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Direct formation of HONO through aqueous-phase photolysis of organic nitrates

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Abstract. Organic nitrates (RONO₂) are secondary compounds whose fate is closely related to the transport and removal of NO_x in the atmosphere. Despite their ubiquitous presence in submicron aerosols, the photochemistry of RONO₂ has only been investigated in the gas phase, leaving their reactivity in condensed phases poorly explored. This work aims to address this gap by investigating, for the first time, the reaction products and the mechanisms of aqueous-phase photolysis of four RONO₂ (i.e., isopropyl nitrate, isobutyl nitrate, α -nitrooxy acetone, and 1-nitrooxy-2-propanol). The results show that the reactivity of RONO₂ in the aqueous phase differs significantly from that in the gas phase. In contrast to the gas phase, where RONO₂ release NO_x upon photolysis, the aqueous-phase photolysis of RONO₂ leads primarily to the direct formation of nitrous acid (HONO or HNO₂), which was confirmed by quantum chemistry calculations. Hence, the aqueous-phase photolysis of RONO₂ represents both a NO_x sink and a source of atmospheric nitrous acid, a significant precursor of \cdot OH and \cdot NO. These secondary radicals (\cdot OH and \cdot NO) are efficiently trapped in the aqueous phase, leading to the formation of HNO₃ and functionalized RONO₂. This reactivity can thus potentially contribute to the aging of secondary organic aerosol (SOA) and serves as an additional source of aqueous-phase SOA.

1 Introduction

Organic nitrates (RONO₂) are secondary compounds, formed through NO_x + volatile organic compound reactions, which play an essential role in the transport and removal of NO_x in the atmosphere. These compounds can have long lifetimes, lasting from a few hours to several days, which allow them to travel to remote regions (Shepson, 1999). During their long-range transport, they can undergo reactions such as gas-phase photolysis and/or \cdot OH oxidation, which can release NO_x back into the atmosphere. As a result, organic nitrates are responsible for a more uniform distribution of NO_x and are thus indirectly responsible for the transport of other pollutants such as O₃ and secondary organic aerosol (SOA; Perring et al., 2013). Furthermore, RONO₂ also participate in NO_x removal from the atmosphere, which can occur either through its deposition to the Earth's surface or by transformation into a less reactive chemical compound, such as nitric acid (Hu et al., 2011; Nguyen et al., 2015). Therefore, its atmospheric reactivity and fate must be considered to accurately predict pollution transport on a regional scale. This is especially important for world regions experiencing decreasing NO_x levels, such as Europe and North America, where the relative importance of RONO₂ in NO_x transport and removal is growing due to the increase in the overall transformation of NO_x into RONO₂ (Romer Present et al., 2020).

RONO₂ are not only present in the gas phase, as some of them have low volatility and can partition into condensed phases. As a result, RONO₂ account for a significant fraction of submicron organic aerosol, ranging from 5% to 77% (Kiendler-Scharr et al., 2016; Ng et al., 2017). The reactivity of RONO₂ in condensed phases may differ from that in the gas phase and may affect the role of $RONO_2$ as NO_x reservoirs. For instance, it is well established that the hydrolysis of tertiary and allylic RONO₂ serves as a fast and permanent sink of NO_x in the atmosphere, as the nitrate group is transformed into nitric acid (Darer et al., 2011; Rindelaub et al., 2015; Hu et al., 2011). However, only a small fraction of RONO₂ (between 9% and 34% for α - and β pinene-related RONO₂) undergo hydrolysis (Takeuchi and Ng, 2018; Wang et al., 2021). Other aqueous-phase reactions are thus to be considered: photolysis and . OH oxidation. Our previous studies have emphasized the significance of aqueous-phase reactivity for atmospherically relevant RONO₂, such as isoprene and terpene nitrates, with intermediate to high water solubilities (González-Sánchez et al., 2021, 2023a). Under typical cloud/fog conditions (liquid water content (LWC) of 0.35 g m^{-3}), the aqueous-phase photoreactivity can act as a major sink (> 50%) for watersoluble RONO₂ ($K_H > 10^5 \text{ M} \text{ atm}^{-1}$), while at a very low LWC $(3 \cdot 10^{-5} \text{ g m}^{-3})$, it can serve as a major sink for very highly water-soluble RONO₂ ($K_H > 10^9 \text{ M} \text{ atm}^{-1}$). Nevertheless, the fate of the nitrate group during these processes is still unknown, and it is uncertain whether this reactivity acts as a NO_x sink or as an additional transport mechanism.

This work intends to address these questions for the aqueous-phase photolysis of RONO₂. While aqueous-phase photolysis is a minor sink when compared with \cdot OH oxidation (González-Sánchez et al., 2023a), the investigation of the photolysis mechanisms is of high importance as it is a first step towards the fundamental understanding of photooxidation pathways of RONO₂.

To explore the aqueous-phase photolysis of RONO₂, the fate of four molecules (i.e., isopropyl nitrate, isobutyl nitrate, α -nitrooxy acetone, and 1-nitrooxy-2-propanol) was experimentally investigated. These RONO₂ served as proxies to understand the fate of the nitrate group. The two alkyl nitrates are simple molecules, simplifying the comprehension of mechanisms related to the nitrate group's reactivity. Furthermore, the other two investigated RONO₂ are polyfunctional; they combine highly relevant functional groups (hydroxy and carbonyl groups) with the nitrate group, allowing for the assessment of their influence on the reactivity. The aqueous-phase photolysis primary and secondary reaction products were identified and quantified, and the fate of the nitrate group was elucidated with support from theoretical calculations. The atmospheric implications of these findings are discussed.

2 Materials and methods

2.1 Experimental setup

The aqueous-phase photolysis experiments were conducted using the experimental setup previously described in detail by González-Sánchez et al. (2023a). Briefly, a 450 cm³ double-wall Pyrex aqueous-phase photoreactor filled with 400 mL of aqueous solution and covered by a quartz lid was used. It was equipped with four apertures for reagent injections and sampling and an optical IDS dissolved oxygen sensor FDO[®] 925 (WTW), which included temperature monitoring. The reactor was thermostated at 298 K and continuously stirred. Irradiation was provided by an arc light source (LOT Quantum Design) equipped with a 1000 W arc xenon lamp. Wavelengths below 290 nm were removed using an ASTM 892 AM1.5 standard filter (see lamp spectra in Fig. S1 in the Supplement along with the liquidphase absorption cross-sections of the investigated RONO₂) (González-Sánchez et al., 2023). A constant distance of 18.4 cm between the lamp and the water surface was carefully maintained in all experiments.

2.2 Photolysis experiments

Prior to each photolysis experiment, the photoreactor was filled with Milli-Q water, and RONO₂ were added. The solution was stirred for 30 min in the dark to ensure complete dilution of the RONO₂. Meanwhile, the lamp was turned on for 10 min to stabilize the light beam. The first aliquot was sampled when the reactor was placed under the light beam, marking the reaction time as 0. Photolysis reactions were performed for 4 to 7 h at 298.0 ± 0.2 K. The specific experimental conditions of all photolysis experiments are appended to Table 1.

