



# Technical note: Preparation and purification of atmospherically relevant $\alpha$ -hydroxynitrate esters of monoterpenes

Elena Ali McKnight, Nicole P. Kretekos, Demi Owusu, and Rebecca Lyn LaLonde

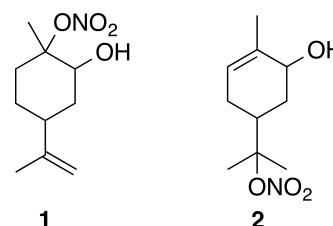
Chemistry Department, Reed College, Portland, OR 97202, USA

**Correspondence:** Rebecca Lyn LaLonde (rlalonde@reed.edu)

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**Abstract.** Organic nitrate esters are key products of terpene oxidation in the atmosphere. We report here the preparation and purification of nine nitrate esters derived from (+)-3-carene, limonene,  $\alpha$ -pinene,  $\beta$ -pinene and perillal alcohol. The availability of these compounds will enable detailed investigations into the structure–reactivity relationships of aerosol formation and processing and will allow individual investigations into aqueous-phase reactions of organic nitrate esters.



**Figure 1.** Two hydroxynitrate esters with available spectral data. Relative stereochemistry is undefined.

## 1 Introduction

Biogenic volatile organic compound (BVOC) emissions account for  $\sim 88\%$  of non-methane VOC emissions. Of the total BVOC estimated by the Model of Emission of Gases and Aerosols from Nature version 2.1 (MEGAN2.1), isoprene is estimated to comprise half, and methanol, ethanol, acetaldehyde, acetone,  $\alpha$ -pinene,  $\beta$ -pinene, limonene, ethene and propene together encompass another  $30\%$ . Of the terpenoids,  $\alpha$ -pinene alone is estimated to generate  $\sim 66 \text{ Tg yr}^{-1}$  (Guenther et al., 2012). These monoterpenes can be oxidized by nitrate radicals that are projected to account for more than half of the monoterpene-derived secondary organic aerosol (SOA) in the US (Pye et al., 2010). Nitrate oxidation pathways have been shown to be important particularly during nighttime. A large portion ( $30\%$ – $40\%$ ) of monoterpene emissions occur at night (Pye et al., 2010). These emissions can then react with  $\text{NO}_3$  radicals, formed from the oxidation of  $\text{NO}_2$  emissions by  $\text{O}_3$  (Pye et al., 2010).

The full role of organic nitrates (ONs) is complicated with many different sources and sinks (Perring et al., 2013). Fully deconvoluting the atmospheric processing of terpene-

derived ON is difficult, particularly due to partitioning into the aerosol phase in which hydrolysis and other reactivity can occur (Bleier and Elrod, 2013; Rindelaub et al., 2014, 2015; Romonosky et al., 2015; Thomas et al., 2016). Hydrolysis reactions of nitrate esters of isoprene have been studied directly (Jacobs et al., 2014) and the hydrolysis of ON has been studied in bulk (Baker and Easty, 1950). These and other studies have shown that the hydrolysis of ON is dependent on structure (Darer et al., 2011). For example, primary and secondary ON are thought to be relatively stable (Hu et al., 2011). In contrast, tertiary nitrates have been shown to hydrolyze on the order of hours (Boyd et al., 2015; Liu et al., 2012) to minutes (Darer et al., 2011). To the best of our knowledge there is only one study of the hydrolysis of an isolated terpene-derived hydroxynitrate (2 in Fig. 1) (Rindelaub et al., 2016a).

Furthermore, fully understanding the atmospheric processes of organic molecules is restricted by the ability to identify these species (Nozière et al., 2015). Part of this challenge is, of course, related to the lack of available standards. While one certainly cannot synthesize all of the atmospherically relevant ON, having access to representative

compounds from monoterpenes would enable key studies. With these molecules in hand, the atmospheric chemistry community could directly study the ON reactivity, such as hydrolysis, and deconvolute the structure–reactivity relationships. Additionally, novel method development would be enabled and validated (Rindelaub et al., 2016b). For example, Nozière and co-authors called attention to “the lack of NMR spectra libraries for atmospheric markers” as a barrier to utilizing nuclear magnetic resonance (NMR) spectroscopy in atmospheric science. While the preparation of  $\alpha$ -hydroxynitrates of terpenes has been alluded to in a handful of reports, we have only been able to identify NMR data for two species (Fig. 1). Two main methods appear to have been attempted, treating an epoxide with either fuming nitric acid (Rollins et al., 2010) or bismuth nitrate (Rindelaub et al., 2016b; Romonosky et al., 2015). The utility of the former is limited by the extreme hazards involved with mixing fuming nitric acid with organic materials (Parker, 1995; Univ. of California, Berkeley, 2019) and only provides characterization data for a single compound. On the surface the latter appears to be a usable method but on closer inspection lacks spectral data, perhaps due to intractable and inseparable mixtures (Romonosky et al., 2015), and has been the subject of a retracted study (Pöschl et al., 2011).

## 2 Experimental

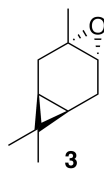
### 2.1 Instrumentation and materials

All starting materials and reagents were purchased from commercial sources and used without further purification unless otherwise noted. Bismuth nitrate (98 %) was purchased from Strem Chemicals and ground to a fine powder prior to use. Isoprene, methyl vinyl ketone (MVK), (+)-3-carene,  $\beta$ -pinene,  $\alpha$ -pinene oxide and 1,2-limonene oxide were purchased from Sigma-Aldrich. Silica gel chromatography was performed on a Teledyne Isco CombiFlash+ Lumen using 25  $\mu$ m SiliCycle spherical silica gel.  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were recorded on a Bruker AV400 spectrometer with chloroform (7.26 ppm) or benzene (7.28 ppm) as internal standards. Infrared (IR) spectra were recorded on a Thermo Nicolet IR100 spectrometer using a Thunderdome attenuated total reflectance (ATR) sample accessory. Melting points were collected on a DigiMelt 160 melting point apparatus. Exact masses were collected at the BioAnalytical Mass Spectrometry Facility at Portland State University using a ThermoElectron LTQ-Orbitrap Discovery high-resolution mass spectrometer with a dedicated Accela HPLC system. Low-resolution masses were collected on a Varian Saturn 2000 gas chromatographer–mass spectrometer (GC–MS) with a Restek Crossbind Rxi-5ms column. Abbreviations include *meta*-chloroperoxybenzoic acid (*m*-CPBA), hexanes (Hex), ethyl acetate (EtOAc), column volume (CV), thin layer chromatography (TLC), retention factor ( $R_f$ ), diastereomeric ra-

tio (d.r.), broad (br), singlet (s), doublet (d), doublet of doublets (dd), doublet of doublets of doublets (ddd), doublet of quartet (dq), triplet (t), multiplet (m), broad (b), weak (w), medium (m), strong (s) and very strong (vs).

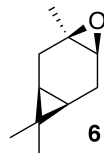
### 2.2 Preparation of epoxides

#### 2.2.1 *trans*-Carene oxide (3)



To a solution of (+)-3-carene (0.86 g, 1 mL, 7.34 mmol) in dichloromethane (25 mL) at 0 °C was added *meta*-chloroperoxybenzoic acid (*m*-CPBA) (1.85 g, 8.1 mmol, 1.1 equiv.). The solution was warmed to 23 °C over 1 h. The solution was poured onto saturated aqueous (sat. aq) sodium bicarbonate (30 mL) and extracted with dichloromethane (3  $\times$  30 mL). The combined organics were washed with sat. aq sodium bicarbonate (2  $\times$  30 mL), dried ( $\text{MgSO}_4$ ) and concentrated to yield a crude clear, colorless oil (1.17 g). The crude oil was purified by column chromatography ( $\text{SiO}_2$ ; 0 %–25 % EtOAc/Hex over 10 column volumes) to yield clear colorless oil (685 mg, 4.5 mmol; 61 % yield):  $^1\text{H}$  NMR (400 MHz, chloroform-*d*)  $\delta$  2.82 (s, 1 H), 2.28 (ddd,  $J$  = 16.4, 9.0, 1.9 Hz, 1 H), 2.13 (dd,  $J$  = 16.2, 9.1 Hz, 1 H), 1.63 (dt,  $J$  = 16.4, 2.3 Hz, 1 H), 1.48 (dd,  $J$  = 16.1, 2.3 Hz, 1 H), 1.24 (s, 3 H), 1.00 (s, 3 H), 0.72 (s, 3 H), 0.52 (td,  $J$  = 9.1, 2.3 Hz, 1 H), 0.44 (td,  $J$  = 9.1, 2.3 Hz, 1 H) ppm. Spectral data are consistent with literature reports (Cabaj et al., 2009).

