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Supplement of

Significant production of ClNO_2 and possible source of Cl_2 from N_2O_5 uptake at a suburban site in eastern China

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26 **Text S1:** CIMS calibrations and data validation

27 **S1.1.** Dependence of the N₂O₅ sensitivity on RH

28 In the ion molecular reaction (IMR) chamber, the reagent ion I⁻ reacts with H₂O to form the
29 iodide water cluster, I(H₂O) which also reacts with N₂O₅ to produce IN₂O₅⁻ (Kercher et al.,
30 2009). Also, N₂O₅ may undergo hydrolysis in the sampling system. Thus, the sensitivity of N₂O₅
31 depends on the RH. In this study, the N₂O₅ signal was normalized to the I(H₂O)⁻ signal (Hz 145)
32 to account for the change of primary ions. During the field measurements, we monitored the
33 RH at the indoor inlet of CIMS. When conducting calibrations, we tested the relationship
34 between the normalized N₂O₅ sensitivity and RH (Fig. S5). A quadratic relationship in Fig. S5
35 ($y = -3.78 \times 10^{-9}x^2 + 1.69 \times 10^{-7}x + 1.72 \times 10^{-5}$) was used to correct the RH effect on the ambient N₂O₅
36 data.

37
38 **S1.2.** Isotopic analysis of ClNO₂ and Cl₂

39 The ClNO₂ signals were recorded at mass 208 and 210 amu, representing ³⁵ClNO₂ and
40 ³⁷ClNO₂, respectively. The relationship between the 208 and 210 signals was examined (Fig.
41 S4). During ambient samplings, the slope of the Hz 210-Hz 208 plot was 0.3135 with R²=0.998
42 (Fig. S4a). And the slope was 0.3154 with R²=0.999 (Fig. S4a) during calibrations. The isotopic
43 analysis was also performed for Cl₂. The correlations between mass 197 amu (³⁵Cl³⁵Cl) and 199
44 amu (³⁵Cl³⁷Cl) were excellent with R²=0.999 for calibration data and R²=0.965 for the ambient

45 data (Fig. S4b). The slope of the plot was 0.599 and 0.558 for the ambient data and calibration
 46 data, respectively, which is similar to previous studies (Liao et al., 2014). These results
 47 confirmed the identity of ClNO₂ and Cl₂ and indicated virtually no interference for ClNO₂ and
 48 negligible interference for Cl₂.

49

50 S1.3 Potential artifact of the inlet

51 When sampling ambient air, ambient particles gradually deposit on the inner wall of the
 52 sampling tubing. After a period of time, N₂O₅ in the ambient air reacts with the deposited
 53 particles, resulting in N₂O₅ loss and ClNO₂ formation. This inlet chemistry may cause
 54 underestimation of N₂O₅ and overestimation of ClNO₂. To minimize the interference from the
 55 sampling inlet, we adopted a virtual impactor design and a by-pass flow. The inlet design
 56 ensured that larger particles were mostly pumped through the by-pass flow. And the increased
 57 total flow (10 Lpm) reduced the residence time of N₂O₅ on the inlet. The sampling line was
 58 replaced daily by a cleansed one just before dusk to achieve minimum artifact on nighttime
 59 measurements.

60 We quantified the percentage of N₂O₅ loss and ClNO₂ formation in the inlet under different
 61 RH. After sampling for 24 hours, the used sampling line was taken indoor and connected to a
 62 zero-air generator with a flow rate of 10 Lpm. Then, we injected N₂O₅ at one end of the
 63 sampling line and measured the outflow at the other end in CIMS. The injected mixing ratios
 64 of N₂O₅ was determined by introducing N₂O₅ directly into the CIMS without passing the
 65 sampling line. The CIMS only inhaled ~1.5 Lpm airflow while the remaining ~8.5 Lpm airflow
 66 was discarded as a by-pass flow. The percentage of N₂O₅ loss and ClNO₂ yield (ClNO₂
 67 production divided by N₂O₅ loss) increased with RH. When RH = 40 %, the N₂O₅ loss was
 68 16.6 %. Thus, we assumed that the inlet artifact caused up to 16.6 % uncertainties for the
 69 ambient measurement of N₂O₅ and ClNO₂. 40 % RH was selected because the average RH
 70 recorded at the inlet of the CIMS was 40 % during the whole campaign. Cl₂ formation on the
 71 sampling tube was negligible in the wall-loss testing.

