



Supplement of

Dithiothreitol activity by particulate oxidizers of SOA produced from photooxidation of hydrocarbons under varied NO_x levels

Huanhuan Jiang et al.

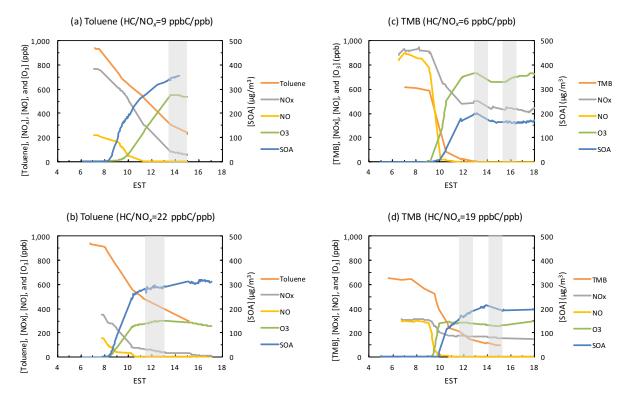
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Section S1. Outdoor chamber experiments.

A non-reactive compound, CCl_4 (500-1000 ppb), was injected into the chamber as a dilution tracer. Gas phase concentrations of hydrocarbons (HCs) and CCl_4 were measured by a gas chromatography-flame ionization detector (GC-FID, HP-5890). The concentration of NO_x was

5 monitored by a chemiluminescence analyzer (Teledyne, Model T201). The concentration of O_3 was monitored by a photometric ozone analyzer (Teledyne, Model 400E). The relative humidity and temperature were monitored by a hygrometer (CS215-L, Campbell Scientific). The typical time profiles of SOA mass concentration, HCs (i.e. toluene and TMB), NO_x, NO and O₃ mixing ratios through the experiments were shown in Fig. S1.



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Figure S1. The time profiles of SOA mass concentration, HCs, NO_x, NO and O₃ mixing ratios through the experiments: (a) toluene (HC/NO_x=9 ppbC/ppb); (b) toluene (HC/NO_x=22 ppbC/ppb); (c) TMB (HC/NO_x=6 ppbC/ppb); (d) TMB (HC/NO_x=19 ppbC/ppb). EST, Eastern Standard Time. The shaded areas indicate the time periods of aerosol sampling.

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Section S2. Chemicals and solution preparation for chemical assays.

Toluene (99.5 %), 1,3,5-trimethylbenzene (99 %), and hydrogen peroxide (50 wt.% aqueous solution) were from Acros Organics (NJ, USA). Isoprene (99 %), α-pinene (98 %), CCl₄ (99.9 %), potassium phosphate buffer (0.1 M), 9,10-phenanthraquinone (99 %), 1,4-naphthoquinone

- 5 (97 %), mesityl oxide (90 %), 2,4-hexadienal (>95 %), sulfanilic acid (99 %), n-(1-naphthyl) ethylenediamine dihydrochloride (≥98 %), potassium hydroxide (≥85 %), sodium nitrite (≥97 %), tert-butyl hydroperoxides (5.0-6.0 M in decane), peracetic acid (32 wt.%), tert-butyl peroxide (98 %), dimethyl sulfoxide (99.9 %) and 4-nitrophenyl boronic acid (≥95 %) were from Sigma-Aldrich (St. Louis, MO, USA). Dithiothreitol (99 %), Tris base (99.8-100.1 %), 5,5'-
- 10 dithiobis-(2-nitrobenzoic acid) (99 %), trichloroacetic acid (99 %), and deionized (DI) water (ACS, ASTM Type I) were obtained from Thermo Fisher Scientific (Pittsburgh, PA, USA). Acrolein (97 %, stabilized by hydroquinone) was from Alfa Aesar (Ward Hill, MA, USA). The preparation details of 9,10-phenanthraquinone (PQN), 1,4-naphthoquinone (NQN), tertbutyl peroxides, and tert-butyl hydroperoxides were shown below:
- 15 PQN stock solution: 5 mM PQN standard stock solution was made by dissolving 20.821 mg of PQN in 20 mL DMSO. The stock solution was kept at 4 °C when not in use. Before chemical analysis, the PQN stock solution was diluted using DI water.

NQN standard: 5 mM NQN standard stock solution was made by dissolving 31.630 mg of NQN in 20 mL DMSO. The stock solution was kept at 4 °C when not in use. Before chemical analysis, the NQN stock solution was diluted using DI water.

analysis, the NQN stock solution was diluted using DI water.
 Tert-butyl peroxide: 26 mM tert-butyl peroxide solution was prepared by adding 0.1 mL tert-butyl peroxide (98 %) into 20 mL DI water. The tert-butyl peroxides stock solution was further diluted to get a 100 μM solution using DI water.