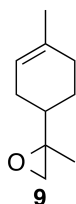
#### 2.2.2 *cis*-Carene oxide (6)



According to the methods of Cocker and Grayson (1969), a round bottom flask was charged with a solution of (+)-3-carene (2.7 g, 20 mmol) in dioxane (20 mL) and water (10 mL). Calcium carbonate (2 g, 20 mmol) and *N*-bromosuccinimide (7 g, 40 mmol) were added to the solution. The internal temperature rose to 50 °C after the initial addition. The mixture was stirred for 2 h then poured onto water (50 mL), filtered and washed with diethyl ether. The filtrate was extracted with ether (2  $\times$  100 mL). The combined extract was washed with water (3  $\times$  100 mL) and sodium thiosulfate (5 % aq, 20 mL), dried ( $\text{Na}_2\text{SO}_4$ ), and concentrated to yield a crude pale yellow oil. Purification by column chromatography ( $\text{SiO}_2$ ; 0 %–50 % EtOAc / Hex) yielded the bromohydrin as a white crystalline solid (4.6 g, 99 % yield):  $^1\text{H}$  NMR (400 MHz, chloroform-*d*)  $\delta$  4.07 (dd,  $J$  = 11.1, 7.6 Hz, 1 H),

2.50–2.36 (m, 2 H), 2.21 (dd,  $J = 14.6$ , 10.1 Hz, 1 H), 1.41 (dd,  $J = 4.9$ , 1.2 Hz, 1 H), 1.38 (s, 3 H), 1.03 (s, 3 H), 1.00 (s, 3 H), 0.91–0.80 (m, 1 H), 0.70 (m, 1 H). Spectral data are consistent with literature reports (Cocker and Grayson, 1969). Bromohydrin (2.44 g, 10.5 mmol) was dissolved in 100 mL *t*BuOH (warmed with a water bath). Potassium *tert*-butoxide (2.44 g, 21.8 mmol, 2.08 equiv.) was added, and the solution was stirred at rt for 2 h. The solution was poured onto water (50 mL), extracted with diethyl ether (3 × 75 mL), washed with water (3 × 50 mL), dried ( $\text{MgSO}_4$ ) and concentrated to yield 2.5 g crude clear oil. Purification by column chromatography (40 g  $\text{SiO}_2$ ; 0 %–25 % EtOAc / Hex over 10 CVs) yielded a clear colorless oil (1.31 g, 63 % yield):  $^1\text{H}$  NMR (400 MHz, chloroform-*d*)  $\delta$  2.89 (d,  $J = 5.6$  Hz, 1 H), 2.30 (ddd,  $J = 16.7$ , 9.0, 5.6 Hz, 1 H), 2.08 (dd,  $J = 16.4$ , 8.9 Hz, 1 H), 1.82 (s, 1 H), 1.78 (s, 1 H), 1.32 (s, 3 H), 0.98 (s, 3 H), 0.94 (s, 3 H), 0.63–0.49 (m, 2 H). Spectral data are consistent with literature reports (Cocker and Grayson, 1969).

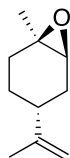
### 2.2.3 8,9-Limonene oxide (9)



Methyl vinyl ketone (10 mL, 90 mmol), isoprene (10 mL, 75 mmol) and dichloromethane (DCM) (90 mL) were added to a round bottom flask with stir bar. The flask was purged with inert atmosphere and chilled to 0 °C with stirring.  $\text{AlCl}_3$  (1.2 g, 9 mmol) was added in three portions over 10 min. The ice bath was removed and the reaction mixture was stirred for 1.5 h. The crude reaction mixture was filtered through a 1.5 in. (3.8 cm) silica gel pad (350 mL, 9 cm diameter) and washed with 8 % EtOAc : Hex (4 × 160 mL). The filtrate was concentrated to yield 1-(4-methyl-3-cyclohexene) ethenone (8 in Fig. 4) as a clear yellow liquid (9.68 g, 88 % yield):  $^1\text{H}$  NMR (400 MHz, chloroform-*d*)  $\delta$  5.44–5.38 (m, 1 H), 2.59–2.49 (m, 1 H), 2.15–2.20 (s, 4 H), 2.06–1.95 (m, 3 H), 1.67 (s, 3 H), 1.61 (m, 2 H). Spectral data are consistent with literature reports (Buss et al., 1987). Sodium hydride (1.766 g, 44.16 mmol) was suspended in dry dimethylsulfoxide (DMSO) (25 mL). A solution of trimethylsulfonium iodide (10.356 g, 47.06 mmol) in DMSO (50 mL) was added via cannula. The solution was stirred at rt for 30 min and then heated to 50 °C until gas evolution ceased (1 h). To this solution was added 1-(4-methyl-3-cyclohexen-1-yl) ethenone (8 in Fig. 4) (5 g, 36 mmol) and heated to 70 °C for 2 h. The reaction mixture was cooled to room temperature and poured onto sat. aq.  $\text{NH}_4\text{Cl}$  (100 mL), extracted with methyl *tert*-butyl ether (MTBE) (4 × 100 mL). The combined organics were washed with water (4 × 50 mL), dried ( $\text{MgSO}_4$ ) and concentrated to yield 5.1 g crude oil. Purification by

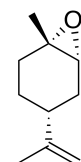
column chromatography (80 g  $\text{SiO}_2$ ; 0 %–20 % EtOAc / Hex over 10 CVs) yielded 8,9-limonene oxide (1 : 1 d.r.) as a clear, colorless oil (3.76 g, 68 % yield):  $^1\text{H}$  NMR (400 MHz, chloroform-*d*)  $\delta$  5.43–5.33 (br s, 1 H), 2.67 (t,  $J = 5.3$  Hz, 1 H), 2.58 (dd,  $J = 10.9$ , 4.8 Hz, 1 H), 2.17–1.73 (m, 5 H), 1.66 (s, 3 H), 1.44 (m, 2 H), 1.29 (d,  $J = 4.5$  Hz, 3 H). Spectral data are consistent with literature reports (Almeida and Jones Jr., 2005).

### 2.2.4 *cis*-1,2-Limonene oxide (12-*cis*)



According to the methods of Steiner et al. (2002), a 20 mL scintillation vial was charged with 1,2-limonene oxide, pyrrolidine and water. The vial was sealed with a Teflon-lined cap and heated to 88 °C for 24 h. The vial was cooled to rt, transferred to a separatory funnel with 100 mL pentane, and washed with sat. aq. ammonium chloride (3 × 30 mL) and water (1 × 50 mL). The organics were dried ( $\text{MgSO}_4$ ) and concentrated to yield a light yellow oil. Purification by column chromatography (80 g spherical  $\text{SiO}_2$ ; 0 %–15 % EtOAc / Hex over 10 CVs) yielded *cis*-1,2-limonene oxide as a clear, colorless oil. (1.52 g, 64 %):  $^1\text{H}$  NMR (400 MHz, chloroform-*d*)  $\delta$  4.77–4.71 (m, 1 H), 4.69 (s, 1 H), 3.07 (s, 1 H), 2.22–2.07 (m, 2 H), 1.90–1.84 (m, 2 H), 1.75–1.64 (m, 5 H), 1.33 (s, 3 H), 1.28–1.15 (m, 1 H) ppm. Spectral data are consistent with literature reports (Steiner et al., 2002).