During the reaction, aliquots were regularly sampled for offline analyses. The pH of the reaction mixture was measured using a 9110DJWP pH probe (Thermo Scientific). Ultrahigh-performance liquid chromatography ultraviolet (UHPLC-UV) detector analyses were performed to monitor the decay of RONO₂ or to identify and quantify carbonyl compounds after 2,4-dinitrophenylhydrazine (DNPH) derivatization. High-performance ionic chromatography conductivity detector (HPIC-CD) analyses were conducted to quantify HNO₂, HNO₃, and organic acids. At the end of the reaction, the remaining volume was used to perform liquidliquid extraction and gas chromatograph-mass spectrometer (GC–MS) analyses to identify the formed oxidized RONO₂. In experiment 1, the headspace of the reactor was monitored with a NO_x analyzer to investigate the possible formation of these compounds.

2.3 Analytical instruments

2.3.1 UHPLC-UV

An ultrahigh-performance liquid chromatography (UH-PLC) ultraviolet (UV) detector (Thermo Scientific Accela) equipped with a Hypersil Gold C18 column ($50 \times 2.1 \text{ mm}$) with a particle size of 1.9 µm and an injection loop of 5 µL was used to quantify carbonyl compounds after 2,4dinitrophenylhydrazine (DNPH) derivatization and RONO₂.

No.	RONO ₂	[RONO ₂] ₀ (mM)	Reaction time (h)	UHPLC-UV	DNPH*	HPIC	GC-MS	NO_x analyzer
1	Isopropyl nitrate	1.00	4					×
2	Isopropyl nitrate	0.93	7	×	×	×	×	
3	Isopropyl nitrate	1.81	7	×	×			
4	Isopropyl nitrate	1.71	5	×	×	Х		
5	Isobutyl nitrate	0.60	7	×				
6	Isobutyl nitrate	0.59	7	×				
7	Isobutyl nitrate	0.53	7	×			×	
8	Isobutyl nitrate	0.55	7	×	×	×		
9	Isobutyl nitrate	0.49	7	×	×			
10	α -Nitrooxy acetone	1.18	7	×	×	×	×	
11	1-Nitrooxy-2-propanol	0.72	7	×	×	×	×	
12	1-Nitrooxy-2-propanol	0.38	7	×	×	×		

Table 1. Initial conditions and analytical instruments used during the photolysis experiments of four individual RONO₂.

* UHPLC-UV analyses after DNPH derivatization of the sample were used to identify and quantify carbonyl compounds. All experiments were performed at 298.0 ± 0.2 K.

Measurements of RONO₂

A binary eluent of H₂O and CH₃CN was used for all analyses, with a flow rate of 400 μ L min⁻¹. Two gradients were used depending on the polarity of the compounds. For isopropyl nitrate and isobutyl nitrate, the gradient started at H₂O/CH₃CN 80/20 (v/v) and was gradually adjusted to 50/50 (v/v) over 3 min; it was held at this proportion for 1 min and then set back to 80/20 (v/v) within 10 s until the end of the run at 5 min. For more polar compounds, i.e., α nitrooxy acetone and 1-nitrooxy-2-propanol, a similar gradient was employed, but the initial and final proportions were adjusted to H₂O/CH₃CN 90/10 (v/v) to optimize their retention times (rt's). All RONO₂ were detected at their maximum absorbance wavelength at 200 nm (González-Sánchez et al., 2023a).

Calibration curves were linear (as determined by the Mandel test) between 5×10^{-5} and $1 \times 10^{-3} \text{ mol L}^{-1}$ with $R^2 > 0.9995$. Aliquots with expected concentrations higher than $1 \times 10^{-3} \text{ mol L}^{-1}$ were diluted before analyses. The retention times were 0.9, 1.2, 2.4, and 3.3 min for 1-nitrooxy-2-propanol, α -nitrooxy acetone, isopropyl nitrate, and isobutyl nitrate, respectively (Fig. S2). Limits of detection (LODs) were $9 \times 10^{-6} \text{ mol L}^{-1}$ for isopropyl nitrate and $1 \times 10^{-5} \text{ mol L}^{-1}$ for the three other compounds.

Measurements of carbonyl compounds

To derivatize the carbonyl compounds, $500 \,\mu\text{L}$ of the aqueous sample was mixed with $450 \,\mu\text{L}$ of $0.005 \,\text{M}$ DNPH and $50 \,\mu\text{L}$ of $0.1 \,\text{M}$ HCl. The mixture was allowed to react for 24 h to achieve high yields. A specific method was developed for separating and quantifying the formed hydrazones. The gradient, with a flow rate of $400 \,\mu\text{L} \,\text{min}^{-1}$, started from H₂O/CH₃CN 80/20 (v/v) for 1 min and was then gradually adjusted to $30/70 \,(v/v)$ over 6 min; it was held at this pro-

portion for 1 min and then set back to 80/20 (v/v) within 10 s until the end of the run at 9 min.

The resulting hydrazones from formaldehyde, acetaldehyde, acetone, hydroxyacetone, and isobutyraldehyde were identified and quantified at 360 nm. External calibrations were performed to quantify carbonyl compounds with concentrations ranging from 5×10^{-6} to 1×10^{-3} M and $R^2 > 0.9995$. Their retention times were 4.4, 5.1, 5.8, 3.8, and 6.8 min, respectively. LODs were 4.1, 2.1, 1.5, 4.2, and 5.3×10^{-6} M, respectively.

2.3.2 HPIC-CD

The formation of HNO₂ and HNO₃ and organic acids, such as formic acid and acetic acid, was quantified using the Dionex ICS-3000 high-performance ionic chromatography (HPIC) system with a Dionex IonPacTM AG11-HC precolumn (4×50 mm) and a Dionex IonPacTM AS11-HC column (4×250 mm) coupled with a CD25 conductivity detector.

A binary eluent gradient method composed of H₂O and NaOH 0.1 mol L⁻¹ aqueous solution was optimized to separate the formed organic acids at relatively short retention times. At a flow rate of 1 mL min⁻¹, the gradient started at H₂O/NaOH 0.1 mol L⁻¹ 96/4 (v/v) for 10 min, then gradually went to 50/50 (v/v) at 12 min, then went back to 96/4 (v/v) within 1 min and was held at this proportion until the end of the analyses at 25 min. The injection volume was 200 µL, and a constant flow of H₂SO₄ 0.05 M continuously passed through the suppressor at a flow rate of 3 mL min⁻¹.

The retention times of acetate, formate, NO₂⁻, and NO₃⁻ were 5.9, 7.2, 17.3, and 21.9 min, respectively. Calibration curves were optimized to obtain good linearity and low LODs (within the concentrations range expected). LODs were 4.3×10^{-6} , 3.5×10^{-6} , 6×10^{-7} , and 5×10^{-7} M for acetic acid, formic acid, NO₂⁻, and NO₃⁻, respectively.

