

**Second-generation
products of
 β -caryophyllene
ozonolysis**

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**Second-generation products of
 β -caryophyllene ozonolysis are the
dominant contributors to particle mass
concentration**

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Abstract

The production of secondary organic aerosol (SOA) by the dark ozonolysis of gas-phase β -caryophyllene was studied. The experiments were conducted in a continuous-flow environmental chamber for organic particle mass concentrations of 0.5 to $30 \mu\text{g m}^{-3}$ and with ozone in excess, thereby allowing the study of second-generation particle-phase products under atmospherically relevant conditions. The particle-phase products were characterized by an ultra-performance liquid chromatograph equipped with an electrospray ionization time-of-flight mass spectrometer (UPLC-ESI-ToF-MS). Fragmentation mass spectra were used for the structural elucidation of each product, and the structures were confirmed as consistent with the accurate m/z values of the parent ions. In total, fifteen products were identified, three of which are reported for the first time in this study. The structures showed that 9 out of 15 particle-phase products were second generation, including all three of the new products. The relative abundance of the second-generation products was approximately 90% by mass among the 15 observed products. The O:C and H:C elemental ratios of the 15 products ranged from 0.13 to 0.50 and from 1.43 to 1.60, respectively. Fourteen of the products contained 3 to 5 oxygen atoms. A singular product, which was one of the three newly identified ones, had 7 oxygen atoms, including 1 carboxylic group, 2 carbonyl groups, and 3 hydroxyl groups. It was identified as 2,3-dihydroxy-4-[2-(4-hydroxy-3-oxobutyl)-3,3-dimethylcyclobutyl]-4-oxobutanoic acid ($\text{C}_{14}\text{H}_{22}\text{O}_7$). The estimated saturation vapor pressure of this product is sufficiently low (3.3×10^{-13} Pa) that it can contribute to new particle formation in the atmosphere.

1 Introduction

The molecules produced by the oxidation of sesquiterpenes ($\text{C}_{15}\text{H}_{24}$) have a high potential to form condensable products and contribute substantially to the secondary organic material of atmospheric particles, especially in areas of elevated biological

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emissions (Hoffmann et al., 1997; Griffin et al., 1999; Jaoui et al., 2003). Compared to the more extensively studied monoterpenes (C₁₀H₁₆) (Hallquist et al., 2009), sesquiterpenes have five additional carbon atoms. The longer carbon chain decreases the vapor pressures of the oxidation products (Pankow and Asher, 2008). Sesquiterpenes also typically have more than one reactive double bond, thereby providing several points of potential molecular connections for the types of oxygenated organic functional groups that reduce vapor pressure. Under atmospheric conditions, the ozonolysis reactions of sesquiterpenes typically dominate compared to competing reaction pathways, such as those with hydroxyl or nitrate radicals (Shu and Atkinson, 1995). For endo-cyclic alkenes like many sesquiterpenes, ozonolysis reactions (1) favor the formation of carboxylic acid groups and (2) minimize the breaking of the carbon backbone structure (Kroll and Seinfeld, 2008). These reaction features both favor the formation of products that have lower vapor pressures compared to the reaction pathways of hydroxyl or nitrate radicals.

The sesquiterpene β -caryophyllene has received the most attention in the literature (Jaoui et al., 2003; Nguyen et al., 2009; Winterhalter et al., 2009). Its structure, which includes one endo-cyclic and one exo-cyclic double bond, is shown in Fig. 1a. It is emitted by many trees and agricultural plants (Duhl et al., 2008) and in some cases is the dominant sesquiterpene emission, such as for *Citrus* varieties (Ciccioli et al., 1999). Given its prevalence, several studies have focused on its role in the formation of secondary organic aerosol (SOA). The molecular products found in the particle phase following oxidation have been characterized, both for atmospheric sampling (Pio et al., 2001; Jaoui et al., 2007; Parshintsev et al., 2008) and in laboratory experiments (Griffin et al., 1999; Jaoui et al., 2003). Laboratory studies have investigated the particle mass yield, defined as the particle mass concentration after reaction divided by the mass concentration of β -caryophyllene consumed (Grosjean et al., 1993; Hoffmann et al., 1997; Griffin et al., 1999; Jaoui et al., 2003; Lee et al., 2006; Ng et al., 2006). The oxidation reaction mechanisms have also been investigated, especially those related to first-generation products (Jaoui et al., 2003; Nguyen et al., 2009; Winterhalter et al.,

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2009).

First-generation products, some of which are illustrated in panel B of Fig. 1, are those resulting from the reaction of one double bond of β -caryophyllene (Jaoui et al., 2003; Winterhalter et al., 2009). An unreacted double bond remains in these products. Further reactions involving this second double bond produce second-generation products. Several examples are shown in panel C of Fig. 1. The examples selected for panels B and C of Fig. 1 are, by way of conclusion, the ones we detect and report in this study. Compared to their first-generation counterparts, the second-generation products are more oxygenated and therefore have lower saturation vapor pressures. They are therefore expected to make larger relative contributions to particle-phase organic mass concentrations. These oxygenated molecules on a partial molar basis also have greater hygroscopicity, contributing both to water uptake at elevated relative humidity (RH) and cloud condensation nuclei (CCN) activity for supersaturated RH (Hartz et al., 2005; Asa-Awuku et al., 2009).

The formation rates of second-generation products, as well as the associated chemical mechanisms, have been little investigated. For the most part, laboratory studies have been conducted under ozone-limited conditions, meaning that the total concentration of double bonds associated with β -caryophyllene exceeded the stoichiometric ozone equivalence. For example, Kanawati et al. (2008) used excess β -caryophyllene (300 ppbv) compared to ozone (200 ppbv) and observed products containing from 2 to 4 oxygen atoms. Jaoui et al. (2003) used 601 ppbv β -caryophyllene and 640 ppbv ozone (i.e., approximately stoichiometric quantities with respect to the first double bond) and observed products having up to 5 oxygen atoms. In contradistinction to these conditions, in the atmosphere ozone concentrations are in excess and second-generation products are expected to dominate.

In the present study, experiments were conducted for the dark ozonolysis of β -caryophyllene in the continuous-flow Harvard Environmental Chamber (HEC) under conditions of excess ozone. The concentration of ozone was maintained constant by feedback control, and excess ozone facilitated the formation of second-generation ox-

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idation products. Therefore, the conditions of the present study can be considered as ozone-rich compared to those of previous studies. An ultra performance liquid chromatograph, coupled to an electrospray ionization time-of-flight mass spectrometer (UPLC-ESI-ToF-MS), was used to characterize the particle-phase products.