Tert-butyl hydroperoxides: 22 mM tert-butyl hydroperoxide solution was prepared by adding

25 0.08 mL tert-butyl hydroperoxide (5.0-6.0 M in decane) into 19.92 mL methanol. The tert-butyl hydroperoxides stock solution was further diluted to get a 96 μM solution using methanol.

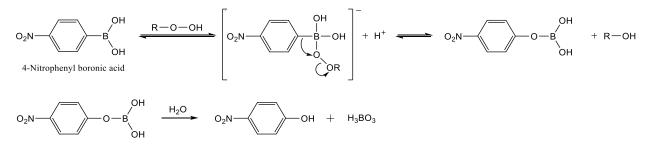
Section S3. Reaction schemes for chemical assays.

DTT assay. DTT assay was employed to quantify the oxidative potential of SOA (Cho et al., 2005; Jiang et al., 2016). DTT was oxidized when mixed with SOA. The remaining DTT can be quantified using 5,5'-dithiobis-(2-nitrobenzoic acid), which reacts with DTT forming 2-nitro-5-

5 thiobenzoic acid, as shown in Scheme S1.



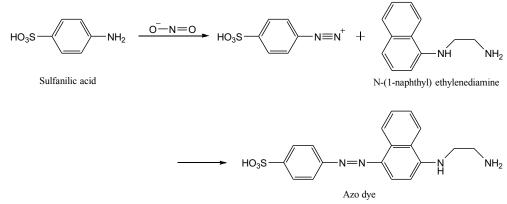
NPBA assay. Organic hydroperoxides react with 4-nitrophenylboronic acid (NPBA) producing 4-nitrophenol (Scheme S2) which has a large absorption coefficient at 406nm.



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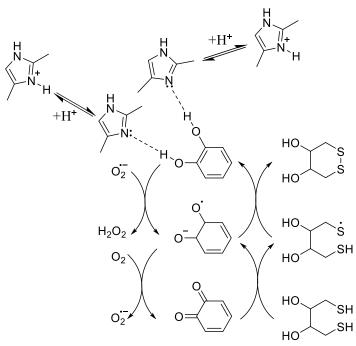
Scheme S2. The reaction between NPBA and organic hydroperoxides (Kuivila, 1954; Kuivila and Armour, 1957).

15 **Griess assay.** PAN is hydrolysed by KOH and forms nitrites. A nitrite reacts with the Griess reagent (sulfanilic acid and n-(1-naphthyl) ethylenediamine dihydrocloride) to form an azo dye, as shown in Scheme S3.



Scheme S3. The reaction between Griess reagent (sulfanilic acid and n-(1-naphthyl 20 ethylenediamine) and nitrites (Giustarini et al., 2008).

DTT enhancement assay. By forming H-bonds with hydroquinones, 2,4-dimethylimidazole facilitated the electrons transfer from hydroquinones to molecular oxygen, accelerating the redox cycling of quinones and enhancing the oxidation of DTT, as shown in Scheme S4 (Dou et al., 2015).



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Scheme S4. The mechanism of DTT enhancement assay (Dou et al., 2015).

Section S4. QC/QA for the chemical assays

S4.1. DTT assay.

Limit of detection (LOD). LOD is defined as three times the standard deviation (σ) of blank controls. The LOD of DTT assay (DTT_t) was 0.23 nmol min⁻¹.

- 5 Blank control. To estimate the effect of radicals produced by sonication on DTT assay, the DTT loss rate in blank control during sonication was compared to that during shaking (Edison Environmental Incubator Shaker G24, low speed, 37 °C). As shown in Fig. S2, the DTT loss rate in blank control was 0.162 nmol min⁻¹ during sonication and 0.153 nmol min⁻¹ during shaking. Tested using the statistical method based on the Student t-test (Andrade and Estévez-Pérez,
- 10 2014), the slope of DTT loss vs. time with the sonicator was not significantly different from the one with the shaker (significance level $\alpha = 0.05$); therefore, in this study, the influence of free radicals generated by sonication on DTT measurement was insignificant.

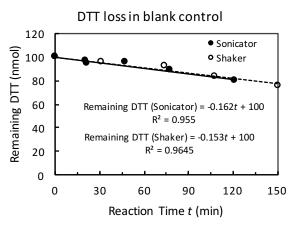


Figure S2. DTT loss in blank control (deionized water) during sonication or shaking (37 °C).

15 Positive controls. The DTT consumption of positive controls (PQN and NQN) was measured. As shown in Fig. S3, the DTT consumption of quiones increased linearly with reactin time due to the catalytic reactions between quinones and DTT.

