Technologies). The capillary column was a 30 m × 0.25 mm ²⁵ ID × 0.25 µm film thickness HP-5MS (Agilent Technologies). One µl was introduced into the GC via an auto-injector (HP model 7673, Agilent Technologies) in splitless mode injection (purge delay = 60 s, purge flow = 60 ml min⁻¹, 25 psi pulse during 60 s). The injector temperature was 270 °C and the oven temperature was held at 60 °C during 2 min, then programmed from 90 °C to 330 °C at a rate of 10 °C min⁻¹ and held at



330 °C during 5 min. Helium (99.9996 % purity, Linde Gas) was used as carrier gas at the constant flow of 1.3 ml min^{-1} . The interface temperature was kept to 290 °C during analysis. Ionization was realized by electron impact mode (70 eV) and mass detection was performed in selected ion monitoring (SIM) mode (dwell time = 60 ms, electron multiplier voltage = (2900 ± 50) V). The ion molecular *m*/*z* were chosen to native PAHs (compounds indexed with * in Table 2) and perdeuterated PAHs (only used in the case of the validation of analytical method with SRM 2975). This second GC/MS method was validated by the application of our analytical procedure on SRM 2975.

2.5 Determination of ¹³C/¹²C isotopic compositions of PAHs

10 2.5.1 HPLC fractionation

An additional purification step was required to eliminate the remaining aromatic unresolved complex mixture (UCM) that could interfere during the GC/C/IRMS analysis. As a consequence, after silica purification, the extract was concentrated in a volume of 200 µl of isooctane and the aromatic fraction was fractionated by high performance liquid chromatography (HPLC) on aminosilane phase (Dynamax, 5 µm, 15 250 mm × 10 mm ID, Varian) using the following solvent program: pentane (100%) during 25 min, then pentane/dichloromethane (90/10, v/v) during 25 min and finally pentane (100%) during 10 min. The flow-rate was 4 ml min⁻¹. The HPLC fractionation based on ring number allowed the collection of five different fractions by detecting the UV signal at 254 nm: monoaromatic and diaromatic (naphthalenes and diben-20 zothiophenes for example), phenanthrenes, fluoranthene+pyrene, benz(a)anthracene + chrysene + pentaaromatics and hexaaromatics. In order to focus the fractionation on the native PAHs, HPLC fractionation was performed in the conditions described in a previous work by Guillon et al. (2012b). The objective was to collect only na-

tive PAHs and to eliminate remaining compounds by collecting fine fractions (Fig. 1) and reducing the time of collection around the time of retention of studied compounds. After each fractionation, the integrity of each fraction was controlled by GC/MS



analysis and finally, the volume of reconcentration was optimized to perform isotopic composition analysis of the different fractions in accurate conditions. Note that under our analytical conditions, benzo(k)fluoranthene (BkF), benzo(b)fluoranthene (BbF) and benzo(j)fluoranthene (BjF) were coeluted and the isotopic composition determined and presented in this work concerned all these isomers (noted BFs).

2.5.2 Measurements of ¹³C/¹²C isotopic compositions by GC/C/IRMS

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Measurements of molecular stable carbon isotopic composition is based on the elution of individual compounds through a gas chromatograph and their conversion to CO_2 and H_2O in a combustion furnace heated at 940 °C. H_2O is then trapped via a Nafion membrane and purified CO_2 is introduced into a magnetic mass spectrometer, continuously monitoring ions having mass ratio (m/z) 44 $({}^{12}C^{16}O_2)$, 45 $({}^{13}C^{16}O_2$ and ${}^{12}C^{17}O^{16}O)$ and 46 $({}^{12}C^{18}O^{16}O, {}^{13}C^{17}O^{16}O$ and ${}^{12}C^{17}O_2)$. The isotopic composition is then calculated using the ratio 44/45 m/z and 44/46 m/z for the correction of ${}^{17}O$ contribution to the 45 m/z signal. The isotopic ratio is reported in terms of $\delta^{13}C$ and expressed in per mil (‰), relative to the Vienna Pee Dee Belemnite standard (VPDB): $\delta^{13}C = [({}^{13}C/{}^{12}C)_{sample}/({}^{13}C/{}^{12}C)_{VPDB} - 1] \times 10^3$.

Methodology of the determination of PAHs molecular isotopic compositions was validated and described in details elsewhere (Guillon, 2011; Guillon et al., 2012b). Briefly, stable-carbon isotopic analyses of individual PAHs were carried out by gas chromatog raphy/combustion/isotope ratio mass spectrometry (GC/C/IRMS) using an HP 5890 Series II Plus gas chromatograph (Hewlett-Packard) interfaced via a CuO furnace (940 °C) and a hygroscopic membrane (Nafion) to a Delta Plus isotopic ratio mass spectrometer (Finnigan MAT Corporation). One μl to 3 μl of solutions were injected into the GC in the splitless mode depending on the concentrations of studied analytes in the

²⁵ fractions. The GC oven temperature program was optimized to reduce PAHs coelutions with undesirable compounds. GC oven programs were used in the following conditions:



- Different fractions containing PAHs except pentaaromatics: 50°C (2 min) to 300°C (15 min) at a rate of 10° C min⁻¹.
- For pentaaromatics fraction : 50 °C (2 min) to 200 °C at a rate of 10 °C min⁻¹ and from 260 °C to 300 °C (15 min) at a rate of 2 °C min⁻¹.
- The injector temperature was 270°C. The carrier gas was helium (99.9996% purity, Linde Gas) (flow rate: 2 ml min^{-1}). The capillary column was a 30 m × 0.25 mm ID \times 0.25 µm film thickness HP-5MS (Agilent Technologies). For calculation purposes, CO₂ reference gas (99.995% purity, Linde Gas) was automatically introduced into the isotopic ratio mass spectrometer in a series of pulses at the beginning of each analysis.