2 Experimental

2.1 Chamber experiments

The dark ozonolysis of β -caryophyllene was conducted in the Harvard Environmental Chamber (HEC) in a continuous-flow steady-state mode. The HEC has been described previously (Shilling et al., 2008; King et al., 2009). For the experiments of this study, reaction conditions were 25 °C and 40% RH, and seed particles of dry quasi-monodisperse ammonium sulfate having an electric mobility-equivalent diameter of 50 nm were used. A liquid solution (1:2500, v/v) of β -caryophyllene (Sigma-Aldrich, $\geq 98.5\%$) in cyclohexane (Sigma-Aldrich, $\geq 99.9\%$) was fed by a syringe pump into a gently warmed glass bulb. A flow of air swept the vapor into the chamber. Cyclohexane served as a scavenger of the OH radicals that are produced by some ozonolysis pathways. The concentration of β -caryophyllene in the chamber prior to reaction (i.e., before ozone injection) was confirmed by gas-phase hydrocarbon measurements using an Ionicon proton-transfer-reaction mass spectrometer (PTR-MS). The total flow rate through the chamber was 21.6 Lpm, corresponding to a mean residence time of 3.6 h.

In a series of experiments, the initial concentration of β -caryophyllene was varied from 1.7 to 46.4 ppbv by adjusting the liquid injection rate from the syringe pump. The ozone concentration was maintained constant by feedback control. We focus on the results of experiments at 50 ppbv. Same products were observed in experiments conducted at 100 and 200 ppbv. The lifetime of β -caryophyllene against ozonolysis (i.e., first-generation reactions on the endo-cyclic double bond) was 70 s based on a bi-

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molecular rate constant of $1.16 \pm 0.43 \times 10^{-14} \text{ molecule}^{-1} \text{ cm}^3 \text{ s}^{-1}$ (Shu and Atkinson, 1995). In turn, the average ozonolysis lifetime of the first-generation products was 7500 s based on an average rate constant of $1.1 \pm 0.4 \times 10^{-16} \text{ molecule}^{-1} \text{ cm}^3 \text{ s}^{-1}$ for the ozonolysis of the exo-cyclic double bond (Winterhalter et al., 2009). Compared to a mean residence time in the chamber of 3.6 h (12 960 s), a small steady-state concentration of β -caryophyllene was therefore expected in the chamber. In agreement, the PTR-MS measurements showed that less than 3% of the β -caryophyllene remained. Much of the reacted hydrocarbon exited the chamber in the particle phase. The organic particle mass concentrations were measured by an Aerodyne high-resolution time-of-flight aerosol mass spectrometer (HR-ToF-AMS) (DeCarlo et al., 2006; Shilling et al., 2008, 2009) and varied from 0.5 to $30 \mu\text{g m}^{-3}$ for the experimental conditions used.

2.2 Sample collection and sample preparation

Aerosol particles were collected for 4 h on quartz filters (Grade QM-A Circles, part number 1851-047, 47 mm, Whatman) by sampling at 5 Lpm. Chamber conditions were at steady state during collection. In front of the filter setup, the particles passed through an ozone scrubber, which consisted of a diffusion tube having an outer annulus filled with ozone destruction catalyst (manganese dioxide/copper oxide, Carus Chemicals, Carulite 200). Samples were stored at 4°C and analyzed by UPLC-ESI-ToF-MS within 3 d. Three consecutive 4-h samples were collected for each experimental condition.

For analysis by UPLC-ESI-ToF-MS, the filters were extracted with a 1:4 (v/v) mixture of HPLC grade acetonitrile (CH_3CN) and dichloromethane in an ultrasonic bath for 15 min. Filter debris was removed by passing the extract through a syringe filter (Spartan, part number 10463100, 13 mm, $0.2 \mu\text{m}$, Whatman). The filtered extract of 5 mL was purged with a gentle flow of ultrapure nitrogen to a volume of less than $100 \mu\text{L}$ and then reconstituted with 1 mL of 0.1% (v/v) formic acid in a mixture of 80:20 (v/v) ultrapure H_2O and HPLC grade methanol (MeOH). The reconstituted solution was passed through a solid-phase extraction (SPE) cartridge (AccuBOND II, part number

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188-1310, ODS-C18, 100 mg, 1 mL, Agilent), which had been preconditioned by 2 mL MeOH and 2 mL H₂O. Inorganic solute (i.e., ammonium sulfate) was removed by washing the SPE cartridges with 1 mL H₂O. The organic compounds were eluted from the cartridges by 1 mL MeOH. A 100- μ L volume of (48 μ M) azelaic acid solution was added as an internal standard.

2.3 UPLC-ESI-ToF-MS analysis

Samples were analyzed by an ultra performance liquid chromatograph (UPLC; Acquity/LCT Premier XE, Waters Micromass, MassLynx 4.1 software) equipped with an electrospray ionization (ESI) interface and coupled to an orthogonal-acceleration time-of-flight mass spectrometer (ToF-MS). The chromatographic conditions are listed in Table S1, including the injection volume, the chromatographic column, the mobile phases, the eluent gradient, and the flow rate. Blank runs with solvent alone and runs with filter blanks both confirmed that the peaks in the chromatograms were not due to artifacts or reactions occurring during sample preparation. The conditions for the operation of the mass spectrometer (i.e., the ESI capillary voltage, the aperture-1 voltage of the ion optics, and so forth) are listed in Table S2 for three modes of ToF-MS measurements.

The three modes included one directed to the determination of the accurate mass of the parent ion and hence its molecular formula, another to the fragmentation of the parent ion for structure determination, and another to the semi-quantification of the products. The recorded mass spectra were corrected for background by subtraction of the baseline mass spectra recorded near the chromatographic peaks. In the accurate-mass mode, the ToF-MS spectral resolution was >12 000, as defined by $m/\Delta m$ where m is the m/z value of a singly charged ion and Δm is the full width at half maximum of the ion peak. Quasi-molecular ions ($[M-H]^-$ in ESI⁻ and $[M+H]^+$ in ESI⁺ modes) as well as adducts with formic acid ($[M+HCOOH-H]^-$, ESI⁻ mode), sodium formate ($[M+NaOOCH-H]^-$, ESI⁻ mode), sodium ($[M+Na]^+$, ESI⁺ mode), and methanol+sodium ($[M+CH_3OH+Na]^+$, ESI⁺ mode) were used for redundant determination of the molecular formulas of the products, where M represents the analyte

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molecule. The same identification approach was used to confirm the empirical formulas of ion fragments. To further confirm the molecular formulas of the products, we substituted acetic acid for formic acid as the additive in the eluent and subsequently identified the acetate adducts ($[M+NaOOCCH_3-H]^-$, ESI⁻ mode) (Table S3). For the mode of semi-quantification, known concentrations of pinic acid in ESI⁻ analysis and pinonic acid in ESI⁺ analysis were used as the calibration compounds, meaning that product concentrations were expressed on pinic-acid or pinonic-acid equivalency basis in the ESI⁻ and ESI⁺ modes, respectively.

