



Supplement of

Heterogeneous interactions between \mbox{SO}_2 and organic peroxides in submicron aerosol

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S1. Quantification of sulfate formation for SOA experiments

Particulate sulfate formation upon the reactive uptake of SO₂ onto different types of SOA was 22 monitored using a compact time-of-flight aerosol mass spectrometer (C-ToF-AMS) (Aerodyne 23 24 Research Inc., Billerica, USA). In this study, the ToF-AMS was operated in the Mass Spectrum 25 (MS) mode with a chopper to regulate particle transmission for aerosol sulfate quantification 26 (DeCarlo et al., 2006). The chopper where the aerosol beam passes is controlled to be 27 alternatively blocked in order to have background corrections for quantification. An aerodynamic lens system selects the particles in a size range of ~35-1000 nm into the vacuum system. The 28 sampling flow rate was 1.26 cm³ s⁻¹. After the sampling stream passing the sizing chamber, 29 30 aerosol is vaporized on a heated porous tungsten surface (600 °C) and immediately ionized in 31 electron impact (EI) mode at 70 eV. 300 nm ammonium nitrate (99.999%, Sigma-Aldrich) was 32 used for AMS ionization efficiency (IE) calibration. The collection efficiency (CE) under humid 33 condition was assumed to be 1. The relative ionization efficiency (RIE) of inorganic sulfate was determined to be 1.05 using ammonium sulfate (Canagaratna et al., 2007). The ions are extracted 34 35 by an orthogonal extractor into the ToF-MS (V-mode), and the MS spectra acquisition was performed under positive mode. Data were analyzed by software "SQUIRREL 1.63" and "PIKA 36 37 1.23" in Igor Pro 6.37 (Wavemetrics, Oregon, USA). For γ_{SO2} measurements of toluene SOA, a strong hydrocarbon interference was observed with the 38 39 SO₂ analyzer, possibly stemming from the high concentrations of gas-phase aromatic compounds. As a result, the uptake rate was instead estimated by measuring the sulfate 40 production rate using AMS. However, it should also be noted that the sulfate collection and 41 42 ionization efficiencies of AMS are highly uncertain, since the yield of organosulfate is significant from these reactions (Wang et al., 2019), and AMS has a lower sensitivity towards 43

44	organosulfates especially under humid conditions in the current study as indicated by ion
45	chromatography and SMPS measurements (Fig.S10-12) (Chen et al., 2019;Farmer et al., 2010).
46	Different RIE for different types of organosulfates were also observed in this study (Fig. S11).
47	As a result, γ_{SO2} of toluene SOA was estimated from rate of sulfate formation measured by AMS
48	and corrected based on the ratio between sulfate formation and SO ₂ consumption of limonene
49	SOA (Fig. S12).
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65 S2. Supplemental Table and Figures

66 Table S1. Summary of experime	ents in this study.
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- Figure S1. Schematic of measuring γ_{SO2} onto ammonium sulfate or malonic acid mixed with peroxides.
- 69 Figure S2. Experimental schematic of measuring γ_{SO2} onto SOA.
- Figure S3. Potential SO₂ loss by peroxides accumulated on the filter before the SO₂ analyzer.
- Figure S4. Investigating potential SO₂ decay by losses inside the SO₂ analyzer.
- Figure S5. Contribution of wet ammonium sulfate aerosol to the observed SO₂ decay.
- Figure S6. Contribution of organic peroxide vapor to the observed SO₂ decay.
- Figure S7. Relationship between γ_{SO2} and peroxide characteristics.
- Figure S8. Comparison between measured γ_{SO2} and predicted γ_{SO2} in experiments.
- Figure S9. Residuals (a) and residual distribution (b) for the multilinear regression.
- Figure S10. (a) IC calibration curves for S (VI). (b) Comparison of sulfate quantified
 by AMS and IC.
- Figure S11. Comparison of SMPS measured sulfate and AMS measured sulfate under dry and
 humid conditions (RH 80%) for both inorganic sulfate and organosulfates.
- Figure S12. Time series of sulfate formation monitored by AMS for SOA experiments.