Quality insurance 2.6 10

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For GC/MS analysis, a calibration solution was prepared with non-deuterated PAH standard solution, internal standard solution and "syringe" standard solution for each type of analysis. This solution was injected before and after each sequence of analysis in order to calculate the PAH response factors with respect to the corresponding

internal standards as well as the internal standard response factors with respect to the 15 corresponding "syringe" standards. The response factor values range between 0.48 and 1.6 depending on the molecule. Variations of the response factors are within 2 to 6%. The injections were carried out only if the conditions required for the analysis were fulfilled (good recovery yields of native and internal PAHs and response factors consistent with control monitoring), the calibration solution being also used to check 20 and monitor the conditions of the chromatographic and detection systems.

Moreover, in order to validate the integrity of analytical system and to monitor the conditions of the chromatographic and detection systems for GC/C/IRMS analysis, a solution containing ${}^{13}C/{}^{12}C$ certified alkanes (*n*-C₁₅, *n*-C₂₀, *n*-C₂₅, Chiron AS) was analysed prior to any analysis sequence of samples. Repeatability and reproducibility of the analytical method were validated by measuring isotopic compositions of





Interactive Discussion

(compounds indexed with * in Table 2) prior to any sequence of analysis. The reproducibility of the individual isotopic measurements is $\pm 0.50 \%$. Precision reported in this study is based on multiple analyses of each sample (at least three analyses). Therefore, as described by Guillon et al. (2012a), the analysis of PAH solutions from 20 to 450 ng ul^{-1} allowed determining the operational linear range of the method and the limit

⁵ 450 ng µl^{-'} allowed determining the operational linear range of the method and the limit of quantification, corresponding to the quantity of injected carbon required to measure signal amplitude of ion 44 peak up to 1 V. Table 3 summarises the quantities of PAHs required to perform GC/C/IRMS analysis in accurate conditions.

Therefore, blanks filters and extraction blanks were extracted and analysed in order to check levels of PAHs during the application of analytical protocol (see Sect. S2 in the Supplement).

3 Results and discussion

3.1 Validation on standard materials

As explained before, three standard reference materials were used for the validation of analytical procedure, described in details in Sect. S1 (Supplement). Briefly, SRM 2975 was used for the validation of analytical protocol applied on natural particles for the determination of ¹³C/¹²C. Analytical protocol of PAH quantification was applied on SRM 1649a and SRM 1650b in order to certify the good extraction of PAH in the sequence of wood particle extraction and to validate the quantification of 31 PAHs by

- GC/MS, respectively. Recovery yields of internal standard range from (46 ± 13) % to (69 ± 12) %, from (59 ± 15) % to (84 ± 12) % and from 47 % to 91 % for SRM 2975, SRM 1650b and SRM1649a, respectively (Fig. A1 in the Supplement). Uncertainties vary between 10 % and 15 %. PAH concentrations determined in this study for the different standard materials are in good agreement with reference and certified values
 reported in the literature (Table A1 and Fig. A2). Concerning the quantification of PAHs
- extracted from SRM 1650b, some values are lower than certified ones but this gap



could be explained by the conditions of extraction which were softer than the ones used for the certification of this standard material. The results obtained on SRM 2975, SRM 1650b and SRM 1649a traduce the good reproducibility and repeatability of our analytical procedure. Analyses performed in accurate conditions were validated and allow the determination of the concentrations and the isotopic compositions of PAHs of generated particles.

3.2 Molecular approach

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3.2.1 Concentrations of PAHs

Concentrations of 31 PAHs were determined in PM_{2.5} sampled during wood combustion experiments. Internal standard recovery yields range from 30 to 97 %. Note that the lowest values corresponded to DBT-d8 recovery yield which is a low molecular weight compound and may be lost easily during evaporation steps. Because of the method of quantification, native PAHs and internal standards were affected by the same physicochemical processes during the various steps of the analytical procedure. The use of

- internal standards allows us correcting the potential losses of native PAHs. Table 4 presents the concentrations of particulate-PAHs sampled in the chamber during the combustion of wood species compared to the global concentrations of organic carbon (OC). Due to the non-availability of OC, concentrations are not reported for four experiments: cork oak twigs 2, juniper tree, heather 2 and cypress 2.
- First, PAH concentrations depend on the compounds and on the wood species. Particulate-PAH concentrations (ΣPAH_{part}) range from 1.83 to 11.40 mg g⁻¹ OC for Cypress 1 and alep pine 2 respectively, meaning a variation of about one order of magnitude between wood species. These values are in good agreement with PAH concentrations reported in the literature concerning controlled wood burnings for the
 characterization of particulate emissions (Fine et al., 2001, 2002; Schmidl et al., 2008;
- Gonçalves et al., 2010) but slightly lower than those reported by Alves et al. (2010).



Therefore, levels of PAH concentrations are significantly different from the same wood family. For example, combustion of oak woods generate PAH levels from 3.6 to 7.98 mg g⁻¹ of OC. It could be explained by the adaptation of the different species of oaks against the Mediterranean climate (humidity, temperature). Concerning cork ⁵ oak burnings, the nature of wood affects the level of PAH concentrations with higher concentration emissions, when the wood was cut in twigs.

In addition, the type of wood (hardwood vs. softwood) is not correlated with the levels of PAH concentrations. This tendency is not consistent with the previous results of Gonçalves et al. (2010), who differentiate Pinus Pinaster combustion with high concentrations (Σ PAH_{part} = 75.5 mg g⁻¹ OC) from three hardwoods emissions with low PAH concentrations (Σ PAH_{part}<10 mg g⁻¹ OC). On the contrary, Fine et al. (2001, 2002) did not significantly differentiate PAH concentrations between hardwood and softwood combustions from North-eastern and Southern United States, confirming the observa-

¹⁵ Considering the concentrations, PAH repartitions for the different studied woods are quite similar: fluoranthene and pyrene are the dominant compounds for all woods, especially for softwoods. This tendency was already reported in the literature (Fine et al., 2001, 2002; Alves et al., 2010; Gonçalves et al., 2010), fluoranthene and pyrene being considered as characteristic of PM_{2.5} from wood combustion. Alves et al. (2010)
 ²⁰ reported a different molecular profile for PM₁₀ with benz(a)anthracene as the most abundant PAH.

