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Secondary Organic Aerosol formation from phenolic compounds in the absence of NO_x

S. Nakao^{1,2}, C. Clark^{1,2}, P. Tang^{1,2}, K. Sato^{2,*}, and D. Cocker III^{1,2}

¹University of California, Riverside, Department of Chemical and Environmental Engineering, Riverside, California, USA ²College of Engineering – Center for Environmental Research and Technology (CE-CERT), Riverside, California, USA

currently at: National Institute for Environmental Studies, Japan

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Correspondence to: D. Cocker III (dcocker@engr.ucr.edu)

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Abstract

SOA formation from benzene, toluene, m-xylene, and their corresponding phenolic compounds were investigated using the UCR/CE-CERT Environmental Chamber to evaluate the importance of phenolic compounds as intermediate species in aromatic

- ⁵ SOA formation. SOA formation yield measurements coupled to gas-phase yield measurements indicate that approximately 20% of the SOA of benzene, toluene, and *m*-xylene could be ascribed to the phenolic route. The SOA densities tend to be initially as high as approximately 1.8 g/cm³ and eventually reach the range of 1.3–1.4 g/cm³. The final SOA density was found to be independent of elemental ratio (O/C) indicating
- that applying constant density (e.g., 1.4 g/cm³) to SOA formed from different aromatic compounds is a reasonable approximation. Results from a novel on-line PILS-ToF (Particle-into-Liquid Sampler coupled with Agilent Time-of-Flight) are reported. Major signals observed by the on-line/off-line ToF include species consistent with bicyclic hydroperoxides. To the authors' best knowledge, this is the first possible detection of bicyclic hydroperoxides in aromatic SOA.

1 Introduction

Secondary organic aerosol (SOA) is formed from oxidative processing of volatile organic compounds in the atmosphere. SOA has been suggested to contribute to climate change (IPCC, 2007; Kanakidou et al., 2005), adverse human health effects (Davidson

- et al., 2005; Pope and Dockery, 2006), and a reduction in visibility (Eldering and Cass, 1996). Previous researchers have estimated approximately 70% of organic aerosols are secondary in nature (Hallquist et al., 2009 and references therein). Aromatic hydrocarbons comprise ~20% of nonmethane hydrocarbons in the urban atmosphere and are considered to be one of the major precursors to urban SOA (Calvert et al., 2002).
- ²⁵ A number of studies have investigated gas-phase photooxidation of aromatic hydrocarbons (e.g., Calvert et al., 2002; Olariu et al., 2002; Volkamer et al., 2002; Takekawa



et al., 2003; Johnson et al., 2004, 2005; Coeur-Tourneur et al., 2006; Arey et al., 2009). Previously identified first generation products of aromatic compound photooxidation explain approximately 50% of carbon balance (Calvert et al., 2002). Although multi-generational reactions have been suggested to contributed to aromatic SOA formation (Hurley et al., 2001; Ng et al., 2007; Sato et al., 2007), the extent of the contribution

6 (Hurley et al., 2001; Ng et al., 2007; Sato et al., 2007), the extent of the contribution from the second or further reaction products to SOA is poorly understood.

Phenolic compounds are one of the major first generation products of OH reaction with aromatic hydrocarbons (Calvert et al., 2002). Previous studies suggested the major gas-phase reaction products from OH reaction of phenolics are dihydroxy com-

- pounds (e.g., 80% catechol formation from phenol, Olariu et al., 2002). However, the mechanisms of SOA formation from phenolic compounds are highly uncertain. In addition, since phenolic compounds are significant products in wood smoke (Schauer et al., 2001; Simoneit, 1999; Hawthorne et al., 1989; Hawthorne et al., 1992), the reaction mechanism of phenolic compounds leading to SOA formation is of a great interest (Chang and Thompson, 2010; Crasiaan, 1084; Sup et al., 2010; Jinuma et al., 2010;
- ¹⁵ (Chang and Thompson, 2010; Grosjean, 1984; Sun et al., 2010; linuma et al., 2010; Henry et al., 2008; Coeur-Tourneur et al., 2010a, b).