3 Results and discussion

3.1 Product identification

3.1.1 Molecular formulas of the products

Figure 2 shows the chromatograms recorded in ESI⁻ and ESI⁺ modes for the particle-phase products of β -caryophyllene ozonolysis. In ESI⁻ mode, products having acidic sites, such as carboxylic acids, are readily ionized via deprotonation to form $[M-H]^-$, while in ESI⁺ mode those having neutral functional groups such as carbonyls are ionized via proton attachment to form $[M+H]^+$ (Kanawati et al., 2008). There are 17 peaks labeled in panels A (ESI⁻ mode) and B (ESI⁺ mode) of Fig. 2, representing 15 detected products. Depending on the functional groups present, some molecules can be ionized in both modes.

For each of the labeled products, the accurate m/z values recorded for the quasi-molecular ions and their corresponding adduct ions are listed in Table 1. Based on the accurate mass, molecular formulas are deduced and are also listed in Table 1. The differences between measured and expected m/z values are less than 5 mDa. A new finding compared to literature is that the product P252-4 has a molecular formula of $C_{14}H_{20}O_4$ (accurate mass of 252.1362). Previous studies assigned a molecular

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formula of $C_{15}H_{24}O_3$ to all isomers observed at a nominal mass of 252 (Jaoui et al., 2003; Winterhalter et al., 2009).

Figure 2 shows that at some retention times chromatographic peaks overlay on one another. Because the ordinate value of Fig. 2 is the sum of measured ion intensities, some deconvolution of the chromatograms is possible by examination of the m/z -resolved chromatograms, shown as extracted ion chromatograms (EICs) in Fig. S1. This additional information is also used by us for product identification.

3.1.2 Structures of the products

Fragmentation of molecular ions by use of increased instrument voltage yields mass spectra that are useful for inferring the molecular structures of the products. Figure 3 shows the fragmentation mass spectra for three products not previously identified in the literature of β -caryophyllene ozonolysis. Figures S2 and S3 show the fragmentation mass spectra observed for products already reported in the literature. For ESI-ionization, fragments were observed at -18 Da (loss of H_2O), -28 Da (loss of CO or C_2H_4), -30 Da (loss of CH_2O), and -44 Da (loss of CO_2 , typically a carboxylic acid). For ESI+ ionization, fragments having consecutive losses of -18 Da (i.e., multiple H_2O loss) were commonly observed. The observed fragmentation mass spectra are in general agreement with those of Kanawati et al. (2008), who used a triple quad-ToF instrument to study the products of β -caryophyllene ozonolysis, both by ESI ionization in negative mode and by atmospheric pressure chemical ionization in positive mode.

From the observed fragmentation mass spectra, we propose the 15 product structures illustrated in Fig. 1 and listed in Table 2. Twelve of the proposed structures confirm those of earlier studies (Jaoui et al., 2003; Kanawati et al., 2008; Winterhalter et al., 2009), and those structures are shown in black in Fig. 1 (panels b and c). In some cases, specific isomeric structures cannot be uniquely attributed to the chromatographic retention times because the fragmentation mass spectra recorded by our methods are similar within the isomer family. In these cases, families such as P252- x , P252- y , and P252- z are used to label the chromatographic peaks in Fig. 2, where x ,

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y, and z are some permutation of 1, 2, and 3 with respect to the specific structures shown in Fig. 1. In comparison, Winterhalter et al. (2009) list six isomers with a molecular weight of 252, namely P2a, P2b, P4, P5, P8 and P9 in Table 5 of that study. Our observed fragmentation mass spectra indicate that the family P252-x, P252-y, and P252-z corresponds to some permutation of P5, P8, and P9 (corresponding to P252-1, P252-3, and P252-2 of our study).

In Fig. 1c, three products P198, P252-4, and P302 not reported previously are shown in red. The mass spectra of these products, as well as the structures proposed to explain the spectra, are shown in Fig. 3. Of special note, product P302, which is 2,3-dihydroxy-4-[2-(4-hydroxy-3-oxobutyl)-3,3-dimethylcyclobutyl]-4-oxobutanoic acid, contains 7 oxygen atoms and is the most oxygenated product reported to date for β -caryophyllene ozonolysis.

In the spectrum of P198 (Fig. 3a), fragments at m/z 153.1286 and 179.1081 correspond to losses of CO_2 and H_2O , respectively, from the quasi-molecular ion ($[\text{M}-\text{H}]^-$, m/z 197.1188), indicating the presence of a carboxylic acid functional group. Compared to the two oxygen atoms in the identified carboxylic acid group, the molecular formula of $\text{C}_{11}\text{H}_{18}\text{O}_3$ indicates the presence of one further oxygen atom. The double-bond equivalency (DBE) is a quantity that includes both double bonds and rings, and the DBE of $\text{C}_{11}\text{H}_{18}\text{O}_3$ is 3, suggesting in addition to the carboxylic acid group the further presence of one ring and a third oxygen atom as a carbonyl group.

For the spectrum of P252-4 (Fig. 3b), the fragment at m/z 207.1390 forms by loss of 44 Da (CO_2) from the quasi-molecular ion ($[\text{M}-\text{H}]^-$, m/z 251.1295), indicating a carboxylic moiety. Loss of H_2O from the quasi-molecular ion forms a fragment at m/z 233.1190 and can occur from either a carboxylic or a carbonyl group. The ion peak at m/z 165.1292 ($\text{C}_{11}\text{H}_{17}\text{O}^-$) suggests a consecutive loss of $\text{C}_2\text{H}_2\text{O}$ from m/z 207.1390 ($[\text{M}-\text{H}-\text{CO}_2]^-$), indicating that a carbonyl group is present in P252-4 as an acetyl group. The fragment at m/z 149.0974 is believed to form via the cleavage of the ring structure.