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Table S1. Summary of chamber experiments in this study

Expt.#	Aerosol	Initial SO ₂ (ppb)	SO ₂ decay (ppb)	Surface area concentration (µm ² cm ⁻³)	γso2 ^a	RH %
1	25 mM ammonium sulfate	218	-1	7.8×10 ³	$(-1.9 \pm 0.08) \times 10^{-7}$	52
2	50 mM tert-Butyl hydroperoxide + 25 mM ammonium sulfate	240	3	8.9×10 ³	$(2.7 \pm 0.09) \times 10^{-6}$	44
3	50 mM tert-Butyl hydroperoxide + 25 mM ammonium sulfate	215	10	7.5×10 ³	$(1.4 \pm 0.07) \times 10^{-5}$	56
4	50 mM tert-Butyl hydroperoxide + 25 mM ammonium sulfate	250	47	1.4×10 ⁴	$(2.8 \pm 0.7) \times 10^{-5}$	67
5	50 mM Cumene hydroperoxide +25 mM ammonium sulfate	201	10	2.9×10 ³	$(1.2 \pm 0.09) \times 10^{-4}$	47
6	50 mM Cumene hydroperoxide +25 mM ammonium sulfate	167	7	2.0×10 ³	$(1.5 \pm 0.3) \times 10^{-4}$	52
7	50 mM Cumene hydroperoxide +25 mM ammonium sulfate	207	39	4.4×10 ³	$(2.4 \pm 0.3) \times 10^{-4}$	61
8	50 mM 2-Butanone peroxide +25 mM ammonium sulfate	108	7	1.6×10 ³	$(1.3 \pm 0.1) \times 10^{-3}$	25
9	50 mM 2-Butanone peroxide +25 mM ammonium sulfate	201	107	7.4×10 ³	$(3.1 \pm 0.2) \times 10^{-3}$	47
10 ^b	50 mM 2-Butanone peroxide +25 mM ammonium sulfate	153	91	3.5×10 ³	$(7.9 \pm 0.4) \times 10^{-3}$	53
11 ^b	50 mM 2-Butanone peroxide +25 mM ammonium sulfate	107	32	1.7×10 ³	$(6.2 \pm 0.7) \times 10^{-3}$	54

12 ^b	50 mM 2-Butanone peroxide +25 mM ammonium sulfate	98	33	1.1×10 ³	$(1.0 \pm 0.05) \times 10^{-2}$	55
13	50 mM 2-Butanone peroxide +25 mM ammonium sulfate	176	73	1.8×10 ³	$(9.3 \pm 0.7) \times 10^{-3}$	62
14 ^c	50 mM 2-Butanone peroxide +25 mM ammonium sulfate	151	133	2.1×10 ³	$(2.7 \pm 0.4) \times 10^{-2}$	71
15	100 mM 2-Butanone peroxide +25 mM ammonium sulfate	100	72	8.1×10 ²	$(4.8 \pm 1) \times 10^{-2}$	52
16	25 mM 2-Butanone peroxide +25 mM ammonium sulfate	175	76	4.1×10 ³	$(4.2 \pm 0.4) \times 10^{-3}$	52
17	5 mM 2-Butanone peroxide +25 mM ammonium sulfate	138	16	4.6×10 ³	$(8.0 \pm 0.4) \times 10^{-4}$	53
18	0.5 mM 2-Butanone peroxide +25 mM ammonium sulfate	180	4	7.3×10 ³	$(8.3 \pm 0.4) \times 10^{-5}$	57
19	100 mM 2-Butanone peroxide +25 mM malonic acid	113	68	3.3×10 ³	$(1.3\pm0.2)\times10^{-2}$	52
20	50 mM 2-Butanone peroxide +25 mM malonic acid	157	49	2.3×10 ³	$(5.1\pm0.7) imes10^{-3}$	54
21	25 mM 2-Butanone peroxide +25 mM malonic acid	186	32	2.0×10 ³	$(2.8\pm0.2)\times10^{-3}$	54
22	5 mM 2-Butanone peroxide +25 mM malonic acid	236	6	1.8×10 ³	$(4.6\pm0.5) imes10^{-4}$	54
23	5 mM 2-Butanone peroxide +25 mM ammonium sulfate +0.00002 M HCl	157	25	4.5×10 ³	$(1.2\pm0.2) \times 10^{-3}$	53