 NO_x concentration impacts SOA formation from aromatic hydrocarbons (Song et al., 2005, 2007; Ng et al., 2007). SOA formation under low NO_x conditions has been observed to be higher, which is generally ascribed to increased contribution of organic

hydroperoxides (ROOH) (Song et al., 2007; Ng et al., 2007). This study focuses on low NO_x conditions where ROOH species are expected to strongly contribute to SOA formation. It evaluates the importance of phenolic compounds as intermediate species of aromatic SOA formation.



2 Experimental

2.1 Smog chamber

Most of the experiments were conducted in the UC Riverside/CE-CERT environmental chamber described in detail in Carter et al. (2005). In short, this facility con-

sists of dual 90 m³ Teflon[®] reactors suspended by rigid frames in a temperature controlled enclosure (27 ± 1 °C) continuously flushed with purified air. The top frames are slowly lowered during the experiments to maintain a slight positive differential pressure (0.03"H₂O) between the reactors and enclosure to minimize dilution and possible contamination of the reactors. 272 115 W Sylvania 350 black lights are used as the light source for all the experiments reported herein.

Some of the results of dimethylphenols (DMPs) were acquired in the CE-CERT mezzanine chamber. The CE-CERT mezzanine chamber is a $2.5 \text{ m} \times 3 \text{ m} \times 7.8 \text{ m}$ enclosure covered with reflective aluminum sheet and is illuminated with 170 of 40 W blacklights with peak intensity at 350 nm (SYLVANIA, 350 BL) with the NO₂ photolysis rate

of 0.6 min⁻¹. Within this enclosure is a 12 m³ volume 2 mil FEP Teflon[®] film reactor. A minimum of 1 m space between the reactor surface and blacklights avoids excessive heating at the surface of the film. Six fans are used to mix the air inside the enclosure with room air to minimize heating in the enclosure. Prior to each experiment, the bag is flushed by pure air (Aadco 737 series air purification system (Cleves, Ohio)) until the background particle concentration is below detection (<0.2 cm⁻³).

2.2 Gas and particle analysis

The Agilent 6890 Gas Chromatograph – Flame Ionization Detector was used to measure concentrations of reactants and products. All phenolic compounds were analyzed by a GC equipped with a thermal desorption system (CDS analytical, ACEM9305, Sorbent Tube MX062171 packed with Tenax-TA/Carbopack/Carbosieve S111). Aromatic hydrocarbon measurements were calibrated using a dilute gas



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cylinder (SCOTT-MARIN, Inc.) or by introducing known amount of liquid hydrocarbons into the reactor. Calibration for phenolic compounds was performed by impregnation of the glass cartridges with known quantities of phenolic compounds in acetonitrile. The Ionicon Proton Transfer Reaction – Mass Spectromer (PTR-MS) was also used to observe gas phase compounds with higher proton affinity than water.

Particle size distribution between 28 nm and 700 nm was monitored by a custom built Scanning Mobility Particle Sizer (SMPS) similar to that described by Cocker et al. (2001). Particle effective density was measured with an Aerosol Particle Mass Analyzer (APM, Kanomax) (Ehara et al., 1996) and SMPS in series. The APM is located upstream of the SMPS for improved time resolution and sensitivity (S/N) over the more common configuration of Differential Mobility Analyzer (DMA) – APM (Khalizov et al.,

2009; McMurry et al., 2002; Xue et al., 2009). A detailed description of the APM-SMPS system and data algorithms are described elsewhere (Malloy et al., 2009).

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The HR-ToF-AMS (DeCarlo et al., 2006) was operated in high resolution W mode. ¹⁵ Elemental analysis (EA) was used to determine the atomic ratio (O/C) of non-refractory organic aerosols (Aiken et al., 2008).