In the fragmentation spectrum of P302 (Fig. 3c), fragments at 103.0040 and 197.1185 correspond to $\text{C}_3\text{H}_3\text{O}_4^-$ and $\text{C}_{11}\text{H}_{17}\text{O}_3^-$, respectively. The fragment $\text{C}_3\text{H}_3\text{O}_4^-$

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is proposed to form from the α,β -dihydroxyl- γ -keto-carboxylic moiety (Fig. 3c) via a McLafferty-type rearrangement (Grossert et al., 2006). Loss of 44 Da (CO₂) is not observed in the spectrum, possibly due to the presence of a hydroxyl group in the α -position to the carboxylic group (Bandu et al., 2004). Instead, consecutive losses of 2 H₂O and 1 HCHO, a phenomenon which is common in carboxyl acids having multiple hydroxyl and carbonyl groups, result in an ion peak at m/z 235.0954 (formula C₁₃H₁₅O₄⁻) (Warscheid and Hoffmann, 2001).

A comparison of our product assignments with those of Winterhalter et al. (2009) can be found in Table 2. Winterhalter et al. (2009) identified 14 products, labeled P1 through P12b. We have in common 6 products (P1, P4, P5, P8, P9, and P10). Of the remaining 8 products, two (P2a and P2b) are excluded from further discussion because these esters are believed to form at low yields (Nguyen et al., 2009), suggesting that their presence is below our limit of detection. The four first-generation products P3, P6, P7, and P11 are anticipated based on the mechanism of Winterhalter et al. (2009) (cf. Fig. 11 therein), yet in our data ion peaks with molecular weights of 208 (P3), 250 (P6/P7), and 222 (P11) are not present, possibly due to low yields of those products (Nguyen et al., 2009) or alternatively due to our detection methods. The second-generation product P12b with molecular weight 210 is also not present in our data. On the other hand, compared to Winterhalter et al. (2009), our data summarized in Table 2 indicate eight additional second-generation products.

3.2 Product formation

Mechanistic studies in the literature of β -caryophyllene ozonolysis have focused mainly on the first-generation products formed by reactions at the endo-cyclic double bond (Jaoui et al., 2003; Winterhalter et al., 2009). Product formation proceeds in large part along one of four reaction competing channels. The four channels are via (1) a stabilized Criegee intermediate (SCI), (2) a vinyl hydroperoxide (VHP), (3) isomerization (ISO), or (4) a dioxirane intermediate to esters or acids (ester channel). Section 5 of the supporting material describes each pathway in greater detail, and the reaction chan-

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nels of β -caryophyllene ozonolysis that lead to the first-generation products shown in Fig. 1b are illustrated in detail in Figs. S4, S5, and S6.

In the case of excess ozone, as in the present study, the first-generation products react further at the second (i.e., exo-cyclic) double bond. The proposed pathways that lead to the formation of the second-generation products detected in the particle phase are shown in Fig. 4. Ozonolysis of the first-generation product P236 along SCI, VHP, and ISO channels results in the second-generation products P238, P252-4, and P198, respectively (Scheme A). A detailed reaction scheme is given in Fig. S7. Ozonolysis of the first-generation products P252-2 and P252-3 via SCI and VHP channels forms the second-generation products P254-2, P254-3, P270-2, and P270-3 (Schemes B and C, respectively). A detailed reaction scheme is given in Fig. S8. The formation of the second-generation products P270-1 and P302 is proposed via VHP and ISO channels from P252-5 and P252-1, respectively (Schemes D and E, Fig. 4). Detailed reaction schemes are given in Fig. S6. The mechanism of P302 formation (Scheme B, Fig. S6) follows multiple VHP or ISO channels to introduce several hydroxyl groups into the molecule. In addition, the six identified first-generation products can in principle form many more second-generation products than represented by the schemes of Fig. 4; only pathways to observed products are shown in the figure. Pathways to other products might occur in low yields (Nguyen et al., 2009), some of the products might not partition favorably to the particle phase, or alternatively some of those products might not be detected by our methods.

The sum of the surrogate concentrations of the identified products detected by the UPLC-MS analysis exceeds the total organic particle mass concentrations measured by the HR-ToF-AMS that was also sampling the chamber outflow (Chen and Martin, 2010). The surrogate approach using a pinic-acid and pinonic-acid equivalency basis for the UPLC-MS analysis is believed to introduce an error of $\pm 30\%$ (Jaoui et al., 2003). In addition, a positive sampling artifact by filter methods can be as high as 60% (Kim et al., 2001; Kirchstetter et al., 2001; Mader and Pankow, 2001). Overestimates by the UPLC-MS method compared to the AMS method are in agreement once these

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uncertainties are taken into account. The comparison therefore suggests that a substantial fraction of the total organic particle mass concentration is represented by the 15 identified products.

3.3 Product volatilities

The reversible partitioning of an organic molecule between the gas and particle phases of an aerosol is regulated to first approximation by the molecule's saturation vapor pressure P_{vap} (Seinfeld and Pankow, 2003). In this study, we used the *SPARC Performs Automated Reasoning in Chemistry* (SPARC) on-line calculator (Hilal et al., 2003) to estimate vapor pressures for molecular structures. Results are listed in Table 2. Saturation vapor concentrations C^* are also listed in Table 2 using the ideal gas law at 298 K to convert from vapor pressures. In Fig. 5, values of P_{vap} and C^* are plotted against the O:C elemental ratios of β -caryophyllene and its ozonolysis products. Black lines show the chemical links from first- to second-generation products.

The first-generation product P254-1 (β -caryophyllinic acid, $C^* = 1.3 \times 10^{-3} \mu\text{g m}^{-3}$) is expected to have a strong presence in the particle phase and has been used for ambient samples as an atmospheric tracer of β -caryophyllene SOA (Jaoui et al., 2007; Hu et al., 2008). Three of the six first-generation products (i.e., P236, P252-2, and P252-3) have relatively high saturation vapor concentrations (i.e., $C^* > 100 \mu\text{g m}^{-3}$). They are therefore predicted to partition dominantly to the gas phase and be minor contributors to the particle phase for the organic particle mass concentrations of 0.5 to $30 \mu\text{g m}^{-3}$ employed in the present study. Even with their high C^* values, however, these products still do partition to some extent to the particle phase, as evidenced in this study's measurements and as explained thermodynamically by the effects of an organic mixture on the activities of the constituent organic species (i.e., having the effect of reducing the vapor pressure of individual species) (Pankow, 1994). In comparison to two first-generation products of low volatility (P252-5 and P254-1), four second-generation products (P270-1, P270-2, P270-3, and P302) have $C^* < 1 \mu\text{g m}^{-3}$, and these products are therefore expected to partition mainly to the particle phase.