2.3.3 GC-MS

The Clarus[®] 680 gas chromatograph (GC; PerkinElmer) equipped with the Elite-5MS capillary column (PerkinElmer) with a length of 30 m, diameter of 0.25 mm, and film thickness of 0.25 µm coupled with an AxION[®] iQTTM quadrupole time-of-flight mass spectrometer (MS; PerkinElmer) was used to qualitatively detect and identify oxidized RONO₂ formed during the aqueous-phase photolysis experiments. RONO₂ were extracted and preconcentrated from the remaining solution after the end of each photolysis experiment. The remaining solutions were stored at ~4 °C for up to 48 h before the analyses.

A total of 100 mL of the remaining solution was extracted using 3×20 mL of dichloromethane in a separatory funnel. UHPLC-UV analyses of the aqueous phase before and after the extraction confirmed that all RONO₂ efficiently partitioned to dichloromethane. The extracts were washed with 20 mL of Milli-Q water and were concentrated in a TurboVap II system (Biotage). The concentration workstation used a nitrogen flow at 11 psi and a water bath at 30 °C to evaporate dichloromethane until a 500 µL sample was obtained.

A total of 1 μ L of the concentrated extract was then injected into the GC–MS. The carrier gas was helium at a flow rate of 1 mL min⁻¹. A split of 20/1 was used due to the high concentration of the compounds. The injector temperature was set to increase from 60 to 200 °C within 1 min to prevent thermolysis of RONO₂. The following program was set in the oven: hold the temperature for 10 min at 30 °C, increase it to 300 °C at a rate of 15 °C min⁻¹, and hold it for 10 min at 300 °C before the end of the analyses.

The analytes were detected with a time-of-flight mass spectrometer using electron impact ionization with an electron energy of 70 eV and an ion source temperature of 250 °C. The ion source was turned on 5–7 min after the analysis started in order to avoid the saturation of the source due to the solvent signal. The detector performed full scan measurements from m/z = 30 to 300 amu. The mass-to-charge ratio of the ion NO₂⁺ (m/z = 46), specific to RONO₂, was extracted to detect these compounds. Seven known RONO₂ were analyzed by GC–MS to investigate their retention times and fragmentation patterns (Sect. S1 in the Supplement).

2.3.4 NO_x analyzer in the reactor's headspace

The CLD 88 p Eco Physics NO_x analyzer was used to determine if $\cdot NO$ and $\cdot NO_2$ were formed and partitioned into the gas-phase headspace of the solution during the photolysis of isopropyl nitrate. Indeed, both $\cdot NO$ and $\cdot NO_2$ are highly volatile compounds ($K_H = 1.8 \cdot 10^{-3} \text{ M atm}^{-1}$ and $K_H = 2.0 \cdot 10^{-2} \text{ M atm}^{-1}$, respectively; Sander, 2015), i.e., from 30 to 10^7 times more volatile than the investigated RONO₂. Therefore, if any $\cdot NO$ or $\cdot NO_2$ was formed during the aqueous-phase photolysis, it would have partitioned to the reactor's headspace.

As the NO_x analyzer monitored the headspace of the reactor, a specific experimental setup consisting of a hermetic 1 L three-neck round-bottom flask was used (Fig. S3). It was irradiated by the lamplight beam on its side. Note that since the reactor's headspace was also illuminated, photolysis of isopropyl nitrate could occur in the reactor's headspace. However, although isopropyl nitrate is highly volatile, most of the compound remained in the aqueous phase in the timescale of the experiments (only 3 % of isopropyl nitrate partitioned into the reactor's headspace after 7 h) (González-Sánchez et al., 2023a).

The NO_x analyzer LOD is 0.1 ppbv for both \cdot NO and \cdot NO₂. Considering the gas-phase dilutions performed downward of the reactor, \cdot NO_x could be detected, if formed, at concentrations higher than \sim 2 ppbv using this setup. Although the CLD 88 p Eco Physics NO_x analyzer uses a photolytic converter, interferences with the RONO₂ were observed. A slight proportion (less than 0.6%) of gas-phase isopropyl nitrate was detected as \cdot NO₂. Further details are given in Sect. S2.

In addition, a control experiment was performed to test the efficiency of the gas-phase \cdot NO₂ photolysis and conversion to \cdot NO under our experimental conditions by bubbling gas-phase \cdot NO₂ into the reactor's aqueous phase and photolyzing it with the lamplight. The experimental setup is depicted in Fig. S4.

2.4 Molar yield determinations

The molar yields of the primary reaction products were determined by plotting their concentrations against Δ [RONO₂], which represents the consumption of the parent organic nitrate (Eq. 1).

yield(%) =
$$\frac{[\text{product}]}{\Delta[\text{RONO}_2]} \cdot 100\%$$
 (1)

Since the reaction products were susceptible to undergoing photolysis over time, the yields were calculated for the initial aliquots, sampled during the first 1–2 h of reaction. The evaporation rate of some RONO₂ could be non-negligible compared with photolysis (González-Sánchez et al., 2023a), and Δ [RONO₂] was thus systematically corrected from evaporation using Eqs. (2)–(3):

$$[\text{RONO}_2]_{\text{reac}} = [\text{RONO}_2]_0 e^{(-k_{\text{exp}} + k_{\text{vap}}) \cdot t}, \qquad (2)$$

$$\Delta[\text{RONO}_2] = [\text{RONO}_2]_0 - [\text{RONO}_2]_{\text{reac}}, \qquad (3)$$

where $[\text{RONO}_2]_0$ is the initial concentration and k_{exp} the pseudo-first-order decay, determined by fitting the RONO₂ concentrations ($[\text{RONO}_2]_t$) versus time (*t*) following Eq. (4).

$$[\text{RONO}_2]_t = [\text{RONO}_2]_0 e^{(-k_{\text{exp}}) \cdot t}$$
(4)

The evaporation rate constant, k_{vap} , was determined by control experiments reported in González-Sánchez et al. (2023a).

2.5 Theoretical calculations

Theoretical simulations of the photolysis reaction of isopropyl nitrate photolysis were performed in a model of aqueous solution and in the gas phase. To build the model in the gas phase, snapshots from 10 ps ab initio molecular dynamics (MD) dynamics were performed using a thermostat at 300 K. For the static calculations of minima, conical intersections, and transition states we used B3LYP/6-31G* for the groundstate calculations and TDDFT using the Tamm-Dancoff approximation for the excited-state calculations, with water treated as an implicit solvent via a polarizable continuum model (PCM). To perform the quantum dynamic simulations, we built a model of isopropyl nitrate in aqueous solution by first constructing a water box of 11.39 nm³ equilibrated using Amber99 TIP3P water force field parameters. Isopropyl nitrate was then soaked in the water box and re-equilibrated with the following protocol (see Sect. S3): (1) a canonical ensemble molecular mechanics (MM) MD at a fixed isopropyl geometry during 125 ps; (2) an isobaricisothermal MM MD at a fixed isopropyl during 1 ns; and (3) an isobaric-isothermal B3LYP/6-31G*//Amber99 quantum mechanics/molecular mechanics (QM/MM) MD in period boundary conditions, relaxing the full system for 12 ps (Bonfrate et al., 2023). The snapshots were taken from the latter, discarding the first 2 ps. For each snapshot, a water droplet of 1 nm was extracted, including a spherical wall potential to avoid evaporation of water during the excited-state dynamics. In each snapshot (gas phase and aqueous solution), non-adiabatic excited-state molecular dynamics were operated using Tully's fewest-switch surface-hopping algorithm (Huix-Rotllant et al., 2023). The trajectories were started from the second excited state (S_2) . Excited states were computed using mixed-reference time-dependent densityfunctional theory, which can describe the multi-configuration character of wave functions during photolysis at the cost of a density-functional theory calculation (Lee et al., 2018; Huix-Rotllant et al., 2023).