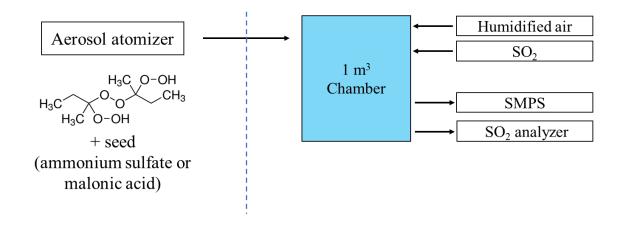
	5 mM 2-Butanone peroxide					
24	+25 mM ammonium sulfate	152	21	3.4×10 ³	$(1.3\pm0.09)\times10^{-3}$	54
	+0.0001 M HCl					
	5 mM 2-Butanone peroxide					
25	+25 mM ammonium sulfate	136	30	3.1×10 ³	$(2.5\pm0.4) \times 10^{-3}$	53
	+0.001 M HCl					
26	α-Pinene SOA	213	12	1.4×10 ³	$(5.7\pm0.4) imes10^{-5}$	51
27	Limonene SOA	340	12	4.6×10 ³	$(2.4\pm0.1)\times10^{-4}$	54
28	Toluene SOA	-	-	2.7×10 ³	(8.3±0.4) ×10 ⁻⁴	60

^a Uncertainty of γ_{SO2} in this study was estimated from SO₂ and SMPS measurements. ^b The

87 reproducibity of the γ_{SO2} measurements were estimated from Expt.10-12 to be 26 %. $^{\circ}\gamma_{SO2}$

88 measured under high relative humididy condition was corrected by the SO₂ repartioning rate (Fig.

89 S6).



- **Figure S1.** Experimental schematic of measuring SO₂ reactive uptake coefficient onto
- 92 ammonium sulfate or malonic acid mixed with organic peroxide. Aerosol was generated from the
- 93 atomizer before being introduced into the chamber.

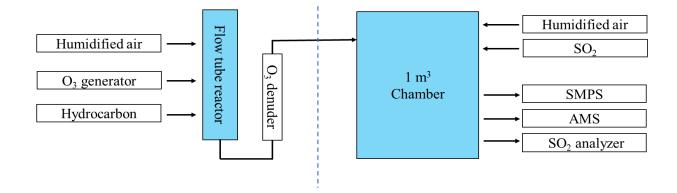




Figure S2. Experimental schematic of measuring SO₂ reactive uptake coefficient onto different

97 types of SOA. Secondary organic aerosol was generated in the flowtube reactor by

98 photooxidation (toluene) or ozonolysis (limonene and α-pinene). Excess amount of O₃ was

99 removed using an O_3 denuder to avoid SO_2 consumption caused by O_3 inside the smog chamber.

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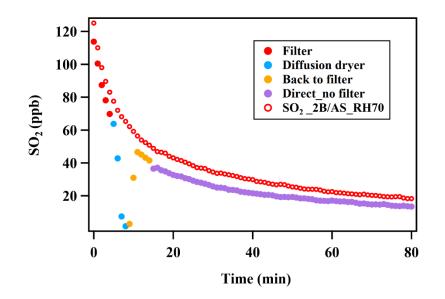


Figure S3. To test whether the observed SO_2 decay is caused by reactions with organic peroxide accumulated on the inline PTFE filter installed in front of the SO₂ analyzer, the first experiment (solid red circle) was conducted under the same condition of the second experiment (empty red circle), but the SO₂ measurement was taken with filter (red), through diffusion dryer (blue), back to filter (orange) and eventually direct measurement without filter (purple). Measurements taken with/without filter in front of the SO₂ analyzer show the same time series of SO₂ decay during the experiment, except when a diffusion dryer was placed inline (causing SO₂ loss by diffusion). These trends demonstrate that the observed SO₂ decay is not likely caused by reactions with organic peroxides accumulated on the PTFE filter in front of the SO₂ analyzer.

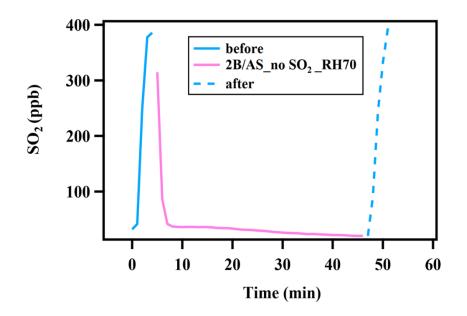


Figure S4. To test whether the observed SO₂ decay is caused by the organic peroxide trapped inside the analyzer during monitoring, SO₂ analyzer (Model 43i, Thermo Scientific) was directly connected to a stream of standard SO₂ gas with a constant SO₂ mixing ratio, and then switched to a chamber filled with organic peroxide-containing ammonium sulfate aerosol (no SO₂) under RH 70% for 40 minutes. Eventually, the SO₂ analyzer was switched back to the standard SO₂ gas stream. The response of the SO₂ analyzer towards standard SO₂ gas shows similar rates before and after measuring organic peroxide-containing ammonium sulfate from the chamber, indicating the significant SO₂ decay observed in our study is not caused by any reaction inside the SO₂ analyzer.