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The mass fractions of first- and second-generation products are shown in Fig. 6 for increasing organic particle mass concentration from 0.5 to $30\ \mu\text{g m}^{-3}$. Across this range, the second-generation products are dominant by a factor of approximately 10. Toward the low-end of the concentration range, the second-generation products are increasingly dominant. This observation is consistent with the representation in Fig. 5 that second-generation products are typically of lower volatility than their first-generation precursors. Lower volatility compounds increasingly dominate the particle phase at lower mass concentrations (Pankow, 1994).

Bonn and Moortgat (2003) suggested that sesquiterpene-ozone reactions might initiate new particle formation in boreal forests. They estimated a threshold saturation vapor pressure of 1.2×10^{-10} Pa for the nucleating compounds, as represented by the red dashed line in Fig. 5. The product P302 has an estimated saturation vapor pressure of 3.3×10^{-13} Pa, suggesting that P302 is a candidate compound for involvement in atmospheric new particle formation.

4 Conclusions

The results of this study suggest that most first-generation products of sesquiterpene ozonolysis, to the extent that β -caryophyllene is representative of that class, are sufficiently volatile to remain in the gas phase. These products, however, are still reactive toward ozone. Further oxidation products are formed by the ozonolysis of the second double bond, and the present study's results show that many of these second-generation products partition to the particle phase. The results suggest that for most of the troposphere, specifically at times and locations for which ozone-rich conditions prevail with respect to sesquiterpene concentrations, second-generation gas-phase chemistry is expected to drive the sesquiterpene contribution to oxygenated organic material and its subsequent appearance in the particle phase. Some of these products also have very low volatilities, as evidenced in a monomeric second-generation product P302 having 7 oxygen atoms that was identified in this study. Its estimated saturation

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vapor concentration suggests that it is a candidate compound for new particle formation in forested environments. The second-generation compounds also have multiple polar functional groups (mainly OH, C=O, and COOH) that favorably interact with water, suggesting that organic material formed under first-generation conditions (e.g., in a laboratory setting) has significantly different hygroscopic properties, including as related to cloud activation, than organic material formed under atmospherically relevant second-generation conditions. The importance of accurately reflecting atmospheric conditions in laboratory experiments has previously been emphasized for the influence of organic particle mass concentration on CCN activity and particle mass yield (Shilling et al., 2008, 2009; King et al., 2009).

Supplementary material related to this article is available online at:
<http://www.atmos-chem-phys-discuss.net/10/17699/2010/acpd-10-17699-2010-supplement.pdf>.

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Table 1. Particle-phase products of the dark ozonolysis of β -caryophyllene, as detected by UPLC-ToF-MS with electrospray ionization (ESI). Retention times (rt) are listed. Measured and calculated mass-to-charge (m/z) values as well as their difference are shown for both quasi-molecular and adduct ions. Labels ESI⁻ and ESI⁺ represent negative and positive modes, respectively, of electrospray ionization. Product label extenders “x”, “y”, and “z” are as discussed in the caption of Fig. 2.

ESI- product	rt (min)	formula	measured	[M-H] ⁻			[M+HCOOH-H] ⁻			[M+NaOOCH-H] ⁻		
				calculated	Δ (mDa)	measured	calculated	Δ (mDa)	measured	calculated	Δ (mDa)	
P198	2.45	C ₁₁ H ₁₈ O ₃	197.1186	197.1178	0.8	243.1246	243.1232	1.4	265.1064	265.1052	1.2	
P252-4	2.33	C ₁₄ H ₂₀ O ₄	251.1291	251.1283	0.8	297.1351	297.1338	1.3	319.1171	319.1158	1.3	
P252-5^a	3.55	C ₁₅ H ₂₄ O ₃	251.1673	251.1647	2.6	297.1683	297.1702	-1.9	–	–	–	
P254-1	2.51	C ₁₄ H ₂₂ O ₄	253.1453	253.1440	1.3	299.1505	299.1495	1.0	321.1330	321.1314	1.6	
P270-1	2.05	C ₁₄ H ₂₂ O ₅	269.1400	269.1389	1.1	315.1458	315.1444	1.4	337.1278	337.1263	1.5	
P302	3.07	C ₁₄ H ₂₂ O ₇	301.1303	301.1287	1.6	347.1376	347.1342	3.4	369.1186	369.1162	2.4	
ESI+ product	rt (min)	formula	measured	[M+H] ⁺			[M+Na] ⁺			[M+CH ₃ OH+Na] ⁺		
				calculated	Δ (mDa)	measured	calculated	Δ (mDa)	measured	calculated	Δ (mDa)	
P236	3.58	C ₁₅ H ₂₄ O ₂	237.1867	237.1855	1.2	259.1688	259.1674	1.4	291.1933	291.1936	-0.3	
P238	2.61	C ₁₄ H ₂₂ O ₃	239.1654	239.1647	0.7	261.1470	261.1467	0.3	293.1725	293.1729	-0.4	
P252-x^b	2.33	C ₁₅ H ₂₄ O ₃	253.1801	253.1804	-0.3	– ^d	–	–	–	–	–	
P252-y^b	2.95		253.1815		1.1	–	–	–	307.1882	307.1885	0.4	
P252-z^b	3.11		253.1822		1.8	–	–	–	307.1889		0.4	
P254-y	3.65	C ₁₄ H ₂₂ O ₄	255.1599	255.1596	0.3	277.1409	277.1416	-0.7	309.1710	309.1678	3.2	
P254-z	3.71		255.1597		0.1	277.1414		-0.2	309.1709		3.1	
P270-y^c	3.24	C ₁₄ H ₂₂ O ₅	271.1558	271.1545	1.3	293.1361	293.1365	-0.4	325.1609	325.1627	-1.8	
P270-z^c	3.32		271.1553		0.8	293.1363		-0.2	325.1639		1.2	

^a see extracted ion chromatograms (EICs) in Fig. S1

^b fragmentation mass spectra did not allow unique assignments; P252-x in this table and Fig. 2 is tentatively assigned as P252-1

^c tentative assignments of P270-y to P270-2 and P270-z to P270-3 were made based on (1) the plausible fragmentation of P270-y to an ion peak at m/z 239 and (2) the ionization of P270-z under ESI⁻ mode (Fig. S1)

^d adduct ions were not observed

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Table 2. Product labels, molecular formulas, oxygen-to-carbon elemental ratios (O:C), product reaction generation, estimated saturation vapor pressures P_{vap} , and estimated saturation vapor concentrations C^* of molecules produced by the dark ozonolysis of β -caryophyllene and as detected in the present study. Saturation vapor pressures were estimated using the SPARC on-line calculator (<http://sparc.chem.uga.edu/sparc/>) (Hilal et al., 2003). Table entries also include products as labeled by other conventions, including terpene nomenclature (Larsen et al., 1998), the IUPAC system (ACD/Labs, 2010), and that of Winterhalter et al. (2009).