2.6 Reagents

Chemicals were commercially available and were used as supplied: isopropyl nitrate (96%, Sigma Aldrich), isobutyl nitrate (98%, Sigma Aldrich), chloroacetone (95%, Sigma Aldrich), AgNO₃ (99%, VWR Chemicals), KI (98%, Sigma Aldrich), NaBH₄ (98%, Sigma Aldrich), 2,4-dinitrophenylhydrazine (\geq 99%, Sigma Aldrich), acetaldehyde DNPH (\geq 99%, Sigma Aldrich), acetaldehydrich), formaldehyde DNPH (\geq 99%, Sigma Aldrich), hydroxy acetone (95%, Alfa Aesar), acetic acid (\geq 99.7%, Sigma Aldrich), formic acid (\geq 96%, Sigma Aldrich), (34%–37%, TraceMetal Grade, Fisher Chemical), nitrite standard for IC (1000 ± 4 mg L⁻¹, Sigma Aldrich), nitrate standard for IC (1000 \pm 4 mg L⁻¹, Sigma Aldrich), NaOH (46%–51%, analytical reagent grade, Fisher Chemical), and H₂SO₄ (95%–98%, Merck). Acetonitrile (Fisher Optima) and isopropanol (Honeywell) were LC/MS grade and were used as supplied. Acetone (Carlo Erba Reagents), dichloromethane (Fisher Chemical), and ether (Fisher Chemical) were HPLC grade. Tap water was purified with the Millipore Milli-Q system (18.2 M Ω cm and total organic carbon (TOC) < 2 ppb). Gases were used as supplied: synthetic air (Linde, > 99.999 stated purity), helium 5.0 (Linde), and ·NO₂ (2 ppm in He 5.0, Linde).

Non-commercial organic nitrates, i.e., α -nitrooxy acetone and 1-nitrooxy-2-propanol, were synthesized and purified. α -Nitrooxy acetone was synthesized by the nucleophilic substitution reaction of iodoacetone, which was synthesized previously from chloroacetone. The ketone group from α -nitrooxy acetone was reduced to produce 1-nitrooxy-2- propanol (see details in González-Sánchez et al., 2023a).

3 Results

The results of the aqueous-phase photolysis of organic nitrates are presented in a stepwise manner. Since NO_x compounds are the known major primary product formed in the gas-phase photolysis of RONO₂, this process is examined first in Sect. 3.1, which describes the attempt to measure any formation and partitioning of NO_x to the headspace of the reactor. Section 3.2, 3.3, and 3.4 present the identified reaction products in the aqueous phase, including HNO₂, HNO₃, carbonyls, organic acids, and oxidized RONO₂, and their associated yields. All results are reported in Table S1. Finally, Sect. 4 provides a detailed discussion of the mechanisms involved, focusing on the fate of the nitrate group.

3.1 Absence of NO_X in the reactor's headspace

Experiment 1 investigated isopropyl nitrate (1 mM) photolysis by analyzing the reactor's gas-phase headspace with a NO_x analyzer (Fig. 1a). Prior to turning on the lamp, the \cdot NO₂ signal increased up to \sim 150 ppb, corresponding to a fraction of gas-phase isopropyl nitrate that was photolyzed inside the NO_x analyzer photolytic converter (see Sect. S2 for further details). Once the lamp was turned on (shown in shaded blue in Fig. 1a), the aqueous-phase photolysis of isopropyl nitrate started, but no · NO signal was detected, while the \cdot NO₂ signal peaked at 800 ppb within \sim 10 min of photolysis. However, this signal did not correspond to $\cdot NO_2$, as demonstrated by the control experiment where $\sim 800 \text{ ppb}$ of $\cdot NO_{2(g)}$ was bubbled through the same volume of ultrapure water. When the lamp was turned on (shown in shaded blue in Fig. 1b), $\cdot NO_{2(g)}$ was effectively photolyzed, directly forming $\cdot NO_{(g)}$. In this experiment, barely any $\cdot NO_{2(g)}$ partitioned to the aqueous phase (confirmed by the absence of aqueous-phase HNO₂ or HNO₃, measured by HPIC), and thus the photolysis of $\cdot NO_{2(g)}$ exclusively occurred in the re-



Figure 1. Headspace time profiles of $\cdot NO_{(g)}$ and $\cdot NO_{2(g)}$ signals during (a) aqueous-phase photolysis of isopropyl nitrate with $[RONO_2]_0 = 10^{-3} \text{ M}$ (exp. 1 in Table 1) and (b) photolysis of $\cdot NO_{2(g)}$ bubbled in water.

actor's headspace. From this control experiment, it was concluded that if the measured $\cdot NO_2$ signal represented actual $\cdot NO_{2(g)}$ directly formed in experiment 1, it would be photolyzed in the headspace of the photoreactor to produce measurable amounts of $\cdot NO_{(g)}$.

Since no \cdot NO_(g) was observed when the lamp was turned on in experiment 1 (Fig. 1a), one can conclude that no substantial amounts of $\cdot NO_{2(g)}$ were present in the system. The signal detected as $\cdot NO_{2(g)}$ likely represented an interfering reagent. Nitrous acid (HONO or HNO₂) cannot be this interfering reagent since the concentrations of the interference in the reactor decreased as the reaction progressed, while the measured HONO in the aqueous phase continuously increased. It likely corresponded to another volatile Ncontaining compound that was detected by the NO_x analyzer as a \cdot NO₂ signal (as isopropyl nitrate does). Its signal could be higher than that observed for isopropyl nitrate if the compound presented less water solubility and/or if it decomposed more efficiently in the photocatalytic converter of the NO_x analyzer. It is worth noting that the estimated interference for isopropyl nitrate is very low; it would represent 0.01 % if equilibrium was reached.

3.2 Formation of HNO₂ and HNO₃

HNO₂ and HNO₃ were formed during aqueous-phase photolysis of RONO₂. Both compounds were detected as NO₂⁻ and NO₃⁻ using the HPIC-CD, but their formation as acids was inferred by the observed fast decrease in pH (Fig. S5) and was confirmed by theoretical calculations (see Sect. 4.1.1).