72

73 S1.4. Details of Cl₂ calibration

74 The Cl₂ standard was generated from a permeation tube heated to 40 °C and flushed by ultrapure
 75 nitrogen gas (20 sccm) and then diluted in humidified zero air (6 SLPM). During the field
 76 campaign, Cl₂ from the permeation tube was introduced into a KI solution (2% wt) for 1 hour.
 77 The permeation rate of Cl₂ (380 ± 20 ng/min) was calculated from the I₃⁻ concentration in the
 78 KI solution which was measured by ultraviolet–visible spectrophotometry at 351 nm.

79

80 **Table S1.** Measuring technique, detection limit and time resolution of the instruments in the
 81 field study. Detection limits were determined by 3σ of the noise level in 10 min.

Measured species	Techniques	Detection limits	Time resolution
N ₂ O ₅ , ClNO ₂ , Cl ₂ , HOCl, BrCl	Q-CIMS	2~7 pptv	10 s
NO, NO ₂	Chemiluminescence with photolytic converter	0.06 ppbv	1 min
NO _y	Chemiluminescence with MoO converter	0.1 ppbv	1 min
CO	Infrared photometry	4 ppbv	1 min

SO ₂	Pulsed ultraviolet fluorescence	0.1 ppbv	1 min
O ₃	Ultraviolet photometry	0.5 ppbv	1 min
HONO	LOPAP	5 pptv	1 min
HNO ₃	ion chromatography	0.05 ppbv	1 hour
PM _{2.5}	TEOM	1 µg/m ³	1 min
NH ₄ ⁺ , Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻	ToF-ACSM	0.01~0.06 µg/m ³	10 min
jNO ₂	Filter radiometer	4×10 ⁻⁵ s ⁻¹	10 s
VOCs	PTR-TOF-MS	10 pptv	10 min

82

83 **Table S2.** Summary of $\gamma(\text{N}_2\text{O}_5)$, $\varphi(\text{ClNO}_2)$, and $\varphi(\text{Cl}_2)$ (where applicable) in the selected 15

84 nighttime cases.

plume	start	end	$\gamma(\text{N}_2\text{O}_5)$	$\varphi(\text{ClNO}_2)$	$\varphi(\text{Cl}_2)$
1	4/12/18 2:10	4/12/18 3:00	0.0043	0.885	0.037±0.004
2	4/12/18 3:10	4/12/18 3:40	0.0068	0.716	0.021±0.003
3	4/12/18 21:40	4/13/18 0:40	0.0061	0.853	0.036±0.004
4	4/16/18 19:50	4/16/18 20:30	0.0031	0.378	0.013±0.000
5	4/16/18 20:40	4/16/18 21:20	0.0033	0.541	0.012±0.001
6	4/17/18 22:20	4/17/18 23:40	0.0058	0.521	0.010±0.001
7	4/18/18 3:00	4/18/18 3:50	0.0135	0.483	-
8	4/18/18 4:10	4/18/18 4:40	0.0139	0.187	-
9	4/19/18 0:00	4/19/18 0:40	0.0055	0.280	0.014±0.001
10	4/19/18 0:40	4/19/18 1:40	0.0041	0.523	0.018±0.002
11	4/19/18 2:00	4/19/18 3:00	0.0091	0.769	0.006±0.001
12	4/20/18 1:00	4/20/18 2:00	0.0084	0.641	0.013±0.001
13	4/20/18 2:10	4/20/18 2:50	0.0074	0.647	0.013±0.000
14	4/26/18 1:20	4/26/18 2:00	0.0136	0.468	0.006±0.001
15	4/26/18 2:30	4/26/18 3:20	0.0125	0.533	0.013±0.002
Average ± standard deviation			0.008±0.004	0.562±0.197	0.016±0.010