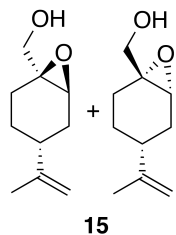
### 2.2.5 *trans*-1,2-Limonene oxide (12-*trans*)



According to the methods of Steiner et al. (2002), a 50 mL flask was charged with 1,2-limonene oxide (4.57 g, 30 mmol), pyrazole (0.34 g, 5 mmol) and water (16.2 mL). The reaction was heated to reflux for 5 h. The reaction mixture was cooled to 80 °C and transferred to a separatory funnel. Warm water (80 °C; 60 mL) was added. The emulsion was extracted with pentane (3 × 50 mL). The combined organic layers were dried ( $\text{MgSO}_4$ ) and concentrated to yield 1.481 g crude oil. Purification by column chromatography (80 g spherical  $\text{SiO}_2$ ; 0 %–20 % EtOAc / Hex over 13.5 CVs) yielded *trans*-1,2-limonene oxide (485 mg, 20 % yield):  $^1\text{H}$  NMR (400 MHz, chloroform-*d*)  $\delta$  4.68 (s, 2 H), 3.01 (d,  $J = 5.3$  Hz, 1 H), 2.11–2.00 (m, 2 H), 1.96–1.82 (m, 1 H), 1.75–1.64 (m, 5 H), 1.45–1.36 (m, 2 H), 1.34 (s, 3 H) ppm. Spec-

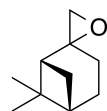
tral data are consistent with literature reports (Steiner et al., 2002).

### 2.2.6 Perillic alcohol epoxide (15)



Perillic alcohol (5 mL, 31.46 mmol) was combined with dichloromethane (62 mL) in a round bottom flask with stir bar and cooled to 0 °C under inert atmosphere. *m*-CPBA (5.97 g, 34.61 mmol) was added in five portions over the course of 10 min. After 30 min the reaction was warmed back up to room temperature and allowed to stir for an hour at room temperature. The reaction mixture was filtered through celite, washed with DCM (3 × 30 mL) and then transferred to a separatory funnel where it was washed with sodium bicarbonate (50 mL) and brine (50 mL). The organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated to yield a crude oil (5.1 g). Purification by column chromatography (80 g spherical SiO<sub>2</sub>; 0 %–100 % EtOAc / Hex over 15 CVs) yielded the epoxide (15) as a clear, colorless oil (3.363 g, 19.8 % yield): <sup>1</sup>H NMR (400 MHz, chloroform-*d*) δ 4.77–4.66 (m, 2 H), 3.78–3.59 (m, 2 H), 3.41–3.31 (m, 1 H), 2.26–2.14 (m, 1 H), 2.09 (doublet of triplet of doublet (dtd) *J* = 14.9, 5.6, 1.8 Hz, 1 H), 1.88–1.83 (m, 1 H), 1.80–1.59 (m, 7 H), 1.56–1.38 (m, 1 H), 1.32–1.16 (m, 1 H). Spectral data are consistent with literature reports (Thomas et al., 2016).

### 2.2.7 β-Pinene oxide



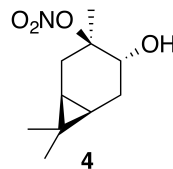
Potassium peroxide monosulfate (4.750 g, 31.20 mmol) was dissolved in deionized water (60 mL). Sodium bicarbonate (4.019 g, 47.84 mmol) was placed into a 125 mL Erlenmeyer flask and acetone (40 mL) was added, followed by β-pinene (1.59 mL, 9.974 mmol). The solution of potassium peroxide monosulfate mixture was slowly added into the β-pinene mixture over the course of 3 min via syringe while the mixture was stirring at 800 rpm. The reaction mixture was stirred for exactly 30 min while stirring at 800 rpm at room temperature. The reaction mixture was transferred to a separatory funnel and extracted with dichloromethane (2 × 40 mL), dried (MgSO<sub>4</sub>) and concentrated to produce a clear oil (1500 g, 96 % yield). <sup>1</sup>H NMR (400 MHz, chloroform-*d*) δ 2.79 (d, *J* = 4.8 Hz, 1 H), 2.62 (d, *J* = 4.8 Hz, 1 H), 2.30 (dtd, *J* = 10.3, 5.9, 1.5 Hz, 1 H), 2.19 (ddd, *J* = 15.1, 10.7, 8.1 Hz, 1 H), 2.06–1.97 (m, 1 H), 1.97–1.79 (m, 2 H), 1.75–

1.65 (m, 2 H), 1.53 (t, *J* = 5.4 Hz, 1 H), 1.27 (s, 3 H), 0.94 (s, 3 H). Spectral data are consistent with literature reports (Charbonneau et al., 2018).

## 2.3 General nitration method

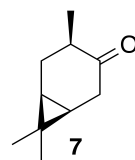
A round bottom flask was charged with a solution of epoxide (2 mmol) in toluene, dioxane or dichloromethane (10 mL, 0.2 M). Bismuth nitrate (1.164 g, 2.4 mmol, 1.2 equiv.) was added to the reaction mixture. The reaction mixture was stirred for 30–60 min. When TLC indicated complete consumption of starting material, the reaction was filtered through a 1 in. (2.5 cm) celite pad and washed with DCM (2 × 15 mL). The filtrate was transferred to a separatory funnel and washed with sodium bicarbonate (3 × 15 mL). The organic layer was dried with sodium sulfate, filtered and concentrated to yield a crude liquid. Desired products were isolated via column chromatography.

### 2.3.1 Nitrate ester (4)



Nitration of *trans*-3-carene oxide (3 mmol) was carried out according to the general method with dioxane. Purification of the crude oil by column chromatography (12 g SiO<sub>2</sub>, 0 %–30 % EtOAc / Hex) yielded nitrate ester (4) as a white crystalline solid (345 mg, 53 % yield): mp = 52.2–54.0 °C; IR (ATR) cm<sup>−1</sup> 3412 (br w, OH), 2944 (w), 2360 (w), 1616 (vs, NO<sub>2</sub>), 1291 (m, NO<sub>2</sub>), 868 (m, NO<sub>2</sub>); high-resolution mass spectrometry (HRMS) electrospray ionization (ESI) + *m/z* calculated for [C<sub>10</sub>H<sub>18</sub>O<sub>4</sub>N]<sup>+</sup> (216.1230, observed; 216.1236, expected) <sup>1</sup>H NMR (400 MHz, chloroform-*d*) δ 3.70 (dd, *J* = 9.6, 7.6 Hz, 1 H), 2.86–2.78 (m, 1 H), 2.16 (dd, *J* = 14.8, 7.5 Hz, 1 H), 1.83 (ddd, *J* = 14.8, 9.6, 7.9 Hz, 1 H), 1.65 (s, 3 H), 1.47 (dd, *J* = 14.3, 4.1 Hz, 1 H), 1.03 (s, 3 H), 1.01 (s, 3 H), 0.83–0.73 (m, 2 H); <sup>13</sup>C NMR (101 MHz, chloroform-*d*) δ 95.68, 71.03, 28.37, 28.34, 27.02, 19.95, 18.98, 17.95, 15.70, 14.39 ppm.

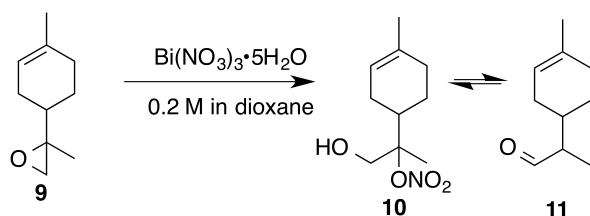
### 2.3.2 *cis*-4-Caranone (7)



Nitration of *cis*-3-carene oxide was attempted according to the general method with dioxane or dichloromethane. Purification of the crude oil by column chromatography (12 g SiO<sub>2</sub>, 0 %–40 % EtOAc / Hex) yielded *cis*-4-

caranone as a clear, colorless oil (53 % yield):  $R_f$  0.51 (5 % EtOAc:Hex; anisaldehyde);  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  2.54 (ddd,  $J = 18.0, 8.4, 1.0$  Hz, 1 H), 2.44–2.25 (m, 3 H), 1.33–1.20 (m, 1 H), 1.14–0.99 (m, 5 H), 0.97 (d,  $J = 6.4$  Hz, 3 H), 0.86 (s, 3 H);  $^{13}\text{C}$  NMR (101 MHz, chloroform- $d$ )  $\delta$  216.79, 41.99, 36.84, 29.77, 27.91, 22.83, 20.34, 19.47, 14.89, 14.11 ppm. Spectral data are consistent with literature reports (Kolehmainen et al., 1993).