The Agilent 6210 Accurate-Mass Time-of-Flight equipped with multimode ionization source for electrospray and atmospheric pressure chemical ionization (ESI/APCI-ToF) is used to obtain accurate mass of analytes. Soft ionization by ESI and/or APCI pro-

- vides lower fragmentation of the analytes compared to electron impact (EI) used for the HR-ToF-AMS or other conventional GC-MS techniques. Mass accuracy is routinely calibrated by standard compounds before analysis and in most cases mass accuracy of less than 5 ppm is achieved. Occasionally higher mass errors (~30 ppm) were observed during sample analysis, which resulted in a consistent shift of mass throughout
- the mass range of the instrument. Since the extent of the shift can be inferred from repeatedly observed ions (e.g., pyruvid acid, reference mixture), formulas were carefully assigned based on tendency of shift and repeat experiments. Filter samples were collected onto Teflo[®] filters (2 μm, 47 mm, PALL Life Sciences) at 25 L/min for 1~4 h. After collection, filters were stored in a freezer until extraction. Extractions were achieved



done by sonicating the filter in 5mL of acetonitrile. The extract volume was reduced under a gentle stream of N₂ until near dryness and reconstituted by 300 μL of acetonitrile/water/acetic acid (50/50/0.1 v). A: Water (0.1 v% acetic acid) and B: acetonitrile were used for LC eluents (B 50%, 0.5 mL/min). Mixed mode ionization (simultaneous APCI and ESI) was used with vaporizer temperature 200 °C, nebulizer pressure 40 psig, corona current 2 μA, fragmentor voltage 100 V. All the Agilent ToF data was acquired in negative polarity mode.

A Particle-Into-Liquid-Sampler (PILS) (Weber et al., 2001; Orsini et al., 2003) was interfaced with the Time-of-Flight to provide a on-line accurate mass analysis of water soluble organic compounds (Bateman et al., 2010). The PILS-ToF system will be described in detail in an upcoming publication (Clark et al., 2011). The negative ESI was used as the ionization method (similar condition as the filter analysis without the corona current).

2.3 Chamber experiments

- ¹⁵ The experimental test matrix is summarized in Table 2. A known volume of high purity liquid aromatic hydrocarbon (All purchased from Sigma-Aldrich: \geq 99% or \geq 99.5%, except 2,4-DMP \geq 98%) was injected through a heated glass injection manifold system and flushed into the chamber with pure N₂. Since phenolic compounds are less volatile than hydrocarbons typically used for chamber experiments, injection into the chambers
- ²⁰ were carefully performed using a heated oven (50~80 °C) through heated transfer line maintained at a temperature higher than oven. The glass manifold inside the oven was packed with glass wool to increase the mass transfer surface area. Since reasonable agreement between the calculated concentration of phenolics (based on injected amount) and observed concentrations were confirmed, loss of phenolics in transfer
- ²⁵ line and to the wall was assumed to be negligible. H_2O_2 50 wt% solution was injected through the same oven system. Since the H_2O_2 solution did not spread through glass wool, the glass wool was processed with an acid/base bath and cleaned by water and acetone which enabled the H_2O_2 solution to spread. Initial H_2O_2 concentration was not



measured, but is estimated to be 1~5 ppm based on amount injected and hydrocarbon decay rate. To investigate the role of organic peroxides, additional high NO experiments were also performed and the chemical composition of SOA was compared to low NO_x condition. Initial ratio of approximately 50/50/500 ppb for phenolic/methyl nitrite/NO
 ⁵ was used to ensure that excess NO was present to suppress RO₂ + HO₂ reaction.