Figure 2 shows an example of HNO₂ and HNO₃ time profiles during the photolysis experiments of isopropyl nitrate (Fig. 2a), isobutyl nitrate (Fig. 2b), α -nitrooxy acetone (Fig. 2c), and 1-nitrooxy-2-propanol (Fig. 2d). HNO₂ could not be quantified during the aqueous-phase photolysis of α nitrooxy acetone due to its fast hydrolysis in the HPIC system



Figure 2. Photolysis experiments of RONO₂: time profiles of RONO₂, HNO₂, and HNO₃ for (**a**) isopropyl nitrate (exp. 4), (**b**) isobutyl nitrate (exp. 8), (**c**) α -nitrooxy acetone (exp. 10), and (**d**) 1-nitrooxy-2-propanol (exp. 11). HNO₂ and HNO₃ were detected as NO₂⁻ and NO₃⁻ in the HPIC-CD.

that used high-pH eluents, where the molecule decomposes into lactate and NO_2^- (Brun et al., 2023).

The figure shows that HNO₂ was efficiently formed as a primary product during all aqueous-phase photolysis reactions of RONO₂. HNO₂ formation slowed down over time due to its fast oxidation to HNO₃, whose time profiles present exponential growth due to its secondary formation. Since this conversion is fast, HNO₃ formation of the first aliquots has been included in the HNO₂ primary yields, assuming that all HNO₃ was formed via HNO₂ oxidation. The detailed chemistry of HNO₂ and HNO₃ that validates this approach is discussed in Sect. 4.1.2. The HNO₂ yields ranged from 40 % to 59 % for isopropyl nitrate (experiments 2 and 4) and from 59 % to 62 % for 1-nitrooxy-2-propanol (experiments 11 and 12), and they were 31 ± 7 % for isobutyl nitrate (experiment 8) and higher than 28 % for α -nitrooxy acetone (experiment 10).

3.3 Formation of carbonyl compounds and organic acids

The formation of primary and secondary carbonyl compounds and organic acids was observed during the aqueousphase photolysis of RONO₂ (Fig. 3).

For isopropyl nitrate (Fig. 3a), the main primary reaction product was acetone, with yields ranging from 32% to 88% (experiments 2–4), and acetaldehyde was formed primarily with lower yields (5%). Additionally, hydroxyacetone, formic acid, and acetic acid were formed as secondary products. These compounds were likely formed via acetone photooxidation (Poulain et al., 2010). For isobutyl nitrate



Figure 3. Photolysis experiments of RONO₂: time profiles of RONO₂, carbonyl compounds and organic acids for (**a**) isopropyl nitrate (Exp. 4), (**b**) isobutyl nitrate (Exp. 8), (**c**) α -nitrooxy acetone (Exp. 10), and (**d**) 1-nitrooxy-2-propanol (Exp. 11). Plain markers are used for primary products.

(Fig. 3b), formaldehyde and acetone were the main nonnitrogen-containing photolysis products (primary yields of 37 %-39 % and 20 %-32 %, respectively, experiments 8-9). Additionally, isobutyraldehyde was detected as a minor product (with a primary yield of 4%-5%). For α -nitrooxy acetone (Fig. 3c), formaldehyde and formic acid appeared as primary products, while hydroxyacetone and acetaldehyde were likely secondary products. The formation yields were found to be $96\pm5\%$ and $79\pm3\%$ for formic acid and formaldehyde, respectively. Other reaction products, such as acetic acid and methylglyoxal, were identified but not quantified due to interferences in the analyzers caused by hydrolysis of α -nitrooxy acetone or oligomerization of methylglyoxal (see Sect. S4). For 1-nitrooxy-2-propanol (Fig. 3d), formaldehyde and acetaldehyde were identified as the main primary reaction products with yields of 63 %-71 % and 50 %-70 %, respectively. Furthermore, lactaldehyde was detected as a primary product with a minor yield of 8 %-14 %. Formic acid and acetic acid were observed as secondary products, likely formed via the photooxidation of formaldehyde and acetaldehyde.

3.4 Secondary formation of oxidized RONO₂

GC–MS analyses at the end of each reaction were performed to look for nitrogen-containing organic products. Figure 4a compares the gas chromatograms obtained for the sample analyzed after 7 h of photolysis with one obtained during a control experiment of isopropyl nitrate in the dark. In both chromatograms, m/z = 46 (which corresponds to a NO₂⁺ fragment) was extracted to display chromatographic peaks related to compounds of RONO₂. The figure shows the formation of at least five oxidized RONO₂ (IP 1, IP 2, IP 3, IP 4, and IP 5), with IP 3 presenting an intensity of 1 order of magnitude higher than the others.

The observed compounds were less volatile than isopropyl nitrate (which had a retention time of 6 min, not shown in Fig. 4a) given their higher retention times and were thus probably oxidized species. The mass spectra of IP 1 to IP 5 confirm that all compounds were RONO₂ with similar chemical structures as isopropyl nitrate (shown in the bottom right of Fig. 4b for comparison). Apart from the NO_2^+ fragment, other fragments observed for isopropyl nitrate were detected. Fragments such as $C_3H_7^+$ (m/z = 43) and $C_2H_4ONO_2^+$ (m/z = 90) were observed in IP 2, IP 3, and IP 5 (and also in IP 1 for m/z = 43). Note that m/z = 43can also correspond to an oxygenated fragment ($C_2H_3O^+$), but the resolution of 1 amu did not allow for separation from $C_3H_7^+$ fragments. Additionally, a specific fragment of a molecule of RONO₂ bearing its nitrate group on a primary carbon atom (CH₂ONO₂⁺ at m/z = 76) was observed for IP 1, IP 3, IP 4, and IP 5. Since IP 3 and IP 5 combine this fragment with a fragment that is specific for the secondary nitrate group (C₂H₄ONO₂⁺ at m/z = 90), these compounds might be dinitrates. This is the case for the most intense chromatographic peak (IP 3). IP 3 was thus assigned to the 1,2-propyl dinitrate molecule due to its mass spectra. Additionally, IP 2 was assigned to 2-nitrooxy-1-propanol due to the C₃H₇O⁺ and C₂H₅ONO₂⁺ fragments (m/z = 59 and m/z = 90, respectively). These identifications are consistent with the proposed mechanism (see Sect. 4.2). However, the absence of standards prevented the precise identification and quantification of these compounds.

Hints of the formation of an oxidized molecule of RONO₂ were also observed in the non-derivatized UHPLC-UV analyses. An unidentified peak was detected at a retention time close to isopropyl nitrate (2.7 vs. 2.4 min). The peak presented similar UV absorption spectra to the standards of RONO₂ (Fig. S6) and was thus assigned to be IP 3 (1,2-propyl dinitrate) due to its major concentrations. The compound was a secondary product since its occurrence started after 2 h of reaction. A rough estimation of its concentration was performed using average calibration curve parameters obtained for isopropyl nitrate, isobutyl nitrate, α -nitrooxy acetone, and 1-nitrooxy-2-propanol. Assuming that IP 3 was a dinitrate, it represented 9% of the consumed nitrogen at the end of the reaction.