3.2.2 Molecular fingerprints of particulate-PAHs

tions reported in this work.

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Different tools for source apportionment may be used to differentiate emissions from combustion of wood species. The determination of molecular fingerprints was per-

formed on all the different woods presented in Table 1. Due to the non-availability of OC values for cork oak-twigs 2, juniper tree, cypress 2 and heather 2 and the fact that these data are not required for the application of molecular approach, these four wood



species are added in that part of the discussion. Figure 2 presents PAHs molecular repartitions of the different woods studied here. Note that percentages of contribution of each PAH were determined considering the total of particulate PAHs from m/z 228 to 302 (from benz(a)anthracene to dibenzoperylenes). The choice of these high molecular

weight compounds is based on the fact that they are as a majority adsorbed on particulate matter (Odabasi et al., 1999; Ré-Poppi et Santiago-Silva, 2005) and the molecular profiles reported in this work can be compared to those reported in the literature.

No significant particularities could be underlined meaning that PAH molecular profiles are conserved for the variety of all wood species. Considering Fig. 2 only, benz(a)anthracene and chrysene represent the dominant contributions in the sum of

- benz(a)anthracene and chrysene represent the dominant contributions in the sum of particulate-PAHs (from BaA to DahP) followed by benzo(a)pyrene. Benz(a)anthracene and chrysene contributions vary from 12.9 and 17.1% and from 14.5 to 25.4% respectively. Note that preponderance of both compounds was already reported in the literature as molecular markers of wood combustion (Marchand et al., 2004; Ravindra et al., 2008). Our results confirm the interest of chrysene measurements in source
- assessment methodologies.

Main differences regard compounds with low concentrations such as perylene, DBacA, DBahA, DBbkF, DalP, DaeP and DahP involving that these compounds could not be used as molecular tracers because of too low concentrations in ambient sam-

- 20 ples. Previous works reported the interest of molecular fingerprints in order to differentiate PAH sources (Sun et al., 2003; Zhang et al., 2009; El Haddad et al., 2011) and some models of source apportionment are based on these measurements to determine the origins of particulate matter. In this study, we show that the use of a single molecular fingerprint based on PAH concentrations could traduce the origin of Mediterranean
- wood burnings whatever the wood species because of the similarities of molecular fingerprints of the eleven woods. Nevertheless, this methodology also does not allow differentiating the variety of different wood species because of the conservation of such fingerprints. In addition, by comparing (1) and (2) samples for cork oak, heather and



cypress species, the impact of combustion conditions is not underlined as they also present similar molecular fingerprints.

The application of the methodology for determining PAH molecular fingerprints on the results of Fine et al. (2001, 2002) by choosing the common measured compounds reveals some similarities and some particularities for the large variety of studied species (Table 5).

This molecular approach shows that some compounds may allow differentiating combustion of woods from North America and Mediterranean Basin. As shown in Table 5, BkF, BjF, IP, BghiP, Antha and Cor contributions to the sum of PAH concentrations (from BaA to Car) are significantly different between the two sets of wood burnings. Note that

- BaA to Cor) are significantly different between the two sets of wood burnings. Note that wood species are characteristic from the two areas and that these values are calculated as a mix of both softwoods and hardwoods, meaning that whatever the type of wood, the molecular signature is conserved from a region. Such observations confirms the importance of the choice of fingerprints in source apportionment models depend-
- ¹⁵ ing on the area of interest and the necessity to study a large panel of particulate-PAHs characteristic from sampling sites. Nevertheless, the conservation of molecular fingerprints does not allow differentiating different kinds of wood burnings in the same area, showing the necessity to use other methodologies of source apportionment regarding wood species from United States of America and from the Mediterranean Basin.

20 3.2.3 Diagnostic ratios

Molecular approach of source apportionment may be based on the use of molecular fingerprints as described in the previous section but also on the use of molecular diagnostic ratios. They are largely used in the literature for source tracking (Yang and Chen, 2004; Li et al., 2009; Yin et al., 2010). This methodology is based on the measure²⁵ ments of PAH concentrations and the determination of concentration ratios which may be specific from each origin and considered as constant during the atmospheric transport of particulate matter. Concerning biomass burning, several reference values of diagnostic ratios are reported in the literature: Ravindra et al. (2008) demonstrates that



[BaP]/[BghiP] value up to 1.25 and [IP]/([IP]+[BghiP]) close to 0.62 correspond to PAHs emitted from wood burning. Li and Kamens (1993) determined reference values between 1 and 1.5 for [BaA]/[BaP] and around 0.8 for [BghiP]/[IP]. [Phen]/([Phen]+[Anth]) value is evaluated at 0.84 ± 0.16 by Galarneau (2008). Table 6 presents diagnostic ratios determined in this work. First some diagnostic ratios are up a similar for the difference of the second se

- tios determined in this work. First, some diagnostic ratios are very similar for the different species, such as [Phen]/([Phen]+[Anth]) and [Fluo]/([Fluo]+[Pyr]). On the contrary, some molecular ratios allow differentiating some variety of wood species. For example, [IP]/([IP]+[BghiP]), [BaA]/[BaP], [BghiP]/[IP] and [Pyr]/[BaP] are significantly different for eucalyptus Moroccan Coal than for other woods. Note that eucalyptus Moroccan
- ¹⁰ Coal is a coal and particulate-PAHs are not considered as wood burning signature, contrary to other materials studied in this work. Moreover, [BaA]/[BaP] and [Pyr]/[BaP] could be used for the apportionment of cypress trees. Despite of a large range of variations of diagnostic ratios, no tendency is observed concerning concerning both softwood and hardwood families. Nevertheless, variations of diagnostic ratios are not
- ¹⁵ sufficient to differentiate all woods, meaning that others factors may involve variations of PAH ratios as conditions of combustion, temperature, elemental carbon normalization. In the literature, several diagnostic ratios are used as reference ratios for PAH emitted during wood burnings, as mentioned before. The different diagnostic ratios measured in this study are consistent with these reference diagnostic ratios ex-
- ²⁰ cept for [IP]/([IP]+[BghiP]), [BghiP]/[IP] and [Fluo]/([Fluo]+[Pyr]). Thus, concerning [IP]/([IP]+[BghiP]) and [BghiP]/[IP], diagnostic ratios from this study are respectively lower and higher than reported reference values. Nevertheless, [IP]/([IP]+[BghiP]) varies in a large range depending on the sources. More, the values reported by Ravindra et al. (2008) for Diesel exhaust, coal and wood combustion are in the range of
- those measured for Diesel exhaust (0.35–0.7) reported by Rogge et al. (1993). The variation of combustion conditions in the different works and the impact of physico-chemical processes on diagnostic ratios during the transport of particulate matter may possibly affect the reference values explaining the gap observed in several studies performed in different conditions. [Fluo]/([Fluo]+[Pyr]) range between 0.45 and 0.50 in this