3 Results and discussion

3.1 SOA formation

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SOA yield (*Y*) is defined as the mass of aerosol formed (wall-loss-corrected) (M_o) divided by mass of hydrocarbon reacted (Δ HC)

$$Y = \frac{M_0}{\Lambda HC}$$
(1)

Its dependence on aerosol mass loading has been traditionally parameterized by as shown below: Odum et al. (1996)

$$Y = \sum Y_i = M_o \sum_i \frac{\alpha_i K_i}{1 + M_o K_i}$$

where α_i is the mass-based stoichiometric coefficient for the reaction generating prod-

- uct *i*, K_i is the partitioning coefficient of product *i*. Equations (1) and (2) were applied for the SOA yields obtained (Fig. 1). Particle density of 1.4 g/cm³ was used. 2,4-DMP had the highest SOA formation potential of the three DMP isomers, with a SOA yield approximately twice as high as the other DMP isomers. SOA yield from phenol was higher than benzene.
- ²⁰ SOA from aromatic hydrocarbons formed in low NO_x condition was previously reported to be effectively non-volatile (Ng et al., 2007), in which case SOA yield would be independent of particle mass concentration (flat SOA yield curves). However, for

(2)

benzene, *o-/m*-cresol, and possibly DMPs, the SOA yield was observed to be slightly dependent on particle concentration. Therefore, in this study, constant SOA yield was not assumed and a one product model fit was applied to perform calculation of the contribution of phenolic route in the following section of the contribution of phenolic ⁵ route.

3.2 Formation of phenolic compounds from aromatic hydrocarbons

The formation yields of phenolics from aromatics were obtained. An example for *o*-cresol and *m/p*-cresol formation from toluene is shown in Fig. 2. The cresol formation yields from toluene were calculated from measured cresol by correcting for the further
reaction of cresols with OH. Equations in Atkinson et al. (1982) and rate constants from Calvert et al. (2002) were used for the correction. Phenol, cresols, and DMPs formation yields are in reasonable agreement with previous studies(Table 2) (Smith et al., 1998; Berndt and Boge, 2006; Volkamer et al., 2002; Atkinson, 1989; Klotz et al., 1998; Smith et al., 1999; Atkinson et al., 1991).

15 3.3 Contribution of phenolic route

Contributions of the phenolic route to aromatic SOA formation in the low NO_x system were estimated by combining SOA yield measurement (Fig. 1), phenolic yields, and consumption of phenolics (e.g., Fig. 2). The amount of phenolics (as products) reacted is calculated as the gap between the observed concentration of phenolic compounds and the concentration corrected for the secondary reaction. Phenolic route SOA is calculated by multiplying reacted phenolics and their SOA yield at appropriate mass loading using the phenolic SOA yield curves in Fig. 1. SOA yields from cresol isomers (*o*-, *m*-, and *p*-) were assumed to be the same (Henry et al., 2008). The ratio of 2,4-DMP and 2,6-DMP produced from OH reaction of *m*-xylene was assumed to be the same as that reported by Smith et al. (1999). Formation of 3,5-DMP is assumed to be insignificant (Smith et al., 1999). Contribution of the phenolic route in the no



 NO_x system is summarized in Table 3. The contribution was approximately 20% for benzene, toluene, and *m*-xylene.

3.4 SOA density and elemental ratio

Real-time SOA density measured by the APM-SMPS is shown in Fig. 3. The SOA densities were observed to be initially high (~1.8 g/cm³), which could be due to high density of nucleating species. The final SOA densities were in the range of 1.3–1.4 g/cm³, in reasonable agreement with previous studies (Ng et al., 2007; Sato et al., 2010). Although Bahreini et al. (2005) observed weak correlation between effective density and relative contribution of *m/z* 44 signal to total organic signal of AMS, the final SOA densities were found to be independent of elemental composition of SOA (Fig. 4). Therefore a constant SOA density (1.4 g/cm³) is applied to all the experiments in this study.

3.5 Gas phase analysis

The PTR-MS detected several gas phase products with higher m/z than reactants. A m/z 171 was detected in *m*-xylene experiments, but not in DMPs experiments (Fig. 5).