Product	Formula	Terpene nomenclature (Larsen et al., 1998)	IUPAC nomenclature (ACD/Labs, 2010)	Label in Winterhalter et al. (2009)	O:C	Generation	P_{vap} (Pa)	C^* ($\mu\text{g}/\text{m}^3$)
P198	$\text{C}_{11}\text{H}_{18}\text{O}_3$	–	3,3-dimethyl-2-(3-oxobutyl)cyclobutanecarboxylic acid	–	0.27	2	1.2×10^{-3}	9.6×10^1
P236	$\text{C}_{15}\text{H}_{24}\text{O}_2$	β -caryophyllon aldehyde	4-[3,3-dimethyl-2-(3-oxobutyl)cyclobutyl]pent-4-enal	P1	0.13	1	4.2×10^{-2}	4.0×10^3
P238	$\text{C}_{14}\text{H}_{22}\text{O}_3$	β -nocaryophyllon aldehyde	4-[3,3-dimethyl-2-(3-oxobutyl)cyclobutyl]-4-oxobutanal	na	0.21	2	8.5×10^{-3}	8.2×10^2
P252-1 ^b	$\text{C}_{15}\text{H}_{24}\text{O}_3$	β -14-hydroxycaryophyllon aldehyde	4-[2-(4-hydroxy-3-oxobutyl)-3,3-dimethylcyclobutyl]pent-4-enal	P5	0.20	1	4.3×10^{-4}	4.3×10^1
P252-2	$\text{C}_{15}\text{H}_{24}\text{O}_3$	β -5-hydroxycaryophyllon aldehyde	4-[3,3-dimethyl-2-(3-oxobutyl)cyclobutyl]-2-hydroxypent-4-enal	P9	0.20	1	1.2×10^{-3}	1.2×10^2
P252-3	$\text{C}_{15}\text{H}_{24}\text{O}_3$	β -8-hydroxycaryophyllon aldehyde	4-[2-(2-hydroxy-3-oxobutyl)-3,3-dimethylcyclobutyl]pent-4-enal	P8 ^b	0.20	1	1.6×10^{-3}	1.6×10^2
P252-4	$\text{C}_{14}\text{H}_{20}\text{O}_4$	–	3-(4-acetyl-7,7-dimethyl-3-oxabicyclo[4.2.0]oct-4-en-2-yl)propanoic acid	–	0.29	2	2.4×10^{-4}	2.4×10^1
P252-5	$\text{C}_{15}\text{H}_{24}\text{O}_3$	β -caryophyllonic acid	4-[3,3-dimethyl-2-(3-oxobutyl)cyclobutyl]pent-4-enoic acid	P4	0.20	1	8.5×10^{-6}	8.7×10^{-1}
P254-1	$\text{C}_{14}\text{H}_{22}\text{O}_4$	β -caryophyllinic acid	4-[2-(2-carboxyethyl)-3,3-dimethylcyclobutyl]pent-4-enoic acid	P10	0.29	1	1.3×10^{-8}	1.3×10^{-3}
P254-2	$\text{C}_{14}\text{H}_{22}\text{O}_4$	β -8-hydroxynocaryophyllon aldehyde	4-[2-(2-hydroxy-3-oxobutyl)-3,3-dimethylcyclobutyl]-4-oxobutanal	–	0.29	2	2.9×10^{-4}	3.0×10^1
P254-3	$\text{C}_{14}\text{H}_{22}\text{O}_4$	β -5-hydroxynocaryophyllon aldehyde	4-[3,3-dimethyl-2-(3-oxobutyl)cyclobutyl]-2-hydroxy-4-oxobutanal	–	0.29	2	3.1×10^{-4}	3.2×10^1
P270-1	$\text{C}_{14}\text{H}_{22}\text{O}_5$	β -4-hydroxynocaryophyllonic acid	4-[3,3-dimethyl-2-(3-oxobutyl)cyclobutyl]-3-hydroxy-4-oxobutanoic acid	–	0.36	2	6.2×10^{-8}	6.8×10^{-3}
P270-2	$\text{C}_{14}\text{H}_{22}\text{O}_5$	β -4,8-dihydroxynocaryophyllon aldehyde	3-hydroxy-4-[2-(2-hydroxy-3-oxobutyl)-3,3-dimethylcyclobutyl]-4-oxobutanal	–	0.36	2	3.8×10^{-6}	4.1×10^{-1}
P270-3	$\text{C}_{14}\text{H}_{22}\text{O}_5$	β -4,5-dihydroxynocaryophyllon aldehyde	4-[3,3-dimethyl-2-(3-oxobutyl)cyclobutyl]-2,3-dihydroxy-4-oxobutanal	–	0.36	2	7.3×10^{-6}	8.0×10^{-1}
P302	$\text{C}_{14}\text{H}_{22}\text{O}_7$	–	2,3-dihydroxy-4-[2-(4-hydroxy-3-oxobutyl)-3,3-dimethylcyclobutyl]-4-oxobutanoic acid	–	0.50	2	3.3×10^{-13}	4.0×10^{-8}

^a listed but not labeled

^b see Table 5 of Winterhalter et al. (2009); version displayed in Fig. 11 therein is not accurate (R. Winterhalter, personal communication, 2010)

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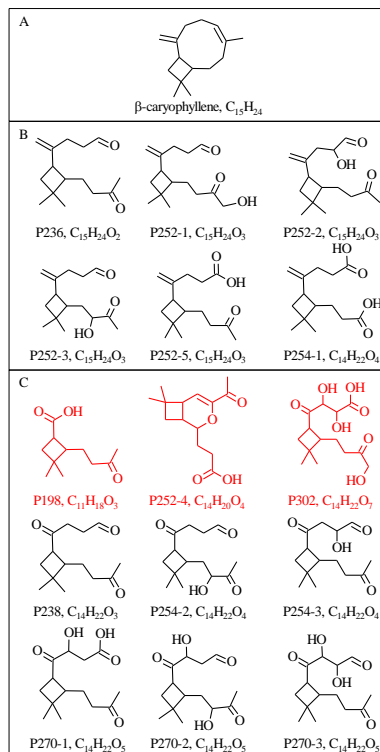


Fig. 1. Molecular structures of **(A)** β -caryophyllene, **(B)** detected first-generation ozonolysis products, and **(c)** detected second-generation ozonolysis products. The fifteen structures represented in panels (B) and (C) are found in the particles after ozonolysis. Twelve of them have been identified previously (Calogirou et al., 1997; Jaoui et al., 2003; Kanawati et al., 2008; Winterhalter et al., 2009). Three of them (viz. P198, P252-4, and P302) are reported herein for the first time and are highlighted in red. Label “Pxxx” represents a product *P* having a nominal mass of xxx. Labels “Pxxx-1”, “Pxxx-2”, and “Pxxx-3” denote distinct molecules (i.e., of different chromatographic retention times) that have the same nominal mass.