study but these values also correspond to gasoline exhaust diagnostic ratios reported by Dallarosa et al. (2005) confirming the limitation of this method for source apportionment. Another example is the [BaA]/[BaP] ratio for which values for the different woods studied in this work are consistent with the reference value (Li and Kamens, 1993) and

with the values reported for other wood combustion studies (Table 6) but also similar to the diagnostic ratios of Diesel exhaust (0.9–1.7) reported by the same study of Li and Kamens (1993). Note that diagnostic ratios are linked to the conditions of formation of PAHs at the emission point and could be affected by the variation of environmental conditions during the transport of particles (photochemistry, oxidative processes, mixing of air masses...).

The application of molecular approach on the sets of data from literature is difficult but allows differentiating wood species from several areas such as North American, Asian or European woods despite of previous noticed differences with reference values. Some diagnostic ratios were calculated by using PAH concentrations reported in previous works of wood burnings from these different areas and are presented in Table 6.

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Depending on the compounds, some diagnostic ratios, such as [Fluo]/([Fluo]+[Pyr]), [Pyr]/[BaP] and [BaP]/[BghiP], are constant in the range of 0.4–0.6, up to 1.24 and up to 1.49 respectively, whatever were the wood and the area of interest, except for the results reported by Gonçalves et al. (2008). For the most of the studied PAH ratios, measurements are not in good agreement because of either high variations of diagnostic ratios or different conditions of sampling. Consequently, diagnostic ratios of this study are significantly different despite the area of interest was similar (Mediterranean Basin).

²⁵ Moreover, [Pyr]/[BaP] is constant except for eucalyptus Moroccan coal which value is lower than any reported values. This may be due to the nature of burned material (coal vs. dried wood) and this is observed considering [BaA]/[BaP], [BghiP]/[IP] and [IP]/([IP]+[BghiP]). In the case of [Pyr]/[BaP], the trend is the same as for [Phen]/([Phen]+[Anth]) and [BaA]/[BaP] with lower ratios for Asian woods than for



American and European species, except for sugarcane combustion. Indeed, this plant is a " C_4 plant" including a photosynthetic pathway different from other studied woods which are " C_3 plants". This difference may be explained by the way of PAH formation during the combustion of both plant families.

- Finally, [IP]/([IP]+[BghiP]) and [BghiP]/[IP] are significantly different between various areas of interest. A large variation is observed for wood combustions in Portugal (Gonçalves et al., 2008) as described before. Works performed in the USA are significantly different presenting diagnostic ratios reported by Fine et al. (2001, 2002) and Rogge et al. (1998) in the ranges of 0.6–0.7 and 0.2–0.3 respectively. Note that
 combustion studies were not performed in the same conditions: Rogge et al. (1998) sampled particulate matter along the chimney during wood combustion and fuelled regularly the fire with wood logs whereas Fine et al. (2001, 2002) performed the sampling four meters above the fire and injected cold air to cool the smoke allowing organic
- vapours to condense. Our values are between 0.45 and 0.5 showing that differences in ¹⁵ European and American wood burnings are significant and could be useful to differentiate the different varieties.

To conclude, variations of diagnostic ratios are not significant to apportion combustion of various wood species. Differences observed between the different reported studies may arise from the nature of woods but also from the combustion conditions (flam-

- ing or smoldering fires, time of burning, moisture content...), type of fireplace facilities (domestic fireplace or laboratory experiments) or conditions at the beginning of fire (addition of fuel, newspapers...). Including all these considerations, it appears sometimes difficult to compare results from different works, especially because of variations of gas-particles partitioning and PAH profiles. Nevertheless, whatever the study, diag-
- nostic ratios are not significantly different for the woods from the same area involving that this approach is not sufficient to clearly differentiate the origins of particulate matter. Therefore, it was decided in this work to complete the simple molecular approach by the determination of molecular isotopic compositions of particulate-PAHs.



3.3 Molecular stable-carbon isotopic compositions of PAHs

Molecular isotopic compositions of PAHs were determined for the panel of woods considered in this work (Table 7). Due to the low quantities of particulate-PAHs on remaining filters after the GC/MS analysis and the limits of quantification of GC/C/IRMS
measurements, especially for high molecular weight compounds, some molecular isotopic compositions could not be determined, explaining the differences of isotopic profiles of wood species (Table 7 and Fig. 3). Note that uncertainties correspond to three replicates of analysis and range from 0.1 to 0.9‰, being in good agreement with uncertainties determined with standard solutions (see previous sections) and reference materials (Guillon et al., 2012b). Considering all the panel of PAHs, two groups of woods could be differentiated by molecular isotopic compositions. The first including rosemary.