A *m/z* 171 is consistent with a bicyclic ketone (Wyche et al., 2009; Birdsall et al., 2010). The absence in DMP experiments is consistent with its formation mechanism- DMPs should form analog products with additional -OH (not detected). *m/z* 139 was detected in the cresol experiments. Also *m/z* 153, additional 14 (-CH₂), was confirmed in DMPs and *m*-xylene experiments. These products are tentatively assigned as dihydroxy benzaldehydes (Fig. 6, molecular weight 138) and dihydroxytolualdehydes (molecular weight 152), which can be formed by OH addition to form dihydroxy compounds (Olariu et al., 2002) and subsequent H abstraction of the methyl substitute.

3.6 Particle chemical composition

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Chemical composition of SOA was analyzed by off-line filter analysis. Mass spectrum obtained by ESI/APCI-ToF is shown in Fig. 7. High mass accuracy measurements



enabled determination of empirical formula. Some of the ions in the three spectrums are consistent with $-CH_2$ shift, such as $C_6H_8O_6$ and $C_6H_8O_7$ in phenol SOA (Fig. 7a'). $C_6H_8O_6$ is consistent with bicyclic hydroperoxides formed from phenol. The presence of bicyclic species as gas-phase products of OH reaction with aromatic hydrocarbons

- ⁵ has been suggested experimentally and theoretically (Glowacki et al., 2009; Andino et al., 1996; Huang et al., 2008; Wyche et al., 2009; Birdsall et al., 2010). Johnson et al. (2004, 2005) predicted bicyclic hydroperoxides as one of the predominant aerosol species from aromatic hydrocarbons. To the authors' best knowledge, bicyclic species in particle phase have not been observed previously.
- ¹⁰ The presence of peroxides was investigated by conducting experiments under excess NO. In excess NO condition, the (bicyclic) peroxy radicals rapidly react with NO to form the alkoxy radical, which is suggested to undergo decomposition (Calvert et al., 2002; Atkinson, 2000). The ESI/APCI-ToF mass spectrum of SOA in high NO condition is shown in Fig. 8. $C_6H_8O_6$ (*m*/*z* 175) and $C_6H_8O_7$ (*m*/*z* 191) observed in Fig. 7a were not significantly observed. The highest signal of *m*/*z* 154 is consistent with nitrocatechol which is consistent with the previous model prediction (Johnson et al., 2005; Kelly et al., 2010) that the major product from OH reaction with toluene under high NO_x

condition include methylnitrocatechol formed from methylcatechols, and observation by linuma et al. (2010) that significant methylnitrocatechol was formed from *m*-cresol in high NO_x condition.

The absence of m/z 175 and m/z 191 in Fig. 8 suggests that peroxy radicals are key precursors for these compounds (m/z 175, 191). A possible formation pathway for C₆H₈O₆ formation is shown in Fig. 9.

3.7 PILS-ToF

²⁵ Mass spectrum obtained by PILS-ESI-ToF of SOA formed from OH reaction with phenol, catechol, and *o*-cresol are shown in Fig. 10.

Major signals observed in filter analysis were also observed by the PILS-ToF such as m/z 175 and 191 in phenol SOA mass spectrum (Fig. 10a). Since the PILS-ToF



was operated without a denuder upstream, water soluble gas-phase compounds can potentially be collected by the PILS system. The highest signal in Fig. 10a (mass spectrum when SOA was highest), m/z 109 is catechol ($C_6H_6O_2$), which is consistent with previous gas-phase studies (Olariu et al., 2002). From *o*-cresol, methylcatechol (m/z 123) was observed. Although further reaction mechanisms of catechol species is highly uncertain, a series of signals consistent with -OH addition to catechol was observed ($C_6H_6O_2$, $C_6H_6O_3$, $C_6H_6O_4$, $C_6H_6O_5$), which could imply successive addition of -OH group to the aromatic ring.