### 2.3.3 Reaction of 8,9-limonene oxide (9)



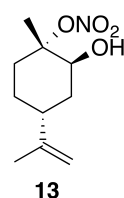
Nitration of 8,9-limonene oxide (9) was attempted according to the general method with acetonitrile or dioxane. Dioxane yielded a mixture of putative nitrate ester (10) and aldehyde (11) (1:0.25) as a crude oil. Nitrate (10) (1:1 d.r.):  $R_f$  0.25 (15 % EtOAc:Hex; anisaldehyde); IR (ATR)  $\text{cm}^{-1}$  3417 (br), 2913 (w), 1613 (vs,  $\text{NO}_2$ ), 1439 (w), 1289 (m,  $\text{NO}_2$ ), 865 (s,  $\text{NO}_2$ );  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  5.39–5.35 (m, 1 H), 4.06–3.86 (m, 2 H), 2.41–2.26 (m, 1 H), 2.09–1.82 (m, 5 H), 1.66 (s, 3 H), 1.49 (s, 1.5 H), 1.48 (s, 1.5 H), 1.43–1.31 (m, 2 H) ppm;  $^{13}\text{C}$  NMR (101 MHz, chloroform- $d$ )  $\delta$  134.31, 134.01, 119.57, 119.31, 97.40, 97.17, 64.62, 64.27, 38.24, 38.19, 30.58, 30.47, 26.42, 25.87, 24.19, 23.32, 23.25, 23.22, 17.17, 16.78 ppm. Purification by column chromatography (12 g  $\text{SiO}_2$  gold column, 0 %–50 % EtOAc / Hex over 30 CVs) yielded acetaldehyde (11) (1:1 d.r.) as a clear, colorless oil (105 mg, 35 % yield):  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  9.69 (d,  $J = 2.4$  Hz, 0.5 H), 9.68 (d,  $J = 2.7$  Hz, 0.5 H), 5.42–5.35 (m, 1 H), 2.35–2.25 (m, 1 H), 2.10–1.73 (m, 6 H), 1.68–1.62 (br s, 3 H), 1.46–1.32 (m, 1 H), 1.10 (s, 1.5 H), 1.08 (s, 1.5 H) ppm;  $^{13}\text{C}$  NMR (101 MHz, chloroform- $d$ )  $\delta$  205.55, 205.47, 134.09, 134.05, 119.95, 119.88, 51.00, 50.68, 34.32, 34.25, 30.15, 29.99, 29.67, 28.05, 27.31, 25.43, 23.43, 10.34, 10.24 ppm. Spectral data are consistent with literature reports (Uehara et al., 2017; Reid and Watson, 2018).



Acetonitrile yielded oxazoline (10a) (1:1 d.r.) as a clear, colorless oil (76 % yield):  $R_f$  0.46 (75 % EtOAc:Hex; anisaldehyde); IR (ATR)  $\text{cm}^{-1}$  3323 (br), 2963 (m), 2913 (m), 1674 (vs), 1437 (m), 1384 (m), 1265 (m), 991 (vs),

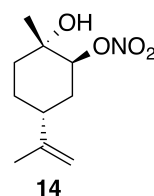
915 (m);  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  5.38 (s, 1 H), 4.10 (d,  $J = 6.2$  Hz, 0.5 H), 4.08 (d,  $J = 6.1$  Hz, 0.5 H), 3.82 (d,  $J = 8.3$  Hz, 0.5 H), 3.79 (d,  $J = 8.3$  Hz, 0.5 H), 2.16–1.91 (m, 6 H), 1.89–1.75 (m, 1 H), 1.73–1.56 (m, 5 H), 1.29–1.14 (m, 4 H);  $^{13}\text{C}$  NMR (101 MHz, chloroform- $d$ ) 162.73, 162.64, 134.23, 133.89, 120.28, 120.23, 77.22, 76.08, 75.39, 72.75, 72.60, 42.55, 42.28, 30.65, 30.57, 26.75, 26.39, 25.03, 24.58, 23.79, 23.75, 23.37, 13.88, 13.87 ppm; GC–MS (100–275  $^{\circ}\text{C}$ ; 15  $^{\circ}\text{C min}^{-1}$ ): 6.691 min,  $m/z$  193.8 ( $\text{M}^+$ , 100 %), 135.0 ( $\text{M} - [\text{C}_2\text{H}_3\text{NO}]$ , 10 %).

### 2.3.4 Nitrate ester (13)



Nitration of *cis*-1,2-limonene oxide was carried out according to the general method with dioxane or dichloromethane. Purification by column chromatography (12 g  $\text{SiO}_2$  gold column, 0 %–50 % EtOAc / Hex over 30 CVs) yielded nitrate ester (13) as a clear, colorless oil (53 % yield in dioxane; 62 % yield in DCM): IR (ATR)  $\text{cm}^{-1}$  3660 (w, alcoholic OH), 3340 (bd w, alcoholic OH), 2980 (m), 1618 (s,  $\text{NO}_2$ ), 1291 (m,  $\text{NO}_2$ ), 860 (vs,  $\text{NO}_2$ ); HRMS ESI+  $m/z$  calculated for  $[\text{C}_{10}\text{H}_{17}\text{O}_4\text{NNa}]^+$  (238.1052, observed; 238.1055, expected);  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  4.76 (dd,  $J = 8.1, 1.5$  Hz, 2 H), 4.13 (s, 1 H), 2.35 (doublet of doublet of triplet (ddt),  $J = 11.6, 8.4, 4.1$  Hz, 1 H), 2.24 (dtd,  $J = 14.8, 3.7, 1.4$  Hz, 1 H), 1.98–1.78 (m, 3 H), 1.76–1.73 (m, 4 H), 1.68–1.58 (m, 4 H), 1.49 (triplet of doublet of doublet (tdd),  $J = 13.3, 11.6, 3.6$  Hz, 1 H);  $^{13}\text{C}$  NMR (101 MHz, chloroform- $d$ )  $\delta$  148.49, 109.48, 91.34, 69.22, 36.84, 33.86, 29.90, 25.77, 20.98, 20.93 ppm.

### 2.3.5 Nitrate ester (14)



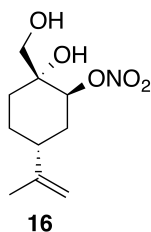
Nitration of *trans*-1,2-limonene oxide was carried out according to the general method with dioxane or dichloromethane. Purification by column chromatography (12 g  $\text{SiO}_2$ , 0 %–50 % EtOAc / Hex over 30 CVs) yielded nitrate ester (14) as a clear, colorless oil (63 % in dioxane; 54 % in DCM):  $R_f$  0.39 (15 % EtOAc:Hex; anisaldehyde); IR (ATR)  $\text{cm}^{-1}$  3414 (bd w, alcoholic OH), 2940 (w), 1627 (s,  $\text{NO}_2$ ), 1279 (m,  $\text{NO}_2$ ), 876 (m,  $\text{NO}_2$ ), 851 (m,  $\text{NO}_2$ ); HRMS

ESI+  $m/z$  calculated for  $[\text{C}_{10}\text{H}_{17}\text{O}_4\text{NNa}]^+$  (238.1048, observed; 238.1055, expected);  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  5.01 (s, 1 H), 4.76 (d,  $J = 6.6$  Hz, 2 H), 2.19 (tt,  $J = 10.9, 3.1$  Hz, 1 H), 2.03 (ddd,  $J = 14.7, 12.1, 2.6$  Hz, 1 H), 1.92 (dq,  $J = 14.4, 3.5, 2.7$  Hz, 1 H), 1.74 (s, 3 H), 1.69–1.56 (m, 4 H), 1.32 (s, 3 H);  $^{13}\text{C}$  NMR (101 MHz, chloroform- $d$ )  $\delta$  148.27, 109.57, 84.43, 69.55, 37.96, 34.78, 29.91, 26.96, 20.89 ppm.