- alep pine 1 and 2 and heathers is characterised by δ^{13} C depleted in 13 C with isotopic compositions between -27.6 and -24.1 ‰. The second group composed of the different oaks, Mediterranean false-brome, eucalyptus Moroccan coal, juniper tree and
- ¹⁵ cypresses is characterised by isotopic compositions in the range of -32 to -27.1 ‰. Whatever the PAH and the conditions of combustion, molecular isotopic signatures of heathers are depleted in ¹³C whereas those of Mediterranean false-brome are depleted in ¹²C, meaning that isotopic measurements could differentiate these wood species from others. No similarities are observed for the isotopic profiles and each compound
- ²⁰ could be analysed separately in order to differentiate wood species (Fig. 3). It is also clearly underlined on Fig. 3 that molecular fingerprints are not sufficient to differentiate the panel of woods contrary to molecular isotopic compositions which are specific for each species. Regarding the small uncertainties of measurements, it is possible in this study to focus the analysis on some compounds of interest, and not on the panel of
- ²⁵ PAHs. Indeed, molecular isotopic compositions of each compound allow differentiating at least the half of wood species signature (Table 7). For example, phenanthrene isotopic compositions are significantly different between all woods studied in this work and isotopic signatures of cypress and oaks are characterised by δ^{13} C lower than those of



heathers, pines and rosemary. Regarding the results for benzo(e)pyrene, the isotopic compositions of heathers and alep pine 2 are depleted in ¹³C contrary to those of other woods. As underlined with phenanthrene and benzo(e)pyrene data, isotopic compositions do not allow separating hardwoods from softwoods and the variations of δ^{13} C

- are probably not induced by this repartition of species (Fig. 3). Nevertheless, the uncertainties on these measurements of isotopic compositions allow to differentiate the origin of particulate-PAHs between the different wood species, by combining the results of several PAH isotopic compositions.
- Several reference values of molecular isotopic compositions of particulate-PAHs from different sources are presented in Fig. 4 and compared with the results of this study. The application of the methodology of source apportionment based on isotopic measurements involves the knowledge of isotopic compositions of PAHs from various sources contributing to the particulate pollution at the site of interest. First, note that all PAH isotopic measurements reported in this work are consistent with previous results reported by O'Malley et al. (1994, 1997) and Okuda et al. (2002), concerning C_3 plant combustion, fireplace and woodburning. The variations of isotopic signatures of particulate-PAHs are reported in the range of -26.6 to -12.9‰ for vehicular exhausts (O'Malley et al., 1994; Okuda et al., 2002; Peng et al., 2006). Significant differences between δ^{13} C of PAHs from woodburning emissions and vehicular exhaust
- ²⁰ confirm the interest of this approach and the possibility of the method to differentiate these sources by isotopic measurements. Most of PAH isotopic compositions allow differentiating wood combustions and vehicular exhaust, except phenanthrene and indeno(1,2,3-c,d)pyrene whose isotopic compositions are almost the same for these two kinds of origins. Nevertheless, coal combustion signatures are difficult to differ-
- ²⁵ entiate from woodburnings, especially for benzofluoranthenes, benzo(a)pyrene and indeno(1,2,3-c,d)pyrene. The main reason is based on the fact that results from literature are not consistent and the range of isotopic compositions for coal combustion is large: δ^{13} C between –22 and –31.2% depending on the origin of coal and the conditions of preparation (temperature, process...).



Consequently, it is necessary to focus the measurement on particulate-PAHs whose isotopic compositions are specific for each source and to continue the characterization of PAH sources at the emission by studying other sources such as vehicular exhausts in European countries. Therefore, regarding the PAH molecular isotopic compositions of coal, values are similar with one of others sources (such as vehicular exhaust) and involves that no apportionment of sources is possible when coal combustion contributes

to the particulate pollution. Consequently, prior to any application of this isotopic approach, a partial knowledge of potential sources involved in the pollution is often necessary and especially if any coal combustion is involved in the particulate pollution.

10 4 Conclusions

In this study, the methodology of source apportionment by the determination of molecular isotopic compositions (δ^{13} C) of particulate-PAHs was applied on different samples of particles arising from the combustion of ten different woods and coal from the Mediterranean Basin. Combustions of woods were performed under controlled conditions in a burning chamber in order to compare nature of woods (hardwood vs. 15 softwood), wood species and conditions of burning (flaming vs. smoldering). After the validation of the full analytical procedure, concentrations of thirty-one PAHs were determined by GC/MS analysis and molecular isotopic compositions of ten PAHs were measured for all the panel of wood particles. Molecular and isotopic approaches were applied on this set of samples in order to identify the relevant tools for differentiating 20 wood species. Molecular approach confirms that molecular fingerprints are constant whatever the wood species and the conditions of combustion. Comparing our results with previous works reported on particulate matter from wood burnings at the emission (Rogge et al., 1998; Fine et al., 2001, 2002; Oros et al., 2006; Gonçalves et al., 2008; Wang et al., 2009), some reference diagnostic ratios are constant in the different

25 2008; Wang et al., 2009), some reference diagnostic ratios are constant in the different areas of interest ([Fluo]/([Fluo]+[Pyr]), [Pyr]/[BaP] and [BaP]/[BghiP]) whereas some allow to apportion wood burnings from Asia to European and North American origins



(for instance [Phen]/([Phen]+[Anth]) and [BaA]/[BaP]). Nevertheless, the values of diagnostic ratios of wood combustion are often in the same range as those of vehicular exhausts, revealing the limitation of this methodology.

- Therefore, it is preferred to use molecular isotopic compositions of particulate-PAHs
 ⁵ for all the panel of woods, wood species being characterised by different isotopic signatures depending on the PAH of interest and on the conditions of combustion. The isotopic signatures of wood burnings from Mediterranean Basin are in good agreement with previous wood combustion data from the literature (O'Malley et al., 1997; Okuda et al., 2002) and significantly different from those reported for vehicular exhaust or fire¹⁰ place soot, confirming the interest of the isotopic methodology for the apportionment of the various sources of particulate matter. Our results also confirm the necessity to measure isotopic compositions of particulate-PAHs at the emission for a larger variety
- of sources, alike wood species in this study, but also for others origins such as vehicular exhausts (Diesel and gasoline) or coal combustion. The large variability of isotopic
- ¹⁵ compositions for wood combustion particles represent the large variety of wood species and of combustion conditions that may occur in the nature during opened fires largely common in this arid area.

Supplementary material related to this article is available online at: http://www.atmos-chem-phys-discuss.net/12/20631/2012/ acpd-12-20631-2012-supplement.pdf.