The secondary formation of oxidized RONO₂ was also confirmed for isobutyl nitrate and 1-nitrooxy-2-propanol. For isobutyl nitrate, two unidentified peaks assigned to oxidized RONO₂ (IB 1 and IB 2) were observed by UHPLC-UV. Both compounds present UV–Vis absorption spectra identical to isobutyl nitrate at lower retention times (1.6 min for IB 1 and 3.1 min for IB 2 vs. 3.4 min for isobutyl nitrate) related to a higher polarity of the molecules. Their time profiles show



Figure 4. Isopropyl nitrate photolysis. (a) Gas chromatogram (extracted for m/z = 46, NO₂⁺) of the end of the reaction of exp. 2 in Table 1 (after 7 h of photolysis) and the control experiment (isopropyl nitrate in water). (b) Mass spectra of the detected peaks.

that both compounds were formed through secondary reactions (Fig. S7). GC–MS analyses (performed after preconcentration of the sample) allowed for the detection of up to nine oxidized RONO₂. For 1-nitrooxy-2-propanol, four oxidized RONO₂, including α -nitrooxy acetone, were observed by GC–MS. The chromatograms and mass spectra as well as comments on the identification of the formed molecules are presented in the Supplement (Sect. S5).

For α -nitrooxy acetone, no oxidized RONO₂ were found in UHPLC-UV analyses or in GC–MS analyses.

4 Discussion

4.1 N budget during aqueous-phase photolysis of RONO₂

Gas-phase photolysis of RONO₂ is known to induce homolytic rupture of the RO–NO₂ bond, releasing \cdot NO₂ into the atmosphere with yields close to 100 % (Talukdar et al., 1997; Carbajo and Orr-Ewing, 2010). This reactivity turns RONO₂ into NO_x reservoirs and shifts pollution transportation from the local to the regional scale. Our results show that, in the aqueous phase, a primary formation of HNO₂ (with yields ranging from 31 % to 62 %) is followed by a secondary formation of HNO₃. Therefore, one of the main questions about the aqueous-phase photolysis of $RONO_2$ is whether (or not) it can regenerate NO_x that would partition to the gas phase.

To address this question, we explored the viability of two different chemical pathways that lead to NO₂⁻ / HNO₂ and NO_3^- / HNO_3 in the aqueous phase. The first explored pathway was the direct formation of $\cdot NO_{2,(aq)}$ followed by its known aqueous reactivity (i.e., hydrolysis and reactivity towards other radicals). This pathway was rejected since · NO and $\cdot NO_2$ should be observable in the system in this scenario (see details in Sect. S6). The second explored pathway was the direct formation of HNO₂ in the aqueous phase. This pathway was confirmed by theoretical calculations for isopropyl nitrate aqueous-phase photolysis. Herein, the Discussion section focuses on this pathway and the secondary chemistry of the photolysis products in our system. Finally, a conclusion is given with proposed mechanisms of aqueousphase photolysis reactions of isopropyl nitrate, isobutyl nitrate, 1-nitrooxy-2-propanol, and α -nitrooxy acetone, including a detailed discussion for isopropyl nitrate.

4.1.1 Direct formation of HNO₂ in the aqueous phase

Theoretical calculations were performed to evaluate if the direct formation of HNO₂ is possible in the aqueous phase. The static calculations showed that the formation of HNO₂ is thermodynamically favorable. Figure 5 represents the potential energy surfaces of a potential reaction pathway of isopropyl nitrate aqueous-phase photolysis, showing that it is indeed a possible reaction. Upon photon absorption, isopropyl nitrate is in the first excited state and can relax rapidly to the minimum of this state. From the excited state, it undergoes a non-radiative internal conversion back to the ground state through a degenerated point between the excited state and the ground state, a conical intersection. The conical intersection has a sloped topology. -ONO₂ presents a pyramidal structure (instead of triangular) in the conical intersection. After reaching the conical intersection, the isopropyl nitrate accumulates all the photon energy as excess vibrational energy. Indeed, this energy is in principle sufficient to cross the large barrier to form HNO₂ as a product. The transition state is a concerted RO-NO2 dissociation and a proton transfer, with an imaginary frequency of 1241.3 cm^{-1} , which leads directly to the formation of acetone and HNO₂ as final products.

To confirm this pathway, we performed excited-state nonadiabatic dynamics in an atomistic model of isopropyl nitrate in water using QM/MM methodology. The dynamics in such a model allow us not only to determine the reaction hypothesis described in Fig. 5, but also to estimate the timescales at which this reaction can happen. In Fig. 6, an example of a reactive trajectory in the excited state is depicted. Initially, the R–ONO₂ is in a trigonal planar conformation. Once the photon is absorbed, the group displays a pyramidal conformation that allows a non-radiative conversion from the excited to the ground state via a conical intersection. This leads directly to



Figure 5. Relative energy potential energy surface of isopropyl nitrate photolysis. The structures of the key points of the reaction, marked with green dots, are also depicted in the figure. The grey arrow shows the hypothetical reaction: (1) the molecule is in an excited state after photon absorption, traveling first to the minimum energy structure of the excited state (2) and accessing a conical intersection (3) through a barrierless surface. The molecule then goes back to the reactant (4) with excess vibrational energy, which allows it to cross a transition state (5) up to the products (6). The reactant's energy has been arbitrarily placed at 0.0 eV. Calculations were performed using a density functional theory (DFT)/time-dependent DFT (TDDFT) model in implicit solvent.

the dissociation of \cdot NO₂, which diffuses towards water. This is at odds with the found transition state, which corresponded to a concerted \cdot NO₂ and proton transfer. Therefore, we can rather claim that the reaction happens in two steps. The interaction of \cdot NO₂ with water favors the 180 ° twist, in which nitrogen is pointing towards water molecules, thus favoring a conformation in which a proton transfer is favored, occurring in less than 1 ps. Despite the fact that this happens to be the main reaction channel, other reactions, in which direct formation of acetaldehyde or dissociation of HNO₂ in \cdot OH and \cdot NO is observed, are possible. This is due to the excess vibrational energy of the photoproducts encapsulated in a water cavity with a diameter of around 0.7 nm, which prevents their diffusion. Still, in longer timescales the photoproducts will either react with water or dissipate the energy to the solvent.

The gas-phase photolysis of isopropyl nitrate was also studied using the same type of dynamics. In the gas phase, the photochemical pathway through a conical intersection until the formation of the \cdot NO₂ radical happens in a similar manner to the aqueous solution. In the gas phase, however, the absence of a water cavity prevents the resulting fragments from reorienting and reacting. Indeed, similar to an explosion, the two resulting fragments \cdot NO₂ and RO· diffuse in opposite directions before any proton transfer can occur between them. This explains the observed direct formation of \cdot NO₂,(g) (Talukdar et al., 1997). In contrast, in the cavity, collisions with the solvent are frequent, and thus the pho-



Figure 6. Representative reactive trajectories for the formation of acetone and HNO_2 with the main reaction steps depicted. Timescales are just indicative.