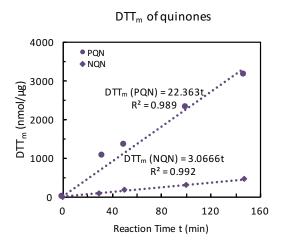
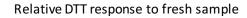
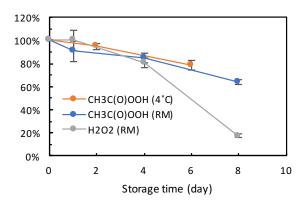


Figure S3. DTT_m of quinones (PQN: 9,10-phenathraquinone, NQN: 1,4-naphthoquinone).

The stability of peroxides. To investigate the impact of the stability of organic compounds on 5 DTT actitivities, the DTT consumptions (ΔDTT at reaction time=100 min) by 400 µL H₂O₂ (100 µM) and 300 µL peracetic acid (CH₃C(O)OOH; 100 µM) were measured as a function of storage time. As shown in Fig. S4, ΔDTT of H₂O₂ and CH₃C(O)OOH stored at room temperature (RM) decreased upto 80 % and 36 % after an 8-day storage, respectivley. ΔDTT of CH₃C(O)OOH stored at 4 °C also decreased 22% after a 6-day storage.



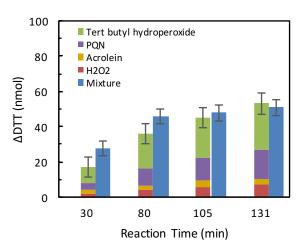


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Figure S4. The DTT responses of H_2O_2 and peracetic acid stored at room temperature (RM) or 4 °C. The errors was estimated using the standard deviation of three replicates.

The additivity of DTT activity by chemical species. To investigate the additivity of DTT response from different types of chemical species, ΔDTT of the blend of several compounds was compared with the sum of ΔDTT originating from individual compounds. Fig. S5 proves that DTT consumption is additive by showing that ΔDTT of the mixture of four model compounds

(i.e., acrolein, PQN, H_2O_2 , and tert-butyl hydroperoxides) was consistent with the sum of individuals.



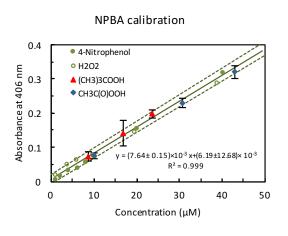
 ΔDTT of a mixture of model compounds

Figure S5. ΔDTT of individual model compounds and the compounds mixture (54 nmol acrolein,
5 15 nmol H₂O₂, 280 nmol tert-butyl hydroperoxides, 0.025 nmol PQN). The error bar associated with ΔDTT of a stacked column was estimated by the sum of the error bars from individuals.

Section S4.2. NPBA assay.

Limit of detection. The LOD of NPBA assay was $0.47 \mu M$.

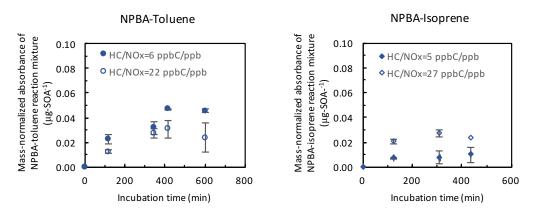
Calibration. The NPBA method was calibrated using aqueous 4-nitrophenol solutions ranging from $1-40 \mu$ M, as shown in Fig. S6.



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Figure S6. The calibration curve of NPBA assay was obtained at the 90 % confidence level. The calibration curve was applied to test the feasibility of using NPBA assay to quantify the concentration of H_2O_2 , tert-butyl hydroperoxides ((CH₃)₃COOH), and peracetic acid (CH₃C(O)OOH).

10 The time dependence of NPBA assay. To investigate the reaction time needed to quantify the organic hydroperoxides concentration using NPBA assay, the toluene SOA and isoprene SOA samples were incubated with NPBA at 85 °C for different reaction time. As shown in Fig. S7, the reaction of organic hydroperoxides with NPBA completed within 7 h for toluene SOA and 2h for isoprene SOA.



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Figure S7. Time profile of the absorbance (normalized by SOA mass) of NPBA-SOA reaction mixture at 85 °C. The error bar of each data point represents the standard deviation of three replicates.

The influence of multi-alcohols and 4-nitrophenol on NPBA assay. It has been reported that boronic acid can react with multi-alcohols to form colorful products (Kim et al., 2007). SOA products are usually multifunctional and contain multi-alcohols. To examine the possible

- 5 interference by NPBA-alcohol adducts, the glycerol aqueous solution was applied to the NPBA method. However, no measurable absorption appeared in the UV spectrum. 4-Nitrophenol, a NPBA assay product, can be found in toluene SOA and potentially influences on NPBA data. However, the light absorption of the SOA sample collected using PILS was negligible at 406 nm, suggesting that NPBA data is not influenced by the light absorbing materials in SOA products.
- 10 The stability of peroxides. To investigate the impact of the stability of peroxides on NPBA assay, the NPBA repsonses for 400 μ L H₂O₂ (100 μ M) and 300 μ L peracetic acid (100 μ M) were measured as a function of storage time. Fig. S8 illustrates that the NPBA response of H₂O₂ and CH₃C(O)OOH stored at room temperature decreased upto 70 % and 27 % after an 8-day storage, respectivley. The decrease of the NPBA response of CH₃C(O)OOH stored at 4 °C was similar to
- 15 that stored at room temperature.