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Common name	Botanical name	Wood family	condition of combustion	Volume of sampling (m ³)	Time of sampling (min)	ОС (µg m ⁻³)	ЕС (µg m ⁻³)
Cork oak-twigs 1	Quercus Suber L.	hardwood	smoldering	8.90	21	3902	67
Cork oak-twigs 2	Quercus Suber L.	hardwood	flaming	10.95	24	n.a.	n.a.
Cork oak	Quercus Suber L.	hardwood	flaming	9.43	20	304	16
Juniper tree	Juniperus	softwood	smoldering	8.97	20	n.a.	n.a.
Rosemary	Rosmarinus Officinalis	hardwood	flaming	11.01	24	1318	165
Alep pine 1	Pinus Halepensis	softwood	flaming	9.5	21	2923	223
Alep pine 2	Pinus Halepensis	softwood	flaming	9.05	20	3433	226
Green oak	Quercus llex	hardwood	flaming	8.94	20	3395	188
"kermes" oak	Quercus Coccifera	hardwood	flaming	10.33	24	515	57
Cypress 1	Cupressus Sempervirens	softwood	smoldering	8.13	21	12416	390
Cypress 2	Cupressus Sempervirens	softwood	flaming	11.05	25	n.a.	n.a.
Mediterranean False-Brome	Brachypodium Retusum	hardwood	flaming	9.95	24	2298	22
Heather 1	Erica Multiflore	hardwood	flaming	9.53	21	1126	174
Heather 2	Erica Multiflore	hardwood	smoldering	13.19	29	n.a.	n.a.
Eucalyptus Moroccan coal	-	-	smoldering	36.64	77	121	< 1

n.a. not available indicated numbers are the identification number of samples.

Table 1. Description of wood species studied in this work.

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Native PAHs	Abbreviations	Internal Standards	" Syringe" Standards
Naphthalene	Naph	Naphthalene d8	
Acenaphtylene	Асу		
Acenaphtene	Ace		
Fluorene	FI	Phenanthrene d10	
Phenanthrene*	Phen		
Anthracene	Anth	Anthracene d10	
Dibenzothiophene	DBT	Dibenzothiophene d8	Pyrene d10
Fluoranthene*	Fluo	Eluoranthene d10	
Pyrene*	Pyr		
Benzo(ghi)fluoranthene	BghiF		
Benzo(c)phenanthrene	BcP		
Benz(a)anthracene	BaA	Chrysene d12	
Chrysene + Triphenylene	Chrys		
Cyclopenta(cd)pyrene	CcdP		
Benzo(b)fluoranthene*	BbF		
Benzo(k)fluoranthene*	BkF		
Benzo(j)fluoranthene*	BjF	Ranza(a) Rurana d12	
Benzo(a)fluoranthene*	BaF	Delizo(e)i yrene urz	
Benzo(e)pyrene [*]	BeP		
Benzo(a)pyrene [*]	BaP	Benzo(a)pyrene d12	
Perylene*	Per	Benzo(e)Pyrene d12	
Dibenzo(ac)anthracene	DBacA		
Indeno(1,2,3-cd)pyrene*	IP		Benzo(b)fluoranthene d12
Dibenzo(ah)anthracene	DBahA	Benzo(abi)pervleno d12	
Benzo(ghi)perylene*	BghiP	Denzo(gin)peryiene d12	
Anthanthrene	Antha		
Coronene	Cor		
Dibenzo(bk)fluoranthene	DbkF		
Dibenzo(al)perylene	DalP	Coronene d12	
Dibenzo(ae)perylene	DaeP		
Dibenzo(ah)perylene	DahP		

Table 2. List of studied PAHs with the corresponding internal and "syringe" standards.

* PAHs used for the validation of HPLC fractionation.





Table 3. Limits of quantification for the analysis of PAHs by GC/C/IRMS.

	Quantity of injected carbon required to obtain a 1 V signal	Quantity of injected compound required to obtain a 1 V signal
Phenanthrene	77 ng	82 ng
Fluoranthene	100 ng	105 ng
Pyrene	90 ng	95 ng
Benz(a)anthracene	125 ng	132 ng
Chrysene	125 ng	132 ng
Benzo(k)fluoranthene	165 ng	173 ng
Benzo(a)pyrene	165 ng	173 ng
Indeno(1,2,3-c,d)pyrene	325 ng	340 ng
Benzo(ghi)perylene	500 ng	523 ng