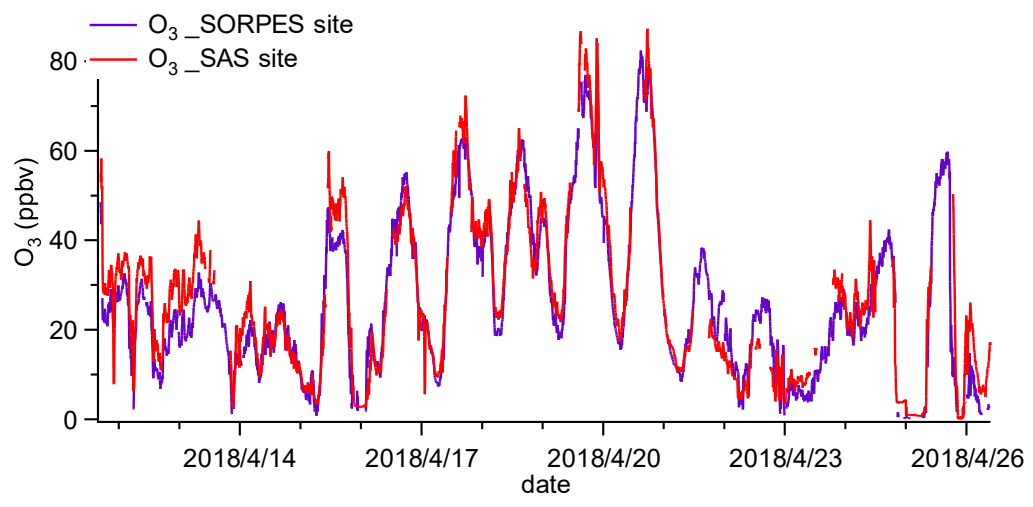
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86 **Table S3.** Uncertainty analysis of the measured and deduced parameters. The uncertainty of87 Cl⁻, NO₃⁻, SO₄²⁻, and S_a were referred to previous studies (Tham et al., 2016;Tham et al., 2018).

Parameter	sources of uncertainty (if any)			Propagated error	Reference
N ₂ O ₅ , ClNO ₂	Signal precision	calibration	inlet interference	18.8 %	This study
	3.0 %	8.3 %	16.6 %		
Cl ₂	Signal precision	calibration	inlet interference	10.4 %	This study
	3.0 %	10.0 %	neglected		
[Cl ⁻], [NO ₃ ⁻], [H ₂ O]	ACSM		E-AIM model	18.0 %	This study
	10.0 %		15 % (assumed)		
[H ⁺]	Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻ , NH ₄ ⁺		E-AIM model	25.0 %	This study
	10 %, 10 %, 10 %, 10 %		15 % (assumed)		
$\gamma(\text{N}_2\text{O}_5)$	N ₂ O ₅ , ClNO ₂	NO ₃ ⁻	S _a	34.2 %	This study

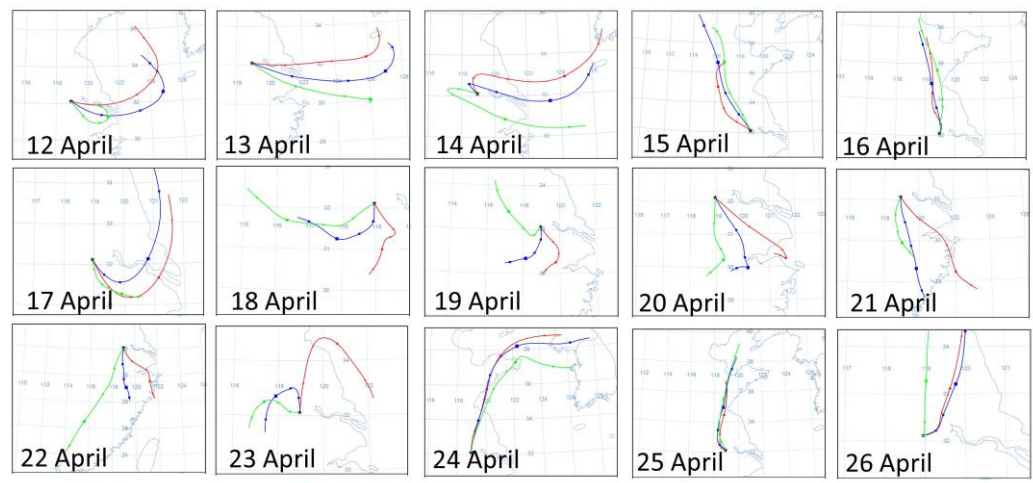
	18.8 %, 18.8 %	10.0 %	19.0 %		
$\varphi(\text{ClNO}_2)$	ClNO_2 18.8 %	NO_3^- 10.0 %		21.3 %	This study
$\varphi(\text{Cl}_2)$	$\gamma(\text{N}_2\text{O}_5)$ 34.2 %	$\text{N}_2\text{O}_5, \text{Cl}_2$ 18.8 %, 10.4 %	S_a 19.0 %	44.6 %	This study
$\text{Cl}^-, \text{NO}_3^-, \text{H}_2\text{O}$				10.0 %	Tham et al. 2016
S_a				19.0 %	Tham et al. 2018