### 2.3.6 Nitration of perillil alcohol epoxide (15)

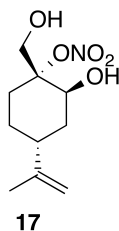
Nitration of 4-(1-methylethenyl)-7-oxabicyclo[4.1.0]heptane-1-methanol (15) was carried out according to the general method with dichloromethane. Purification by column chromatography (12 g  $\text{SiO}_2$ , 0%–40% EtOAc/Hex over 30 CVs) yielded two regioisomers and nitrate esters (16) and (17) in 23% and 32% yield, respectively.

#### Nitrate ester (16)



Nitrate ester (16) was isolated as a white, crystalline solid (106 mg, 23% yield): mp = 72–75 °C;  $R_f$  0.31 (30% EtOAc:Hex; anisaldehyde); IR (ATR)  $\text{cm}^{-1}$  3285 (bd w, alcoholic OH), 2941 (w), 1627 (vs,  $\text{NO}_2$ ), 1280 (s,  $\text{NO}_2$ ), 880 (s,  $\text{NO}_2$ ), 850 (s,  $\text{NO}_2$ ); HRMS ESI+  $m/z$  calculated for  $[\text{C}_{10}\text{H}_{17}\text{O}_4\text{NNa}]^+$  (238.1054, observed; 238.1055, expected);  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  5.23 (s, 1 H), 4.80–4.73 (m, 2 H), 3.74 (dd,  $J = 10.9, 5.0$  Hz, 1 H), 3.44 (dd,  $J = 11.0, 5.0$  Hz, 1 H), 2.59 (d,  $J = 1.0$  Hz, 1 H), 2.27–2.16 (m, 1 H), 2.03–1.97 (m, 2 H), 1.79 (t,  $J = 5.2$  Hz, 1 H), 1.71–1.61 (m, 3 H), 1.58 (d,  $J = 0.8$  Hz, 4 H);  $^{13}\text{C}$  NMR (101 MHz, chloroform- $d$ )  $\delta$  148.24, 109.58, 79.89, 70.98, 66.45, 38.47, 29.70, 29.55, 25.10, 20.77 ppm.

#### Nitrate ester (17)



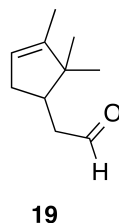
Nitrate ester (17) was isolated as a clear, colorless oil (149 mg, 32% yield):  $R_f$  0.20 (30% EtOAc:Hex; anisaldehyde); IR (ATR)  $\text{cm}^{-1}$  3286 (bd w, alcoholic OH), 2941 (w), 1623 (vs,  $\text{NO}_2$ ), 1288 (s,  $\text{NO}_2$ ), 863 (vs,

$\text{NO}_2$ ); HRMS ESI–  $m/z$  calculated for  $[\text{C}_{10}\text{H}_{16}\text{O}_4\text{N}]^-$  (214.10813, observed; 214.10848, expected);  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  4.76 (d,  $J = 8.5$  Hz, 2 H), 4.31 (s, 1 H), 4.27 (d,  $J = 13.0$  Hz, 1 H), 3.99 (d,  $J = 13.0$  Hz, 1 H), 2.41 (tt,  $J = 12.1, 3.6$  Hz, 1 H), 2.27–2.19 (m, 1 H), 1.98–1.78 (m, 3 H), 1.75 (s, 3 H), 1.72–1.65 (m, 1 H), 1.51–1.38 (m, 1 H);  $^{13}\text{C}$  NMR (101 MHz, chloroform- $d$ )  $\delta$  148.58, 109.43, 92.51, 65.67, 62.91, 37.47, 33.53, 25.33, 25.26, 20.86 ppm.

### 2.3.7 Nitration of $\alpha$ -pinene oxide

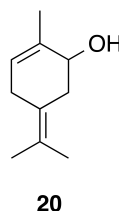
Nitration of  $\alpha$ -pinene oxide was carried out according to the general method. Crude oil from a 10 mmol reaction was purified by column chromatography (40 g  $\text{SiO}_2$ , 0%–40% EtOAc/Hex over 30 CVs) yielded the following isomerization products and nitrate esters.

#### $\alpha$ -Campholenic aldehyde (19)



$\alpha$ -Campholenic aldehyde (19) was isolated as a clear, colorless oil:  $R_f$  0.59 (10% EtOAc:Hex; anisaldehyde);  $^1\text{H}$  NMR (400 MHz, benzene- $d_6$ )  $\delta$  9.51 (t,  $J = 2.1$  Hz, 1 H), 5.23 (s, 1 H), 2.41 (doublet of doublet of doublet of triplet (dddt),  $J = 13.8, 6.0, 2.7, 1.6$  Hz, 1 H), 2.25–2.16 (m, 1 H), 2.11 (ddd,  $J = 15.9, 4.4, 1.8$  Hz, 1 H), 2.00 (ddd,  $J = 15.9, 10.3, 2.3$  Hz, 1 H), 1.80 (doublet of doublet of pentet (ddp),  $J = 15.8, 9.2, 2.5$  Hz, 1 H), 1.58 (dt,  $J = 3.0, 1.6$  Hz, 3 H), 0.90 (s, 3 H), 0.68 (s, 3 H). Spectral data are consistent with literature reports (Thomas et al., 2016).

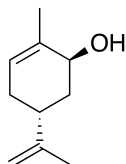
#### 5-Isopropylidene-2-methyl-2-cyclohexen-1-ol (20)



Compound (20) was isolated as a white, crystalline solid:  $R_f$  0.47 (20% EtOAc:Hex; anisaldehyde);  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  5.49 (ddd,  $J = 4.4, 3.2, 1.5$  Hz, 1 H), 3.99 (t,  $J = 4.2$  Hz, 1 H), 2.86 (d,  $J = 19.6$  Hz, 1 H), 2.69–2.56 (m, 2 H), 2.35 (d,  $J = 15.5$  Hz, 1 H), 1.78–1.75 (m, 3 H), 1.74 (s, 3 H), 1.68 (s, 3 H);  $^{13}\text{C}$  NMR (101 MHz, chloroform- $d$ )  $\delta$  135.94, 125.57, 124.23, 123.29, 70.34,

35.92, 29.83, 20.31, 20.14, 19.91 ppm. Spectral data are consistent with literature reports (Motherwell et al., 2004).

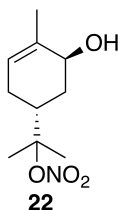
#### *trans*-Carveol (21)



21

*trans*-Carveol was isolated as a clear, colorless oil:  $R_f$  0.37 (20 % EtOAc : Hex; anisaldehyde);  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  5.62 (dd,  $J = 5.2, 1.5$  Hz, 1 H), 4.78–4.73 (m, 2 H), 4.06–4.03 (m, 1 H), 2.34 (ddd,  $J = 13.7, 9.8, 2.9$  Hz, 1 H), 2.21–2.12 (m, 1 H), 1.96 (dq,  $J = 13.6, 2.3$  Hz, 1 H), 1.93–1.85 (m, 1 H), 1.85–1.81 (m, 3 H), 1.77 (s, 3 H), 1.64 (dd,  $J = 13.2, 4.0$  Hz, 1 H), 1.61–1.57 (m, 1 H) ppm. Spectral data are consistent with literature reports (Motherwell et al., 2004).

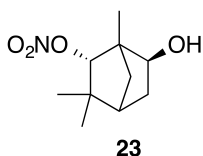
#### *trans*-Carveol nitrate ester (22)



22

Nitrate ester (22) was isolated as a clear, colorless oil: IR (ATR)  $\text{cm}^{-1}$  3353 (bd w, alcoholic OH), 2917 (w), 1614 (vs,  $\text{NO}_2$ ), 1293 (s,  $\text{NO}_2$ ), 868 (m,  $\text{NO}_2$ );  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  5.32 (ddd,  $J = 9.9, 3.5, 2.1$  Hz, 1 H), 4.12 (ddd,  $J = 10.6, 4.6, 1.9$  Hz, 1 H), 2.68–2.49 (m, 2 H), 1.88 (t,  $J = 4.8$  Hz, 1 H), 1.43 (dd,  $J = 14.1, 3.7$  Hz, 1 H), 1.24 (dd,  $J = 13.8, 4.8$  Hz, 1 H), 1.11 (s, 3 H), 0.97 (s, 3 H), 0.94 (s, 3 H) ppm. Spectral data are consistent with literature reports (Rindelaub et al., 2016b).