	HARDWOOD								COAL		
	Cork oak twigs 1	Rosemary	Green oak	Kermès oak	Cork oak	Mediterranean False-Brome	Heather 1	Alep pine 1	Alep pine 2	Cypress 1	Eucalyptus Moroccan coal
Combustion condition	smold	Flame	flame	flame	flame	flame	flame	flame	flame	smold	smold
Naph	0.074	0.001	0.004	0.017	0.103	nd	5.747	0.006	0.824	0.026	1.072
Acy	0.038	Nd	0.015	nd	nd	0.012	0.004	0.093	0.094	0.078	0.004
Ace	0.004	0.004	0.003	nd	0.005	0.001	nd	0.016	0.023	0.012	nd
FI	0.073	0.046	0.034	0.010	0.014	0.016	0.019	0.083	0.122	0.110	0.003
Phen	1.385	1.164	0.564	0.188	0.179	0.236	0.449	1.286	2.971	0.524	0.047
Anth	0.323	0.226	0.132	0.021	nd	0.043	0.072	0.212	0.479	0.123	nd
DBT	0.003	0.003	0.002	nd	nd	nd	nd	0.006	0.005	0.002	nd
Fluo	1.340	1.661	0.745	0.589	0.452	0.433	0.691	1.323	1.528	0.259	0.246
Pyr	1.450	1.771	0.778	0.655	0.530	0.427	0.737	1.442	1.548	0.221	0.280
BghiF	0.248	0.360	0.151	0.222	0.156	0.088	0.136	0.307	0.269	0.036	0.380
CcdP	0.392	0.670	0.218	0.293	0.215	0.125	0.213	0.480	0.431	0.059	0.579
BcP	0.103	0.181	0.056	0.086	0.057	0.032	0.053	0.111	0.145	0.013	0.096
BaA	0.410	0.777	0.214	0.340	0.285	0.137	0.250	0.377	0.454	0.059	0.634
Chrys	0.437	1.146	0.278	0.514	0.380	0.143	0.343	0.392	0.547	0.075	0.795
BbF	0.206	0.306	0.126	0.229	0.188	0.090	0.135	0.218	0.255	0.032	0.511
BkF	0.115	0.174	0.062	0.107	0.086	0.045	0.067	0.128	0.129	0.014	0.249
BjF	0.169	0.278	0.092	0.165	0.118	0.061	0.105	0.183	0.203	0.022	0.336
BaF	0.119	0.184	0.068	0.113	0.091	0.054	0.071	0.115	0.141	0.018	0.208
BeP	0.147	0.223	0.094	0.174	0.131	0.056	0.101	0.161	0.175	0.023	0.389
BaP	0.292	0.431	0.164	0.271	0.208	0.102	0.176	0.318	0.307	0.035	0.581
Per	0.049	0.068	0.028	0.045	0.035	0.018	0.028	0.053	0.051	0.006	0.092
DBacA	0.021	0.031	0.015	0.020	nd	0.009	0.013	0.022	0.023	0.003	0.032
IP	0.152	0.235	0.093	0.151	0.092	0.058	0.090	0.184	0.165	0.020	0.281
DBahA	0.022	0.037	0.015	0.020	nd	0.007	0.013	0.021	0.023	0.003	0.034
BghiP	0.159	0.248	0.109	0.167	0.110	0.056	0.105	0.213	0.181	0.023	0.389
Antha	0.094	0.138	0.067	0.069	0.053	0.034	0.059	0.113	0.104	0.014	0.116
DBbkF	0.018	0.036	0.013	0.008	nd	0.011	0.012	0.026	0.025	0.004	nd
DalP	0.043	0.064	0.024	0.036	nd	0.018	0.034	0.043	0.048	0.002	0.054
DaeP	0.020	0.026	0.012	nd	nd	0.006	0.008	0.021	0.023	0.002	nd
Cor	0.073	0.135	0.056	0.117	0.117	0.029	0.061	0.125	0.096	0.013	0.199
DahP	nd	Nd	nd	nd	nd	nd	nd	nd	0.006	Nd	nd
ΣPAH _{part}	7.98	10.62	4.23	4.63	3.60	2.35	9.79	8.08	11.40	1.83	7.61
OC ($\mu g m^{-3}$)	3902	1318	3395	515	304	2298	1126	2923	3433	12416	121
EC (µg m ⁻³)	67	165	188	57	16	22	174	223	226	390	< 1

Table 4. PAH concentrations in $PM_{2.5}$ from wood combustion (mg g⁻¹ of OC).

n.d. = non detected compounds numbers are the identification numbers of samples.

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PAH contribution (%)	USA woods Fine et al. (2001, 2002) Min–Max	Mediterranean Woods this study Min–Max
BaA	13.4–19.4	13.8–18.5
Chrys	13.7–20.3	15.8–27.3
BbF	6.3–10.4	7.3–11.1
BkF	8.0–11.3	3.3–5.4
BjF	2.4-4.6	5.3–7.5
BeP	4.8–6.6	5.3–8.4
BaP	8.9–11.4	9.6–12.8
Per	1.0–1.8	1.5–2.2
IP	6.9–11.8	5.1–7.4
BghiP	4.0–5.3	5.7–8.6
Antha	0.9–2.0	2.5–5.9
DahA	0.4–0.9	0.0–1.1
Cor	8.6–18.7	2.8–6.5

Table 5. PAH molecular fingerprints of wood combustion emissions from USA and Mediterranean Basin.



Table 6. PAH diagnostic ratios of wood combustion.

Wood species	[Phen]/ [Phen]+[Anth]	[Fluo]/ [Fluo]+[Pyr]	[IP]/ [IP]+[BghiP]	[BaA]/[BaP]	[BghiP]/[IP]	[Pyr]/[BaP]	[BaP]/[BghiP]	
Cork oak-twigs 1	0.81	0.48	0.49	1.40	1.05	4.97	1.83	this study
Cork oak-twigs 2	0.79	0.50	0.49	1.58	1.02	4.32	2.01	(Mediterranean
Juniper Tree ^a	0.73	0.48	0.49	1.80	1.05	3.81	1.69	area)
Rosemary	0.84	0.48	0.49	1.80	1.06	4.11	1.74	
Alep Pine 1 ^a	0.86	0.48	0.46	1.18	1.16	4.54	1.49	
Green oak	0.81	0.49	0.46	1.31	1.17	4.74	1.51	
"kermès" oak	0.90	0.47	0.47	1.26	1.11	2.42	1.63	
Cork oak	-	0.46	0.46	1.37	1.20	2.55	1.90	
Cypress 1 ^a	0.81	0.54	0.47	1.66	1.13	6.25	1.57	
Cypress 2 ^a	0.74	0.50	0.48	1.64	1.09	5.04	1.81	
Mediterranean	0.85	0.50	0.51	1.34	0.97	4.17	1.82	
False-Brome								
Eucalyptus	-	0.47	0.42	1.09	1.38	0.48	1.49	
Moroccan coal								
Heather 1	0.86	0.48	0.46	1.42	1.16	4.20	1.68	
Heather 2	0.72	0.50	0.49	1.62	1.02	4.31	1.73	
Alep pine 2°	0.86	0.50	0.48	1.48	1.10	5.04	1.69	
Eucalyptus globulus	0.97	0.51	0.57	1.26	0.76	4.17	1.62	Gonçalves et al. (2008)
Pinus pinaster ^a	0.85	0.64	0.35	1.18	1.89	1.53	0.73	(Portugal)
Quercus suber	0.28	0.78	0.10	1.98	9.47	8.84	0.25	
Acacia longifolia	-	0.54	0.28	1.46	2.60	2.41	0.22	
Pine wood ^a	0.90	0.44	0.21	1.02	3.68	2.56	1.94	Rogge et al. (1998)
Oak wood	0.84	0.43	0.27	0.91	2.77	2.30	1.77	(USA)
Red maple	-	0.43	0.71	1.61	0.40	3.41	1.88	Fine et al.
Red oak	-	0.45	0.61	1.60	0.64	4.07	(2001, 2002) (USA)2.20	
Yellow poplar	0.83	0.46	0.59	1.45	0.70	3.55	2.02	
White ash	-	0.43	0.63	1.18	0.60	2.40	2.13	
White pine ^a	0.83	0.54	0.64	1.41	0.55	5.06	2.53	
Hemlock ^a	0.78	0.48	0.65	1.85	0.55	4.41	2.60	
Balsam fir ^a	0.78	0.48	0.63	1.81	0.58	4.43	2.26	
Loblolly pine ^a	0.86	0.52	0.67	1.64	0.48	5.36	2.47	
Slash pine ^a	0.74	0.51	0.71	1.67	0.40	9.00	1.96	
Bamboo	0.70	0.53	-	1.00	-	1.80	-	Oros et al.
Sugarcane	0.77	0.53	-	2.15	-	5.85	-	(2006) (Malaysia)
Chestnut oak	0.83	0.51	-	0.85	-	1.08	-	Wang et al.
Chinese evergreen	0.71	0.50	-	0.95	-	1.24	-	(2009) (China)
chinkapin								
Common aporusa ^b	0.76	0.60	-	0.77	-	2.70	-	
Cape jasmine ^b	0.71	0.56	-	0.69	-	1.33	_	
Moaning myrtle ^b	0.72	0.54	-	0.93	-	1.60	-	