Figure 7. Concomitant acetone and HNO_2 formation during isopropyl nitrate photolysis at two different initial concentrations, (a) 0.57 mM (exp. 2) and (b) 1.42 mM (exp. 4).

tolysis likely follows the pathway with the minimum-energy barrier, which leads to the formation of HNO₂ and acetone.

These calculations agree with our observations that the photolysis of isopropyl nitrate caused the direct concomitant formation of HNO₂ and acetone as shown in Fig. 7. Furthermore, minor trajectories where the carbon backbone structure of isopropyl nitrate breaks, leading to the formation of acetaldehyde and other species, have been also experimentally observed since acetaldehyde was determined to be a primary product with low yields ($\sim 4 \%$).

4.1.2 Secondary chemistry of HNO₂ in the aqueous phase

Once formed in the solution, HNO_2 was highly reactive, as shown by its time profiles (in Fig. 7). It may disproportionate to yield $\cdot NO$ and $\cdot NO_2$ (R1).

$$2HNO_2 \rightarrow \cdot NO + \cdot NO_2 + H_2O \tag{R1}$$

However, this reaction is quite slow under our experimental conditions (rate constant of $28.6 \text{ M}^{-1} \text{ s}^{-1}$, Vione et al., 2004). Nevertheless, considering the lamp actinic flux, the photolysis/photooxidation of HNO₂ was likely its major sink. The photolysis of HNO₂ and NO₂⁻ is known to form \cdot NO and a \cdot OH radical (R2–4) (Mack and Bolton, 1999; Fischer and Warneck, 1996; Kim et al., 2014). HNO₂ can also decompose due to the additional energy of the photolysis of RONO₂.

$$NO_{2}^{-} + h\nu \to \cdot NO + \cdot O^{-} \tag{R2}$$

$$HNO_2 + h\upsilon \to \cdot NO + \cdot OH \tag{R3}$$

$$\cdot O^{-} + H_3 O^{+} \longleftrightarrow \cdot OH + H_2 O \tag{R4}$$

Additionally, \cdot OH radicals readily react with NO₂⁻ and HNO₂ (rate constants of 1.0×10^{10} and 2.6×10^9 M⁻¹ s⁻¹, respectively) (Mack and Bolton, 1999; Kim et al., 2014).

$$NO_2^- + \cdot OH \rightarrow \cdot NO_2 + OH^-$$
 (R5)

$$HNO_2 + \cdot OH \to \cdot NO_2 + H_2O \tag{R6}$$

HNO₃ can then be formed through \cdot NO₂ hydrolysis (R7) (Finlayson-Pitts and Pitts, 2000).

$$2 \cdot \text{NO}_2 + \text{H}_2\text{O} \rightarrow \text{HNO}_2 + \text{HNO}_3 \tag{R7}$$

Another pathway can be initiated by the reaction of \cdot NO with HO₂ \cdot radicals to yield peroxynitrite (R8). The latter can isomerize into HNO₃ / NO₃⁻ (R9) or decompose, yielding \cdot NO₂ and \cdot OH radicals (R10).

$$\cdot \text{NO} + \text{HO}_2 \cdot \rightarrow \text{HOONO}$$
(R8)

$$HOONO \to NO_3^- + H^+ \tag{R9}$$

$$HOONO \rightarrow \cdot NO_2 + \cdot OH \tag{R10}$$

 $HO_2 \cdot radicals$ were likely formed by the photooxidation of organic compounds. Since $\cdot OH$ radicals were formed through HNO_2 / NO_2^- photolysis, they could attack the organic molecules present in the photoreactor (i.e., the RONO₂, as no scavenger was used). Upon oxygen addition, the $\cdot OH$ attack yielded peroxy radicals. The formation of peroxy radicals was confirmed by the dissolved oxygen time profiles: during each photolysis experiment, dissolved [O₂] underwent slight decay due to the reaction of alkyl radicals ($R \cdot$) and oxygen (Fig. S8).

Peroxy radicals can readily react with \cdot NO to form peroxynitrites (ROONO) that can isomerize to RONO₂ (R11). Additionally, peroxy radicals react with \cdot NO₂, forming peroxynitrates (ROONO₂) (Goldstein et al., 2004).

 $\text{ROO} \cdot + \cdot \text{NO} \rightarrow \text{ROONO} \rightarrow \text{RONO}_2$ (R11)

$$\text{ROO} \cdot + \cdot \text{NO}_2 \to \text{ROONO}_2$$
 (R12)

The rate constants observed for R11 and R12 range, respectively, from 2.8×10^9 to $3.5 \times 10^9 \text{ M}^{-1} \text{ s}^{-1}$ (with *R* being (CH₃)₂CCH₂- and CH₃-) and from 0.7 to $1.5 \times 10^9 \text{ M}^{-1} \text{ s}^{-1}$ (with *R* being (CH₃)₂CCH₂-, CH₃-, and c-C₅H₉-). Therefore, these reactions could readily occur under our experimental conditions but not in the first step, as they would be limited by the formation of \cdot OH radicals to be initiated.



Figure 8. Proposed mechanism of isopropyl nitrate photolysis in the aqueous phase. In red are the measured primary products with their molar yields. In blue are the measured secondary products.

In our experiments, the formation of oxidized RONO₂ during isopropyl nitrate and isobutyl nitrate photolysis was confirmed by GC–MS and UHPLC-UV analyses (Fig. 4 and Sect. S4). The possibility of forming oxidized RONO₂ via the aforementioned reactions is consistent with the substantial number of compounds displaying the NO₂⁺ fragment found by GC–MS analyses (up to six compounds for isopropyl nitrate photolysis and up to eight for isobutyl nitrate). Nevertheless, ROONO₂, if formed, was not detected due to its thermolysis during the analysis.

During isopropyl nitrate photolysis, the main formed oxidized organic nitrate (IP 3 in Fig. 4) was suspected to be a dinitrate (1,2-propyldinitrate) since its mass spectra conjugate mass fragments that correspond to both primary (m/z = 76, CH₂ONO₂⁺) and secondary nitrate groups (m/z = 90, CH(ONO₂)CH₃⁺). The formation of this compound through secondary photochemistry of HNO₂ / NO₂⁻ agrees well with the observed secondary time profile of this product. An equivalent compound was observed during isobutyl nitrate photolysis (IB 6 in Fig. S4.1).

4.2 Proposed chemical mechanisms

4.2.1 Proposed mechanism of isopropyl nitrate aqueous-phase photolysis

Combining all the reactions mentioned in the Discussion, Fig. 8 proposes a complete mechanism of isopropyl nitrate aqueous-phase photolysis.