#### Nitrate ester (23)

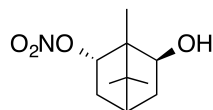


23

Nitrate ester (23) was isolated as a white, crystalline solid: mp = 132–134 °C;  $R_f$  0.29 (25 % EtOAc : Hex; anisaldehyde); IR (ATR)  $\text{cm}^{-1}$  3294 (bd w, alcoholic OH), 2961 (w), 2360 (w), 1624 (m,  $\text{NO}_2$ ), 1282 (m,  $\text{NO}_2$ ), 862 (m,  $\text{NO}_2$ ); HRMS ESI- $m/z$  calculated for  $[\text{C}_{10}\text{H}_{16}\text{O}_4\text{N}]^-$  (214.10797, observed; 214.10848, expected);  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  4.64 (s, 1 H), 4.00 (ddd,  $J = 7.1,$

3.3, 0.9 Hz, 1 H), 2.37 (ddd,  $J = 13.8, 7.1, 2.5$  Hz, 1 H), 1.81 (d,  $J = 4.3$  Hz, 1 H), 1.60–1.51 (m, 3 H), 1.32 (dt,  $J = 13.8, 4.0$  Hz, 1 H), 1.23 (s, 3 H), 1.22 (s, 3 H), 0.89 (s, 3 H);  $^{13}\text{C}$  NMR (101 MHz, chloroform- $d$ )  $\delta$  94.13, 68.52, 53.01, 47.34, 40.49, 37.76, 37.10, 29.24, 19.12, 14.75 ppm.

#### Nitrate ester (24)



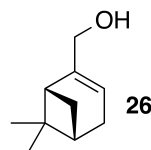
24

Nitrate ester (24) was isolated as a white, crystalline solid: mp = 62–64 °C;  $R_f$  0.24 (15 % EtOAc : Hex; anisaldehyde); HRMS ESI+ $m/z$  calculated for  $[\text{C}_{10}\text{H}_{17}\text{O}_4\text{NNa}]^+$  (238.10459, observed; 238.10498, expected); IR (ATR)  $\text{cm}^{-1}$  3434 (w, alcoholic OH), 2959 (w), 1622 (s,  $\text{NO}_2$ ), 1284 (vs,  $\text{NO}_2$ ), 856 (m,  $\text{NO}_2$ );  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  5.31 (ddd,  $J = 9.9, 3.8, 2.1$  Hz, 1 H), 4.12 (m, 1 H), 2.69–2.48 (m, 2 H), 1.87 (t,  $J = 4.9$  Hz, 1 H), 1.43 (dd,  $J = 14.0, 3.7$  Hz, 1 H), 1.24 (dd,  $J = 13.7, 4.7$  Hz, 1 H), 1.11 (s, 3 H), 0.97 (s, 3 H), 0.94 (s, 3 H);  $^{13}\text{C}$  NMR (101 MHz, chloroform- $d$ )  $\delta$  91.04, 78.09, 52.58, 49.53, 42.73, 39.05, 36.71, 20.20, 19.70, 11.79 ppm.

### 2.3.8 Nitration of $\beta$ -pinene oxide

Nitration of  $\beta$ -pinene oxide (11.1 mmol) was carried out according to the general procedure in toluene. Purification by column chromatography (80 g  $\text{SiO}_2$ , 0 %–30 % EtOAc / Hex over 20 CVs) yielded a 1 : 1 mix of myrtenol (26) and nitrate (29) (344 mg, 9 % combined yield), perillal alcohol (28, 600 mg, 36 % yield) and nitrate (27) (71 mg, 3 % yield). The combined myrtenol and nitrate (29) were separated by column chromatography (40 g  $\text{SiO}_2$ ; 5 %–20 % EtOAc / Hex over 25 CVs).

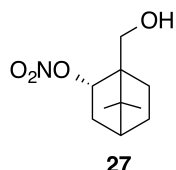
#### Myrtenol (26)



26

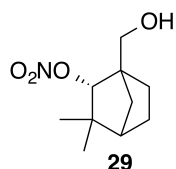
Myrtenol was isolated as a clear, colorless oil:  $R_f$  0.25 (15 % EtOAc : Hex; anisaldehyde);  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  5.49 (dt,  $J = 3.0, 1.5$  Hz, 1 H), 4.00 (t,  $J = 1.8$  Hz, 2 H), 2.42 (dt,  $J = 8.6, 5.6$  Hz, 1 H), 2.37–2.20 (m, 2 H), 2.19–2.09 (m, 2 H), 1.61–1.45 (m, 1 H), 1.31 (s, 3 H), 1.19 (d,  $J = 8.6$  Hz, 1 H), 0.85 (s, 3 H) ppm. Spectral data are consistent with literature reports (Motherwell et al., 2004).

## Nitrate ester (27)



Nitrate ester (27) was isolated as a clear colorless oil: IR (ATR)  $\text{cm}^{-1}$  3362 (bd w, alcoholic OH), 2947 (w), 1624 (s,  $\text{NO}_2$ ), 1281 (vs,  $\text{NO}_2$ ), 864 (m,  $\text{NO}_2$ ); HRMS ESI –  $m/z$  calculated for  $[\text{C}_{10}\text{H}_{16}\text{O}_4\text{N}]^-$  (214.10813, observed; 214.10848, expected);  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  5.57–5.49 (m, 1 H), 3.71 (q,  $J$  = 11.5, 8.5 Hz, 2 H), 2.55 (ddt,  $J$  = 14.1, 9.8, 3.9 Hz, 1 H), 1.95–1.83 (m, 2 H), 1.76 (t,  $J$  = 4.5 Hz, 1 H), 1.52–1.22 (m, 5 H), 1.10 (s, 3 H), 1.01 (s, 3 H);  $^{13}\text{C}$  NMR (101 MHz, chloroform- $d$ )  $\delta$  85.30, 62.08, 53.45, 48.62, 45.59, 35.78, 27.53, 22.88, 20.20, 20.02 ppm.

## Nitrate ester (29)



Nitrate ester (29) was isolated as a white, crystalline solid: mp = 59–61 °C;  $R_f$  0.55 (15 % EtOAc : Hex; anisaldehyde); IR (ATR)  $\text{cm}^{-1}$  3374 (bd w, alcoholic OH), 2915 (w), 1625 (s,  $\text{NO}_2$ ), 1281 (vs,  $\text{NO}_2$ ), 863 (m,  $\text{NO}_2$ ); HRMS ESI +  $m/z$  calculated for  $[\text{C}_{10}\text{H}_{17}\text{O}_4\text{NNa}]^+$  (238.10474, observed; 238.10498, expected);  $^1\text{H}$  NMR (400 MHz, chloroform- $d$ )  $\delta$  4.92 (d,  $J$  = 1.6 Hz, 1 H), 3.75 (d,  $J$  = 11.3 Hz, 1 H), 3.65 (d,  $J$  = 11.3 Hz, 1 H), 1.90–1.80 (m, 3 H), 1.72–1.63 (m, 1 H), 1.56–1.50 (m, 1 H), 1.24–1.20 (m, 4 H), 0.96 (s, 3 H);  $^{13}\text{C}$  NMR (101 MHz, chloroform- $d$ )  $\delta$  89.49, 63.82, 54.38, 48.25, 40.28, 37.03, 29.26, 24.74, 22.11, 19.46 ppm.