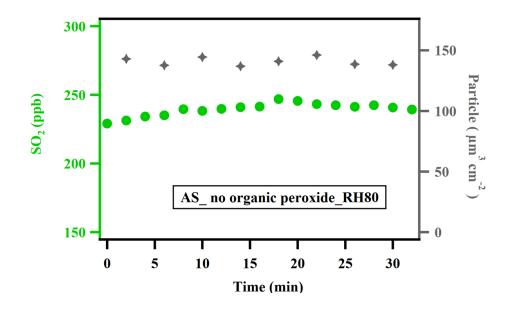


Figure S5. SO₂ time series in a blank control experiment. SO₂ was introduced into a chamber
containing 140 μm³ cm⁻³ of ammonium sulfate aerosol with no organic peroxide at an RH of
80 %. No SO₂ decay was observed. Deliquesced ammonium sulfate aerosol is not a significant
contributor to the decay of SO₂ under RH 80 % observed in chamber experiment, indicating the
dominant sink of SO₂ during the other chamber experiment is the reaction with organic
peroxides.

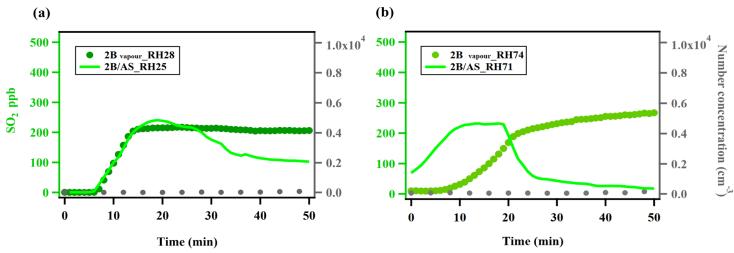


Figure S6. SO₂ consumption by 2-butanone organic peroxide vapor under dry (RH 28 %) and humid (RH 74 %) condition during chamber experiment. Organic peroxide vapor was introduced by placing an inline PTFE filter between the atomizer and chamber in order to remove particlephase peroxides. No significant SO₂ decay was observed when only organic peroxide vapor was present under both dry and humidity conditions. For comparison, the SO₂ time series with particulate organic peroxide-ammonium sulfate mixture showed significant SO₂ decay. However, we noticed there was SO₂ repartitioning from the chamber wall under high RH (74 %), and this SO₂ repartitioning rate was used to correct the γ_{SO2} measured under high RH conditions (above 70 %, Expt.14).

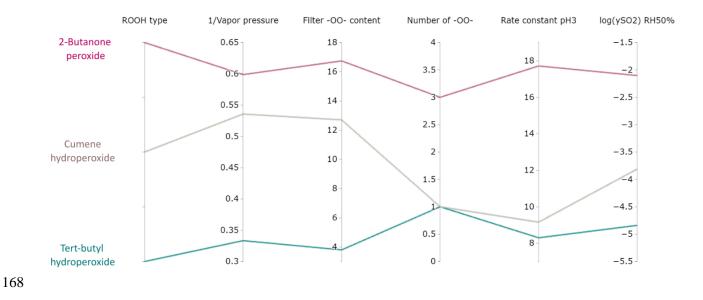


Figure S7. Visualization of organic peroxides' characteristics and γ_{SO2} for chamber experiments at RH 50% and a total peroxide to ammonium sulfate ratio of 2:1. y_{SO2} is positively related with aqueous phase second order reaction rate constant (M⁻¹ s⁻¹) at pH 3 (which is associated with number of -OO- group) and with particulate -OO- content (%) on the filter (which is negatively associated with vapor pressure (kPa)).

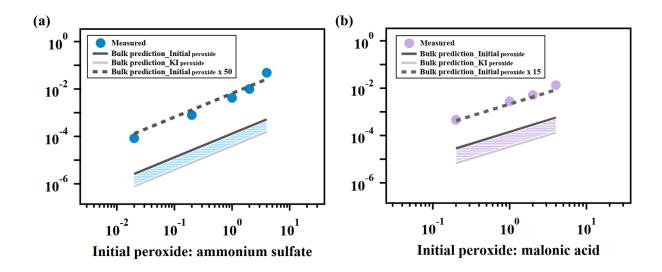




Figure S8. Comparison between measured γ_{SO2} and predicted γ_{SO2} for experiments with different peroxide to ammonium sulfate ratios (a) and peroxide to malonic acid ratios (b). The γ_{SO2} predictions were made based on initial peroxide content assuming no partitioning and off-line KI measured peroxide content, respectively. The shadowed area is the uncertainty in γ_{SO2} prediction in terms of particulate peroxide content. The discrepancy observed for ammonium sulfate aerosol (50 times) is larger than that of malonic acid (15 times) under the same experimental conditions.