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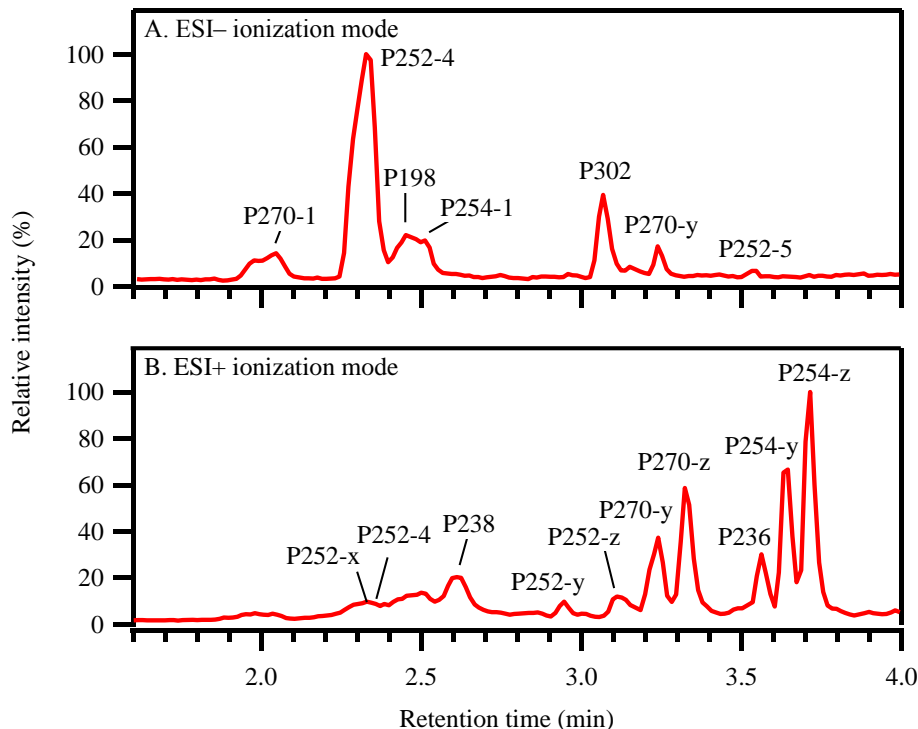


Fig. 2. Chromatograms of the particle-phase products of the dark ozonolysis of β -caryophyllene, as obtained by UPLC-ToF-MS with electrospray ionization (ESI). Ordinate values are the sum of measured ion intensities. Panels A and B show the results from analysis in ESI⁻ and ESI⁺ modes, respectively. Product labels are as described in the caption of Fig. 1. Labels such as “P252-*x*”, “P252-*y*”, and “P252-*z*” have ambiguous molecular assignments, with *x*, *y*, and *z* representing a permutation of 1, 2, and 3.

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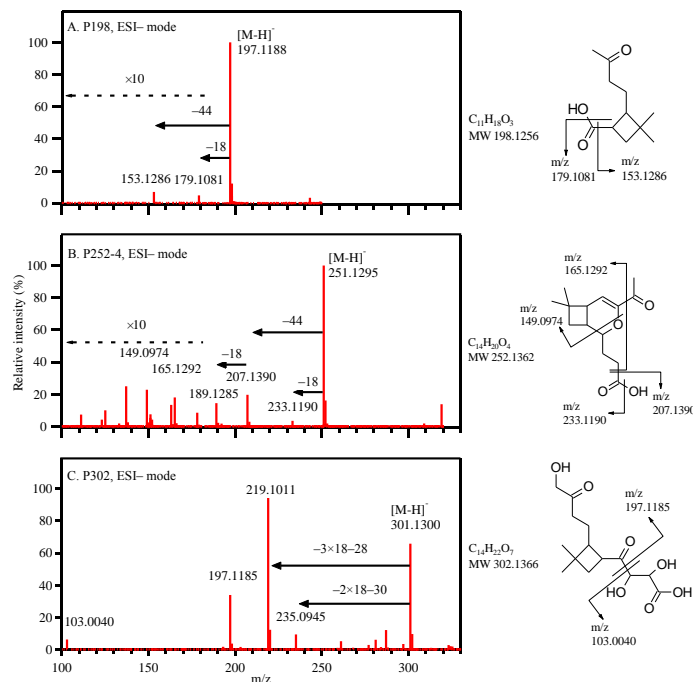


Fig. 3. Fragmentation mass spectra of particle-phase products not previously identified in the literature, including (A) P198, (B) P252-4, and (C) P302. Fragmentation spectra were obtained by increasing the “aperture 1” voltage of the ToF-MS from 5 to 25 V (Table S2), inducing in-source collision and dissociation. Fragmentation achieved in this way is comparable to that obtained by tandem-in-space techniques such as a triple quadrupole analyzer (Gerssen et al., 2008) and provides additional information for structural elucidation. Proposed structure and fragmentation points are shown to the right of each panel. Solid arrows in the main figures indicate the fragmentation pathways that yield the observed ion peaks. Loss of H_2O is indicated by -18 , and loss of CO_2 corresponds to -44 . Dashed arrows indicate regions in which the signal intensities are multiplied by a factor of 10.