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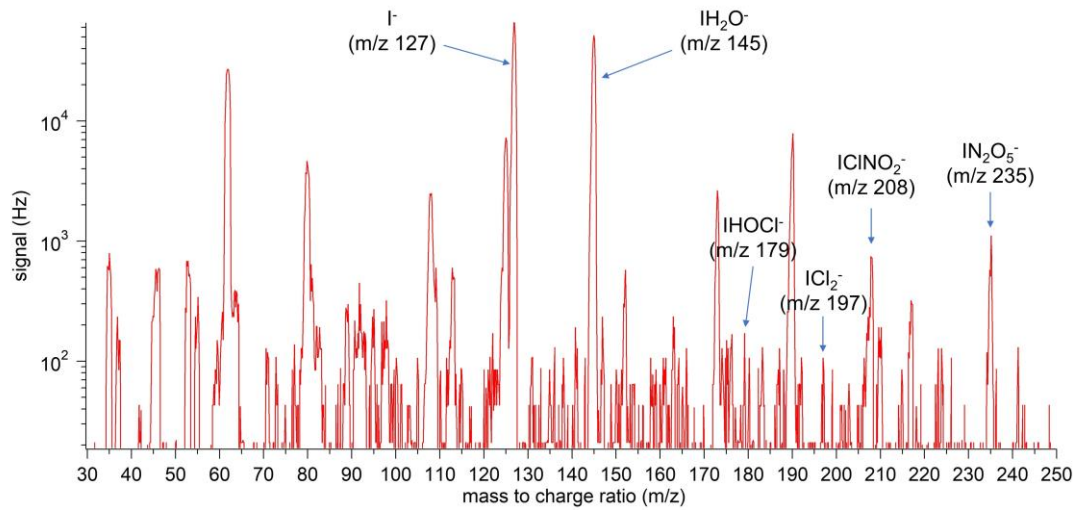
Figure S1. Comparison of O_3 measurements at the SORPES site and the SAS site.



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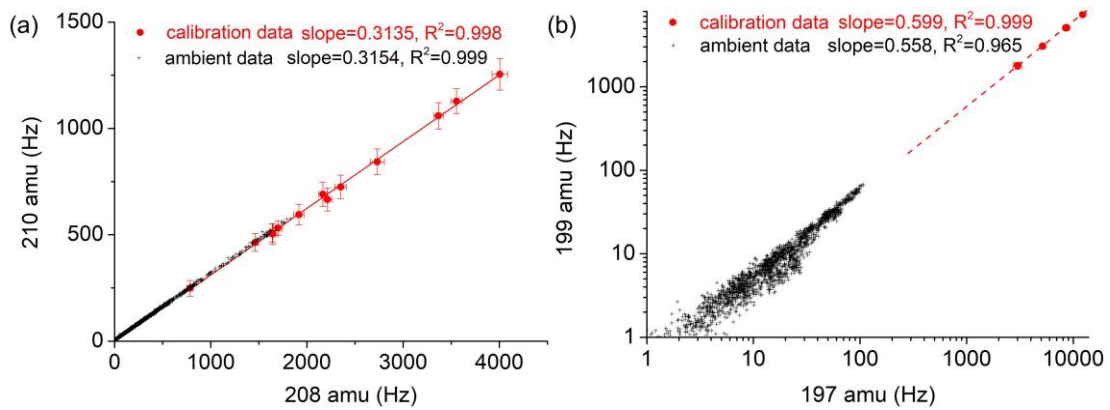
Red: 200m height. Blue: 500 m height. Green: 1000 m height. Local time: 00:00.

Figure S2. Daily backward trajectories arriving at the sampling sites during the field observation period.