Relative NPBA response to fresh sample

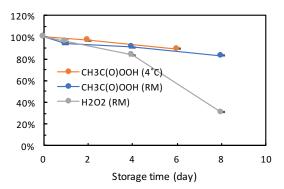


Figure S8. The NPBA responses of H_2O_2 and peracetic acid stored at room temperature or 4 °C. The error bar was estimated using the standard deviation of three replicates.

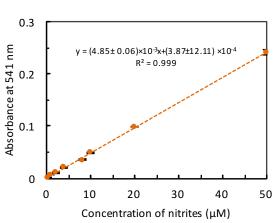
S4.3. Griess assay.

Limit of detection. LOD of Griess assay was 0.60 µM.

Hydrolysis time of PAN products in SOA. In this study, the hydrolysis time for PANs was set to 15 min. To ensure that 15 min is enough to hydrolyse the PANs in SOA products, the Griess response with 15-min hydrolysis was compared to that with 30-min hydrolysis. There was no

5 response with 15-min hydrolysis was compared to that with 30-min hydrolysis. There was no significant difference in Griess response between a 15-min hydrolysis and a 30-min hydrolysis.

Calibration. The Griess assay was calibrated using NaNO₂ solutions with varied concentrations $(1-50 \ \mu\text{M})$, as shown in Fig. S9.



Griess Calibration

10 Figure. S9 Calibration curve of Griess assay. The error bar associated with each data point was calculated by $t_{0.90} \times \sigma / \sqrt{n}$, where $t_{0.90}$ is the t-score (2.920 for n = 3 replicates) with a two-tail 90 % confidence level, and σ is the standard deviation.

Section S5. Lifetime of electron-deficient alkenes under the typical chamber condition.

Electron-deficient alkenes produced from the photooxidation of hydrocarbons can further react with oxidants, such as OH radicals (Finlayson-Pitts and Pitts Jr, 2000). The lifetime of two semi-volatile electron-deficient alkenes (4-oxo-2-butenoic acid, and 2-hydroxy-3-penten-1,5-dial) that

5 were reported in the previous study (Jang and Kamens, 2001) was estimated using (Finlayson-Pitts and Pitts Jr, 2000):

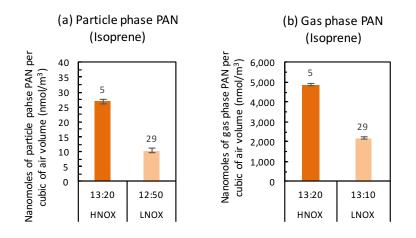
$$\tau = \frac{1}{k[\text{OH}]} \tag{1}$$

where [OH] is the concentration of OH radicals, and k is the rate constant (Finlayson-Pitts and Pitts Jr, 2000; Jang and Kamens, 2001). The typical concentration of OH radicals during the

photooxidation of toluene was about 2×10⁻⁴ ppb around noontime, estimated using the Master Chemical Mechanism (Saunders et al., 2003, 1997). The rate constants (k_{4-oxo-2-butenoic acid} (298 K) =2.53×10⁻¹¹ cm³ molecule⁻¹ s⁻¹, k_{2-hydroxy-3-penten-1,5-dial} (298 K) =7.78×10⁻¹¹ cm³ molecule⁻¹ s⁻¹) were calculated using the structure-reactivity relationship (Kwok and Atkinson, 1995). The lifetime of 4-oxo-2-butenoic acid and 2-hydroxy-3-penten-1,5-dial was estimated to be 134 min and 43 min, respectively.

Section S6. The concentration of gas-phase and particle-phase PAN in isoprene SOA.

For the isoprene experiment (22 Jul. 2016), the concentrations of gas phase and particle phase PANs were measured using the Griess assay. The collection efficiency of gaseous compounds using the impinger sampling method was not determined in this study. However, the concentrations of gaseous PAN products were at least 200 times higher than those of particulate PAN products, as shown in Fig. S10. The *x*-axis represents the mid-collection time (Table 1).



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Figure S10. (a) The nanomoles of particle phase PANs (collected by a particle-into-liquid sampler) per cubic of air volume. (b) The nanomoles of gas phase PANs (collected by an impinger) per cubic of air volume. The error bar of each column is the standard deviation of three replicates. The number above each column represents the initial HC/NO_x ratio (ppbC/ppb).

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