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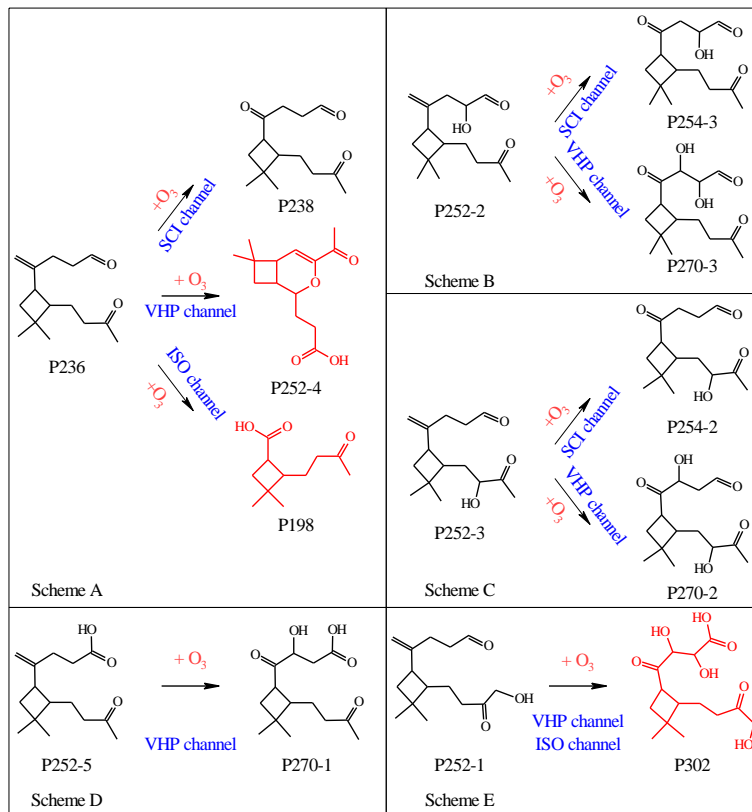


Fig. 4. Ozonolysis pathways from first- to second-generation products. The three new products identified in this study are shown in red. Reaction channels (blue): SCI, stabilization of Criegee intermediate (CI) by collision with gas-phase molecules (M) and subsequent reaction with H₂O followed by loss of H₂O₂ (cf. Fig. S4); VHP, formation and subsequent rearrangement of a vinyl hydroperoxide (cf. Fig. S4); and ISO, formation and subsequent isomerization of a product (cf. Fig. S5).

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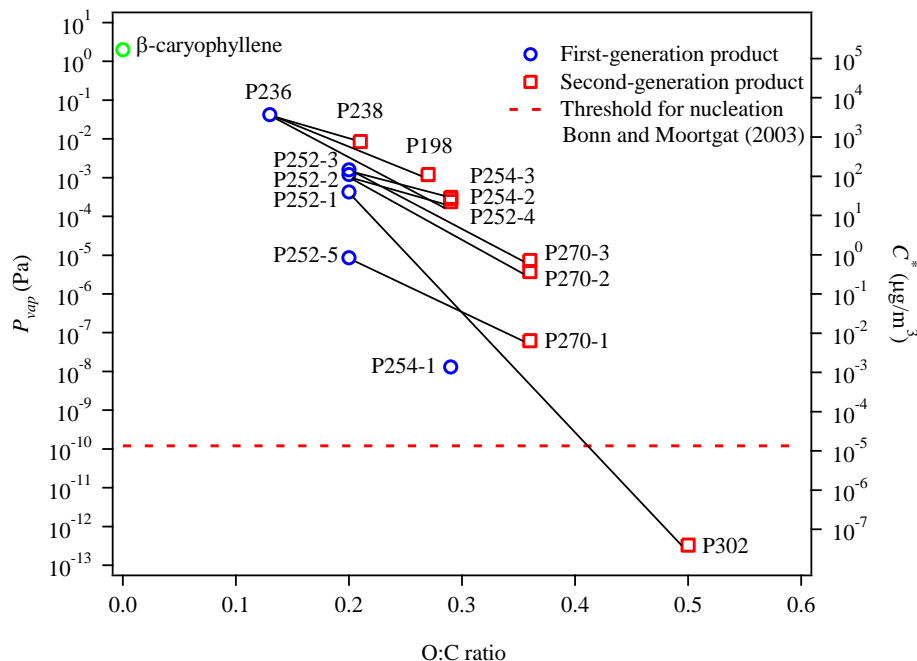


Fig. 5. Estimated saturation vapor pressures P_{vap} and saturation vapor concentrations C^* plotted against the O:C elemental ratio of β -caryophyllene and its ozonolysis products. The dashed red line is the threshold saturation vapor pressure at which new particle formation occurs, as suggested by Bonn and Moortgat (2003). Black lines connect first-generation products to their second-generation counterparts. Although each first-generation product can be expected to produce multiple possible second-generation products (e.g., such as three products from P236), for clarity of presentation only those products actually observed in the particle phase on the basis of the present study's results are represented in the figure.

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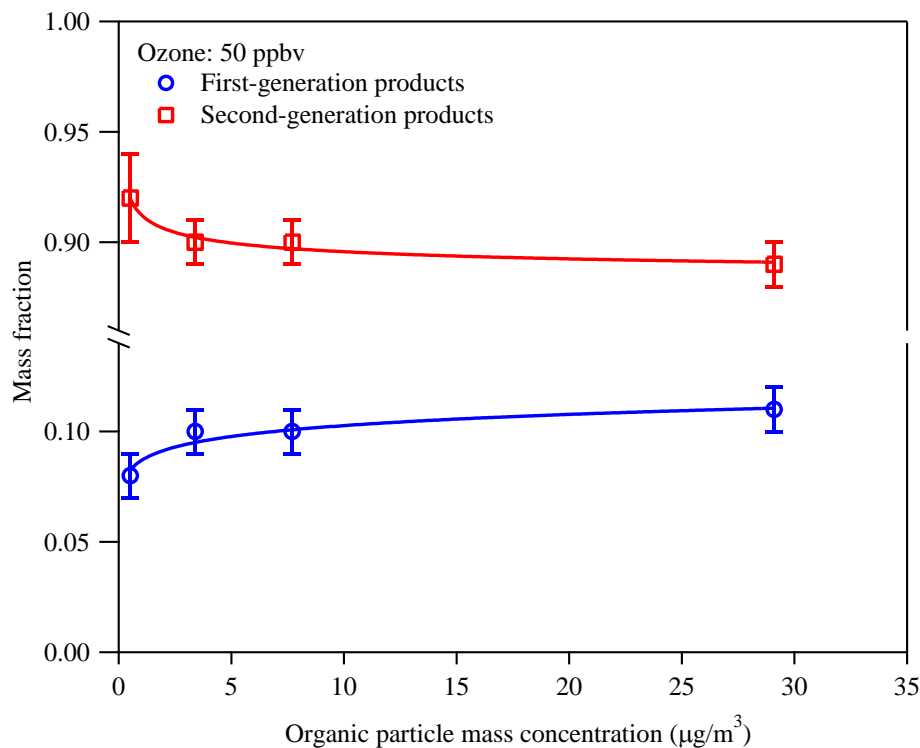


Fig. 6. Mass fractions of first- (blue) and second-generation (red) products for increasing organic particle mass concentration. The ozone concentration was 50 ppbv. Error bars represent ± 1 standard deviation of the mass percentages of three replicate experiments. Lines are shown to guide the eye.

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