^a Softwoods

^b Shrubs numbers are the identification numbers of samples.



¹³ C (‰)	Cork oak twigs (1)	Cork oak twigs (2)	Juniper Tree	Rosemary	Alep pine (1)	Alep pine (2)	Heather (Heather 1) (2)
Phen	-29.7 ± 0.2	-27.6 ± 0.1	-	-26.3 ± 0.1	-25.3 ± 0.2	-25.5 ± 0.3	-24.9 ± 0	$.1 -24.4 \pm 0.3$
Fluo	-30.3 ± 0.9	-28.7 ± 0.1	-30.0 ± 0.4	-27.3 ± 0.4	-26.3 ± 0.1	-26.8 ± 0.3	-25.8 ± 0	$.5 - 25,8 \pm 0.3$
Pyr	-29.4 ± 0.4	-28.4 ± 0.1	-27.8 ± 0.2	-26.9 ± 0.2	-26.3 ± 0.2	-26.0 ± 0.2	-24.7 ± 0	$-25,0\pm0.3$
BaA	-28.9 ± 0.2	-28.1 ± 0.1	-27.1 ± 0.3	-26.9 ± 0.4	-	-25.9 ± 0.4	-24.4 ± 0	.2 -24,1 ± 0.2
Chrys	-29.0 ± 0.1	-30.0 ± 0.3	-27.7 ± 0.8	-26.6 ± 0.6	_	-26.7 ± 0.6	-25.0 ± 0	.3 -24,8±0.2
BkF	-29.9 ± 0.1	-29.6 ± 0.3	-	-27.5 ± 0.2	_	-26.6 ± 0.1	-25.7 ± 0	.4 -25,0±0.1
BeP	-29.5 ± 0.3	-28.7 ± 0.1	_	-27.3 ± 0.3	-	-25.7 ± 0.2	-25.1 ± 0	.5 -24,9±0.9
BaP	-29.8 ± 0.1	-29.7 ± 0.5	_	-27.6 ± 0.4	-	-26.4 ± 0.2	-25.5 ± 0	.4 -24,6±0.6
IP	-29.9 ± 0.1	-	_	-	-	-25.4 ± 0.4		$26,0 \pm 0.4$
BghiP	-29.0 ± 0.1	-	-	-	-	-25.6 ± 0.4		- -25,2 ± 0.2
¹³ C (‰	Green o	ak kermès d	oak Cork o	oak Cypres	s Cypres (1)	s Euca (2) Morocca	lyptus Me an coal F	editerranean False-Brome
Phe	en –28.2±0	0.1	_	27.7±	0.8 -28.8 ±	0.4	_	_
Flue	-31.3 ± 0	$-29.6 \pm$	0.1 -30.3 ±	0.7	_	_	_	-32.0 ± 0.1
Pvr	-30.4 ± 0	$-28.7 \pm$	0.1 -29.0 ±	0.4 –27.7±	0.1	28.	7 ± 0.3	-31.4 ± 0.1
Ba/	A −29.4±0	$-28.7 \pm$	0.3 –28.7 ±	0.3 –27.5±	0.4 –28.3±	0.5 –28.	4 ± 0.4	-30.2 ± 0.2
Chr	ys –29.8±0	$-29.1 \pm$	0.4 –29.4 ±	0.4 –28.3±	0.2 -30.0 ±	0.7 –28.	8±0.5	-31.7 ± 0.3
BkF	-30.0±0	$-29.9 \pm$	0.3 –29.3 ±	0.5	_	_	_	-31.3 ± 0.3
BeF	-28.9 ± 0	0.1 –29.1 ±	0.3 –29.1 ±	0.5	_	_	_	-30.9 ± 0.1
BaF	-30.3 ± 0	0.2 –29.8 ±	0.3 –29.4 ±	0.4	_	_	_	-31.2 ± 0.3
IP		_	_	-	_	-	_	_
Bgh	ηiΡ	_	_	_	_	_	_	_

Table 7. Molecular isotopic compositions of PAHs.



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Fig. 2. Molecular fingerprints of particulate-PAHs from wood combustion.





Fig. 3. Stable-carbon isotopic compositions of particulate-PAHs from different wood combustions compared with their molecular fingerprints.





Fig. 4. Comparison of molecular isotopic compositions of particulate-PAHs from wood combustion with results from the literature.