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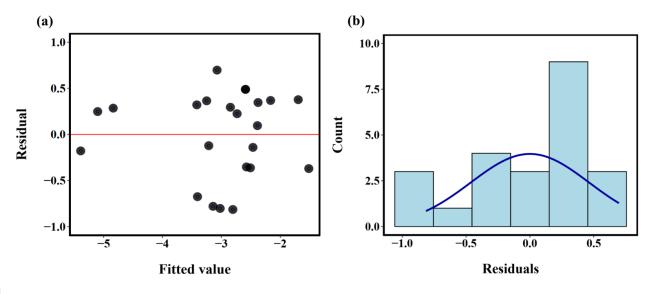




Figure S9. Residuals for individual data points of log γ_{SO2} (a) and the residual distribution (b)

196 from the multilinear regression follows a normal distribution.

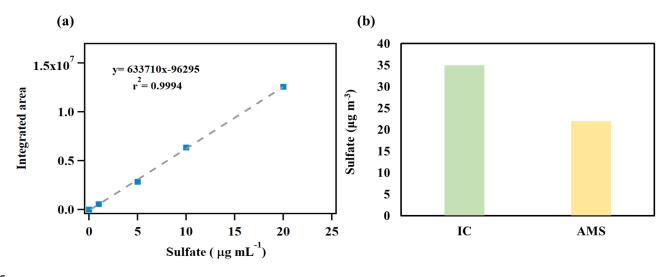




Figure S10. Comparison of sulfate measured by AMS and by ion chromatography (IC). Aerosol from the chamber SO₂ uptake experiments was collected, extracted and measured by an IC (Perkin Elmer, USA). (a) IC calibration curve for SO_4^{2-} . (b) Sulfate quantification comparison between the off-line IC measurement and on-line AMS measurement. Results from AMS presents a lower sensitivity than IC in terms of quantifying total aerosol sulfate during the chamber experiment in this study.

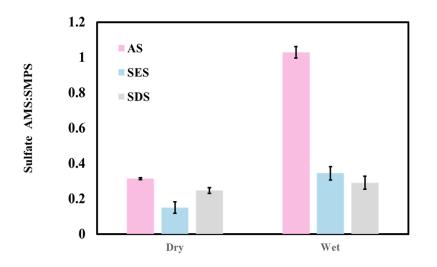




Figure S11. Comparison of SMPS measured sulfate and AMS measured sulfate under dry (with diffusion drier) and humid conditions (RH 80 %) for ammonium sulfate (AS), sodium ethyl sulfate (SES) and sodium dodecyl sulfate (SDS). AMS collection efficiencies for sulfate were considered to be 0.5 and 1 under dry and humid conditions, respectively(Matthew et al., 2008;Middlebrook et al., 2012). Under humid condition, the AMS quantified sulfate can be 3-5 times lower than the SMPS quantified sulfate in terms of organosulfate. The less effective response of AMS sulfate quantification towards organosulfate was also investigated in the work by Chen et al.(2019).

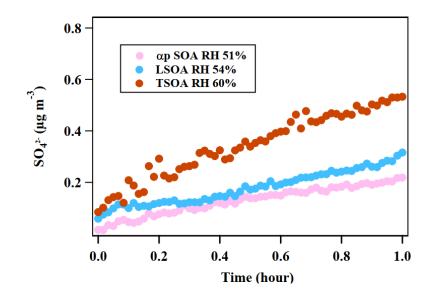


Figure S12. Time series of sulfate formation monitored by AMS during chamber experiments for the α pinene SOA (α p SOA), limonene SOA (LSOA) and toluene SOA (TSOA). The γ _{SO2} of TSOA was estimated from the γ_{SO2} of LSOA, and corrected by the sulfate formation ratio between the two SOA systems, where $\frac{d_{SO2_TSOA}}{d_{SO2_LSOA}} = \frac{AMS_{SO4_TSOA}}{AMS_{SO4_LSOA}}$.

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