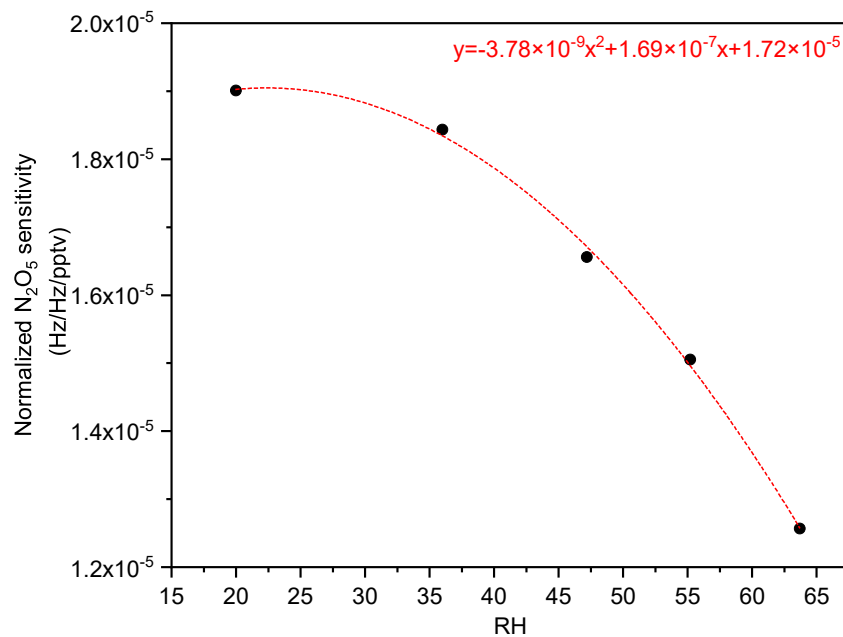
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Figure S3. An example of the CIMS spectra taken at 18 April 01:00 LT.



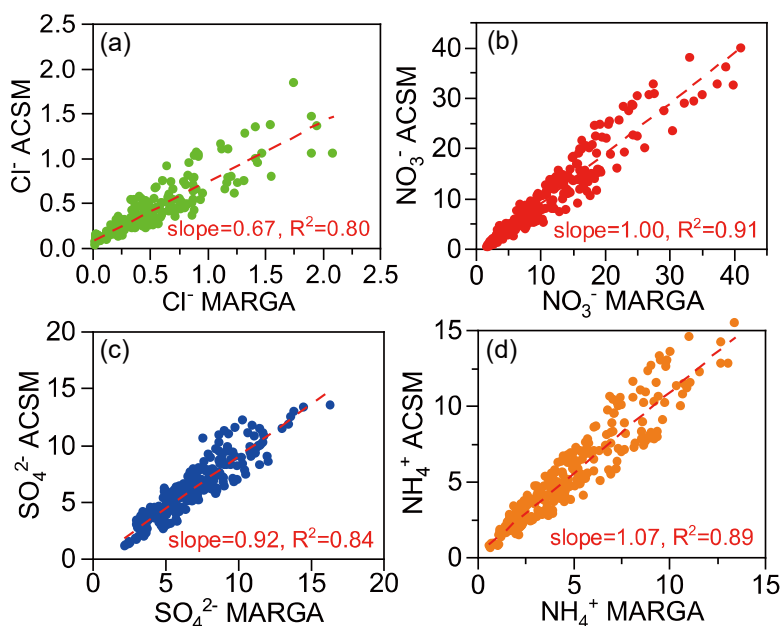
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Figure S4. Isotopic analysis of (a) ClNO_2 and (b) Cl_2 . The red dots and corresponding fitting lines represent calibration data, while the black crosses “+” denote ambient data.