3.8 Implication

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The potential role of bicyclic hydroperoxides as a key intermediate species of phenolic SOA formation could be useful for modeling studies to predict aromatic SOA formation. Conventional techniques such as GC-MS can not observe this species, due to weak bonding of the peroxides. The quantification of bicyclic hydroperoxides may improve the current poor understanding of aromatic SOA composition (typically up to 10%).
 ¹⁵ Further structural studies of phenolic SOA using soft ionization techniques and, for example, ion trap mass spectrometry is recommended.

4 Conclusions

Significance of phenolic compounds as intermediate species of aromatic SOA and possible SOA formation mechanism in the absence of NO_x was investigated using the UCR/CE-CERT Environmental Chamber. SOA formation yield measurements coupled to gas-phase yield measurements indicate that approximately 20% of the SOA of benzene, toluene, and *m*-xylene could be ascribed to the phenolic route. Initial SOA densities were as high as approximately 1.8 g/cm³ and eventually reached the range of 1.3–1.4 g/cm³, independent of elemental ratio (O/C) of SOA. Major signals from on-



soluble gas phase products observed by PILS-ToF such as catechol. Major signals observed by ESI/APCI-ToF included species consistent with bicyclic hydroperoxides. To the authors' best knowledge, this is the first detection of bicyclic hydroperoxides in aromatic SOA.

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Table 1. Experimental test matrix (low NO_x).

	Run ID	HC _i , (ppb)	HC _f , (ppb)	ΔHC (µg/m ³)	Mo (µm/cm ³)	SOA yield
\bigcirc	$benzene + H_2O_2 \\ EPA1141A \\ EPA1149A \\ EPA1161A \\ EPA1161B \\ EPA1225A \\ EPA1225B \\ ePa125B \\ $	491 490 1031 528 953 959	434 429 929 478 833 882	182 195 326 160 383 246	25 26 64 21 73 36	0.19 0.18 0.28 0.19 0.27 0.20
P H →	phenol + H ₂ O ₂ EPA1206A EPA1206B EPA1217A EPA1217B	51 52 138 76	21 20 49 28	115 122 338 184	35 40 96 51	0.43 0.45 0.40 0.38
ů.	<i>catechol</i> + H ₂ O ₂ EPA1293A	71 ^a	0	318	89	0.39
\Diamond	$toluene + H_2O_2$ EPA1266A EPA1251B EPA1141B EPA1290B	104 84 85 432	77 59 57 326	100 93 108 397	12 12 15 65	0.17 0.19 0.20 0.23
↓ at	<i>o-cresol</i> + <i>H</i> ₂ <i>O</i> ₂ EPA1251A EPA1252A EPA1252B EPA1266B	75 54 45 101	27 12 8 41	210 184 166 263	74 52 41 87	0.49 0.39 0.35 0.46
ar an	$\begin{array}{l} \textit{m-cresol+} \ \textit{H}_{2}\textit{O}_{2} \\ \text{EPA1255A} \\ \text{EPA1255B} \end{array}$	67 55	18 16	212 174	41 38	0.27 0.31
¢	m-xylene + H ₂ O ₂ EPA1244A EPA1180B EPA1209A EPA1209B EPA1212A EPA1212B EPA1212B EPA1248A EPA1248B	124 234 229 177 114 52 315 121	47 90 77 59 21 6 262 48	333 621 656 509 401 197 229 318	57 134 126 95 91 45 16 48	0.24 0.30 0.27 0.26 0.32 0.32 0.10 0.21



Table 1. Continued.