Isopropyl nitrate photolyzes into acetone and nitrous acid. Nitrous acid undergoes equilibrium with nitrite in the aqueous phase. Both HNO₂ and NO₂⁻ can undergo photolysis, yielding \cdot NO and \cdot OH radicals (R5 to 7). \cdot OH radicals can react with isopropyl nitrate, yielding an alkyl radical that upon oxygen addition forms a peroxy radical. The peroxy radical can decompose into products (i.e., acetone, formic acid, acetic acid, hydroxy acetone, and acetaldehyde, which could also be issued from acetone photooxidation), or react with \cdot NO or \cdot NO₂, to form a dinitrate or a peroxynitrate. The dinitrate likely corresponds to the compound detected by GC–MS (IP 3 in Fig. 4) and is formed secondarily, in agreement with the proposed mechanism. Additionally, IP 3 was estimated to account for 18 % of the reactive N at the end of the reaction, in agreement with the 20 % of isopropyl nitrate estimated to undergo \cdot OH oxidation. Furthermore, HNO₃ is formed through secondary reactions such as \cdot NO₂ hydrolysis (R1) or peroxynitrite isomerization (R11).

4.2.2 Proposed mechanism of isobutyl nitrate and 1-nitrooxy-2-propanol aqueous-phase photolysis

The primary formation of HNO₂ was also observed during the photolysis of isobutyl nitrate and 1-nitrooxy-2propanol in the aqueous phase (Fig. 2). The determined yields were 31 % and 59 %–62 % for isobutyl nitrate (Fig. 9a) and 1-nitrooxy-2-propanol (Fig. 9b), respectively. Although no DFT calculations were performed specifically for these molecules, they likely undergo a similar photolysis process to the one detailed for isopropyl nitrate, where an adjacent hydrogen atom is captured by the $-NO_2$ -leaving moiety (as shown in Fig. 6).

Nevertheless, the formation of carbonyl products concomitant to HNO₂ was different from that expected from the main isopropyl nitrate mechanism. The corresponding carbonyl compounds were only observed in minor proportions: yields of 5 % isobutyraldehyde and 8 %-10 % lactaldehyde were obtained for isobutyl nitrate and 1-nitrooxy-2-propanol, respectively. The major carbonyl products were formed after the breakdown of the organic chain, probably due to the excess energy the molecules have after light absorption. This pathway has been observed during the isopropyl nitrate calculations, although as a minor pathway, leading to the formation of acetaldehyde. Figure S9 clearly shows that the carbonyl products formed concomitantly to HNO₂ were acetone and formaldehyde (yields of 20 %-32 % and 37 %-39 %, respectively) during isobutyl nitrate photolysis and formaldehyde and acetaldehyde (yields of 63 %–71 % and 50 %–70 %, respectively) during 1-nitrooxy-2-propanol photolysis.

The proposed pathways for their photolysis are given in Fig. 9. Further studies should be conducted to understand the breakdown of the organic chain.

During isobutyl nitrate photolysis, two oxidized RONO₂ were observed; these compounds were likely formed through the \cdot OH oxidation of isobutyl nitrate initiated by HNO₂ / NO₂⁻ aqueous-phase photochemistry (as it occurs in isopropyl nitrate photolysis). During 1-nitrooxy-2-propanol photolysis, the secondary formation of α -nitrooxy acetone was observed (Fig. 9b).



Figure 9. Proposed mechanisms of aqueous-phase photolysis of **(a)** isobutyl nitrate and **(b)** 1-nitrooxy-2-propanol. In red are the measured primary products and in blue the detected secondary products.

4.2.3 Proposed mechanism of α -nitrooxy acetone aqueous-phase photolysis

During α -nitrooxy acetone photolysis, NO₂⁻ could not be measured due to its base-catalyzed hydrolysis in the HPIC system (at a pH of 12, Brun et al., 2023), but NO₃⁻ was quantified and showed a secondary formation. Primary formation of HNO₂ was therefore expected with a minimum yield of 28 %. The complete mechanism could not be elucidated for this molecule, since some of its primary products could not be quantified (i.e., methylglyoxal and acetic acid). Nevertheless, the mechanism appears to be similar to that of isobutyl nitrate and 1-nitrooxy-2-propanol. The identification of methylglyoxal indicates one pathway, leading to its direct formation along with HNO₂ (homologous mechanism as in Fig. 5). However, the detection of significant quantities of formic acid and formaldehyde and the identification of acetic acid suggest that the molecule tends to break at different points (similar to isobutyl nitrate and 1-nitrooxy-2-propanol). Further analytical efforts or modeling should be conducted to clearly identify how the molecule undergoes rupture by photolysis.

5 Conclusions and atmospheric implications

This work investigated the fate of the nitrate group during the aqueous-phase photolysis of four species of RONO₂: isopropyl nitrate, isobutyl nitrate, 1-nitrooxy-2-propanol, and α nitrooxy acetone. Our findings suggest a completely different reactivity from the gas phase one. While RONO₂ release NO_x back into the atmosphere upon photolysis in the gas phase, HNO_2 is directly formed in the aqueous phase.

 HNO_2 was detected as a primary reaction product along with others such as carbonyl compounds or organic acids. The direct formation of HNO_2 through aqueous-phase photolysis was confirmed by DFT theoretical calculations and was supported by the absence of direct $\cdot NO_2$ formation due to solvent cage effects.

Therefore, aqueous-phase photolysis of RONO2 represents both a sink of NO_x and a source of atmospheric HNO_2 (or HONO). The latter is an important precursor of ·OH and . NO radicals. During our experiments, these secondarily formed radicals were shown to be trapped in the aqueous phase, producing HNO₃ and functionalized RONO₂. In the atmosphere, this reactivity can potentially contribute to the sink of NO_x , a source of $\cdot OH$ radicals in condensed phases, and an additional source of SOA_{aa}. Aqueous-phase photolysis has been reported to be negligible in the sinks of RONO₂ in the atmosphere due to the hindering effect of the solvent cage (González-Sánchez et al., 2023a). Nevertheless, the mechanisms of this reactivity might be relevant to more significant reactions such as the aqueous-phase · OH oxidation of RONO₂ or potentially their heterogeneous photolysis. Therefore, further work should be done to better assess the role of RONO₂ in NO_x sink and transport in the formation of atmospheric HONO and SOA.

Code and data availability. Data related to this article are available at https://doi.org/10.7910/DVN/USWU6V (González-Sánchez et al., 2023b). Data related to the theoretical calculations can be requested from Miquel Huix-Rotllant (miquel.huixrotllant@univ-amu.fr).

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Author contributions. JMGS performed all experiments and treated all experimental data. MHR developed the quantum chemistry model and conducted the calculations. JMGS, NB, and CD developed the HPIC-CD method. JM built the NO_x analyzer experimental setup. JMGS and SR developed the UHPLC-UV method. JMGS and AD developed the GC–MS method. JMGS and JLC performed the organic synthesis of RONO₂. AM and JLC coordinated the work. JMGS, MHR, and AM wrote the article with inputs from all co-authors.

Competing interests. The contact author has declared that none of the authors has any competing interests.

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