## 3 Results and discussion

## 3.1 Synthesis

Previous reports have described the opening of epoxides and aziridines using bismuth nitrate in acetonitrile (Das et al., 2007). Others report optimal conditions in dichloromethane (Rindelaub et al., 2016b), or 1,4-dioxane with undesired side reactions in acetonitrile (Pinto et al., 2007). Thus, we began our investigations into the nitration of *trans*-carene oxide (3) using bismuth nitrate in various solvents (Fig. 2). In our hands, dioxane clearly outperformed dichloromethane, yielding 53 % of the desired product after 45 min (entry 3). A diol was isolated as a major (16 %) byproduct. Attempts to mitigate hydrolysis by the use of base or molecular sieves

were ineffective. Acetonitrile produced a complex intractable mixture with a high amount of diol and caronaldehyde (entry 2). Other solvents, such as tetrahydrofuran (THF) and nitromethane, also produced complex mixtures with no trace of desired nitrate. Methanol, interestingly, only produced an undesired methyl ether. It is unclear if this product was generated through methanolysis of the nitrate ester or direct substitution of the epoxide. The regiochemistry of the nitration was easily elucidated with gHSQC NMR data. A doublet of doublets at 3.69 ppm correlated with a carbon shift at 71.1 ppm and was thus assigned to the alcohol methyne carbon. A tetrasubstituted carbon, with no correlations in the gHSQC at 95.6 ppm was consistent with the tertiary nitrate ester. Finally, the IR of compound (4) displayed the expected strong absorbances at 1616, 1291 and 868  $\text{cm}^{-1}$ .

Next, we aimed to investigate the impact of relative stereochemistry on the nitration. We hypothesized that the secondary nitrate would predominate due to classical stereoelectronic effects. (+)-3-Carene was treated with *N*-bromosuccinimide and calcium carbonate to produce a bromohydrin which subsequently cyclized under basic conditions to produce *cis*-carene oxide (Cocker and Grayson, 1969). Interestingly, we never observed the desired nitrate ester. Instead a 1,2-hydrogen shift generated *cis*-4-caranone (7) in 53 % yield.

The preparation of 8,9-limonene oxide (9) began with a Diels–Alder reaction to form 1-(4-methyl-3-cyclohexene) ethenone (8) in 88 % yield. Corey–Chaykovsky addition of a sulfur ylide cleanly produced 8,9-limonene oxide (9). Nitration of (9) was attempted in a variety of solvents (DCM, benzene, dioxane, acetonitrile, nitromethane). In acetonitrile, oxazoline (10a) (1 : 1 d.r.) was cleanly produced presumably via nucleophilic addition of acetonitrile and subsequent cyclization to the oxazoline. Pinto et al. (2007) observed a similar bismuth nitrate mediated epoxide opening with acetonitrile. In dioxane, aldehyde (11) (1 : 1 d.r.) was observed along with a product consistent with a tertiary nitrate ester (10) (1 : 1 d.r.) in a 0.25 : 1 mixture. Formation of 11-enol presumably proceeds via a facile intramolecular elimination of nitric acid. The IR of the crude mixture showed the expected strong absorbances at 1613, 1288 and 865  $\text{cm}^{-1}$ . A set of signals in the  $^{13}\text{C}$  NMR at 97.40 and 97.17 ppm, with no gHSQC correlations, is consistent with a tertiary nitrate. Attempts to purify this mixture on silica gel gave small amounts of (11) and unidentifiable decomposition products. Furthermore, the fast hydrolysis rates of tertiary nitrate esters hinder the ability to isolate these products (Boyd et al., 2015; Darer et al., 2011; Liu et al., 2012). No reaction was observed when non-polar solvents such as DCM and benzene were employed. We hypothesize that the steric hindrance at the  $\beta$ -carbon severely limits reactivity.

Commercially available 1,2-limonene oxide was resolved into pure samples of *cis*- and *trans*-1,2-limonene oxide by treating with cyclic amine bases (Steiner et al., 2002). The nitration of the *cis* isomer was predicted to produce





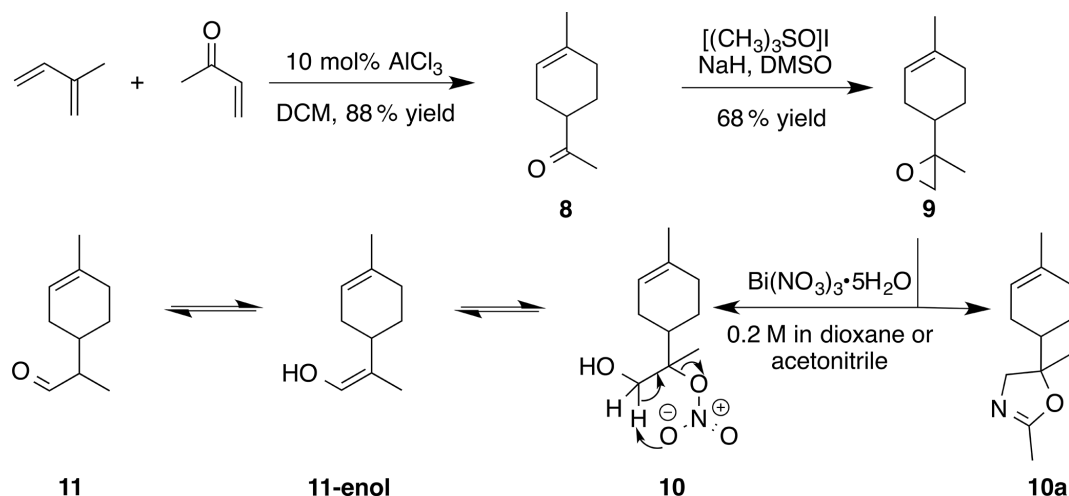


Figure 4. Preparation of 8,9-limonene oxide and attempted nitration.

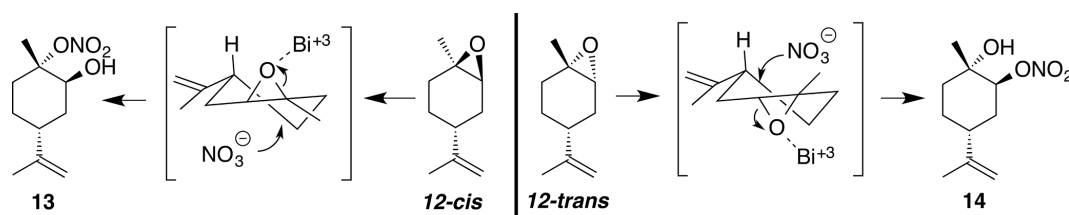


Figure 5. Nitration of *cis*- and *trans*-1,2-limonene oxide.

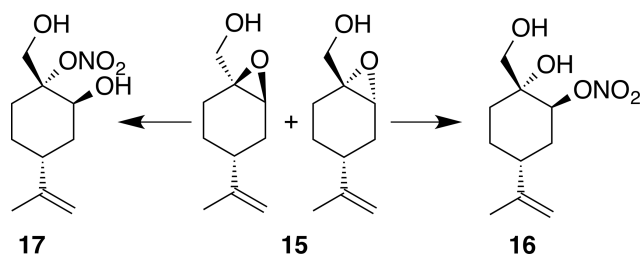


Figure 6. Nitration of perillaldehyde oxide.

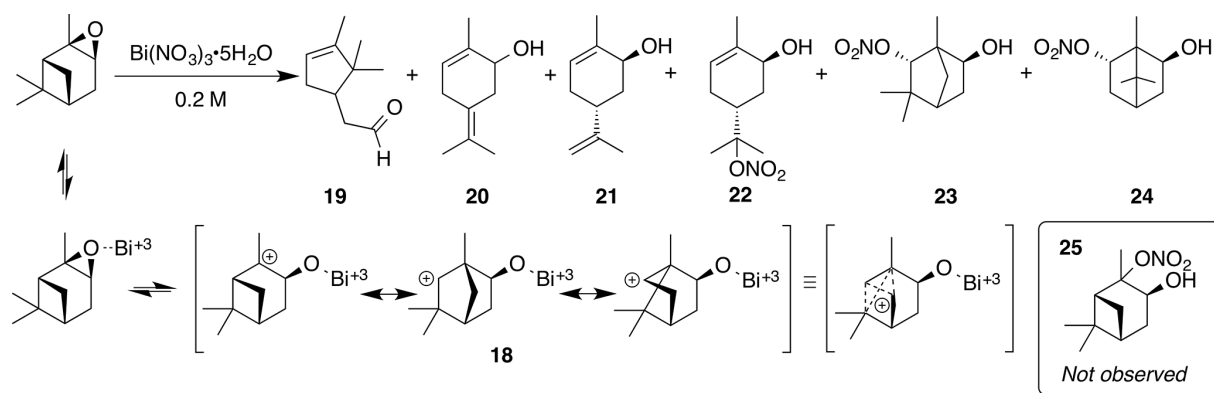
Under all conditions campholenic aldehyde was the major product (20 %–28 % yield). First generation nitrate ester (25) was not found under all conditions. All six components were isolated in small amounts when the reaction was run in dichloromethane (Fig. 7, entry 1). Cooling the reaction to  $-78^{\circ}\text{C}$  completely shut down all reactivity (entry 2). Interestingly, adding one equivalent of tetrabutylammonium nitrate (TBAN) as an external nitrate source in addition to bismuth nitrate at  $-78^{\circ}\text{C}$  generated mostly nitrate (22) (entry 3). More polar solvents, like dioxane (entry 4), generated slightly higher amounts of nitrates at room temperature. Aromatic solvents, such as benzene and toluene, provided the best yield of (23). Interestingly, adding TBAN to the nitration in toluene dramatically increased the amounts of diene (20) and *trans*-carveol (21). Using acetonitrile as a solvent gen-