	Run ID	HC_i ,	HC _f ,	ΔHC	Мо	SOA yield
		(ppb)	(ppb)	(µg/m ³)	(µm/cm ³)	
	2,4-DMP + H ₂ O ₂					
	EPA1238A	83	6	381	199	0.73
Hin L	EPA1238B	62	2	293	140	0.67
	mezzanine chamber runs ^b					
	mez100809	68	2.2	327	222	0.95
~ `	mez100909	131	4.9	628	331	0.74
	mez101009	48	1	234	108	0.65
	mez101109	72	2	349	155	0.62
	mez101309	77	1.9	373	197	0.74
	mez102109	66	1	322	162	0.70
	2,6-DMP + H_2O_2					
	EPA1240A	98	6	457	134	0.41
	EPA1240B	58	2	278	75	0.38
1	mezzanine chamber runs ^b					
	mez081409	64	0	319	79	0.35
	mez081609	66	0	327	86	0.37
	mez081809	67	16	255	24	0.13
	mez100609	64	2.8	305	95	0.44
	mez100709	130	2.7	632	196	0.43
	mez120309	125	0	621	149	0.34
	mez120409	96	0	478	124	0.36
1	3,5-DMP + H ₂ O ₂					
	EPA1243A	90	9	400	83	0.29
	EPA1243B	60	7	262	33	0.17
	mezzanine chamber runs ^b					
HO 🔨 🔪	mez051910	72.24	0	359	61	0.24
	mez052010	85.09	0	422	75	0.25
	mez052310	154.1	0	765	148	0.27
	mez060210	160.5	0	797	174	0.31

^a Initial concentration calculated by amount injected. Injection done with oven temperature ~120 °C.

^b Initial DMPs concentrations calculated based on injection. Final concentration obtained by the PTR-MS.



	Phenolic for	NO _x (ppm)	
	Phenol		
This study		0	
Berndt and Boge (2006)	6	0	
Volkamer et al. (2002)	53	0.002-2	
	o-Cresol	(m+p)-Cresol	
This study	15.8	7.3	0
Atkinson et al. (1989)	20.4 ± 2.7	4.8 ± 0.9	0-10
Klotz et al. (1998)	12.0 ± 1.4	5.9 ± 0.9	0.003–0.3
Smith et al. (1998)	12.3 ± 0.6	5.6 ± 0.4	0.10-0.42
	(2,4 + 2,6		
This study [*]	8.2	0	
Smith et al. (1999)	10.	0.157–1.081	
Atkinson et al. (1991)	21.0 ± 5.6		0–10
Noda et al. (2009)	14.1 ± 2.6		0.01–0.1

Table 2. Phenolic formation yield from OH reaction with benzene, toluene, and *m*-xylene.

* Acquired in mezzanine chamber using PTR-MS.



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Table 3. Estimated contribution of phenolic route to benzene, toluene, and *m*-xylene SOA.

	Benzene	Toluene	<i>m</i> -Xylene
Phenolicroute (%)	$23.5 \pm 4.7^{*}$	$15.8 \pm 3.8^{*}$	$16.9 \pm 3.4 \ (n=4)$

* Error estimated based on repeated m-xylene experiments and phenolic route calculations.



Interactive Discussion

Fig. 1. SOA yield of benzene, toluene, m-xylene, and their phenolic compounds in the absence of NO_v (Dotted lines are one product model fit for phenolic compounds). * SOA yields from cresols isomers are combined. ** Open symbols are data acquired in CE-CERT mezzanine chamber.

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Fig. 3. Time series of the density of SOA formed from aromatic hydrocarbons and phenolic compounds.











Fig. 5. Time series of m/z 171 observed by PTR-MS.









Fig. 7. ESI/APCI-ToF mass spectrums of SOA formed by OH reaction with phenolic compounds (the formulas inside boxes is consistent with bicyclic hydroperoxides formed from phenolic compounds and dihydroxy compounds).





Fig. 8. ESI/APCI-ToF mass spectrum of SOA formed by OH reaction with phenol with the presence of excess NO.





Fig. 9. Possible formation pathway of a bicyclic hydroperoxide $(C_6H_8O_6)$ from OH reaction with phenol in the absence of NO_x.