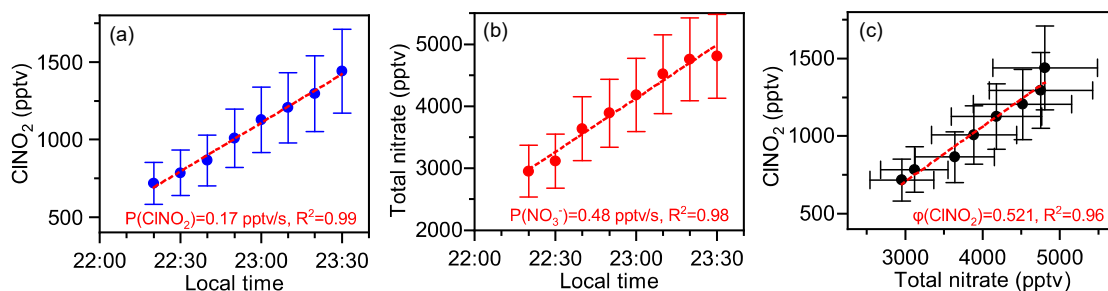
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105 **Figure S5.** Dependence of the N_2O_5 sensitivity on RH. The normalized sensitivity of N_2O_5 (y
 106 axis) was fitted as a quadratic function of RH (x axis), which is $y=-3.78\times 10^{-9}x^2+1.69\times 10^{-7}x+1.72\times 10^{-5}$ ($R^2=1$).
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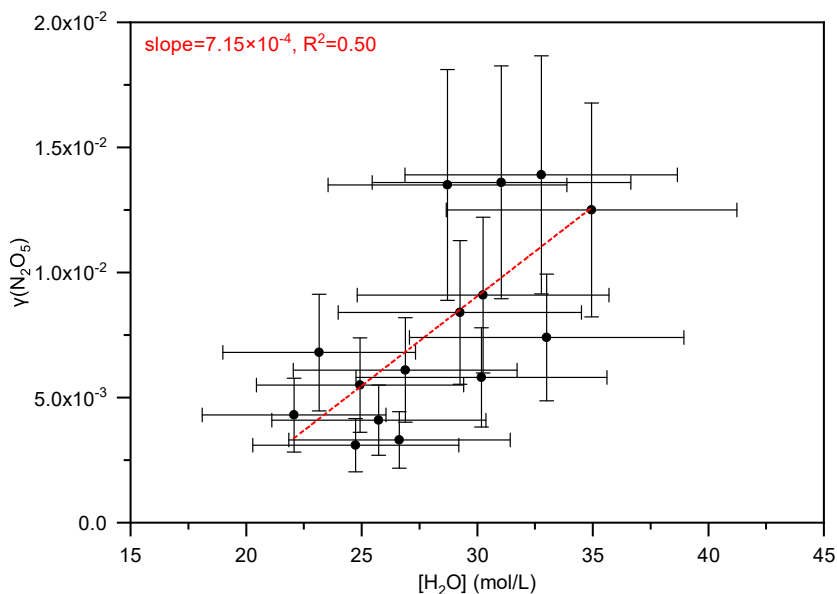
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Figure S6. Comparison of ACSM and MARGA data. (a), (b), (c), and (d) showed the comparison of Cl^- , NO_3^- , SO_4^{2-} , and NH_4^+ during the whole campaign, respectively. Since the resolution of the MARGA data was 1 hour, the ACSM data was averaged to 1 hour. Units of the ions are all $\mu\text{g}/\text{m}^3$.



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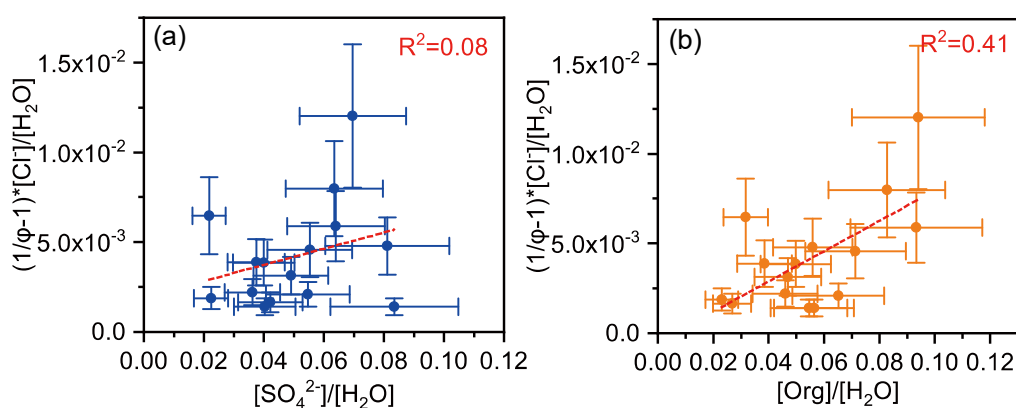
Figure S7. An example showing the calculation of $\gamma(\text{N}_2\text{O}_5)$ and $\phi(\text{CINO}_2)$. (a) and (b) show the increasing rate of CINO_2 and total nitrate observed on the night of Apr 17. (c) displays the relationship between CINO_2 and total nitrate shown in (a) and (b).



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122 **Figure S8.** Dependence of $\gamma(\text{N}_2\text{O}_5)$ on $[\text{H}_2\text{O}]$.

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124

125 **Figure S9.** Investigation of the potential of sulfate and total aerosol organics to consume NO_2^+ .

126 **(a)** and **(b)** represent sulfate and total organic aerosols, respectively.

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