erated a complex mixture that appeared to be mainly diols. Zirconium nitrate produced a similar distribution of products (Das et al., 2006), but other metal nitrate complexes ( $\text{Y}(\text{NO}_3)_3 \cdot 6\text{H}_2\text{O}$ ,  $\text{Co}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ ) resulted in no observable reaction.

The nitration of  $\beta$ -pinene was similarly complicated by isomerization pathways through the corresponding non-classical carbocation (Fig. 8). Nitrate (27) initially co-eluted with myrtenol (26) but was separable upon a second purification by column chromatography. The methyne proton of (27) at 5.51 ppm is correlated with a carbon at 85.3 ppm in the gHSQC NMR. These data are consistent with the secondary nitrate ester. Similarly, the methyne proton in nitrate (29) is a doublet at 4.91 ppm ( $J^4 = 1.6\text{ Hz}$ ; W coupling). This signal correlates with a carbon at 89.5 ppm. Interestingly, nitrates (30) and (31) were not observed.

### 3.2 Spectral data

As shown in Table 1, all nitrate esters displayed the expected strong nitrate ester absorbances at  $\sim 1600$ , 1300 and  $900\text{ cm}^{-1}$ . Methyne protons next to the secondary nitrate esters appeared between 4.64 and 5.51 ppm. Carbon chemical shifts were found over a broader range than anticipated from 80.1 to 95.6 ppm. Finally, we observed masses consistent with methanolysis products ( $185.1\text{ m/z M}^+$ ;



Entry	Solvent	Temp (°C)	Additive	%19	%20	%21	%22	%23	%24
1	$\text{CH}_2\text{Cl}_2$	23	--	21	2	5	2	1	4
2	$\text{CH}_2\text{Cl}_2$	-78	--	--	--	--	--	--	--
3	$\text{CH}_2\text{Cl}_2$	-78	1 equiv TBAN	20	--	6	13	3	--
4	Dioxane	23	--	25	--	--	5	5	2
5	Toluene	23	--	22	1	4	4	8	--
6	Toluene	23	1 equiv TBAN	28	8	13	6	4	2

Figure 7. Nitration of  $\alpha$ -pinene oxide.

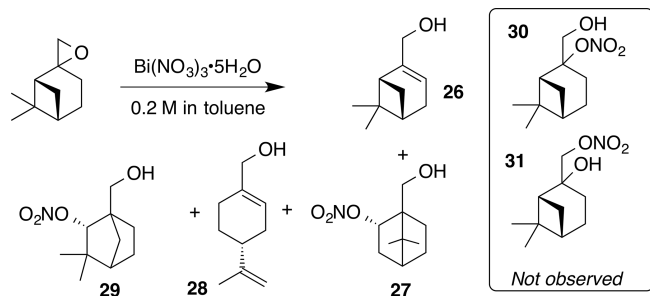


Figure 8. Nitration of  $\beta$ -pinene oxide.

223.1  $m/z$  M + K) when nitrate esters were analyzed (GC–MS) as solutions in methanol. Further experimentation is necessary to evaluate the implications of methanolysis.

### 3.3 Stability and storage

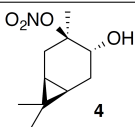
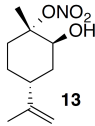
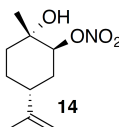
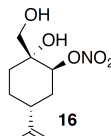
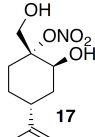
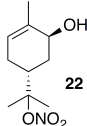
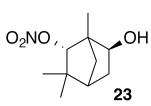
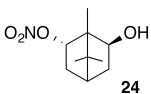
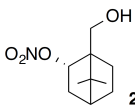
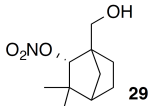
We were surprised to find that five of the nitrate esters (4, 16, 23, 24 and 29) are solids. We have stored these compounds at 0 °C for up to 9 months with no noticeable decline in purity. The remaining nitrate esters are stable at 0 °C for 2–4 weeks. Interestingly, some of these compounds appear to deviate from the expected stability patterns (*vide supra*). For example, tertiary nitrate ester (4) is particularly stable stored as a solid at 0 °C. In contrast, when secondary nitrate ester (14) was stored as a neat oil at 0 °C, it decomposed to a complex mixture within a few weeks. Freezing the sam-

ples as a solution in benzene is recommended for longer-term (~ 6 months) storage.

## 4 Conclusions

We have clearly delineated successful methods required to synthesize and purify nine nitrate esters derived from monoterpene. Seven of these compounds are undescribed in the literature and the remaining two had gaps in their characterization. Using our methods it is possible to cleanly isolate 50–100 mg even of the least prevalent isomers from the nitration reactions of  $\alpha$ -pinene and  $\beta$ -pinene. Interestingly, we did not observe the formation of  $\alpha$ -pinene oxide and  $\beta$ -pinene oxide products that retained their bicyclic ring structures. This is consistent with the solution-phase synthetic literature (Kaminska et al., 1992) but in contrast to many atmospheric reports (for example, see Rindelaub et al., 2016b; Duporte et al., 2016). We believe that the availability of these compounds will enable further study of the structure–reactivity relationships. For example, comparing the specific hydrolysis rates for tertiary versus secondary nitrate esters in (13) and (14) as well as (17) and (16) could help deconvolute the fates of each terpene in the atmosphere. We also believe that these compounds will assist in confirming the identities of organic nitrates that have previously been limited to detection by MS-based methods (Rindelaub et al., 2016b). A forthcoming report will describe our investigation into the behavior of these authentic compounds in mass spectrometers. In par-

**Table 1.** Collated  $^1\text{H}$  and  $^{13}\text{C}$  NMR data with IR absorbances for nitrate esters.

Compound	$^1\text{H}$ NMR (ppm)	$^{13}\text{C}$ NMR (ppm)	IR ( $\text{cm}^{-1}$ )		
 4	—	95.6	1616	1291	868
 13	—	91.4	1618	1291	860
 14	5.00 (s, 1 H)	84.4	1627	1279	876
 16	5.21 (s, 1 H)	80.1	1627	1280	880
 17	—	92.5	1623	1288	863
 22	—	94.2	1614	1293	868
 23	4.64 (s, 1 H)	94.13	1624	1282	862
 24	5.31 (ddd, $J = 9.9, 3.8, 2.1$ Hz, 1 H)	91.04	1622	1284	856
 27	5.51 (m, 1 H)	85.3	1624	1281	864
 29	4.91 (s, 1 H)	89.5	1625	1281	863

ticular, the detailed MS–MS data, impact of various ionization conditions (ESI, chemical ionization (CI), etc.) and sample preparation will be described. Finally, the availability of these compounds is important for further studies into the influence of terpene structure on the fate and roles of organic nitrates in SOA formation.

**Data availability.** All spectral data for new compounds are available in the Supplement to this article.

**Supplement.** The supplement related to this article is available online at: <https://doi.org/10.5194/acp-20-4241-2020-supplement>.

**Author contributions.** RLL designed the experiments and prepared the manuscript with contributions from all co-authors. All co-authors carried out experiments and contributed to reviewing and editing the manuscript.

**Competing interests.** The authors declare that they have no conflict of interest.

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